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Identification of Histamine H₃ Receptor Ligands Using a New Crystal Structure Fragment-based Method

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Virtual screening offers an efficient alternative to high-throughput screening in the identification of pharmacological tools and lead compounds. Virtual screening is typically based on the matching of target structures or ligand pharmacophores to commercial or in-house compound catalogues. This study provides the first proof-of-concept for our recently reported method where pharmacophores are instead constructed based on the inference of residue-ligand fragments from crystal structures. We demonstrate its unique utility for G protein-coupled receptors, which represent the largest families of human membrane proteins and drug targets. We identified five neutral antagonists and one inverse agonist for the histamine H₃ receptor with potencies of 0.7–8.5 μM in a recombinant receptor cell-based inositol phosphate accumulation assay and validated their activity using a radioligand competition binding assay. H₃ receptor antagonism is of large therapeutic value and our ligands could serve as starting points for further lead optimisation. The six ligands exhibit four chemical scaffolds, whereof three have high novelty in comparison to the known H₃ receptor ligands in the ChEMBL database. The complete pharmacophore fragment library is freely available through the GPCR database, GPCRdb, allowing the successful application herein to be repeated for most of the 285 class A GPCR targets. The method could also easily be adapted to other protein families.

G protein-coupled receptors (GPCRs) are membrane proteins activated by many diverse ligands including photons, ions, neurotransmitters, lipids, carbohydrates, nucleotides, amino acids, peptides (e.g. hormones) and proteins¹. The GPCR family is with its ~800 genes among the largest families in the human genome² and involved in most physiological processes^{3,4}. Their regulation of pathophysiology in diverse disease areas and accessibility at the cell surface have earned them a key role in medicine: more than 30% of the drug on the market target GPCRs⁵. 59 GPCRs have been drugged with small molecules⁶; the vast majority binding within a structurally conserved pocket within the transmembrane heptahelical bundle (7TM)⁷. Modelling of ligand-receptor complexes can now be performed with higher accuracy as a result of the increasing number of GPCR crystal structures, and has been evaluated in three community-wide 'GPCR Dock' assessments^{8–10}.

Virtual screening is an efficient alternative to high-throughput screening to identify new ligands. This entails a preceding screening *in silico* of commercial or in-house compound catalogues against a target structure^{11–13} or ligand pharmacophore^{14,15}. We recently described the development of a new chemogenomics method to generate pharmacophores for GPCRs¹⁶. The method is based on the extraction of a reference library of crystal structure fragments, which are interacting moiety-residue pairs. The complete library of moiety-residue pairs has been made available through the GPCR database, GPCRdb^{17,18}. Although specific for GPCR ligand discovery, this is analogous to general resources that extract the residue-ligand fragment information from crystal structures^{19–21}. Here, we describe the first application of this method to identify new histamine H₃ receptor ligands.

The H₃ receptor is found mainly in the CNS and is implicated in cognition, sleep regulation, feeding, memory, nociception and the sleep/wake cycle^{22–24}. It functions both as a presynaptic autoreceptor, as well as a regulator of

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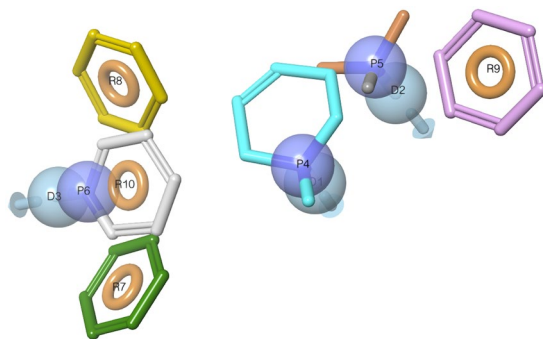


Figure 1. Histamine H₃ receptor pharmacophore model used in the virtual screening for new ligands. The pharmacophore was based on a receptor residue – ligand moiety fragment library derived from GPCR crystal structures (Supplementary Table 1), and built with Phase⁶¹. The pharmacophore elements include three hydrogen bond donor (D1-3, light blue), three cation (P4-6, blue), three aromatic (R7, R9 and R10; orange) and one dual aromatic/hydrophobic (R8, orange) features.

the release of other neurotransmitters; such as serotonin, dopamine and acetylcholine; which may be co-released with GABA in some neurons to control wakefulness²⁵. H₃ has a remarkably high level of constitutive activity²⁶ also *in vivo*²⁷, and many of the classical H₃ receptor antagonists have recently been found to be inverse agonists²². H₃ is coupled to the G_{1/0} class of G proteins, leading to inhibition of the adenylyl cyclase and decreased cAMP generation^{28,29}. The H₃ receptor has shown a potential therapeutic use in a number of CNS indications; ADHD, cognitive disorders, epilepsy, narcolepsy, neurodegeneration and pain^{30,31}; as well as in treating alcohol³² and eating³³ behaviours (for general reviews see refs 22–24).

Results

Pharmacophore model and virtual screening hits. The transmembrane domain of GPCRs can be aligned both in structure and sequence to identify the corresponding residue positions, which are below indexed using the GPCRdb scheme³⁴. This is an evolution of the Ballesteros-Weinstein scheme³⁵ modified to take into account helical bulges and constrictions observed in GPCR crystal structures. Matching of the H₃ receptor sequence against our library of crystal structure fragments identified five conserved residues: W3.28, D3.32, Y3.33, Y6.51 and W6.48. A sixth residue, E5.46x461, could be added from reference ligands matched to the pharmacophore and docked into a H₃ structure model. Apart from the residue, each fragment also has an interacting ligand moiety onto which the pharmacophore elements are placed. Figure 1 shows the final pharmacophore constructed from all the retrieved fragments, which are listed in Supplementary Table 1. The pharmacophore contains three hydrogen bond donor (D1-3), three cation (P4-6), three aromatic (R7, R9 and R10) and one dual aromatic/hydrophobic (R8 and R10) elements. The screening of this pharmacophore and subsequent hit clustering resulted in 44 compounds selected for purchase (Supplementary Chart 1).

The majority of the pharmacophore hits have long linkers between the two essential elements, the cationic P5 and aromatic R10, which is typical for antagonistic rather than agonistic aminergic ligands. Apart from the essential match to P5 (the site of the archetypical aminergic ligand interacting with the conserved D3.32), as many as 31 hits also matched P6, whereof only two matched the third element P4. The additional match to P6 is significant, as this feature, or rather its interacting residue E5.46x461, is not shared by the H₁₋₂ receptors and therefore is likely to contribute to selectivity. Apart from the essential match to the aromatic element R10, about half (21) and a third (15) of the hits also matched R7 and R9, respectively (5 matched both). In contrast, R8 that is a dual hydrophobic/aromatic element matched only 3 aromatic moieties, but 23 hydrophobic. Finally, the number of matches for the hydrogen bonding elements reflected that of their adjacent cationic features, with 39, 33 and 5 matches for D3, D2 and D1, respectively.

Histamine H₃ receptor assay validation. As shown in Fig. 2, the co-expression of H₃ receptor and the chimeric G protein Gqi5 in tsA cells allowed for the generation of a histamine concentration-response curve in the IP-One assay with an EC₅₀ value of 150 nM (pEC₅₀ = 6.8 ± 0.4; n = 12). Furthermore, the reference antagonist thioperamide gave an IC₅₀ value of 62 nM (pIC₅₀ = 7.2 ± 0.3; n = 4), in the presence of EC₈₀ of histamine (data not shown). Previous studies also using Gqi5 co-expression in HEK-derived cells reported values of 21.8 nM³⁶ and 151 nM³⁷ for histamine and thioperamide in mouse and human H₃ receptors, respectively. The differences in EC₅₀ and IC₅₀; about 7-fold and 2-fold, respectively; may be attributed to differences in assay conditions (IP₁ accumulation versus Ca²⁺ release), species used, and/or differences in expression system. To validate the suitability of the assay for high-throughput screening, the Z'-factors³⁸ were determined, as described in methods. The Z'-factors determined were 0.5 and 0.3 for agonism and antagonism modes, respectively, which is indicative of an acceptable separation between the positive and the negative controls³⁸. Thus, the assay was found to have adequate sensitivity, reproducibility and accuracy for the intended compound screening.

Novel H₃ receptor ligands. The first pharmacological assaying evaluated the selected 44 pharmacophore screening hits, and resulted in four antagonist hits; 21, 34, 35, 42. The second pharmacological assaying concerned 32 analogues of the four first hits (21: 45–48, 34: 49–54, 35: 55–72 and 42: 73–76), and resulted in four

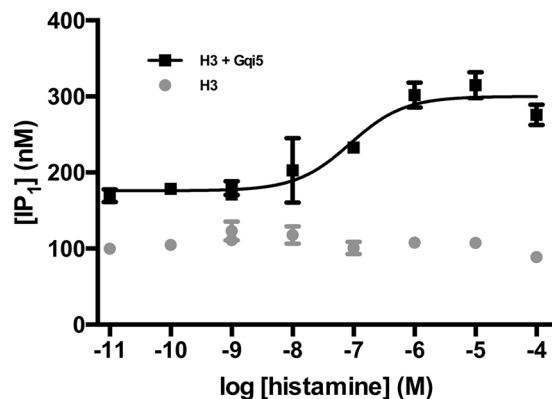


Figure 2. Assessment of histamine H_3 receptor G_{q1} -coupled pharmacological assay using the IP_1 accumulation assay measured by the HTRF IP-One assay. Concentration-dependent responses to histamine in H_3 receptor expressing tsA201 cells transfected with (black) and without (grey) Gq15. Data are means \pm S.D. of a single representative experiment performed in triplicate. Two additional experiments gave similar results.

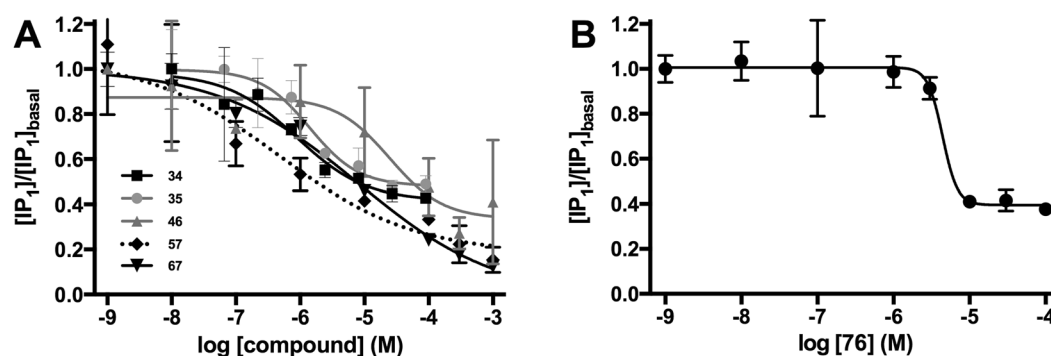


Figure 3. IP-One assay concentration-response curves. (A) Concentration-inhibition curves of the five identified neutral antagonists in the presence of histamine (EC_{80} concentration). (B) Inverse agonism of compound 76 (no histamine added). Data are normalized to the basal level of IP_1 (fold response) in ligand buffer and are shown as means S.D. of a single representative experiment performed in triplicate. Due to solubility issues the lower plateau for the concentration-response curve of compounds 46, 57 and 67 was constrained to the buffer value during curve fitting.

new antagonists 46, 57, 67 and 76. Three seemingly active compounds from the first screen were excluded due to borderline activity (20) or an azaindolizine substructure (9 and 44), which can be fluorescent leading to a false positive response. Furthermore, after the secondary screen, compounds 21, 42, 64, 68 and 75 were excluded because of the more stringent IC_{50} cut-off of $10\ \mu\text{M}$ (Supplementary Table 2). The three antagonists 34, 67 and 76 displayed inhibition below the basal level, and were therefore also tested as inverse agonists. Inverse agonistic activity was shown for 76 with an IC_{50} of $4.2\ \mu\text{M}$ but not for 34 and 67. In summary, we identified five antagonists (34, 35, 46, 57 and 67) (Fig. 3A) and one inverse agonist (76) (Fig. 3B), with IC_{50} values ranging 0.66 – $8.5\ \mu\text{M}$ (Table 1).

To confirm the results obtained from the functional studies, the six compounds (34, 35, 46, 57, 67 and 76) were also examined in a [^3H]N- α -methylhistamine radioligand competition binding assay (Fig. 4A). Their K_i values ranged between 1.59 – $6.07\ \mu\text{M}$ (Table 1) and were generally in good agreement with the potency observed in the functional study (Fig. 4B).

We assessed the novelty of the ligand structures by identifying the most similar H_3 ligands in ChEMBL (Table 1). Ligand structures can be considered novel if their fingerprints have a Tanimoto Coefficient below 0.4^{39} . 35 is just above this cut-off and 57 and 67 come close, whereas 34, 46 and 76 are quite dissimilar to the known H_3 ligands. The three least novel structures, share a fused triple ring core that in 35 contains a central imidazole, which is shared by histamine and many surrogate ligands. In contrast, the three novel ligands 34, 46 and 76, contain scaffolds that are distinct also among the herein described hit compounds. Apart from fused ring systems, all six ligands contain an amine that is positive ionisable, which was an essential match in the pharmacophore screening. Furthermore, 76 contains an amine β -hydroxyl and 46 a 4-aryl-piperidine that are known to be common functionalities in many aminergic and GPCR 40,41 ligands, respectively.

Histamine receptor binding site selectivity hotspots. Whereas experimental ligand selectivity was outside of the scope of this study, we compared the sequences and structures of the four histamine receptor subtypes to provide a rationale for future optimisation efforts. This showed that the H_1 , H_2 and H_4 anti-targets

Cmpd	Structure	IC ₅₀ (μM)	pIC ₅₀ ± SEM	n	Ki (μM)	Affinity (pKi)	n	Most similar ChEMBL ligand	ECFP-4* similarity	Affinity (pKi)
34		0.66	6.2 ± 0.13	5	1.97	5.71 ± 0.17	5	ChEMBL 237514	0.26	6.1
35		3.3	5.5 ± 0.27	5	1.75	5.76 ± 0.05	4	ChEMBL 1923529	0.42	6.2
46		6.6	5.2 ± 0.37	3	6.07	5.22 ± 0.09	3	ChEMBL 2048591	0.24	7.3
57		5.2	5.3 ± 0.37	4	1.59	5.80 ± 0.14	3	ChEMBL 460313	0.37	6.9
67		8.5	5.1 ± 0.25	6	3.52	5.45 ± 0.19	3	ChEMBL 186456	0.36	5.5
76		2.7	5.6 ± 0.14	2	1.59	5.80 ± 0.05	3	ChEMBL 65849	0.26	8.0
	Inverse agonism activity	4.2	5.4 ± 0.068	3						

Table 1. Identified H₃ receptor ligand structures, potencies and most similar ChEMBL ligand. *ECFP-4 2D topological similarity.

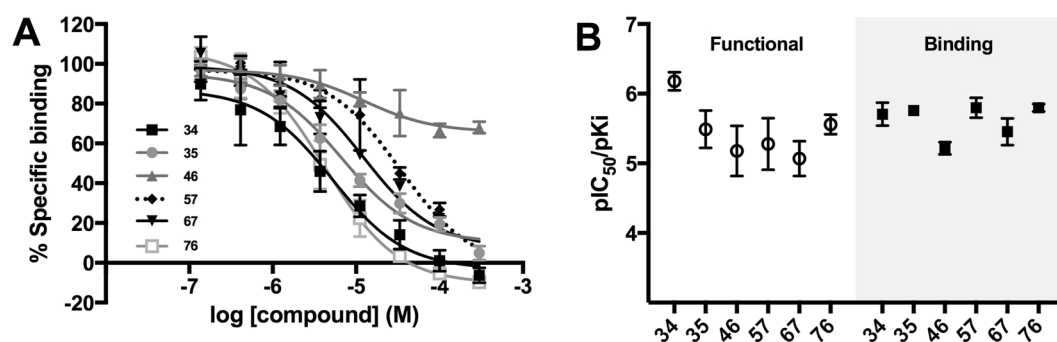


Figure 4. (A) Radioligand competition binding assay validates hits at the H₃ receptor. Representative curves of the competitive binding of [³H]N-α-methylhistamine (0.3 nM) in the presence of various concentrations of compounds. Data points are shown as the mean ± S.D. of triplicate measurements. (B) The obtained affinities correlate with the potencies obtained in the functional assay. The potencies of the six most potent compounds obtained from the IP-One assay (*Functional*) shown as pIC₅₀ compared with their affinities as determined in the radioligand competition binding assay (*Binding*). All data points are shown as the mean ± S.E.M, n = 2–5.

the extent to which diverse and novel scaffolds are selected. What sets this method apart is instead the advantage that ligands can be identified without the requirement for previously known ligands. If such data would have been incorporated (beyond the E5.46x461 fragment) or the structural novelty not enforced as a filter, it can be expected that the hit rate and affinity would have been higher. Another advantage observed in the application of the crystal structure-based method to the H₃ receptor target is that its inference from other (crystallized) receptors yielded a larger number of pharmacophore elements – ten instead of four¹⁴ and five⁴⁷, respectively. This offers unique opportunities to target medicinal chemistry substitutions in the ligand optimisation phase on the basis of crystallographic evidence.

The unique ability to infer pharmacophore elements suggests that the crystal structure-based pharmacophore method is likely to offer a bigger advantage when applied to other GPCR targets that lack known ligands or a closely homologous crystal structure. Klabunde *et al.* succeeded in the identification of C3a receptor ligands using a method based on fragments and pharmacophores⁴⁸. A larger variety of studies have inferred whole scaffolds or ligands between targets, based on chemogenomic techniques that detect local similarities within the transmembrane binding pocket^{49–56}. The ability to perform such binding pocket comparisons have greatly increased as an effect of the many high-quality templates in the form of GPCR crystal structures, many of which are in complex with ligands. In this analysis, the H₃ receptor was used as a template to place residue backbone atoms onto transmembrane helices. This receptor displays a sequence similarity of 59% and 57% to the H₁ receptor transmembrane bundle and the doxepin binding site (5 Å proximity), respectively. The fragments, defining the residue sidechains and ligand moieties, were instead inferred from a variety of aminergic receptors. For such SAR-borrowing it is crucial to use structure-based sequence alignments to ensure that the equivalent residues are correctly identified, but this is greatly complicated by a number of frequent structural distortions in GPCR helices, bulges and constrictions. Specifically, one residue, E5.46x461, in the applied fragments is located at the tip of a bulge in many aminergic GPCRs. We used the GPCRdb alignments, which take this into account by defining the corresponding residue structure/sequence positions by superposition of crystal structures, and appends a second correct generic residue number to the Ballesteros-Weinstein number³⁴.

In the present study, we utilized co-transfection of the human histamine H₃ receptor with the chimeric Gq15 protein to channel receptor signalling into the Gq pathway. Others have successfully applied this strategy in the attempt of developing robust Gq-directed pharmacological assays for GPCRs in general⁵⁷, as well as for the mouse³⁶ and human⁵⁸ H₃ receptor. Instead of utilizing Gq coupling for measuring calcium release we chose to develop an HTRF-IP₁ accumulation assay that offers high sensitivity in an efficient 384-well format⁵⁹. This assay was successfully established and showed satisfactory Z'-factors taken into consideration that it was a transient expression of receptor and G protein. It is to our knowledge the first demonstration of applying the IP-One Tb Cisbio assay on the histamine H₃ receptor. Hence, with these assays we were able to confirm histamine agonism and identify several novel antagonists and one inverse agonist, demonstrating a broad detection range and versatility.

The subsequent characterization of the most potent compounds in the radioligand competition binding assay provides evidence that the inhibitory activity seen in the IP-One assay was mediated via an interaction with the H₃ receptor, as the potencies/affinities obtained in both assays generally correlate well (Fig. 4B). This was however not true for all the compounds, as compounds **21** and **68** were unable to compete with binding of [³H]N- α -methylhistamine in the binding assay despite having previously shown inhibitory activity in the IP-One assay. This conflicting data may be due to an interference with the IP-One assay or an unspecific event leading to an apparent inhibition, such as cytotoxicity. This underlines the value of validating hits after a screening with an assay based on another principle. Additionally, the binding assay allowed for a more precise determination of the compound potency rank order, as the variations in the pIC₅₀ values from the IP-One assay was greater than the corresponding variation in the pKi values for the binding assay (Fig. 4B). Interestingly, some compounds (**42**, **44** and **46**) were unable to lower the binding of [³H]N- α -methylhistamine to non-specific binding levels, as would otherwise be expected for ligands competing for binding at the orthosteric site. It cannot be ruled out that the unprecedented number of (10) pharmacophore elements, whereof three cationic elements and four aromatic, allow for compounds to bind simultaneously with the radioligand, [³H]N- α -methylhistamine, which is relatively small. However, future studies would be needed to firmly establish if **42**, **44** and **46** bind in an allosteric binding site or there is another explanation (e.g. a secondary binding site for the radioligand).

Conclusions and Future Perspectives. A variety of alternative approaches exist for virtual screening for new lead drugs and pharmacological tool compounds. This study provided the first proof-of-concept for our recent pharmacophore method based on crystal structure fragments. This method will benefit from more fragments as the number of GPCR crystal structures increase even further, also for the same receptor in complex with a diversity of ligands. We are currently working to implement this method on targets with less structural information, including orphan receptors, and the application in the wider community is facilitated by free matching of the fragments to any class A GPCR target in GPCRdb. Both H₃ receptor inverse antagonists and (neutral) antagonists are of proven therapeutic value. The ligands identified herein are drug-like molecules with novel scaffolds that could serve as starting points for further lead optimization.

Methods

Construction of a H₃ pharmacophore. A structure-based sequence alignment of the H₁ and H₃ receptors was downloaded from GPCRdb¹⁷, and a model of the H₃ receptor transmembrane domain was built in Modeller⁶⁰ using the H₁ crystal structure (PDB: 3RZE)⁴⁴ template. The pharmacophore was designed using our previously described crystal structure-based pharmacophore method¹⁶, which is based on the manual annotation of a library of structural fragments, pairs of a receptor residue and interacting ligand moiety, from GPCR crystal structure complexes. Herein, we uploaded our H₃ receptor model to GPCRdb¹⁷ to identify conserved residues represented

in this library, and to superpose the backbones of the corresponding fragments. The pharmacophore elements were placed using Phase⁶¹ at the highest density of the (multiple) fragment moieties. The vectors of the hydrogen bonding features were defined after optimization of ligand moiety – receptor residue interactions. Furthermore, the H₃ pharmacophore was extended with an additional element not covered by the fragment library, but instead defined by matching reference ligands to the pharmacophore as well as docking them into a H₃ structure model. The additional pharmacophore element represents a cationic ligand functionality that interacts with a Glu residue in position 5.46 × 461. This residue has been shown by mutagenesis studies be important for binding of imidazole- and pyridine-containing ligands, including histamine^{62,63}.

Preparation of reference ligand and screening databases. Histamine H₃ receptor reference ligands were downloaded from ChEMBL⁶⁴ and the IUPHAR guide to pharmacology databases⁴. We used only the ligands with submicromolar dose-response affinity or activity values (K_i , pK_i , EC_{50} , pEC_{50} , IC_{50} and pIC_{50}), and the highest assay confidence scores: 8 or 9. The screening database, eMolecules plus⁶⁵, was prepared with LigPrep⁶⁶ to desalt, add hydrogen atoms and generate tautomers, stereoisomers (max 32) and 3D conformations (max 10 ring conformations). Epik and the OPLS 2005 force field were applied to generate charge states at pH: 7.0 ± 1.0 ⁶⁶. LigFilter was used to remove structures with reactive functional groups and match the properties (Supplementary Table 4) of the reference ligands⁶⁷.

Pharmacophore screening and hit selection. The Phase database, containing both reference ligands and screening compounds, was prepared with 100 maximum conformers, up to 10 conformations per rotatable bond, thorough conformational sampling, conformational variation of amide bonds and a maximum relative energy difference of 6.0 kcal/mol. A minimum of four matching pharmacophore elements was required and a preference was set for partial matches involving more sites. Hits were sorted by fitness score and clustered with Canvas⁶⁸ to select diverse representative structures. As a secondary assessment of compound structures, we used SiteMap⁶⁹ on a H₃ structure model. After the first assaying round small structure-activity relationship analyses were conducted and the compounds sorted into lead ligand series. The selections of analogues were based on substructures drawn in MarvinSketch and queried in the eMolecules database loaded into Instant JChem (Marvin 5.12.3, 2014 and Instant JChem 6.2.0, 2014, ChemAxon, www.chemaxon.com).

Histamine receptor binding site comparison. The H₁ crystal structure and updated H₂₋₄ homology models were downloaded from GPCRdb^{17,18}. An initial alignment of all class A GPCR ligand binding pocket residues was retrieved, also from GPCRdb. This alignment was first filtered to pinpoint the receptor subtype-differing positions, and subsequently by structural investigation to extract only those residues that may be accessible to ligand substitutions (Fig. 5A).

Pharmacological assaying materials. Buffers and media for cell culturing were all purchased from Invitrogen (Paisley, United Kingdom) whereas non-enzymatic dissociation solution, HEPES (4-(2-hydroxyethyl) piperazine-1-ethanesulfonic acid) and additional assay buffer supplements were from Sigma-Aldrich (St. Louis, MO, USA). The IP-One Tb assay kit was purchased from Cisbio (Codolet, France). The human histamine H₃ DNA (Genbank accession no. AF321910.1) in a pSI expression vector was identical to a previously used one⁵⁸. The G protein Gqi5 was a kind gift from Dr. Evi Kostenis, University of Bonn, Germany. Histamine hydrochloride and thioperamide maleate were obtained from Sigma-Aldrich and Abcam Biochemicals (Cambridge, UK), respectively.

Cell culture and transfections. Human tsA201 cells, a transformed HEK-293 cell line⁷⁰, were cultivated in Dulbecco's Modified Eagle's Medium (DMEM) with GlutaMAX, supplemented with 10% foetal bovine serum, penicillin 100 U/ml and streptomycin 100 µg/ml. Subconfluent cells grown in a 100 mm dish were transfected with 5 µg DNA using the Polyfect transfection reagent using the protocol of the manufacturer (Qiagen, West Sussex, UK), however with half of the recommended PolyFect volumes. Co-transfections (ratio 4:1) were performed with either constructs expressing the human histamine H₃ receptor together with an empty vector and chimeric G protein Gqi5, respectively.

IP-One assay. We applied the highly sensitive high-throughput homogeneous time-resolved fluorescence (HTRF) technology⁷¹ for detection of IP-One generation using the Cisbio IP-One Tb assay kit, exactly as previously described⁷². In brief, on the day of assay, ligand solutions were prepared in ligand buffer (Hank's Balanced Salt Solution (HBSS) containing 20 mM HEPES pH 7.4, 1 mM CaCl₂, 1 mM MgCl₂, 40 mM LiCl) and added to the wells of a 384-well OptiPlate (PerkinElmer, Waltham, USA) in triplicates. When testing for antagonist activity the ligand buffer was supplemented with an EC₈₀ concentration of histamine, and we used a final compound concentration of 20 µM and 2 µM for the first (virtual hits) and second (analogues) screening, respectively. Cell suspensions were added and incubated with ligands for 1 hour, followed by the addition of detection solution (IP-One Tb conjugate & Lysis Buffer +2.5% anti-IP₁ cryptate Tb conjugate +2.5% IP₁-d2 conjugate. 38:1:1) and incubation in the dark for one hour at room temperature. The plate was read on an EnVision 2104 Multilabel Reader (PerkinElmer) by exciting the wells with light of 340 nm and measuring the emission at 615 nm and 665 nm. The fluorescence resonance energy transfer ratios (665 nm/615 nm) were converted to IP₁ concentrations by interpolating values from an IP₁ standard curve generated from an IP₁ calibration stock, provided by the manufacturer (Cisbio).

[³H]N-α-methylhistamine radioligand competition binding assay. HEK-239T cells were grown to 70% confluency in 150 mm cell culture dishes and transfected with 8 µg human H₃ receptor using the Polyfect transfection reagent as previously described⁷². After 48 hours, cells were washed from each cell culture dish with

Dulbecco's phosphate buffered saline and harvested using a cell scraper. The resulting cell suspension was then centrifuged and resuspended in 500 μL lysis buffer (50 mM Tris-HCl, pH 7.4). The cell lysate was centrifuged at 16,000 g for 20 minutes at 4 °C. The resultant membrane pellets were stored at $-80\text{ }^{\circ}\text{C}$ until use. Each pellet was resuspended in 500 μL binding buffer (50 mM Tris-HCl, 0.5 mM EDTA, pH 7.4), homogenized and centrifuged at 16,000 g for 15 minutes at 4 °C, and subsequently resuspended in 15 mL of binding buffer to a desired protein concentration of approximately 0.2 mg/mL as determined by the method of Bradford⁷³. The membrane solution (30 μg protein per well) was incubated for 90 minutes in 250 μL of binding buffer, with or without compound, together with 0.3 nM [³H]N- α -methylhistamine at room temperature. Non-specific binding was defined as the radioligand bound to membrane solution incubated with 10 μM histamine. The binding reaction was terminated by rapid filtration through Whatman GF/C unifiers (PerkinElmer), and washed four times with ice-cold wash buffer (50 mM Tris-HCl, 10 mM MgCl₂, 0.1 mM EDTA, pH 7.4) using a 96-well Packard FilterMate cell harvester (PerkinElmer). Finally, Microscint 0 scintillation liquid (PerkinElmer) was added to the dried filters, and the radioactivity quantified in a Packard TopCount NXT microplate Scintillation Counter (PerkinElmer).

Data analysis. In the first (virtual) screening, hits were selected if they displayed inhibition values greater than 3 standard deviations above the inhibition mean of the set of tested compounds. In the second screening, we applied a more stringent hit criterion requiring an IC₅₀ value lower than 10 μM . Data were analysed using GraphPad Prism 6 (GraphPad Software, San Diego, CA, USA). Concentration-response curves were fitted by nonlinear regression to equation (1):

$$R = R_{\max} + \frac{R_{\max} - R_{\min}}{1 + 10^{(\log IC_{50} - X) \cdot n_H}} \quad (1)$$

where X is the logarithm of the concentration, R is the response, R_{max} is the maximal response, R_{min} is the minimal response, IC₅₀ is the concentration giving half-maximum reduction of the response, and n_H is the Hill coefficient, which describes the steepness of the curve. IP-One assay results are reported either as raw FRET in arbitrary units, as IP₁ concentration (nM) or as fold over basal (normalized to the basal level of IP₁ in ligand buffer [IP₁]/[IP₁]_{basal}). The Z'-factors were calculated using equation (2):

$$Z' = 1 - \frac{3\sigma_{c+} + 3\sigma_{c-}}{|\mu_{c+} - \mu_{c-}|} \quad (2)$$

Z' = 1 - ((3 σ_{c+} + 3 σ_{c-})/| μ_{c+} - μ_{c-} |), where σ_{c+} and σ_{c-} are the standard deviation of the positive control and the negative control, respectively, and μ_{c+} and μ_{c-} are the mean of the positive control and the negative control, respectively. Values were determined for both agonist mode (histamine alone) and antagonist mode (histamine + thioperamide) using buffer as negative control. The Z' determination was performed on a separate plate (n = 10–14 for each condition) and not on the library screening plates themselves, which, however, all contained controls that confirmed that the assay worked.

The binding data was fitted by nonlinear regression to equation (3):

$$Y = Y_{\max} + \frac{Y_{\max} - Y_{\min}}{1 + 10^{X - \log IC_{50}}} \quad (3)$$

where X is the logarithm of the concentration, Y is the specific binding, Y_{max} is the maximal specific binding, Y_{min} is the minimal specific binding, and IC₅₀ is the concentration giving half-maximum reduction in specific binding. The obtained IC₅₀ values were converted to K_i values by the Cheng-Prusoff equation, using a published K_d value, 0.15 nM for this receptor-radioligand pair^{58,74}.

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Author Contributions

D.E.G. conceived the study, purchased ligands analogues and supervised the modelling. K.F. and A.S.H. gathered crystal structure fragments, and K.F. made the virtual screening. V.I. made fragments available in GPCRdb. I.O.F. and M.W.B. conducted the functional and binding pharmacological experiments under the supervision of P.W. and H.B.-O., respectively. A.S.H., D.E.G., I.O.F., M.W.B., and P.W. drafted the manuscript, and all authors have approved the final version.

Additional Information

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