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# Expanding the Landscape of Diterpene Structural Diversity through Stereochemically Controlled Combinatorial Biosynthesis

Johan Andersen-Ranberg, Kenneth Thermann Kongstad, Morten Thrane Nielsen, Niels Bjerg Jensen, Irini Pateraki, Søren Spanner Bach, Britta Hamberger, Philipp Zerbe, Dan Staerk, Jörg Bohlmann, Birger Lindberg Møller, and Björn Hamberger\*

**Abstract:** Plant-derived diterpenoids serve as important pharmaceuticals, food additives, and fragrances, yet their low natural abundance and high structural complexity limits their broader industrial utilization. By mimicking the modularity of diterpene biosynthesis in plants, we constructed 51 functional combinations of class I and II diterpene synthases, 41 of which are “new-to-nature”. Stereoselective biosynthesis of over 50 diterpene skeletons was demonstrated, including natural variants and novel enantiomeric or diastereomeric counterparts. Scalable biotechnological production for four industrially relevant targets was accomplished in engineered strains of *Saccharomyces cerevisiae*.

Diterpenoids are a diverse class of structurally complex natural products with broad therapeutic applications. Plants provide the major source of these chemicals and present a rich reservoir of potentially new pharmaceuticals. Exploitation of this diversity is severely limited by difficulties in extracting pure compounds from natural sources on an industrial scale. Alternatives include semisynthesis from naturally occurring precursors, plant cell cultures, and chemical synthesis.<sup>[1]</sup> Stereospecific synthetic routes for complex diterpenoids often remain commercially impracticable owing to their low efficiency.<sup>[2]</sup> Combinatorial metabolic engineering of micro-

bial or plant hosts, using the enzymes of the biosynthetic pathways, offers an alternative environmentally benign access to structurally complex diterpenoids with specific and novel stereochemistry, which often defines the therapeutic value.<sup>[3]</sup> The carbon skeletons of bi- and polycyclic labdanes and clerodanes are formed in a two-step enzyme reaction catalyzed by bifunctional class I/II diterpene synthases (diTPSSs) or pairs of distinct monofunctional diTPSSs defined by their catalytic activity as class I or class II enzymes.<sup>[4]</sup> Class II diTPSSs catalyze the initial cyclization of (*E,E,E*)-geranylgeranyl diphosphate (GGPP) into bicyclic diphosphate intermediates with a characteristic decalin core (Scheme 1). The structure and specific stereochemistry at the C9 and C10 carbon atoms of the intermediate are determined by the class II diTPSS catalyzed reaction. The subsequent class I diTPSS reaction completes the formation of the diterpene skeleton through dephosphorylation of the class II product and specific rearrangements, additional cyclizations, or water addition, thereby resulting in the introduction of additional chiral centers (Scheme 1).<sup>[4b]</sup>

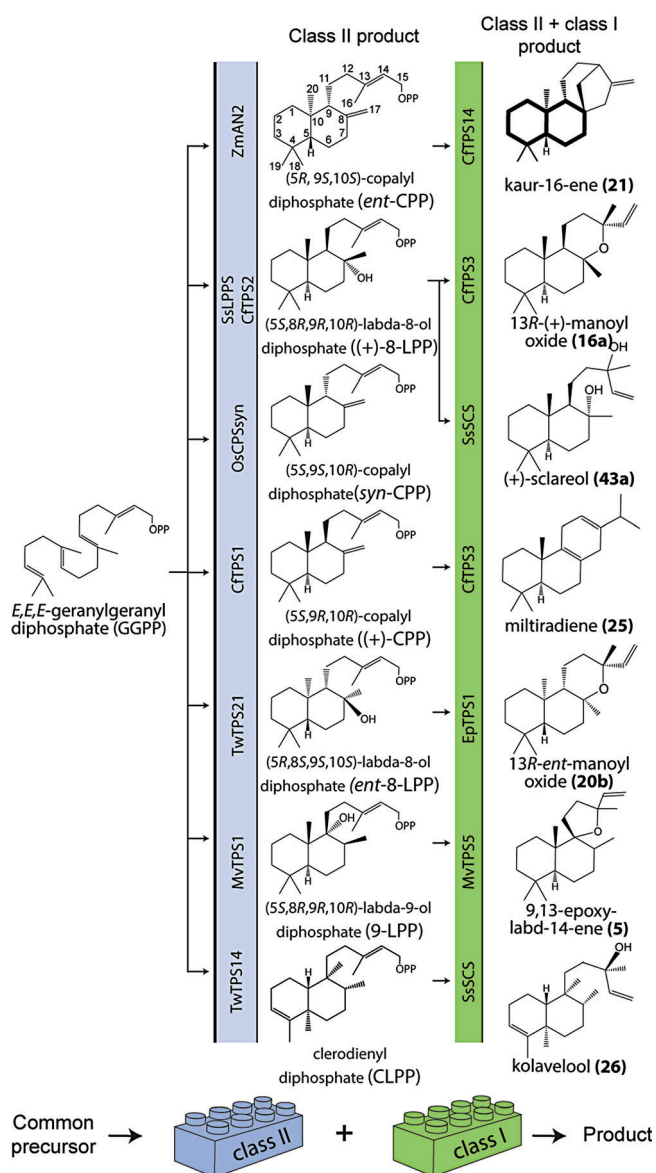
We have recently identified a diverse suite of 40 class II and class I diTPSSs from ten plant species, each harboring structurally exceptional diterpenoids.<sup>[5]</sup> In plants, specific pairs of class I and class II diTPSSs form distinct diterpene skeletons, which underpins the structural diversity of native diterpenoids.<sup>[6]</sup> To explore the inherent metabolic potential and plasticity of modular combinations of diTPSSs, we systematically paired diTPSSs through co-expression in heterologous hosts. Specifically, we probed the substrate and product plasticity for every combination of eleven class II and nine class I diTPSSs representative of diverse plant species and potential functionalities. Eight functionally uncharacterized diTPSSs were included in this approach (Table S1 in the Supporting Information).

We report herein the stereoselective formation of more than 50 labdane- and clerodane-type skeletons through the assembly of combinations of diTPSS pairs. These skeletons are labelled throughout (**1–47**) according to their Kovats retention indices (Figure S1–4 and Table S2 in the Supporting Information). The a and b suffixes indicate (+) or *ent* configurations, respectively, as shown by chiral GC–MS analysis, NMR, and optical rotation measurements of selected purified compounds. Three examples illustrate the potential of this approach for control of the stereochemical configuration of diterpenes through specific selection of class II diTPSSs and for extension of the known structural repertoire of skeletons accessible by biosynthesis through combination

[\*] Dr. J. Andersen-Ranberg, Dr. M. T. Nielsen, Dr. I. Pateraki, Dr. S. S. Bach, B. Hamberger, Prof. Dr. B. L. Møller, Prof. Dr. B. Hamberger  
Department of Plant and Environmental Sciences  
Center for Synthetic Biology  
Thorvaldsensvej 40, 1871 København (Denmark)  
E-mail: bjoernh@plen.ku.dk  
Dr. K. T. Kongstad, Prof. Dr. D. Staerk  
Natural Products Research, University of Copenhagen (Denmark)  
Dr. N. B. Jensen  
Evolva AS, København (Denmark)  
Prof. Dr. P. Zerbe  
Department of Plant Biology, University of California Davis (USA)  
Prof. Dr. J. Bohlmann  
Michael Smith Laboratories, University of British Columbia  
Vancouver, V6T 1Z4 (Canada)

Supporting information and ORCID(s) from the author(s) for this article are available on the WWW under <http://dx.doi.org/10.1002/anie.201510650>.

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**Scheme 1.** The modular nature of diterpene biosynthesis. Overview of class II and class I diTPSs and their products. The decalin core is shown in bold. Zm = *Zea mays*; Cf = *Coleus forskohlii*; Ss = *Salvia sclarea*; Os = *Oryza sativa*; Mv = *Marrubium vulgare*; Tw = *Tripterygium wilfordii*.

with class I diTPSs. A list of all of the diterpenes produced is given in Table S2.

DiTPS combinations were tested by *Agrobacterium tumefaciens* mediated transient expression in *Nicotiana benthamiana* and the results were supported by in vitro assays with recombinant diTPS expressed and purified from *E. coli* for each type of class II diTPS. To demonstrate fermentation-based diterpene production, selected diTPS combinations were introduced into engineered *Saccharomyces cerevisiae* to yield target molecules at a scale relevant for industrial consideration.

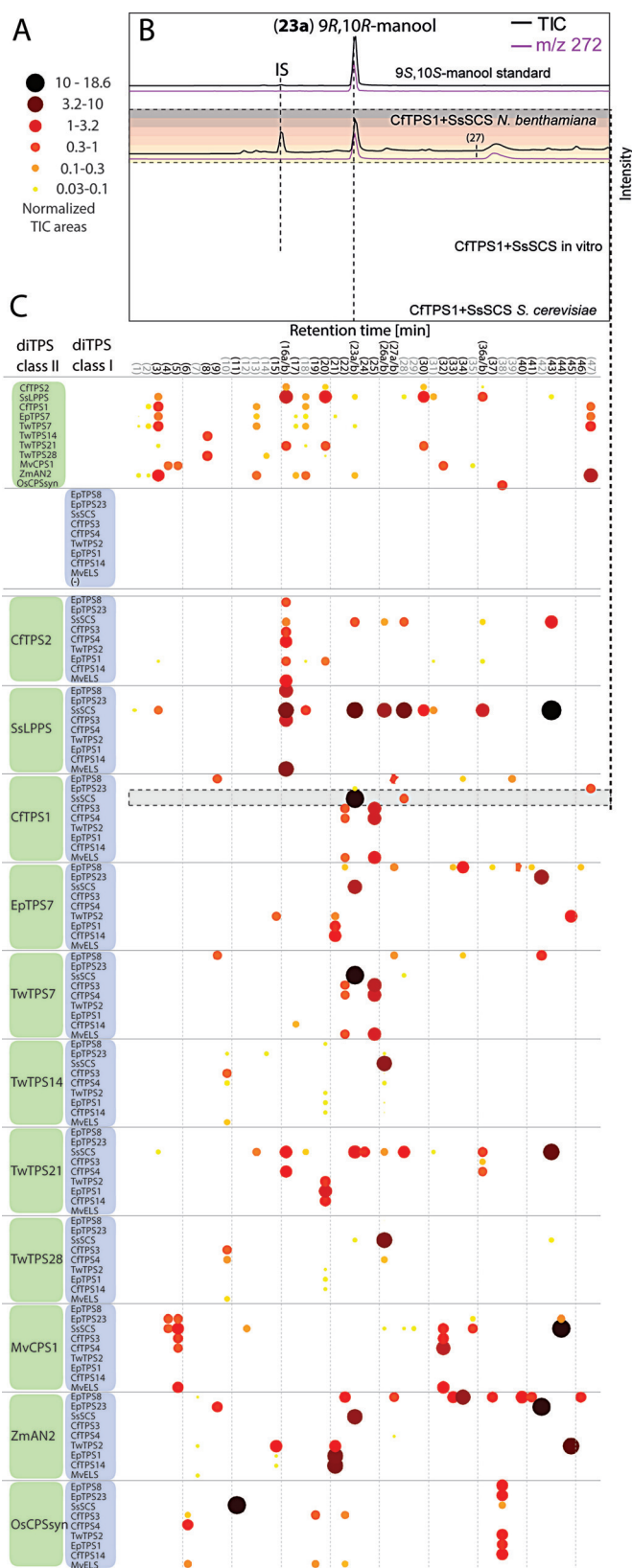
Comparison of substrate and product profiles showed that the class I diTPSs show distinct degrees of substrate and product plasticity with regards to the stereochemistry and hydroxylation pattern of the diphosphate substrate. Specifi-

cally, in a total of 12 combinations, a subset of six class I diTPSs converted the two stereoisomers of labda-13-en-8-ol diphosphate [(+)-8-LPP and *ent*-8-LPP, Figure 1].

The discrete formation of three stereoisomers of the diterpene manoyl oxide highlights how biosynthetic routes can be stereoselectively controlled (Figure 2). (13*R*)-(+)-manoyl oxide (**16a**), a precursor for the diterpenoid pharmaceutical forskolin,<sup>[6a]</sup> was formed from (+)-8-LPP by the combination of *Salvia sclarea* SsLPPS with *Marrubium vulgare* MvTSP5 at 3.7 times higher levels than the native combination of CFTPS2 and CFTPS3 from *Coleus forskohlii*, which may be relevant for future production. Stereoselective access to manoyl oxide stereoisomers is a prerequisite for the biotechnological production of forskolin derivatives with novel stereochemical configurations. In combination with the class II diTPS TwTSP21 from *Tripterygium wilfordii*, CFTPS3 catalyzed the stereoselective formation of (13*R*)-*ent*-manoyl oxide (**16b**). Combination of the class I diTPS TwTSP2 with CFTPS14 or *Euphorbia peplus* EpTSP1, which are unable to convert (+)-8-LPP, enabled the stereoselective biosynthesis of the diastereomer (13*S*)-*ent*-manoyl oxide (**20b**; Figure 2 and Figure S16). The *ent*-configuration of compounds **16b** and **20b** is consistent with a function of the class II diTPS TwTSP21 in providing specifically (5*R*,8*S*,9*S*,10*S*)-labda-13-en-8-ol diphosphate (*ent*-8-LPP), a potential intermediate of bioactive diterpenoids with this stereochemistry found in nature yet unknown in biosynthetic routes.<sup>[8]</sup>

The formation of structurally distinct bicyclic diterpenes, such as sclareol, which is used in high-end fragrance manufacture, is catalyzed in *S. sclarea* by SsLPPS and SsSCS.<sup>[9]</sup> Antibiotic activities have been reported for a compound of this class, (+)-manool,<sup>[10]</sup> which is also recognized as a starting material for the semisynthesis of a wide range of diterpenoids with pharmaceutical properties.<sup>[11]</sup> SsSCS was used as a representative of class I diTPSs that, following rearrangement, facilitate neutralization of the carbocation through water quenching at C13 to give regiospecific introduction of a hydroxy group.<sup>[9]</sup> When offered the three known stereoisomers of copalyl diphosphate (CPP), SsSCS formed products with the stereochemistry preserved at the chiral centers C9 and C10, as defined by the class II diTPSs. Specifically, the pairing of SsSCS with diTPSs forming *ent*-CPP, (+)-CPP, or *syn*-CPP, resulted in the stereoselective formation of *ent*-manool (**23b**), (+)-manool (**23a**), and *syn*-manool (**11**), respectively, both in *N. benthamiana* and in vitro (Figure 2, Figure S16, and Tables S4, S5). Upon identification of the new combination of diTPSs that efficiently yields **23a**, CFTPS1 and SsSCS were codon optimized and stably integrated into the genome of a strain of *S. cerevisiae* engineered for diterpene production to afford **23a** at 310 mg L<sup>-1</sup> (Table S9), thereby effectively tripling previously reported titers.<sup>[12]</sup>

The highly efficient conversion of bicyclic diphosphate intermediates by SsSCS proved instrumental in elucidating the function of an uncharacterized class II diTPS. The combination of TwTSP14 with SsSCS produced a compound consistent with a diTPS function new to plants and only recently reported in the bacterium *Herpetosiphon aurantia*-



*cus*.<sup>[13]</sup> Purification and NMR analysis established the structure as kolavelool (**26**), from the distinct class of clerodane-type diterpenes (Table S6).<sup>[4b]</sup> These diterpenes have been reported from a range of plant species and form the skeleton

**Figure 1.** The diterpene chemoscape. A) Levels of diterpenes accumulating in the *N. benthamiana*/Agrobacterium system ( $n=3$ ). B) GC-MS chromatograms of extracts from the *N. benthamiana* control, CFTPS1/SsSCS expressed in *N. benthamiana* or in *S. cerevisiae*, the in vitro assay, and an authentic standard of (9S,10S)-manool (**23a**). Total ion current (TIC) and extracted-ion chromatogram (EIC) levels are not to scale. C) 2D representation of diterpenes detected in *N. benthamiana* expressing either a single diTPS or combinations of diTPSs from the two classes (y axis). Diterpenes are sorted on the x axis by their Kovats retention indices. Cf = *C. forskohlii*; Ss = *S. sclarea*; Mv = *M. vulgare*; Ep = *Euphorbia peplus*; Os = *O. sativa*; Tw = *T. wilfordii*; Zm = *Z. mays*. Diterpenes in samples expressing solely class II diTPSs are likely the result of endogenous phosphatase activity and non-specific rearrangement (Figure S7).<sup>[6a,7]</sup>

of salvinorin A, a lead drug candidate for the treatment of mental disorders and substance dependence,<sup>[14]</sup> yet the biosynthetic route remains unknown. Similarly, coupling of two functionally uncharacterized class II diTPSs with the nine class I diTPSs and comparison of the product profiles with those of established enzymes enabled the classification of EpTPS7 and TwTPS7 as *ent*-CPP synthases (Figure 2). Furthermore, combining *ent*-CPP synthases with EpTPS8 or EpTPS23 resulted in the formation of *ent*-neoabietadiene and *ent*-dehydroabietadiene, respectively (Figure S14), diterpene scaffolds not previously reported in nature. Likewise, the TwTPS21/SsSCS combination yielded *ent*-sclareol, which constitutes the first biosynthetic route to this specific stereoisomer.

Several of the combinations provided skeletons of high-value diterpenoids, for example, miltiradiene (**25**), the backbone of cardioprotectives (tanshinones)<sup>[15]</sup> and antioxidants (carsoic acid),<sup>[16]</sup> **16a** and **23a** as precursors for the drug forskolin,<sup>[6a]</sup> and the synthetic ambergris substitute (–)-ambrox,<sup>[6a]</sup> which is used in high-end fragrances.<sup>[17]</sup> For stereoselective production of these targets, expression-optimized variants of CftPS1/CftPS3, CftPS2/CftPS3, and TwTPS21/EpTPS1 were introduced stably into the genome of a *S. cerevisiae* strain engineered for diterpene production. Production titers between 250–370 mg L<sup>-1</sup> (Table S9) underline the potential for scale-up production by fermentation.

In summary, we show that engineered diTPS combinations can be used to expand the biosynthetically accessible repertoire of diterpene skeletons with superior stereochemical control. Our approach meets a long-standing desire for control over the stereochemical configuration of synthesized diterpenes, which remains a challenge in formal chemical synthesis. With chiral molecules representing more than half of approved drugs worldwide, collectively valued at over \$200 billion in 2008,<sup>[18]</sup> our findings make an important contribution to accelerate drug development and demonstrate how synthetic biology can complement chemical synthesis for the production of stereochemically complex small molecules.

## Experimental Section

Generation of expression constructs, expression in *N. benthamiana*, *S. cerevisiae* and *E. coli*, extraction analysis of products, and structural elucidation by NMR were done as previously described and are given in supplementary material and methods.<sup>[6a,19]</sup>



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- [1] a) C. J. Paddon, et al., *Nature* **2013**, *496*, 528; b) S. A. Wilson, S. C. Roberts, *Plant Biotechnol. J.* **2012**, *10*, 249; c) R. A. Holton, H. B. Kim, C. Somoza, F. Liang, R. J. Biediger, P. D. Boatman, M. Shindo, C. C. Smith, S. Kim, *J. Am. Chem. Soc.* **1994**, *116*, 1599; d) L. Jørgensen, S. J. McKerrall, C. A. Kuttruff, F. Ungeheuer, J. Felding, P. S. Baran, *Science* **2013**, *341*, 878.
- [2] G. Appendino, *Angew. Chem. Int. Ed.* **2014**, *53*, 927; *Angew. Chem.* **2014**, *126*, 943.
- [3] a) F. Chen, D. Tholl, J. Bohlmann, E. Pichersky, *Plant J.* **2011**, *66*, 212; b) I. Pateraki, A. Heskes, B. Hamberger in *Biotechnology of Isoprenoids, Vol. 148* (Eds.: J. Schrader, J. Bohlmann), Springer International Publishing, **2015**, p. 107.
- [4] a) P. Zerbe, J. Bohlmann, *Trends Biotechnol.* **2015**, *33*, 419; b) R. J. Peters, *Nat. Prod. Rep.* **2010**, *27*, 1521.
- [5] P. Zerbe, et al., *Plant Physiol.* **2013**, *162*, 1073.
- [6] a) I. Pateraki, et al., *Plant Physiol.* **2014**, *164*, 1222; b) A. Cyr, P. R. Wilderman, M. Determan, R. J. Peters, *J. Am. Chem. Soc.* **2007**, *129*, 6684.
- [7] P. Zerbe, A. Chiang, H. Dullat, M. O'Neil-Johnson, C. Starks, B. Hamberger, J. Bohlmann, *Plant J.* **2014**, *79*, 914.
- [8] C. A. Gray, D. E. Rivett, M. T. Davies-Coleman, *Phytochemistry* **2003**, *63*, 409.
- [9] A. Caniard, P. Zerbe, S. Legrand, A. Cohade, N. Valot, J.-L. Magnard, J. Bohlmann, L. Legendre, *BMC Plant Biol.* **2012**, *12*, 119.
- [10] M. R. Moreira, et al., *Rev. Bras. Farmacogn.* **2013**, *23*, 870.
- [11] a) A. W. W. van Wyk, K. A. Lobb, M. R. Caira, H. C. Hoppe, M. T. Davies-Coleman, *J. Nat. Prod.* **2007**, *70*, 1253; b) F. J. Salazar, J. E. Villamizar, *J. Chem. Res.* **2013**, *37*, 63; c) F. J. Salazar, J. E. Villamizar, *J. Chem. Res.* **2013**, *37*, 1.
- [12] C. Ignea, E. Ioannou, P. Georgantea, S. Loupassaki, F. A. Trikka, A. K. Kanellis, A. M. Makris, V. Roussis, S. C. Kampranis, *Metab. Eng.* **2015**, *17*, 91.
- [13] C. Nakano, M. Oshima, N. Kurashima, T. Hoshino, *ChemBioChem* **2015**, *16*, 772.
- [14] L. Kutrzeba, F. E. Dayan, J. L. Howell, J. Feng, J.-L. Giner, J. K. Zjawiony, *Phytochemistry* **2007**, *68*, 1872.
- [15] J. Guo, et al., *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 12108.
- [16] K. Brückner, et al., *Phytochemistry* **2014**, *101*, 52.
- [17] H. R. Schenk, H. Gutmann, O. Jeger, L. Ruzicka, *Helv. Chim. Acta* **1954**, *37*, 543.
- [18] J. Caldwell, *Hum. Psychopharmacol.* **2001**, *16*, S67.
- [19] S. Spanner Bach, J.-E. Bassard, J. Andersen-Ranberg, M. Emil Møldrup, H. Toft Simonsen, B. Hamberger in *Methods of Molecular Biology: Plant isoprenoids, Vol. 1153* (Ed.: J. M. Walker), Springer, Heidelberg, **2014**, p. 330.

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