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# **APPENDICES**

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## Appendix 1

Phang, S. M., 1995. Distribution & abundance of marine algae on the coral reef flats at Cape Rachado, Port Dickson, Peninsular Malaysia. *Malaysian Journal of Science* 16A: 23-32

Phang, S. M., 1988. The effect of siltation on algal biomass production at a fringing coral reef flat, Port Dickson, Peninsular Malaysia. *Wallaceana* 51: 3-5

## Distribution and abundance of marine algae on the coral reef flats at Cape Rachado, Port Dickson, Peninsular Malaysia

Phang

Advanced Studies, University of Malaya, 59100 Kuala Lumpur, Malaysia

**ABSTRACT** The species distribution and abundance of marine algae on three fringing coral reef flats at Cape Rachado, Port Dickson (on the west coast of Peninsular Malaysia) were investigated using line-transect and quadrat-sampling methods. Data collected over twelve months on the species composition, distribution and biomass are discussed in relation to environmental factors. In all sites, 15 species were recorded, the dominant belonging to *Ulva*, *Turbinaria* and *Padina*. Highest biomass (67, 103, 95 g m<sup>-2</sup>) were recorded at sites A, B and C) occurred during the inter-monsoon period, August and March. Biomass and the total number of species increased towards the reef edge. High levels of total suspended matter during exposure periods, rainfall and strong wave action reduced biomass production. Site A, the most silted area had the highest biomass whereas Site B, the area most exposed to human activity, had the highest.

**KEYWORDS** Kepelbagaian spesies alga marin di tiga sempadan terumbu karang di Cape Rachado, Port Dickson (di pantai barat Semenanjung Malaysia) telah dikaji dengan menggunakan kaedah transek dan persampelan kuadrat. Data yang dikumpulkan dalam jangka masa dua belas bulan bagi komposisi spesies, taburan dan biojisim telah dibincangkan dan dikaitkan dengan faktor-faktor persekitaran. Enam puluh sembilan spesies alga telah direkodkan; kebanyakannya ialah *Sargassum*, *Turbinaria* dan *Padina*. Biojisim yang tertinggi (67, 103 dan 95 g m<sup>-2</sup>) untuk Kawasan A, B dan C) didapati di musim antara monsoon Ogos dan Mac. Biomassa dan jumlah spesies bertambah berdekatan tepi terumbu karang. Bilangan jumlah pepejal terapai yang tinggi, masa terdedah kepada hujan dan kesan ombak yang kuat mengurangkan penghasilan biojisim. Kawasan A, di mana terdapat kelodak yang banyak, mempunyai biojisim yang terendah manakala Kawasan B, yang paling banyak terdedah kepada gangguan manusia, mempunyai biojisim yang tertinggi.

Keywords: algae, coral reefs, biomass, environment)

### INTRODUCTION

Fringing coral reefs have been little studied and early research consisted mainly of taxonomic enumerations of species found in Sabah [1-3], Pulau Redang [4], Pulau Paya [5] and the other islands of Peninsular Malaysia [6-8]. The most recent reports dating back more than ten years describe the coral community structure of the fringing coral reef [9] at Cape Rachado, Port Dickson, and its epiphytic algae [10].

The distribution and ecology of marine algae of Peninsular Malaysia [4,10-16] and Sabah [17] have been documented in several reports. Physiological and biochemical studies have also been conducted on selected species of marine algae [18-20], and environmental factors such as light, temperature, nutrients and photoperiod have been shown to control both reproduction and vegetative growth in these algae [21]. The studies also confirmed that seasonal behaviour in other temperate algal species are more marked compared with those of the tropics. However, other observations [22] showed that the macro class of algae, i.e., the seaweeds, that grow on the rocks and coral reefs in Townsville, Australia, exhibit somewhat marked seasonal patterns in growth and reproduction. For this particular tropical locality, the patterns could be attributed to both temporal differences and variations in habitats.

Cape Rachado in Port Dickson (on the west coast of Peninsular Malaysia) is a segment of a stretch of beach that has been earmarked for the construction of tourist resorts. Such construction would, unfortunately, adversely change its coral reef formation. This paper documents the species distribution and biomass (dry weight) estimates of the marine algae found on three fringing coral reef flats, and discusses the effects of silt content in the water, exposure period, wave action and temperature and seasonal growth patterns on the algal distribution and abundance.

### MATERIALS AND METHODS

**Study site** Cape Rachado faces the Straits of Melaka and the fringing coral reef flats (Fig. 1) extend out for about 50 m; beyond this distance, they gently slope towards the reef edge. Here, they slope off to a depth of about 8 m [9]. Three sites were selected for the study. Site A has a reef flat extending to a maximum of about 140 m to the reef edge and is about 200 m wide. The mangroves to the right of the seafront had been partially removed during the construction of a condominium in 1985-1986. The destruction had resulted in heavy

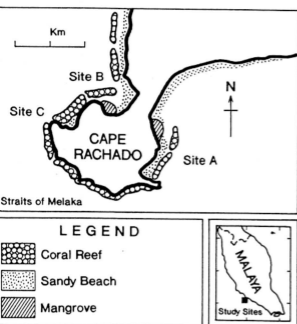


Figure 1. Map of study sites at Cape Rachado, west coast Peninsular Malaysia.

ing over the coral reef, causing damage to the coral and marine algal communities. Site B stretches to a maximum of about 110 m out to sea and is about 350 m wide. This area is popular for picnicking. The bay is bordered by a small stand of mangroves on the left and a rocky shore on the right. Site C is the least exposed to man disturbance as the sea is rather deep. The maximum length is about 110 m and it is about 300 m wide. The bay here is surrounded by a rocky shore.

If the tide level is about 0.3 m or less, three-quarters of the reef flats are exposed. At less than 0.1 m above chart datum, the reef flats are completely exposed. The reefs were exposed 27 times during the sampling period. At extremely low tides, the reef flats are exposed for more than three hours. Tides are at their lowest level around August and between February and March.

An earlier study [9] in 1987 found that the nitrate levels along the coast at Port Dickson ranged from 2.17 to 6.27  $\mu\text{g L}^{-1}$ , ammonia levels between 3.92 to 6.49  $\mu\text{g L}^{-1}$  and phosphate levels between 0.75 and 17.91  $\mu\text{g L}^{-1}$ . Light penetration ranged between 2 m and 7 m as measured with a Secchi disk.

**Sampling procedure** The study period was from May 1987 to March 1988; sampling was carried out in May, August and November 1987, and March 1988 during

the lowest spring tide of the particular month. The beginning of the reef flats that is parallel to the shoreline was regarded as being the baseline for the measurements. Line transects were then laid at intervals of 40 m at Site A (seven) and Site B (four), and at intervals of 60 m at Site C (four). The line transects traversed the flats and were perpendicular to the baseline. Along each transect, poles were marked as stations at 10-m intervals; samplings were carried out in 0.09  $\text{m}^2$  quadrats. The algal material within each quadrat was collected and placed in plastic bags to be brought back to the laboratory and sorted according to species. The dry weight of the total biomass of each quadrat was determined. Dry weights were recorded only on those species that were abundant. Each species was assigned a proportion number, N, according to its percentage of the total wet biomass (Table 1). The proportion numbers were used in calculating the Diversity Index.

Table 1. Distribution of biomass.

Percentage of biomass (%)	N
0 - 10	1
11 - 20	2
21 - 30	3
31 - 40	4
41 - 50	5
51 - 60	6
61 - 70	7
71 - 80	8
81 - 90	9
91 - 100	10

**Data processing** The algal material was dried at 100°C for 72 hours in the *in situ* determination of biomass, but this method would exclude losses arising from detachment and grazing [24]. An Orion pH meter was used to measure the acidity. Dissolved oxygen levels were determined by the Winkler Method. Salinity (measured by a hand refractometer), temperature and the level of total suspended solids (TSS) were determined at each sampling time. The total suspended solids content was determined by filtering a fixed volume of sample onto a preweighed glass fibre filter, and dried to constant weight at 100°C. The acidity and salinity were determined at every quadrat and water samples from every



ere collected for TSS determination. Total biomass (dry weight) was calculated in s for each quadrat. Less quadrats were exam- g high water levels (in November). Number of species in a quadrat was counted the Shannon-Weiner Diversity Index, H was from standard procedures [25]:  $H = - \sum p_i \log p_i$  s = number of species and  $p_i$  = proportion of n the quadrat. The Evenness Index, J, is  $H/\log$  rensen's Quotient of Similarity, S, of the three given by  $S = [2C/(A+B)]$ , where A = number in Site A, B = number of species in Site B, and er of species in the two sites being compared. equencies (F) of each species in the transects sites were calculated by dividing the number ts in which a species was found by the total f quadrats. The dominance of each species, in the ratio of the abundance of the species to the ce of all species in the quadrat, was calculated data of the March 1988 sample, which was representative.

## RESULTS

### Composition and distribution

he species were recorded in the three sites 2 and 3). The three sites displayed similar ions, as suggested by the Sorensen's Quotient; somewhat more similar to Site C (Table 4). e seaweeds found in the three sites, the brown aeophyta (*Sargassum*, *Turbinaria* and *Padina*) st abundant. The red algae Rhodophyta (*Pte- caerulescens*) were less abundant, but they had st number of species. Table 5 gives the more species at the four sampling times. *Lobophora* a was the third most common species. With the ntya family, *Udotea javensis*, *Struvea anasto-*

Number of species belonging to the different divisions in sites.

No. sites	Phaeophyta	Chlorophyta	Rhodophyta	Cyanophyta
	12	15	22	2
	14	10	26	0
	18	8	19	0
	19	18	30	2

*mosans* and *Cladophora vagabunda* were the most common species; however, *Turbinaria* sp. 1, *Valonia* sp., *Udotea flabellum*, *Herposiphonia*, *Gelidium* sp. 3, *Tolypocladia glomerulata*, *Lyngbya majuscula*, *Caulerpa serrulata* and *Caulerpa taxifolia* were only found at Site A, and *Enteromorpha intestinalis*, *Cladophora* sp. 2, *Gelidium* sp. 2, *Galaxaura* sp., *Gelidiella bonetii* only at Site B, and *Hormophysa triquetra*, *Acetabularia* sp., *Laurencia* sp. 3 and *Hydroclathrus clathratus* only at Site C.

The number of species increased away from the shore (Fig. 2), decreasing somewhat at the edge of the reef. The lowest number of species per quadrat in Site A was unity (August and November) and the highest 16 (March). For Site B, this was again unity (all months except March) and 14 (March), whereas for Site C, this was 2 (August) and 12 (March). March appeared to have provided more suitable conditions for the growth of a diverse algal community than November, during which month rain storms were frequent.

### Shannon-Weiner Diversity Index

The Diversity Indices suggest trends similar to those of species number, both values increasing seawards (Fig. 2). The indices for the three sites were lower nearer the shore, but increased from the middle of the flat (around quadrat 6 to 10) to near the reef edge. Site B had the highest diversity indices, the maximum being 1.29 (March) compared with 1.12 in Site A (March) and 1.05 in Site C (March).

### Dominance

Dominance was calculated for the three sites from March 1988 (Table 6) and for each station/quadrat, from the baseline to the reef edge. At Site A, *Gracilaria salicornia* was co-dominant with *Acanthophora* sp. from quadrats 1 to 2 whereas *Caulerpa racemosa*, and *Caulerpa serrulata* were co-dominant at quadrats 2 and 3. The dominant species were *Ceratodictyon* sp. and *Sargassum oligocystum* in quadrats 5 to 7 but nearer the edge (from quadrats 8 to 13), *Sargassum oligocystum* was also co-dominant with *Sargassum baccularia*.

At Site B, *Sargassum oligocystum* was dominant from quadrats 1 to 11, with *Ceratodictyon* being co-dominant at quadrat 1 and *Turbinaria conoides* co-dominant from quadrats 9 to 11. *Sargassum siliquosum*, which was absent from the first few quadrats, was dominant in quadrats 9 and 10.

At Site C, *Sargassum oligocystum* and *Turbinaria*

Table 3. List of algae recorded in the sites.

#### CHAEOPHYTA

*Sargassum oligocystum* Montagne  
*Sargassum baccularia* (Mertens) C. Agardh  
*Sargassum siliquosum* J. Agardh  
*Sargassum* sp.1  
*Sargassum* sp.2  
*Padina commersonii* Bory de Saint-Vincent  
*Padina tetrastromatica* Hauck  
*Arbinaria conoides* (J. Agardh) Kutzing  
*Arbinaria ornata* (Turner) J. Agardh  
*Arbinaria* sp.1  
*Erythrocladia caeruleascens* (Kutzing) Santelices  
*Dictyota bartayresii* Lamouroux  
*Dictyota dichotoma* (Hudson) Lamouroux  
*Dictyota ceylanica* Kutzing  
*Dictyota cervicornis* Kutzing  
*Acanthophora variegata* (Lamouroux) Womersley  
*Polysiphonia triquetra* C. Agardh  
*Hydroclathrus clathratus* (Bory de Saint Vincent) Howe  
*Cladocarpus*

#### PHLOROPHYTA

*Podocladia javensis* (Montagne) A. & E.S. Gepp  
*Podocladia flabellum* (Ellis & Sonder) Howe  
*Chaetomorpha linum* (Muller) Kutzing  
*Cladophora fascicularis* (Mertz) Kutzing  
*Cladophora prolifera* (Roth) Kutzing  
*Cladophora* sp.1  
*Cladophora* sp.2  
*Enteromorpha intestinalis* (Linnaeus) Nees  
*Enteromorpha clathrata* (Roth) Greville  
*Enteromorpha aegagropila* C. Agardh  
*Enteromorpha* sp.1  
*Enteromorpha anastomosans* (Harvey) Piccone & Grunow ex Piccone  
*Enteromorpha* sp.2  
*Enteromorpha* sp.3  
*Enteromorpha* sp.4  
*Enteromorpha* sp.5  
*Enteromorpha* sp.6  
*Enteromorpha* sp.7  
*Enteromorpha* sp.8  
*Enteromorpha* sp.9  
*Enteromorpha* sp.10  
*Enteromorpha* sp.11  
*Enteromorpha* sp.12  
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#### PHODOPHYTA

*Porolithothamnion* sp.1 (cf. *concreta* Cribb)  
*Porolithothamnion* sp.2  
*Porolithothamnion* sp.3  
*Porolithothamnion* sp.4  
*Porolithothamnion* sp.5  
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*Porolithothamnion* sp.100

*Leveillea jungermannioides* (Hering & Martens) Harvey  
*Centroceras*  
*Herposiphonia*  
*Gelidiella acerosa* (Forsskal) Feldmann & Hamel  
*Gelidiella bornetii* (Weber-van Bosse) Feldmann & Hamel  
*Gelidiella* sp.1  
*Gelidium* sp.1  
*Gelidium* sp.2  
*Gelidium* sp.3  
*Acanthophora specifera* (Vahl) Boergesen  
*Acanthophora orientalis* J. Agardh  
*Galaxaura* (cf. *filamentosa* Chou)  
*Tolytiocladia glomerulata* (C. Agardh) Schmitz  
*Erythrotrichia*

#### CYANOPHYTA

*Lygbya majuscula* Harvey  
*Nostoc*

Table 4. Sorensen's Quotient of Similarity (S)% for all sites (March 1988).

Site	A	B	C
A	100		
B	74	100	
C	68	79	100

*conoides* were co-dominant from quadrats 1 to 10, but at quadrat 11, *Sargassum siliquosum* was again abundant and co-dominant with *Sargassum baccularia*.

#### Faunal communities

At the three sites, the coral community was dominated by the hermatypic scleractinians. At least three species of alcyonaceans (soft corals) were found from the middle of the reef flats to the edge. Nearer the shore, species of *Porites* dominated whereas in the middle, *Goniastrea* and *Goniopora* dominated.

Large coral formations were observed at Site A, where the sediment load was highest near the shore; however, the polyps that were directly exposed to the sediments had been destroyed, although the polyps at the vertical planes of the corals were still alive. At Site B, *Porites enidari* Umbgrove that dominated in the first few quadrats had been trampled on by people. At Site C, *Porites lutea* Milne Edwards developed branches and ramose forms, probably as an adaptation to the stress of sedimentation. Although the reef edge had a live coral cover of 60%, the reef flat had a live coral cover of only 27%.

Table 5. Average frequencies (%) of the most commonly occurring species for all sites at all sampling times.

Site	May 1987	August 1987	November 1987	March 1988
A	<i>Padina commersonii</i> (59%) <i>S. oligocystum</i> (55%)	<i>Sargassum oligocystum</i> (58%) <i>P. commersonii</i> (56%)	<i>P. commersonii</i> (35%) <i>Pterocladia caerulescens</i> (28%)	<i>P. commersonii</i> (50%) <i>S. oligocystum</i> (48%)
	<i>Lobophora variegata</i> (45%) <i>S. baccularia</i> (37%)	<i>S. baccularia</i> (38%) <i>Turbinaria conoides</i> (36%)	<i>L. variegata</i> (25%) <i>Udotea javensis</i> (25%)	<i>S. baccularia</i> (48%) <i>Pterocladia caerulescens</i> (41%)
B	<i>S. oligocystum</i> (75%) <i>T. conoides</i> (67%) <i>L. variegata</i> (48%) <i>P. commersonii</i> (43%)	<i>S. oligocystum</i> (82%) <i>T. conoides</i> (68%) <i>P. commersonii</i> (53%) <i>S. baccularia</i> (52%)	<i>P. commersonii</i> (72%) <i>T. conoides</i> (62%) <i>S. oligocystum</i> (61%) <i>Gelidiella acerosa</i> (53%)	<i>S. oligocystum</i> (85%) <i>Champia parvula</i> (84%) <i>T. conoides</i> (77%) <i>Hypnea</i> sp.3 (77%)
C	<i>S. oligocystum</i> (91%) <i>P. commersonii</i> (84%) <i>T. conoides</i> (56%) <i>Laurencia</i> sp.2 (42%)	<i>S. oligocystum</i> (93%) <i>T. conoides</i> (83%) <i>Amphiroa fragillissima</i> (63%) <i>Pterocladia caerulescens</i> (60%)	<i>S. oligocystum</i> (87%) <i>P. commersonii</i> (81%) <i>T. conoides</i> (61%) <i>G. acerosa</i> (50%)	<i>S. oligocystum</i> (87%) <i>T. conoides</i> (83%) <i>P. commersonii</i> (63%) <i>L. variegata</i> (62%)

corals (*Sarcophyton ehrenbergi* Von Marenzeller and *Lobophora pauciflorum*) were common from the mid-reef flat to the edge.

al species of coral fish were encountered, many probably fed directly on the macroalgae as well as the turf algae. Two species of sea cucumber, *Stichopus* sp. and *Stichopus* sp. were observed in the sites, being most abundant in the cleaner waters of

#### Experimental conditions and abundance

lists the average biomass (standing crop) of the sites at the various sampling times. The biomass of 103 g m<sup>-2</sup> is low.

st (1987) and March (1988) were the two periods of maximum biomass production; the latter month had the maximum yield for the three sites. July 1987

Table 6. Dominance of the marine algae at the three study sites.

	Site A	Site B	Site C
Dominant species	<i>Sargassum oligocystum</i> (18.2%)	<i>S. oligocystum</i> (21.6%)	<i>S. oligocystum</i> (28.3%)
Co-dominant species	<i>Sargassum baccularia</i> (9.3%) <i>Ceratodictyon</i> (8.0%)	<i>Turbinaria conoides</i> (13.8%)	<i>Turbinaria conoides</i> (24.7%)
Sub-dominant species	<i>Udotea javensis</i> (0.8%) <i>Laurencia</i> (0.8%)	<i>Gracilaria salicornia</i> (0.2%) <i>Udotea javensis</i> (0.9%)	<i>Acanthophora spicifera</i> (1.0%)

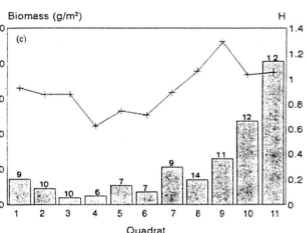
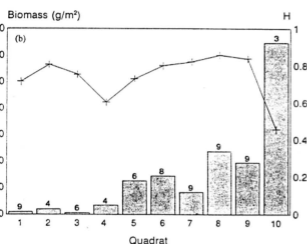
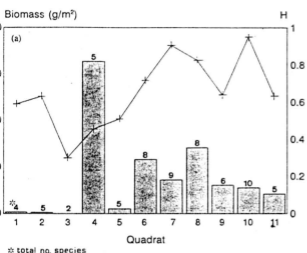


Figure 2. Biomass ( $\text{g}/\text{m}^2$ ) and Shannon-Weiner Diversity Index ( $H'$ ) for Transect A at Site A, March 1988; Transect L at Site B, March 1988; and Transect Q at Site C, March 1988.

Table 7. Average algal biomass ( $\text{g}/\text{m}^2$ ) of all sites at all sampling times.

Date	Site A	Site B	Site C
May 1987	16.9 $\pm$ 9.7	39.1 $\pm$ 25.1	32.1 $\pm$ 2.3
August 1987	41.2 $\pm$ 13.5	83.8 $\pm$ 31.2	73.4 $\pm$ 5.4
November 1987	28.1 $\pm$ 17.8	43.4 $\pm$ 21.8	32.2 $\pm$ 11.4
March 1988	66.6 $\pm$ 44.3	103.5 $\pm$ 29.7	95.2 $\pm$ 32.7

Table 8. Range of the physical-chemical parameters measured in the study sites.

	Site A	Site B	Site C
pH	8.10 - 8.50	8.10-8.50	8.00 - 8.65
salinity ppm	29.0 - 34.0	29.0-33.0	30.0 - 33.0
*temperature C	30.5 - 31.5	30.5-31.0	30.5 - 31.0
*D.O. ppm	7.30 - 8.60	6.40-8.90	7.35 - 8.40
TSS ppm	17.2 - 1502.1	14.3-108.4	7.6 - 100.8

(\* : taken at noon, time of sampling)

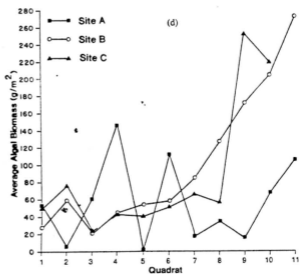
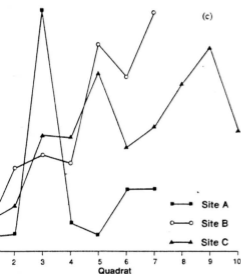
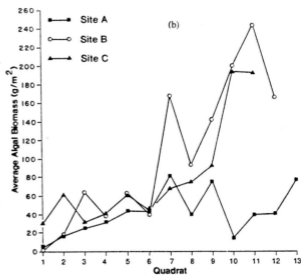
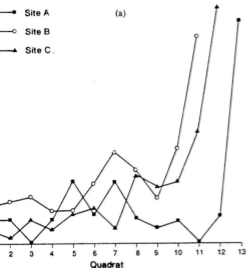
(mean maximum air temperature of 31.5°C) and February 1988 (mean maximum air temperature of 32.5°C) 1988 were hot and dry months. July 1987 and February 1988 had rainfalls of 130.5 mm and 157.4 mm; August 1987 and March 1988 had averages of 227.4 mm and 273.3 mm. A hot dry period preceding a wetter period appeared to have enhanced biomass production. Site B had the highest biomass. Biomass at the three sites increased away from the shoreline (Fig. 3).

The acidity, salinity, temperature, dissolved oxygen and TSS values taken during the sampling times (Table 8) showed no significant trends.

## DISCUSSION

The coral reefs at Cape Rachado have a *Sargassum-Turbinaria* community whose flora is dominated by *Sargassum oligocystum*, *S. baccularia*, *Turbinaria conoides* and *Padina commersonii*. The literature also reports that *Sargassum* spp. were the dominant algae on reef flats at Pulau Bidong Laut [14] and Magnetic Island, Australia [26].

The level of TSS of the reefs points to the high level of pollution. The reefs were completely exposed for about 27 times a year. These two factors probably limited the species that could grow here. *Sargassum*, *Turbinaria* and *Padina*, being physically large and



Average algal biomass ( $\text{g}/\text{m}^2$ ) in quadrats from the land (quadrat 1) to the reef edge (a) May 1987, (b) August 1987, (c) November 1987 and (d) March 1988.

thrived because they could grow above the silt. Species in quadrats nearer to shore included *Enteromorpha* sp., *Enteromorpha* sp., *Lyngbya* sp., *Lyngbya* sp., *Valonia* sp., *Ceratodictyon* sp., *Udotea* sp., *Udotea flabellum* and *Gracilaria salicornia*. The presence of the first two in heavily silted quadrats suggested their tolerance of high sediment load in the water. *Sargassum siliquosum*, *Sargassum siliquosum* and *Amphiroa foliacea* were found mainly near the reef edge. Species found throughout the reef included *S. oligocystum*, *P. commersonii*, *T. conoides* and *Lobophora variegata*. *Lobophora variegata*, which

encrusts over the dead coral surfaces, was dominant because of its tolerance to desiccation and wave force [28]. *Acanthophora* and *Enteromorpha* were also found nearer the shore on the reef flat at Pulau Bidong Laut [14]. However, all algal species were distributed randomly on a rocky shore on Penang island [11]. The apparent zonation observed in the present study can be attributed to sediment load. *Gracilaria salicornia* was more abundant near the baseline possibly because of its ability to withstand the higher sediment load and longer exposure periods.

The most common of the 16 epiphytic algae be-

longed to the Rhodophyta (*Champia parvula* and *Polysiphonia*), followed by *Cladophora fascicularis*. The hosts were mainly *Sargassum* and *Turbinaria*. These epiphytes were found throughout the reef flat but were more common in the middle. High sediment loads and strong wave action at the reef edge probably contributed to the establishment of epiphytic communities. The epiphytes in the three sampling sites were most abundant in March [29]. Their pattern of growth paralleled that of their hosts, as observed previously [30]. Cape Rachado has a salinity that lies at the lower end of the range [31] of Shark Bay, Australia, which would explain its richness of species. *Leveillea jungermanniodes* and *Tolypiocladia glomerulata*, which are typically found at low salinity levels, were however relatively common. Many of the epiphytes, particularly *Polysiphonia* and *Champia*, were fertile during March, at which time biomass abundance peaked [29].

Algae did not grow in those quadrats dominated by the soft corals which secrete defense chemicals that inhibit the growth of other organisms. Their spicules [32] would also prevent other moving organisms from encroaching.

There were more species towards the edge of the reef as there was less sediment load but at the edge, the number decreased owing to strong wave action.

*Gracilaria salicornia*, *S. oligocystum* and *Ceratodictyon* sp. dominated the heavily sedimented areas whereas *S. siliquosum* and *S. baccularia* preferred the cleaner waters near the reef edge. Quadrats in the major part of the reef flats had several co-dominant species but near the edge, only one species was more dominant.

The seaweeds were physically smaller nearer the shore: *Sargassum baccularia*, *Turbinaria conoides* and *Padina* found here were damaged. These consisted largely of holdfasts and stipes with missing fronds; on the other hand, larger complete seaweeds were found near the edge. The difference in appearance was more marked at Site B owing to more frequent human disturbance.

Shorter exposure periods from the middle to the edge would decrease sediment load and improve biomass production. Wave action, detrimental to biomass production of *Gracilaria verrucosa* [36], could account for the decreased biomass at the reef edge. This trend was also noted in studies of algae on a rocky shore in Singapore [33], and on corals at Eilat, Red Sea [34,35]. In this study, trends in biomass production from one side of the reef area to the other were not evident.

The TSS contents were significantly different for the three sites, with Site A (51 species) having the highest sediment load and Site C (45 species) the lowest. Although living organisms would be smothered by sediments that block out sunlight, the algae at Cape Rachado apparently tolerated the high sediment load, growing well even at a TSS of 100 ppm (Site B). A value above 100 ppm was reflected in decreased biomass production (Site A). Biomass production at Site B exceeded that at Site C as frequent trampling by humans, which would stir up nutrients, stimulated the growth of the algae. The acidity and salinity did not vary significantly in the sites at the various sampling times. Other reports [36,37] suggested that salinity did not contribute to the distribution and biomass production of *Gracilaria* species.

The algal biomass in the sites peaked around August and March; these months correspond to the hot and dry inter-monsoon periods. Such conditions favour biomass production [38]. In Guam, *Sargassum* (except *S. cristaefolium*) attains maximum growth during the warmer months in the temperate areas and during the colder months in the tropical and sub-tropical areas. However, in Puerto Penasco, Mexico, *Sargassum* had higher growth rates from October to March (the coldest part of the year), peaking in spring. Since peak abundance [39] coincides with peak fertility [27], the *Sargassum* species in Puerto Penasco peak in spring so as to avoid reproducing during the seasonal extremes. In the tropics, most *Sargassum* species grow most abundantly in the cooler months (from November to March [27,40,41]), whereas *S. siliquosum* and *S. paniculatum* grow well in the warmer months (from July to August). In the Philippines, the growth was the least in the coldest months. The species were fertile at the end of the rapid growth period, about one month after the plants had reached maximum growth. Another study [42] has claimed good correlations between maximum plant growth and high water-phosphate content, and between growth rate and daylight hours. Lower seawater temperatures have been suggested to initiate the onset of fertility; lower temperatures bring about a decline in standing crop but they contribute towards embryo growth and development [27]. Another study on algal succession patterns on the rocky shore on Penang island [13] showed that maximum biomass production was obtained in the hot dry inter-monsoon period (from January to April), but rainstorms caused scanty growth. Strong wave action and high rainfall during the monsoon

sulted in low standing crop of *Gracilaria* in the lines [36]. In the present study, heavy rains and waves during the monsoons (especially in November) resulted in low biomass; the end of the monsoon was followed by a period of active growth. Variations of the microclimatic conditions showed a wet and dry spell at the end of the monsoon period. Growth rates were highest during the warm period following the monsoon. *Polysiphonia* and *Champia* were most abundant in March. Biomass nearer to the shore was lower than at nearer the reef edge, possibly arising from the effect of the longer duration of exposure at low tides, although the total biomass of the reef flat was not significantly different.

Minor differences in temperature and light intensity between monsoons and inter-monsoon periods probably gave rise to the observed patterns of growth and production of all algae. For the epiphytic species, maximum points of biomass and of fertility were observed that could be attributed to abrupt changes in microclimate.

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# THE EFFECT OF SILTATION ON ALGAL BIOMASS PRODUCTION AT A FRINGING CORAL REEF FLAT, PORT DICKSON, PENINSULAR MALAYSIA.

PHANG SIEW-MOI

Institute of Advanced Studies, University of Malaya, Kuala Lumpur

## INTRODUCTION

The marine environment in Malaysia is threatened by various pollutants arising from both land-based sources and from the heavy traffic in its waters. The pollutants include silt in the form of terrestrial and marine sediment, oil, pesticides and detergents (Phang *et al.*, 1987). Silt comes from both inland development schemes and construction along the coastline. Damage to coral reefs at Pulau Tioman on the east coast has been reported as being attributed to sedimentation and the use of explosives for mining (De Silva *et al.*, 1984). On the west coast, heavy land development activities result in higher sediment loads being brought by the rivers into the waters of Melaka. Sheltered by Sumatra, the sedimentation problem is enhanced. Much of the corals on Pulau Besar, Melaka are dead or dying, smothered by a layer of silt (Phang *et al.*, 1987).

Lack of light and excessive sediment deposition are factors which limit but not prohibit coral development (Roy & Smith, 1971). Many coral species have cleaning mechanisms to remove the sediments. Species of *Favia*, *Fungia* and *Porites* have been shown to survive in water containing 800 mg of suspended mud (Marshall & Orr, 1931). High sediment loads were observed to control species diversity and cover of both coral and algal communities on a fringing reef flat at Magnetic Island, Australia (Morrissey, 1980). The sedimentation parameters increased seawards across the reef flat as emersion, sedimentation and water movement decreased.

## METHOD

Cape Rachado at Port Dickson, has fringing coral reefs extending almost all around it. The study site is a fringing reef flat in the bay on the east of the island. It extends to a maximum of 140m to the reef edge. To the right was a stand of mangroves dominated by species of *Rhizophora*. Construction of a condominium complex at the beach just behind the mangroves started in 1985 and was completed in 1987.

The land which slopes down to the sea was cleared and a major portion of the mangroves were removed for 'aesthetic' reasons. This resulted in an excessive amount of silt being washed into the reef flat.



*Sargassum* spp. are the dominant algae on the heavily sedimented reefs at Cape Rachado.

Biomass estimations of the marine algae growing on the coral reef flats around Cape Rachado were carried out in March 1986, and from May 1987 to March 1988. Only results from this particular site and for three sampling periods are discussed. The sampling periods were March 1986, August 1987 and March 1988. They correspond to the early part of the construction, the end and about 7 months after completion of the complex respectively.

Line transects were laid perpendicular to a baseline which ran parallel to the shore at the start of the reef flat. Seven transects at intervals of 40m were used each time. 0.3 m<sup>2</sup> quadrats were placed at 10 m intervals along the transects and all algal material within the quadrats were collected as far as possible, and brought back to the laboratory for sorting. This was done according to species, but only the total dry weight of each quadrat was determined. This was done by drying the algae at 100°C for 72 hours. Many species were too small for individual dry weights to be assessed. However, each species was assigned a proportion number according to its percentage of the total wet biomass in the quadrat as follows:

Percentage Algal Biomass	Proportion Number
— 10	1
— 20	2
— 30	3
— 40	4
— 50	5
— 60	6
— 70	7
— 80	8
— 90	9
— 100	10

Numbers were used for calculating the Shannon-Weaver Diversity Index (Shannon & Weaver,

## RESULTS AND DISCUSSION

gives the average pH, salinity and total dissolved solids (TSS) content and the average algal biomass (g/m<sup>2</sup>) for the different sampling periods. There were no significant differences in pH or salinity. In March 1986, after construction had ceased, the TSS ranged from 17 to 150 mg/l. The highest TSS determined was 1530 mg/l at a depth of 60 m from the baseline. In 1988, the TSS increased, ranging from 40 to 107 mg/l. The average biomass of the algae was observed to be inversely proportional to the TSS content, being 80 g/m<sup>2</sup> in 1986 and lowest (32 g/m<sup>2</sup>) in 1988, increased again in 1988 (67 g/m<sup>2</sup>) following a decrease in TSS content.

Table 2 shows the total number of species per quadrat (S), the Shannon Weaver Diversity Index (H') and the Evenness Index (J) for selected transects at the three sampling periods. The Shannon Diversity Index did not change significantly for the three sampling periods. This may be because the species inhabiting this reef are tolerant of high sediment loads and were able to survive in decreased abundance. The index also followed the same trend as the total number of species per quadrat, both increasing towards the edge but decreasing slightly at the edge. High sediment loads and longer exposure near shore and stronger wave action at the edge are possible reasons for this trend. The

highest number of species per quadrat encountered in 1986 was 13, in 1987 was 10, and in 1988 was 16.

The dominant species on this reef belong to *Sargassum*, *Turbinaria* and *Padina*. Having a much dissected thallus and being flexible in the water probably help to prevent these plants being smothered by the sediments. *Udotea javensis*, *Dictyota* spp., *Gracilaria salicornia* and *Pterocladia* were also frequently observed in the reef flat.

There were no significant differences in the most frequently occurring species between the sampling periods. *Sargassum binderi*, *Padina commersoni* and *Turbinaria conoides* were most frequently occurring at all times. *Avrainvillea* sp., *Leveillea junggermanniodes* and *Microdictyon* sp. which were observed in March 1986 were not observed again. *Tolyptocladia glomerulata* and *Caulerpa racemosa* which was present in March 1986 but disappeared in 1987 was collected again in 1988.

## CONCLUSION

Observations show that increased TSS in the water due to the nearby construction, resulted in decreased algal biomass production at the coral reef flat. Many of the corals belonging to species of *Porites* and *Goniostrea* nearer shore were dead and covered by a layer of sediment. However algal biomass production appeared to recover quite soon after the end of construction. Species diversity did not change significantly throughout the sampling periods, showing that although abundance decreased, most of the original species could have remained throughout the construction period. Species like *Avrainvillea* and *Microdictyon* which are perhaps more vulnerable to heavy sediments loads were however not observed again. This reef flat lies in a protected area which would naturally have a higher sediment load than other reef and coastal areas. Species growing here are quite tolerant of high sediment loads and were able to survive the sudden increase, although with the 'loss' of a few species. Species growing on reefs in 'cleaner' waters may not be able to do the same, and irreversible damage may occur to our coral reefs if our coastline and our many islands are not protected from unnecessary development.

Table 1

The four parameters at the three sampling times

Parameter	March 1986	August 1987	March 1988
pH	8.25	8.25	8.26
Salinity mg/l	31	30	32
TSS mg/l	17 - 150	78 - 1503	40 - 107
Algal Biomass g/m <sup>2</sup>	80	32	67

Table 2

Total number of species per quadrat (S), Shannon Weaver Diversity Index (H) and the Evenness Index (J) for the three sampling periods.

Quadrat	March 1986			August 1987			March 1988		
	S	H	J	S	H	J	S	H	J
0 (baseline)	1	—	—	1	—	—	4	0.589	0.978
1	4	0.565	0.939	6	0.634	0.815	5	0.630	0.901
2	8	0.828	0.917	6	0.614	0.789	2	0.301	0.999
3	6	0.728	0.936	10	0.954	0.954	5	0.454	0.650
4	10	0.941	0.941	7	0.756	0.895	5	0.509	0.728
5	7	0.797	0.662	9	0.886	0.928	8	0.716	0.793
6	7	0.687	0.812	5	0.638	0.913	9	0.905	0.948
7	5	0.549	0.493	5	0.627	0.897	8	0.825	0.914
8	7	0.491	0.630	7	0.784	0.704	6	0.640	0.822
9				6	0.637	0.819	10	0.951	0.951
10							5	0.633	0.906

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# COMPOSTING OF AGRICULTURAL WASTES: A RURAL TECHNOLOGY

J.J. THAMBIRAJAH

*Institute of Advanced Studies, University of Malaya, Kuala Lumpur*

Recently, there has been a widespread interest in utilization of agricultural wastes. This is increasingly evident in countries where agriculture an important part of the economy. These areas face problems with the disposal of post-harvest material such as crop residues and agricultural by-products. These organic wastes which are essentially "resources out-of-place" are potential sources of pollution. Efficient utilization of the waste is often made difficult by the presence of non-cellulosic material which may not be amenable to biodegradation.

In this context composting, which has been practiced since time immemorial has been singled out

as a useful method, for the treatment of organic waste particularly in rural areas. The process involves the natural breakdown of the higher plant polymers by a succession of aerobic thermophilic microorganisms. Inherent in the waste, they ultimately convert it to a nutrient rich and stable humus suitable for crop production. With the increasing cost of chemical fertilizers, particularly in developing countries where there may be a ready supply of labour, the process of composting appears to have good potential. Organic fertilizer derived from agro-industrial waste are known to last longer in the soil; thus releasing nutrients slowly in a sustained manner.

**Appendix 2(a)**  
*In-situ* Physicochemical Measurements

**Site A**

Sampling Point (distance from baseline)	Temperature (°C)	DO (mg L <sup>-1</sup> )	pH	Salinity (ppt)
A1 (0 m)	30.5	7.2	7.32	29
A2 (20 m)	31.0	7.2	7.48	30
A3 (30 m)	32.0	7.0	7.84	32
Mean	31.20 ± 0.76	7.10 ± 0.12	7.50 ± 0.27	30.30 ± 1.53

**Site B**

Sampling Point (distance from baseline)	Temperature (°C)	DO (mg L <sup>-1</sup> )	pH	Salinity (ppt)
B1 (0 m)	32.5	9.5	7.98	31
B2 (20 m)	32.5	9.8	8.19	31
B3 (40 m)	31.5	10.0	8.20	32
B4 (60 m)	31.5	10.5	8.36	32
B5 (80 m)	31.5	10.2	8.37	32
Mean	31.90 ± 0.49	10.00 ± 0.34	8.20 ± 0.14	31.60 ± 0.49

**Site C**

Sampling Point (distance from baseline)	Temperature (°C)	DO (mg L <sup>-1</sup> )	pH	Salinity (ppt)
C1 (0 m)	25	8	7.7	31
C2 (20 m)	31	7.2	7.8	31
C3 (40 m)	32	6.9	7.9	30
C4 (60 m)	32	7.0	7.9	30
C5 (80 m)	31	7.3	7.6	29
Mean	30.20 ± 2.64	7.28 ± 0.39	7.78 ± 0.12	30.20 ± 0.77

**Appendix 2(b)**  
Nutrients (NH<sub>3</sub>-N & O-PO<sub>4</sub>)

**Ammoniacal Nitrogen (NH<sub>3</sub>-N)**

Sampling Point (distance from baseline)	Concentration ( $\mu\text{g L}^{-1}$ ) at site A	Concentration ( $\mu\text{g L}^{-1}$ ) at site B	Concentration ( $\mu\text{g L}^{-1}$ ) at site C
A1/ B1/ C1 (0 m)	10.6	1.44	1.59
A2/ B2/ C2 (20 m)	6.25	2.41	3.55
A3/ B3/ C3 (40 m)	5.84	2.79	5.40
A4/ B4/ C4 (60 m)	missing data	2.02	3.72
A5/ B5/ C5 (80 m)	missing data	0.63	6.23
Mean	7.56 $\pm$ 2.60	1.86 $\pm$ 0.85	4.10 $\pm$ 1.80

**Dissolved Orthophosphate (O-PO<sub>4</sub>)**

Sampling Point (distance from baseline)	Concentration ( $\mu\text{g L}^{-1}$ ) at site A	Concentration ( $\mu\text{g L}^{-1}$ ) at site B	Concentration ( $\mu\text{g L}^{-1}$ ) at site C
A1/ B1/ C1 (0 m)	1.15	0.19	0.04
A2/ B2/ C2 (20 m)	1.85	0.15	0.22
A3/ B3/ C3 (40 m)	1.78	0.10	1.15
A4/ B4/ C4 (60 m)	missing data	0.11	non-detectable
A5/ B5/ C5 (80 m)	missing data	0.08	non-detectable
Mean	1.59 $\pm$ 0.40	0.13 $\pm$ 0.04	0.28 $\pm$ 0.50

**Appendix 2(c)**  
Total Suspended Solids (TSS)

**Site A**

Sampling Point (distance from baseline)	Replicate	W <sub>1</sub> (g)	W <sub>2</sub> (g)	W <sub>2</sub> -W <sub>1</sub> (g)	TSS (mg L <sup>-1</sup> )	Mean TSS (mg L <sup>-1</sup> )
A1 (0 m)	I	0.0936	0.0833	0.0103	103.0	108.0 ± 5.0
	II	0.0923	0.0815	0.0108	108.0	
	III	0.0923	0.0810	0.0113	113.0	
A2 (20 m)	I	0.0935	0.1042	0.0107	107.0	108.3 ± 6.11
	II	0.0932	0.1035	0.0103	103.0	
	III	0.0925	0.1040	0.0115	115.0	
A3 (40 m)	I	0.0928	0.1045	0.0117	117.0	117.1 ± 3.10
	II	0.0917	0.1032	0.0115	115.0	
	III	0.0921	0.1046	0.0121	121.0	
Mean TSS at site A (mg L <sup>-1</sup> )						111.3 ± 5.49

**Site B**

Sampling Point (distance from baseline)	Replicate	W <sub>1</sub> (g)	W <sub>2</sub> (g)	W <sub>2</sub> -W <sub>1</sub> (g)	TSS (mg L <sup>-1</sup> )	Mean TSS (mg L <sup>-1</sup> )
B1 (0 m)	I	0.0924	0.1269	0.0345	34.5	34.5 ± 9.50
	II	0.0925	0.0969	0.0044	44.0	
	III	0.0941	0.0966	0.0025	25.0	
B2 (20 m)	I	0.0929	0.0965	0.0036	36.0	38.0 ± 5.29
	II	0.0929	0.0973	0.0044	44.0	
	III	0.0927	0.0961	0.0034	34.0	
B3 (40 m)	I	0.0929	0.0939	0.0010	10.0	11.3 ± 3.21
	II	0.0919	0.0934	0.0015	15.0	
	III	0.0928	0.0937	0.0009	9.0	
B4 (60 m)	I	0.0927	0.0932	0.0005	5.0	8.3 ± 4.16
	II	0.0931	0.0944	0.0013	13.0	
	III	0.0927	0.0934	0.0007	7.0	
B5 (80 m)	I	0.0930	0.0939	0.0009	9.0	9.0 ± 0.00
	II	0.0917	0.0926	0.0009	9.0	
	III	0.0932	0.0941	0.0009	9.0	
Mean TSS at site B (mg L <sup>-1</sup> )						20.2 ± 14.7

**Appendix 2(c) [Continued]**  
**Total Suspended Solids (TSS)**

**Site C**

Sampling Point (distance from baseline)	Replicate	W <sub>1</sub> (g)	W <sub>2</sub> (g)	W <sub>2</sub> -W <sub>1</sub> (g)	TSS (mg L <sup>-1</sup> )	Mean TSS (mg L <sup>-1</sup> )
C1 (0 m)	I	0.0957	0.0975	0.0018	18.0	16.0 ± 2.0
	II	0.0942	0.0958	0.0016	16.0	
	III	0.0947	0.0961	0.0014	14.0	
C2 (20 m)	I	0.0944	0.0954	0.0010	10.0	11.3 ± 3.21
	II	0.0952	0.0961	0.0009	9.0	
	III	0.0941	0.0956	0.0015	15.0	
C3 (40 m)	I	0.0942	0.0945	0.0013	13.0	10.7 ± 2.52
	II	0.0946	0.0954	0.0008	8.0	
	III	0.0947	0.0958	0.0011	11.0	
C4 (60 m)	I	0.0945	0.0947	0.0002	2.0	7.7 ± 5.13
	II	0.0961	0.0970	0.0009	9.0	
	III	0.0965	0.0977	0.0012	12.0	
C5 (80 m)	I	0.0960	0.0967	0.0007	7.0	6.7 ± 1.53
	II	0.0972	0.0977	0.0005	5.0	
	III	0.0969	0.0977	0.0008	8.0	
Mean TSS at site C (mg L <sup>-1</sup> )						10.5 ± 3.65

### Appendix 3

#### Mean Air Temperature

Month	Temperature					Relative Humidity
	Mean			Extreme		Mly
	Max	Min	Mly	Max	Min	
Jan	32.7	24.2	27.6	35.2	22.8	81.0
Feb	35.1	25.0	29.0	37.3	24.0	74.7
Mar	35.5	25.3	29.3	37.2	22.8	75.4
Apr	35.9	25.9	30.0	37.3	25.2	75.3
May	34.4	25.4	29.1	38.0	23.0	81.7
Jun	32.6	24.7	28.1	34.7	23.0	83.5
Jul	32.5	24.2	27.9	35.7	22.3	82.2
Aug	31.6	23.8	27.1	33.5	22.2	85.7
Sep	31.8	24.0	27.5	33.3	22.8	83.0
Oct	32.7	24.5	28.1	35.6	22.8	79.8
Nov	32.0	24.1	27.4	34.0	23.0	83.0

Source: Malaysian Meteorological Service



## Appendix 4

## Seaweed Composition Checklist for Cape Rachado

Division	Order	Family	Species	Site A	Site B	Site C
Cyanophyta	Oscillatoriales	Oscillatoriaceae	<i>Lyngbya majuscula</i> Harvey	x		
			<i>Lyngbya</i> sp. 1	x		x
			<i>Oscillatoria</i> sp. 1	x		x
			<i>Oscillatoria</i> sp. 2		x	x
			<i>Oscillatoria</i> sp. 3		x	x
			<i>Oscillatoria</i> sp. 4	x	x	x
Total			5	2	4	
Chlorophyta	Ulinales	Ulvaeae	<i>Enteromorpha clathrata</i> (Roth) Greville	x	x	x
			<i>E. intestinalis</i> (Linnaeus) Nees	x	x	x
			<i>Enteromorpha</i> sp. 1			
			<i>Sarvea anastomosans</i> (Harvey) Piccone & Grunow ex Piccone	x		x
			<i>Boodlea composita</i> (Harvey) Brand	x		x
			<i>Valoniaceae</i>			
			<i>Dichosphaeria cavernosa</i> (Forskkaal) Boerg ex Endlicher	x	x	x
			<i>Valonia aegogrophila</i> C. Agardh	x	x	x
			<i>Chaetomorpha</i> cf. <i>antennina</i> (Bory de Saint Vincent) Kutzing			
			<i>Chaetomorpha</i> sp. 1	x		x
			<i>Cladophora fascicularis</i> (Mert.) Kutzing	x	x	x
			<i>Cladophora</i> cf. <i>rupesstris</i> (Linnaeus) Kutzing			
			<i>Cladophora</i> cf. <i>stimpsonii</i> Harvey			
			<i>Cladophora</i> sp. 1	x		x
			<i>Cladophora</i> sp. 2			
<i>Cladophora</i> sp. 3						
<i>Cladophora</i> sp. 4						
<i>Cladophoropsis</i> sp.						
<i>Rhizoclonium</i> sp.						
Caulerpaceae	Caulerpaceae	<i>Caulerpa racemosa</i> (Forskkaal) J. Agardh	x	x	x	
		<i>C. peltata</i> Lamouroux				
		<i>C. verticillata</i> J. Agardh	x	x	x	
Udoteaceae	Udoteaceae	<i>Udotea javensis</i> (Montagne) A & E. S. Gepp	x	x	x	
		<i>Avrainvillea erecta</i> (Berkeley) A & E. S. Gepp	x		x	
Polyphysaceae	Polyphysaceae	<i>Acetabularia</i> sp. 1	x		x	
		<i>Acetabularia</i> sp. 2				
Bryopsidaceae	Bryopsidaceae	<i>Bryopsis pennata</i> Lamouroux	x		x	
Total			17	8	23	

Appendix 4 [Continued]  
Seaweed Composition Checklist for Cape Rachado

Division	Order	Family	Species	Site A	Site B	Site C		
Rhodophyta	Nemalionales	Achrochaetiaceae	<i>Achrochaetium</i> sp.		x	x		
		Gelidiaceae	<i>Gelidium acerosa</i> (Forsk.) Feldmann & Hamel <i>Gelidium cf. pinnosa</i> (Bornet) Feldmann & Hamel <i>Pterocladia caerulea</i> (Kütz.) Stentiles	x x x	x x x	x x x		
	Cryptonemiales	Corallinaceae	<i>Amphiroa fragilisissima</i> (Linnaeus) Lamouroux <i>Amphiroa cf. anceps</i> (Lamarck) Desainse	x x	x x	x x		
			<i>Jania rubens</i> (Linnaeus) Lamouroux	x	x	x		
	Gigartinales	Hypneaecae	<i>Hypnea pinnosa</i> J. Agardh <i>H. cf. esperi</i> Bory de Saint-Vincent <i>H. cf. charoides</i> Lamouroux	x x x	x x x	x x x		
			Champiaecae	<i>Champia parvula</i> C. Agardh	x	x	x	
	Ceramiales	Gracilariaceae	<i>Gracilaria salicornia</i> (C. Agardh) Dawson <i>Gelidiopsis</i> sp.	x x		x x		
			Ceramiaeae	<i>Ceratodicyon spongiosum</i> Zanardini <i>Ceramium tenuissimum</i> (Roth) J. E. Areschoug <i>C. gracillimum</i> (Kütz.) Zanardini <i>Centroceras</i> sp. <i>Anotrictium tenue</i> (C. Agardh) Nageli <i>Griffithsia</i> sp.	x x x x x	x x x x x	x x x x x	
		Rhodomelaceae	<i>Callithamnion</i> sp. <i>Acanthophora spicifera</i> (Vahl.) Bergesen <i>Chondria</i> sp. 1 <i>Chondria</i> sp. 2 <i>Laurencia parvipapillata</i> Tseng <i>Laurencia cf. corymbosa</i> J. Agardh <i>Laurencia</i> sp. 1 <i>Laurencia</i> sp. 2 <i>Levillaea junggerrmannioides</i> (Martens et Heeling) Harvey <i>Polysiphonia cf. nigrescens</i> (Hudson) Greville <i>Polysiphonia cf. violacea</i> (Greville) <i>Polysiphonia</i> sp. 1 <i>Polysiphonia</i> sp. 2 <i>Tolypocladia glomerulata</i> (C. Agardh) Schmitz <i>Herposiphonia</i> sp.		x x x x x x x x x x x x	x x x x x x x x x x x x		
			<i>Hildenbrandia</i> sp.	x	17	31		
			Total			23	17	31

Appendix 4 [Continued]

Seaweed Composition Checklist for Cape Rachado

Division	Order	Family	Species	Site A	Site B	Site C
Phaeophyta	Ectocarpales	Ectocarpaceae	<i>Ectocarpus</i> sp.		X	X
		Dictyotales	Dictyotaceae	<i>Padina commersonii</i> Domantay	X	X
	<i>P. tetrastromatica</i> Hauck			X	X	X
	<i>Lobophora variegata</i> (Lamouroux) Womersley			X	X	X
	<i>Dictyota dentata</i> Lamouroux			X		
	<i>D. dichotoma</i> (Hudson) Lamouroux			X		
	<i>Dictyopteris delicatula</i> Lamouroux			X		
	<i>Sargassum baccharia</i> (Mert.) C. Agardh			X	X	X
	<i>S. binderi</i> Sonder			X	X	X
	<i>S. siliquosum</i> Agardh			X		
Fucales	Sargassaceae	<i>Sargassum</i> sp. 1	X	X	X	
		<i>Turbinaria conoides</i> (J. Agardh) Kützting	X	X	X	
Sphaecelariales	Sphaecelariaceae		X	X	X	
		<i>Sphaecelaria</i> sp.	X	X	X	
Total				9	8	11
Total number of species at each site				54	35	69
Total number of species at Cape Rachado				81		

**Appendix 5**  
Species Richness, Shannon Index ( $H'$ ) and Evenness Index ( $J$ ) for Cape Rachado, 1998.

Quadrat	Site A												Mean Species Richness for each quadrat (S.D.)	Mean $H'$ for each quadrat (S.D.)	Mean $J$ for each quadrat (S.D.)
	Transect A			Transect B			Transect C			Species Richness	$H'$	$J$			
	Species Richness	$H'$	$J$	Species Richness	$H'$	$J$	Species Richness	$H'$	$J$						
0	6	0.53	0.68	2	0.13	0.44	11	0.80	0.77	6 (5)	0.49 (0.34)	0.63 (0.17)			
1	9	0.70	0.73	8	0.69	0.76	3	0.28	0.58	7 (3)	0.56 (0.24)	0.69 (0.10)			
2	3	0.25	0.52	3	0.31	0.66	1	0.00	0.00	2 (1)	0.19 (0.16)	0.39 (0.35)			
3	6	0.51	0.65	1	0.00	0.00	13	0.89	0.80	7 (6)	0.47 (0.45)	0.48 (0.42)			
4	6	0.61	0.79	14	0.99	0.86	20	1.15	0.88*	13 (7)	0.91 (0.27)	0.84 (0.05)			
5	6	0.51	0.65	15	0.91	0.78	18	1.11	0.89	13 (6)	0.85 (0.31)	0.77 (0.12)			
6	14	0.95	0.83	23	1.24	0.91	16	1.05	0.87	18 (5)	1.08 (0.15)	0.87 (0.04)			
7	15	0.96	0.82	11	0.80	0.77	17	1.08	0.88	14 (3)	0.95 (0.14)	0.82 (0.05)			
Transect mean (S.D.)	8 (4)	0.63 (0.24)	0.71 (0.11)	10 (8)	0.63 (0.44)	0.65 (0.3)	12 (7)	0.79 (0.43)	0.71 (0.3)						
Total Species Richness	36			42			30								
Overall mean at site A (S.D.)										10 (6)	0.69 (0.37)	0.69 (0.24)			

### Appendix 5 [Continued]

Species Richness, Shannon Index ( $H'$ ) and Evenness Index ( $J$ ) for Cape Rachado, 1998.

Quadrat	Site B											Mean $H'$ for each quadrat (S.D.)	Mean $J$ for each quadrat (S.D.)	
	Transect A			Transect B			Transect C			Mean Species Richness for each quadrat (S.D.)				
	Species Richness	$H'$	$J$	Species Richness	$H'$	$J$	Species Richness	$H'$	$J$					
0	9	0.90	0.94	1	0.0	0.0	12	0.91	0.84	0.84	7 (6)	0.6 (0.52)	0.59 (0.52)	
1	8	0.72	0.80	5	0.63	0.91	12	0.96	0.89	0.89	8 (4)	0.77 (0.17)	0.87 (0.06)	
2	9	0.82	0.86	15	1.04	0.88	13	0.95	0.85	0.85	12 (3)	0.94 (0.11)	0.87 (0.01)	
3	9	0.82	0.86	8	0.78	0.87	14	1.01	0.88	0.88	10 (3)	0.87 (0.12)	0.87 (0.01)	
4	8	0.69	0.76	7	0.77	0.91	1	0.00	0.00	0.00	5 (4)	0.49 (0.42)	0.56 (0.49)	
5	6	0.60	0.78	11	0.90	0.87	18	1.22	0.97*	0.97*	12 (6)	0.91 (0.31)	0.87 (0.10)	
6	13	1.00	0.90	12	1.03	0.95	18	1.13	0.90	0.90	14 (3)	1.05 (0.06)	0.92 (0.03)	
7				11	0.88	0.84					11	0.88	0.84	
Transect mean (S.D)	9 (2)	0.79 (0.13)	0.84 (0.07)	9 (5)	0.75 (0.36)	0.78 (0.34)	13 (6)	0.88 (0.40)	0.76 (0.34)	0.76 (0.34)				
Total Species Richness	22			22			29							
Overall mean at site B (S.D.)											10 (5)	0.81 (0.30)	0.79 (0.26)	

### Appendix 5 [Continued]

Species Richness, Shannon Index ( $H'$ ) and Evenness Index ( $J$ ) for Cape Rachado, 1998.

Quadrat	Site C												Mean $H'$ for each quadrat (S.D.)	Mean $J$ for each quadrat (S.D.)
	Transect A			Transect B			Transect C			Mean Species Richness for each quadrat (S.D.)				
	Species Richness	$H'$	$J$	Species Richness	$H'$	$J$	Species Richness	$H'$	$J$					
0	8	0.67	0.74	13	0.98	0.88	17	1.03	0.84	13 (5)	0.89 (0.20)	0.82 (0.07)		
1	8	0.80	0.88	12	0.87	0.81	12	0.85	0.78	11 (2)	0.84 (0.04)	0.82 (0.05)		
2	13	0.98	0.88	12	0.91	0.84	20	1.20	0.92	15 (4)	1.03 (0.15)	0.88 (0.04)		
3	15	0.99	0.84	14	0.95	0.83	18	1.06	0.85	16 (2)	1.00 (0.06)	0.84 (0.01)		
4	13	1.05	0.94	14	1.11	0.97	20	1.25	0.96	16 (4)	1.14 (0.10)	0.96 (0.01)		
5	13	0.99	0.89	21	1.17	0.88	20	1.21	0.93*	18 (4)	1.12 (0.12)	0.90 (0.03)		
6	12	0.91	0.84	16	1.10	0.91	17	1.12	0.91	15 (3)	1.04 (0.12)	0.89 (0.04)		
7	18	1.16	0.93	17	1.10	0.90	14	0.93	0.81	16 (2)	1.06 (0.12)	0.88 (0.06)		
8	18	1.08	0.86	15	1.10	0.93	22	1.19	0.89	18 (4)	1.12 (0.06)	0.89 (0.04)		
9	16	1.11	0.92	23	1.29	0.95	20	1.12	0.86	20 (4)	1.17 (0.10)	0.91 (0.05)		
10	17	1.13	0.92	16	1.13	0.94	19	1.16	0.90	17 (2)	1.14 (0.02)	0.92 (0.02)		
11	17	1.15	0.93	22	1.25	0.93	0	0.00	0.00	13 (12)	0.80 (0.69)	0.62 90.54)		
12	16	1.09	0.90	15	1.07	0.91	24	1.28	0.93	18 (5)	1.15 (0.12)	0.91 (0.01)		
13	18	1.11	0.89	20	1.26	0.96	25	1.35	0.97	21 (4)	1.24 (0.12)	0.94 (0.05)		
14	18	1.19	0.95	32	1.49	0.99				25 (10)	1.34 (0.21)	0.97 (0.03)		
15	29	1.39	0.95							29	1.39	0.95		
Transect mean (S.D.)	16 (5)	1.05 (0.17)	0.89 (0.05)	17 (5)	1.12 (0.16)	0.91 (0.05)	18 (6)	1.05 (0.33)	0.82 (0.24)					
Total Species Richness	49			53			50							
Overall mean at site C (S.D.)										17 (5)	1.10 (0.16)	0.90 (0.05)		

## Appendix 6

Categorisation of the seaweeds of Cape Rachado according to Littler's functional morphologic groups.

Division	Family	Species	Group	
Chlorophyta	Ulvaceae	<i>Enteromorpha clathrata</i> (Roth) Greville	SG	
		<i>E. intestinalis</i> (Linnaeus) Nees	SG	
		<i>Enteromorpha</i> sp. 1	SG	
	Boodleaecae	<i>Siruvea anostomosans</i> (Harvey) Piccone & Grunow ex Piccone	SG	
		<i>Boodlea composita</i> (Harvey) Brand	SG	
	Valoniaceae	<i>Dictyosphaeria cavernosa</i> (Forsk.) Boerg ex Endlicher	SG	
		<i>Valonia aegagrophila</i> C. Agardh	SG	
	Cladophoraceae	<i>Chaetomorpha linum</i> (O. F. Muller) Kützting	F	
		<i>Chaetomorpha</i> cf. <i>antennina</i> (Bory de Saint Vincent) Kützting	F	
		<i>Chaetomorpha</i> sp. 1	F	
		<i>Cladophora fascicularis</i> (Mert.) Kützting	F	
		<i>Cladophora</i> cf. <i>rupestris</i> (Linnaeus) Kützting	F	
		<i>Cladophora</i> cf. <i>stimpsonii</i> Harvey	F	
		<i>Cladophora</i> sp. 1	F	
		<i>Cladophora</i> sp. 2	F	
		<i>Cladophora</i> sp. 3	F	
		<i>Cladophora</i> sp. 4	F	
		<i>Cladophoropsis</i> sp.	F	
		<i>Rhizoclonium</i> sp.	F	
		Caulerpaceae	<i>Caulerpa racemosa</i> (Forsk.) J. Agardh	CB
			<i>C. peltata</i> Lamouroux	CB
<i>C. verticillata</i> J. Agardh	CB			
Udoteaceae	<i>Udotea javensis</i> (Montagne) A & E. S. Gepp	CB		
	<i>Avrainvillea erecta</i> (Berkeley) A & E. S. Gepp	TL		
Polyphysaceae	<i>Acetabularia</i> sp. 1	F		
	<i>Acetabularia</i> sp. 2	F		
Bryopsidaceae	<i>Bryopsis pennata</i> Lamouroux	SG		
Achrochaetiaceae	<i>Achrochaetium</i> sp.	F		
Rhodophyta	Gelidiaceae	<i>Gelidiella acerosa</i> (Forsk.) Feldmann & Hamel	CB	
		<i>Gelidiella</i> cf. <i>pannosa</i> (Bornet) Feldmann & Hamel	CB	
		<i>Pterocladia caerulescens</i> (Kützting) Santelices	CB	
	Corallinaceae	<i>Amphiroa fragilissima</i> (Linnaeus) Lamouroux	JC	
		<i>Amphiroa</i> cf. <i>anceps</i> (Lamarck) Decaisne	JC	
		<i>Jania rubens</i> (Linnaeus) Lamouroux	JC	
	Hypneaceae	<i>Hypnea pannosa</i> J. Agardh	CB	
		<i>H.</i> cf. <i>esperii</i> Bory de Saint Vincent	CB	
		<i>H.</i> cf. <i>charoides</i> Lamouroux	CB	
	Champiaceae	<i>Champia parvula</i> C. Agardh	CB	
	Gracilariaceae	<i>Gracilaria salicornia</i> (C. Agardh) Dawson	CB	
		<i>Gelidiopsis</i> sp. <i>Ceratodictyon spongiosum</i> Zanardini	CB CB	
	Ceramiaceae	<i>Ceramium tenuissimum</i> (Roth) J. E. Areschoug	F	
		<i>C. gracillimum</i> (Kützting) Zanardini	F	
		<i>Centroceras</i> sp.	F	
		<i>Anotrichium tenue</i> (C. Agardh) Nageli	F	
		<i>Griffithsia</i> sp. <i>Callithamnion</i> sp.	F F	
	Rhodomelaceae	<i>Acanthophora spicifera</i> (Vahl.) Bergesen	CB	
		<i>Chondria</i> sp. 1	CB	
<i>Chondria</i> sp. 2		CB		
<i>Laurencia parvipapillata</i> Tseng		CB		
<i>Laurencia</i> cf. <i>corymbosa</i> J. Agardh		CB		
<i>Laurencia</i> sp. 1		CB		
<i>Laurencia</i> sp. 2		CB		
<i>Leveillea junggermannioides</i> (Martens et Herling) Harvey		CB		
<i>Polysiphonia</i> cf. <i>nigrescens</i> (Hudson) Greville		F		
<i>Polysiphonia</i> cf. <i>violaceae</i> (Greville)		F		
<i>Polysiphonia</i> sp. 1		F		
<i>Polysiphonia</i> sp. 2		F		
<i>Tolyptocladia glomerulata</i> (C. Agardh) Schmitz		F		
<i>Herposiphonia</i> sp.		F		
Hildenbrandiaceae	<i>Hildenbrandia</i> sp.	C		

### Appendix 6 [Continued]

Categorisation of the seaweeds of Cape Rachado according to Littler's functional morphologic groups.

Division	Family	Species	Site A
Phaeophyta	Ectocarpaceae	<i>Ectocarpus</i> sp.	F
	Dictyotaceae	<i>Padina commersonii</i> Domantay	TL
		<i>P. tetrastromatica</i> Hauck	TL
		<i>Lobophora variegata</i> (Lamouroux) Womersley	TL
		<i>Dictyota dentata</i> Lamouroux	TL
		<i>D. dichotoma</i> (Hudson) Lamouroux	TL
		<i>Dictyopteris delicatula</i> Lamouroux	TL
	Sargassaceae	<i>Sargassum baccularia</i> (Mert.) C. Agardh	TL
		<i>S. binderi</i> Sonder	TL
		<i>S. siliquosum</i> Agardh	TL
		<i>Sargassum</i> sp. 1	TL
		<i>Turbinaria conoides</i> (J. Agardh) Kützing	TL
	Sphacelariaceae	<i>Sphacelaria</i> sp.	F

Abbreviations:

- SG Sheet group
- F Filamentous group
- CB Coarsely branched group
- TL Thick-leathery group
- JC Jointed calcareous group
- C Crustose group



### Appendix 7

Seaweed Dry Weight Biomass for Cape Rachado in 1998.

Site A				
Quadrat	Transect A (g m <sup>-2</sup> )	Transect B (g m <sup>-2</sup> )	Transect C (g m <sup>-2</sup> )	Mean for each quadrat (S.D.)
0	3.24	3.15	2.73	3.04 (0.3)
1	11.21	2.21	0.22	4.55 (5.9)
2	35.73	5.82	0.16	13.9 (19.1)
3	60.52	44.66	1.99	35.72 (30.3)
4	2.05	5.67	100.22	35.98 (55.7)
5	7.50	12.21	96.14	38.62 (49.9)
6	22.96	61.79	103.28	62.68 (40.2)
7	63.48	12.09	102.19	59.25 (45.2)
Transect mean (S.D.)	25.84 (24.93)	18.45 (22.25)	50.87 (53.06)	
Mean seaweed dry weight biomass at site A, Cape Rachado				31.72 (37.38)

Site B				
Quadrat	Transect A (g m <sup>-2</sup> )	Transect B (g m <sup>-2</sup> )	Transect C (g m <sup>-2</sup> )	Mean for each quadrat (S.D.)
0	19.05	3.58	19.45	14.03 (9.05)
1	69.42	31.21	22.64	41.09 (24.91)
2	20.70	15.29	35.38	23.79 (10.39)
3	38.19	55.97	46.88	47.01 (8.89)
4	74.80	86.21	4.74	55.25 (44.11)
5	22.19	47.97	93.00	54.39 (35.84)
6	137.14	116.05	95.11	116.1 (21.01)
7		189.71		189.71
Transect mean (S.D.)	54.5 (43.15)	68.25 (61.28)	45.31 (35.79)	
Mean seaweed dry weight biomass at site B, Cape Rachado				56.58 (47.38)

### Appendix 7 [Continued]

Seaweed Dry Weight Biomass for Cape Rachado in 1998.

Site C				
Quadrat	Transect A (g m <sup>-2</sup> )	Transect B (g m <sup>-2</sup> )	Transect C (g m <sup>-2</sup> )	Mean for each quadrat (S.D.)
0	0.58	25.75	32.39	19.58 (16.78)
1	2.42	27.95	35.81	22.06 (17.45)
2	40.05	12.69	29.99	27.58 (13.84)
3	39.44	30.26	59.78	43.16 (15.11)
4	8.70	5.10	33.69	15.83 (15.57)
5	12.66	93.79	86.85	64.43 (44.97)
6	25.82	34.31	182.78	80.97 (88.27)
7	41.47	31.13	41.05	37.88 (5.85)
8	24.67	30.56	41.77	32.33 (8.69)
9	93.76	106.54	46.06	82.12 (31.88)
10	62.31	36.73	190.88	96.64 (82.61)
11	79.22	36.43	0.00	38.55 (39.65)
12	32.08	96.05	110.21	79.45 (41.63)
13	56.64	62.59	137.61	85.61 (45.13)
14	192.06	84.64		138.35 (75.96)
15	157.44			157.44
Transect mean (S.D.)	54.33 (54.17)	47.63 (32.43)	73.49 (59.71)	
Mean seaweed dry weight biomass at site C, Cape Rachado				58.06 (50.06)



# Appendix 8 (a) [Continued]

Frequency, Dominance and the Importance Value Index (IVI) for seaweed species at site A, Cape Rachado, in 1998.

## Site A

Seaweed species	Number of quadrat in which species was recorded	Frequency value (f) [b] = [a]/24 quadrats	Frequency (%) [c] = [b]*100	Relative frequency (f <sub>r</sub> ) [d] = [b]/total [b]*100	Total N value	Dominance (%) [f] = [c]/total [c]*100	Dominance value (d) [g] = [c]/area sampled [area sampled = 2.16 m <sup>2</sup> ]	Relative dominance (R/d) [h] = [g]/total [g]*100	IVI [i] = [(d) + (h)]/2
<i>Acetabularia</i> sp. 2	1	0.04	4.17	0.43	1	0.24	0.46	0.23	0.33
<i>Byropsis pennata</i>	2	0.08	8.33	0.85	5	1.18	2.31	1.13	0.99
<b>Rhodophyta</b>									
<i>Achrochaetium</i> sp.		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Gelidium acerosa</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Gelidium</i> cf. <i>pinnosa</i>	1	0.04	4.17	0.43	1	0.24	0.46	0.23	0.33
<i>Pterocladia carolinensis</i>	3	0.13	12.50	1.28	12	2.83	5.56	2.71	1.99
<i>Amphiroa fragilissima</i>	5	0.21	20.83	2.13	7	1.65	3.24	1.58	1.85
<i>Amphiroa</i> cf. <i>anceps</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Jania rubens</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Hypnea pinnosa</i>	7	0.29	29.17	2.98	16	3.77	7.41	3.61	3.30
<i>H.</i> cf. <i>esperi</i>	6	0.25	25.00	2.55	7	1.65	3.24	1.58	2.07
<i>H.</i> cf. <i>charoides</i>	1	0.04	4.17	0.43	9	2.12	4.17	2.03	1.23
<i>Champia parvula</i>	4	0.17	16.67	1.70	4	0.94	1.85	0.90	1.30
<i>Gracilaria salicornia</i>	5	0.21	20.83	2.13	49	11.56	22.69	11.06	6.59
<i>Gelidium</i> sp.	1	0.04	4.17	0.43	1	0.24	0.46	0.23	0.33
<i>Ceratodictyon spongiosum</i>	4	0.17	16.67	1.70	25	5.90	11.57	5.64	3.67
<i>Ceramium tenuissimum</i>	11	0.46	45.83	4.68	11	2.59	5.09	2.48	3.58
<i>C. gracillimum</i>	9	0.38	37.50	3.83	9	2.12	4.17	2.03	2.93
<i>Centroceras</i> sp.		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Anotrichium tenue</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Griffithsia</i> sp.	3	0.13	12.50	1.28	3	0.71	1.39	0.68	0.98
<i>Callithamnion</i> sp.	5	0.21	20.83	2.13	5	1.18	2.31	1.13	1.63
<i>Acanthophora spicifera</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Chondria</i> sp. 1		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Chondria</i> sp. 2		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Laurencia parvipapillata</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Laurencia</i> cf. <i>corymbosa</i>	1	0.04	4.17	0.43	1	0.24	0.46	0.23	0.33
<i>Laurencia</i> sp. 1	2	0.08	8.33	0.85	2	0.47	0.93	0.45	0.65
<i>Laurencia</i> sp. 2		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Levellia juggermannioides</i>	1	0.04	4.17	0.43	1	0.24	0.46	0.23	0.33
<i>Polysiphonia</i> cf. <i>nigrescens</i>	6	0.25	25.00	2.55	6	1.42	2.78	1.35	1.95
<i>Polysiphonia</i> cf. <i>violacea</i>	3	0.13	12.50	1.28	3	0.71	1.39	0.68	0.98

# Appendix 8 (a) [Continued]

Frequency, Dominance and the Importance Value Index (IVI) for seaweed species at site A, Cape Rachado, in 1998.

## Site A

Seaweed species	Number of quadrat in which species was recorded	Frequency value (f) [b] = [a]/24 quadrats	Frequency (%) [c] = [b]*100	Relative frequency (f/r) [d] = [b]/total [b]*100	Total N value [e]	Dominance (%) [f] = [e]/total [e]*100	Dominance value (d) [g] = [e]/area sampled [area sampled = 2.16 m <sup>2</sup> ]	Relative dominance (R/d) [h] = [g]/total [g]*100	IVI [i] = [(d) + (h)]/2
<i>Polysiphonia</i> sp. 1		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Polysiphonia</i> sp. 2	4	0.17	16.67	1.70	4	0.94	1.85	0.90	1.30
<i>Tolyposiocladia glomerulata</i>	4	0.17	16.67	1.70	4	0.94	1.85	0.90	1.30
<i>Herposiphonia</i> sp.	6	0.25	25.00	2.55	6	1.42	2.78	1.35	1.95
<i>Hildenbrandia</i> sp.	13	0.54	54.17	5.53	20	4.72	9.26	4.51	5.02
<b>Phaeophyta</b>									
<i>Ectocarpus</i> sp.	10	0.42	41.67	4.26	28	6.60	12.96	6.32	5.29
<i>Padina commersonii</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>P. tetrastrumata</i>	7	0.29	29.17	2.98	7	1.65	3.24	1.58	2.28
<i>Lobophora variegata</i>	2	0.08	8.33	0.85	2	0.47	0.93	0.45	0.65
<i>Dicyota dentata</i>	2	0.08	8.33	0.85	2	0.47	0.93	0.45	0.65
<i>D. dichotoma</i>	5	0.21	20.83	2.13	5	1.18	2.31	1.13	1.63
<i>Diclypteria delicatula</i>	8	0.33	33.33	3.40	48	11.32	22.22	10.84	7.12
<i>Sargassum boccularia</i>	4	0.17	16.67	1.70	12	2.83	5.56	2.71	2.21
<i>S. binderi</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>S. siliquosum</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Sargassum</i> sp. 1	6	0.25	25.00	2.55	6	1.42	2.78	1.35	1.95
<i>Turbinaaria conoides</i>	3	0.13	12.50	1.28	3	0.71	1.39	0.68	0.98
<i>Sphaerocaria</i> sp.									
<b>Total</b>	24 quadrats	9.79	979.17	100.00	443	104.48	205.09	100.00	100.00

# Appendix 8 (b)

Frequency, Dominance and the Importance Value Index (IVI) for seaweed species at site B, Cape Rachado, in 1998.

## Site B

Seaweed species	Number of quadrat in which species was recorded	Frequency value (f) [b] = [a]/22 quadrats	Frequency (%) [c] = [b]*100	Relative frequency (Rf) [d] = [b]/total [b]*100	Total N value [e]	Dominance (%) [f] = [c]/total [c]*100	Dominance value (d) [g] = [c]/area sampled [area sampled = 1.98 m <sup>2</sup> ]	Relative dominance (Rd) [h] = [g]/total [g]*100	IVI [i] = [(d) + (h)]/2
<b>Cyanophyta</b>									
<i>Lyngbya majuscula</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Lyngbya</i> sp. 1		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Oscillatoria</i> sp. 1		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Oscillatoria</i> sp. 2		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Oscillatoria</i> sp. 3		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Oscillatoria</i> sp. 4		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<b>Chlorophyta</b>									
<i>Enteromorpha clathrata</i>	4	0.18	18.18	1.81	4	1.01	* 2.02	1.01	1.41
<i>E. intestinalis</i>	4	0.18	18.18	1.81	4	1.01	2.02	1.01	1.41
<i>Enteromorpha</i> sp. 1		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Struvea anastomosans</i>	1	0.05	4.55	0.45	1	0.25	0.51	0.25	0.35
<i>Boodlea composita</i>	1	0.05	4.55	0.45	1	0.25	0.51	0.25	0.35
<i>Dictyosphaeria cavernosa</i>	1	0.05	4.55	0.45	1	0.25	0.51	0.25	0.35
<i>Valonia aegagophylla</i>	3	0.14	13.64	1.36	7	1.76	3.54	1.76	1.56
<i>Chaetomorpha linum</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Chaetomorpha</i> cf. <i>antennina</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Chaetomorpha</i> sp. 1		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Cladophora fascicularis</i>	12	0.55	54.55	5.43	12	3.02	6.06	3.02	4.22
<i>Cladophora</i> cf. <i>rupesstris</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Cladophora</i> cf. <i>stimpsonii</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Cladophora</i> sp. 1		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Cladophora</i> sp. 2		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Cladophora</i> sp. 3		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Cladophora</i> sp. 4		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Cladophoropsis</i> sp.		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Rhizoclonium</i> sp.	16	0.73	72.73	7.24	75	18.84	37.88	18.84	13.04
<i>Caularia racemosa</i>	1	0.05	4.55	0.45	1	0.25	0.51	0.25	0.35
<i>C. peltata</i> Lamouroux		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>C. verticillata</i>	3	0.14	13.64	1.36	3	0.75	1.52	0.75	1.06
<i>Udotea javensis</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Avicillaea erecta</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Actinobularia</i> sp. 1		0.00	0.00	0.00		0.00	0.00	0.00	0.00

# Appendix 8 (b) [Continued]

Frequency, Dominance and the Importance Value Index (IVI) for seaweed species at site B, Cape Rachado, in 1998.

## Site B

Seaweed species	Number of quadrat in which species was recorded	Frequency value (f) [b] = [a]/22 quadrats	Frequency (%) [c] = [b]*100	Relative frequency (Rf) [d] = [b]/total [b]*100	Total N value [e]	Dominance (%) [f] = [e]/total [e]*100	Dominance value (d) [g] = [e]-area sampled [area sampled = 1.98 m <sup>2</sup> ]	Relative dominance (Rd) [h] = [g]/total [g]*100	IVI [i] = [(d) + (h)]/2
<i>Acetabularia</i> sp. 2		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Bryopsis pinnata</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<b>Rhodophyta</b>									
<i>Achlorochaetium</i> sp.	1	0.05	4.55	0.45	1	0.25	0.51	0.25	0.35
<i>Gelidium acerosa</i>	9	0.41	40.91	4.07	10	2.51	5.05	2.51	3.29
<i>Gelidium</i> cf. <i>pinnosa</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Pterocladia carulescens</i>	11	0.50	50.00	4.98	14	3.52	7.07	3.52	4.25
<i>Amphiroa fragillissima</i>	6	0.27	27.27	2.71	6	1.51	3.03	1.51	2.11
<i>Amphiroa</i> cf. <i>anceps</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Jania rubens</i>	3	0.14	13.64	1.36	3	0.75	1.52	0.75	1.06
<i>Hypnea pinnosa</i>	17	0.77	77.27	7.69	17	4.27	8.59	4.27	5.98
<i>H.</i> cf. <i>esperi</i>	1	0.05	4.55	0.45	1	0.25	0.51	0.25	0.35
<i>H.</i> cf. <i>charoides</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Champia parvula</i>	16	0.73	72.73	7.24	16	4.02	8.08	4.02	5.63
<i>Gracilaria salicornia</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Gelidopsis</i> sp.		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Ceratodictyon spongiosum</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Gelidopsis</i> sp.		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Ceramium tenuissimum</i>	1	0.05	4.55	0.45	1	0.25	0.51	0.25	0.35
<i>C. gracillimum</i>	10	0.45	45.45	4.52	10	2.51	5.05	2.51	3.52
<i>Centroceras</i> sp.		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Antractium tenue</i>	1	0.05	4.55	0.45	1	0.25	0.51	0.25	0.35
<i>Griffithsia</i> sp.		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Callithamnion</i> sp.		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Acanthophora spicifera</i>	2	0.09	9.09	0.90	2	0.50	1.01	0.50	0.70
<i>Chondria</i> sp. 1		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Chondria</i> sp. 2		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Laurencia purvipapillata</i>	3	0.14	13.64	1.36	3	0.75	1.52	0.75	1.06
<i>Laurencia</i> cf. <i>corymbosa</i>	7	0.32	31.82	3.17	7	1.76	3.54	1.76	2.46
<i>Laurencia</i> sp. 1		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Laurencia</i> sp. 2		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Levillaea jaggernamnioides</i>	6	0.27	27.27	2.71	6	1.51	3.03	1.51	2.11
<i>Polysiphonia</i> cf. <i>nigrescens</i>	6	0.27	27.27	2.71	6	1.51	3.03	1.51	2.11
<i>Polysiphonia</i> cf. <i>violaceae</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00

# Appendix 8 (b) [Continued]

Frequency, Dominance and the Importance Value Index (IVI) for seaweed species at site B, Cape Rachado, in 1998.

## Site B

Seaweed species	Number of quadrat in which species was recorded	Frequency value (U) [b] = [a]/22 quadrats	Frequency (%) [c] = [b]*100	Relative frequency (Rf) [d] = [b]/total [b]*100	Total N value [e]	Dominance (%) [f] = [c]/total [c]*100	Dominance value (d) [g] = [e]-area sampled [area sampled = 1.98 m <sup>2</sup> ]	Relative dominance (Rd) [h] = [g]/total [g]*100	IVI [i] = ([d] + [h])/2
<i>Polysiphonia</i> sp. 1		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Polysiphonia</i> sp. 2	7	0.32	31.82	3.17	7	1.76	3.54	1.76	2.46
<i>Tolyocladia glomerulata</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Herposiphonia</i> sp.	6	0.27	27.27	2.71	6	1.51	3.03	1.51	2.11
<i>Hildenbrandia</i> sp.		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<b>Ethecophyta</b>									
<i>Ectocarpus</i> sp.	2	0.09	9.09	0.90	2	0.50	1.01	0.50	0.70
<i>Fadma commersonii</i>	16	0.73	72.73	7.24	16	4.02	8.08	4.02	5.63
<i>P. tetrastrumatica</i>	2	0.09	9.09	0.90	2	0.50	1.01	0.50	0.70
<i>Lobophora variegata</i>	3	0.14	13.64	1.36	3	0.75	1.52	0.75	1.06
<i>Dictyota dentata</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>D. dichotoma</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Dictyoptera delicatula</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Sargassum boecularia</i>	18	0.82	81.82	8.14	76	19.10	38.38	19.10	13.62
<i>S. binderi</i>	9	0.41	40.91	4.07	35	8.79	17.68	8.79	6.43
<i>S. stilloquum</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Sargassum</i> sp. 1		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Turbinaria conoides</i>	13	0.59	59.09	5.88	39	9.80	19.70	9.80	7.84
<i>Sphaerularia</i> sp.		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<b>Total</b>	<b>22 quadrats</b>	<b>10.05</b>	<b>1004.55</b>	<b>100.00</b>	<b>398</b>	<b>100.00</b>	<b>201.01</b>	<b>100.00</b>	<b>100.00</b>



## Appendix 8 (c)

Frequency, Dominance and the Importance Value Index (IVI) for seaweed species at site C, Cape Rachado, in 1998.

Seaweed species	Number of quadrat in which species was recorded	Frequency value (U) [b] = [a]/45 quadrats	Frequency (%) [c] = [b]*100	Relative frequency (βf) [d] = [b]/total [b]*100	Total N value [e]	Dominance (%) [f] = [c]/total [c]*100	Dominance value (d) [g] = [e]/area sampled [area sampled = 4.05 m <sup>2</sup> ]	Relative dominance (βd) [h] = [g]/total [g]*100	IVI [i] = ([d] + [h])/2
<b>Cyanophyta</b>									
<i>Lyngbya majuscula</i>	3	0.07	6.67	0.40	3	0.28	0.74	0.28	0.34
<i>Lyngbya</i> sp. 1		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Oscillatoria</i> sp. 1	3	0.07	6.67	0.40	3	0.28	0.74	0.28	0.34
<i>Oscillatoria</i> sp. 2	2	0.04	4.44	0.26	2	0.19	0.49	0.19	0.23
<i>Oscillatoria</i> sp. 3	1	0.02	2.22	0.13	1	0.09	0.25	0.09	0.11
<i>Oscillatoria</i> sp. 4	2	0.04	4.44	0.26	2	0.19	0.49	0.19	0.23
<b>Chlorophyta</b>									
<i>Enteromorpha clathrata</i>	26	0.58	57.78	3.43	26	2.42	6.52	2.42	2.93
<i>E. intestinalis</i>	12	0.27	26.67	1.58	12	1.12	2.96	1.12	1.35
<i>Enteromorpha</i> sp. 1	1	0.02	2.22	0.13	1	0.09	0.25	0.09	0.11
<i>Stroeva amestomosans</i>	1	0.02	2.22	0.13	1	0.09	0.25	0.09	0.11
<i>Boodlea composita</i>	6	0.13	13.33	0.79	6	0.56	1.48	0.56	0.68
<i>Dictyosphaeria cavernosa</i>	4	0.09	8.89	0.53	3	0.28	0.74	0.28	0.40
<i>Folomia aegrographilla</i>	2	0.04	4.44	0.26	2	0.19	0.49	0.19	0.23
<i>Chaetomorpha linum</i>	23	0.51	51.11	3.03	23	2.14	5.68	2.14	2.59
<i>Chaetomorpha</i> cf. <i>antennina</i>	1	0.02	2.22	0.13	1	0.09	0.25	0.09	0.11
<i>Chaetomorpha</i> sp. 1	2	0.04	4.44	0.26	2	0.19	0.49	0.19	0.23
<i>Cladophora fascicularis</i>	36	0.80	80.00	4.75	36	3.36	8.89	3.36	4.05
<i>Cladophora</i> cf. <i>repustris</i>	1	0.02	2.22	0.13	1	0.09	0.25	0.09	0.11
<i>Cladophora</i> cf. <i>stimpsonii</i>	2	0.04	4.44	0.26	2	0.19	0.49	0.19	0.23
<i>Cladophora</i> sp. 1		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Cladophora</i> sp. 2	1	0.02	2.22	0.13	1	0.09	0.25	0.09	0.11
<i>Cladophora</i> sp. 3	1	0.02	2.22	0.13	1	0.09	0.25	0.09	0.11
<i>Cladophora</i> sp. 4	2	0.04	4.44	0.26	2	0.19	0.49	0.19	0.23
<i>Cladophoropsis</i> sp.		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Rhizoclonium</i> sp.	1	0.02	2.22	0.13	1	0.09	0.25	0.09	0.11
<i>Caulerpa racemosa</i>	2	0.04	4.44	0.26	9	0.84	2.22	0.84	0.55
<i>C. peltata</i> Lamouroux		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>C. verticillata</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Udotea javensis</i>	18	0.40	40.00	2.37	18	1.68	4.44	1.68	2.03
<i>Avrainvillia erecta</i>		0.00	0.00	0.00		0.00	0.00	0.00	0.00
<i>Acetabularia</i> sp. 1	1	0.02	2.22	0.13	1	0.09	0.25	0.09	0.11

# Appendix 8 (c) [Continued]

Frequency, Dominance and the Importance Value Index (IVI) for seaweed species at site C, Cape Rachado, in 1998.

## Site C

Seaweed species	Number of quadrat in which species was recorded	Frequency value (f) [a]	Frequency (%) [b] = [a]/45 quadrats	Frequency (%) [c] = [b]*100	Relative frequency (f <sub>R</sub> ) [d] = [b]/total [b]*100	Total N value [e]	Dominance (%) [f] = [e]/total [e]*100	Dominance value (d) [g] = [e]/area sampled [area sampled = 4.05 m <sup>2</sup> ]	Relative dominance (d <sub>R</sub> ) [h] = [g]/total [g]*100	IVI [i] = [(d) + (h)]/2
<i>Acetabularia</i> sp. 2	4	0.09	8.89	13.33	0.53	4	0.37	0.99	0.37	0.45
<i>Bryopsis pennata</i>	6	0.13	13.33		0.79	6	0.56	1.48	0.56	0.68
<b>Rhodophyta</b>										
<i>Achlorochaetium</i> sp.	1	0.02	2.22		0.13	1	0.09	0.25	0.09	0.11
<i>Gelidium acerosa</i>	28	0.62	62.22		3.69	31	2.89	7.65	2.89	3.29
<i>Gelidium</i> cf. <i>pinnosa</i>		0.00	0.00		0.00		0.00	0.00	0.00	0.00
<i>Pterocladia caerulescens</i>	34	0.76	75.56		4.49	34	3.17	8.40	3.17	3.83
<i>Amphiroa fragillissima</i>	30	0.67	66.67		3.96	30	2.80	7.41	2.80	3.38
<i>Amphiroa</i> cf. <i>ancaps</i>	3	0.07	6.67		0.40	3	0.28	0.74	0.28	0.34
<i>Jania rubra</i>	5	0.11	11.11		0.66	6	0.56	1.48	0.56	0.61
<i>Hypnea pinnosa</i>	38	0.84	84.44		5.01	38	3.54	9.38	3.54	4.28
<i>H.</i> cf. <i>esperi</i>	2	0.04	4.44		0.26	2	0.19	0.49	0.19	0.23
<i>H.</i> cf. <i>charoides</i>	1	0.02	2.22		0.13	1	0.09	0.25	0.09	0.11
<i>Champia parvula</i>	38	0.84	84.44		5.01	38	3.54	9.38	3.54	4.28
<i>Gracilaria salicornia</i>		0.00	0.00		0.00		0.00	0.00	0.00	0.00
<i>Gelidopsis</i> sp.	3	0.07	6.67		0.40	3	0.28	0.74	0.28	0.34
<i>Ceratodicyon spongiosum</i>		0.00	0.00		0.00		0.00	0.00	0.00	0.00
<i>Ceramium tenuissimum</i>	13	0.29	28.89		1.72	13	1.21	3.21	1.21	1.46
<i>C. gracillimum</i>	27	0.60	60.00		3.56	27	2.52	6.67	2.52	3.04
<i>Centroceras</i> sp.		0.00	0.00		0.00		0.00	0.00	0.00	0.00
<i>Anastrichium tenue</i>	21	0.47	46.67		2.77	21	1.96	5.19	1.96	2.36
<i>Griffithsia</i> sp.	3	0.07	6.67		0.40	3	0.28	0.74	0.28	0.34
<i>Callithamnion</i> sp.	1	0.02	2.22		0.13	1	0.09	0.25	0.09	0.11
<i>Acanthophora specifera</i>	20	0.44	44.44		2.64	21	1.96	5.19	1.96	2.30
<i>Chondria</i> sp. 1	1	0.02	2.22		0.13	1	0.09	0.25	0.09	0.11
<i>Chondria</i> sp. 2	1	0.02	2.22		0.13	1	0.09	0.25	0.09	0.11
<i>Laurencia parvipapillata</i>	4	0.09	8.89		0.53	5	0.47	1.23	0.47	0.50
<i>Laurencia</i> cf. <i>corymbosa</i>	16	0.36	35.56		2.11	16	1.49	3.95	1.49	1.80
<i>Laurencia</i> sp. 1	4	0.09	8.89		0.53	4	0.37	0.99	0.37	0.45
<i>Laurencia</i> sp. 2	1	0.02	2.22		0.13	1	0.09	0.25	0.09	0.11
<i>Levellia jurggermannioides</i>	7	0.16	15.56		0.92	7	0.65	1.73	0.65	0.79
<i>Polysiphonia</i> cf. <i>nigrescens</i>	19	0.42	42.22		2.51	19	1.77	4.69	1.77	2.14
<i>Polysiphonia</i> cf. <i>violaceae</i>	1	0.02	2.22		0.13	1	0.09	0.25	0.09	0.11

## Appendix 8 (c) [Continued]

Frequency, Dominance and the Importance Value Index (IVI) for seaweed species at site C, Cape Rachado, in 1998.

### Site C

Seaweed species	Number of quadrats in which species was recorded	Frequency value (f) [b] = [a]/45 quadrats	Frequency (%) [c] = [b]*100	Relative frequency (Rf) [d] = [b]/total [b]*100	Total N value [e]	Dominance (%) [f] = [e]/total [e]*100	Dominance value (d) [g] = [e]/area sampled [area sampled = 4.05 m]	Relative dominance (Rd) [h] = [g]/total [g]*100	IVI [i] = [(d) + (h)]/2
<i>Polysiphonia</i> sp. 1	3	0.07	6.67	0.40	3	0.28	0.74	0.28	0.34
<i>Polysiphonia</i> sp. 2	29	0.64	64.44	3.83	29	2.70	7.16	2.70	3.26
<i>Tolypocladia glomerulata</i>	1	0.02	2.22	0.13	1	0.09	0.25	0.09	0.11
<i>Herposiphonia</i> sp.	14	0.31	31.11	1.85	14	1.30	3.46	1.30	1.58
<i>Hildenbrandia</i> sp.	4	0.09	8.89	0.53	5	0.47	1.23	0.47	0.50
<i>Phaeosiphonia</i>	2	0.04	4.44	0.26	2	0.19	0.49	0.19	0.23
<i>Ectocarpus</i> sp.	36	0.80	80.00	4.75	50	4.66	12.35	4.66	4.70
<i>Padina commersoni</i>	23	0.51	51.11	3.03	41	3.82	10.12	3.82	3.43
<i>P. tetrastrumatica</i>	27	0.60	60.00	3.56	27	2.52	6.67	2.52	3.04
<i>Lobophora variegata</i>	17	0.38	37.78	2.24	18	1.68	4.44	1.68	1.96
<i>Dicyotia dentata</i>	0	0.00	0.00	0.00	0	0.00	0.00	0.00	0.00
<i>D. dichotoma</i>	0	0.00	0.00	0.00	0	0.00	0.00	0.00	0.00
<i>Dicyotiers delicatula</i>	42	0.93	93.33	5.54	225	20.97	55.56	20.97	13.26
<i>Sargassum bacularia</i>	13	0.29	28.89	1.72	37	3.45	9.14	3.45	2.58
<i>S. binderi</i>	1	0.02	2.22	0.13	3	0.28	0.74	0.28	0.21
<i>S. siliquosum</i>	1	0.02	2.22	0.13	1	0.09	0.25	0.09	0.11
<i>Sargassum</i> sp. 1	38	0.84	84.44	5.01	98	9.13	24.20	9.13	7.07
<i>Turbinaria conoides</i>	21	0.47	46.67	2.77	21	1.96	5.19	1.96	2.36
<i>Sphaecularia</i> sp.	21	0.47	46.67	2.77	21	1.96	5.19	1.96	2.36
<b>Total</b>	<b>45 quadrats</b>	<b>16.84</b>	<b>1684.44</b>	<b>100.00</b>	<b>1073</b>	<b>100.00</b>	<b>264.94</b>	<b>100.00</b>	<b>100.00</b>

### Appendix 9

Seaweed Composition Checklist for Cape Rachado, as recorded in the surveys of 1987/88 and 1998

Division	Species	1987/88			1998		
		Site A	Site B	Site C	Site A	Site B	Site C
Cyanophyta	<i>Lyngbya majuscula</i> Harvey	x			x		
	<i>Lyngbya</i> sp. 1				x		
	<i>Oscillatoria</i> sp. 1				x		x
	<i>Oscillatoria</i> sp. 2						x
	<i>Oscillatoria</i> sp. 3				x	x	x
	<i>Oscillatoria</i> sp. 4				x	x	x
	<i>Nostoc</i>	x					
<b>Total</b>		<b>2</b>	<b>0</b>	<b>0</b>	<b>5</b>	<b>2</b>	<b>4</b>
Chlorophyta	<i>Enteromorpha clathrata</i> (Roth) Greville	x	x		x	x	x
	<i>E. intestinalis</i> (Linnaeus) Nees	x	x		x	x	x
	<i>Enteromorpha</i> sp. 1						x
	<i>Struvea anostomosans</i> (Harvey) Piccone & Grunow ex Piccone	x	x	x	x		x
	<i>Boodlea composita</i> (Harvey) Brand				x		x
	<i>Dictyosphaera cavernosa</i> (Forsskaal) Boerg ex Endlicher	x	x	x		x	x
	<i>Valonia aegagrophila</i> C. Agardh	x	x	x	x		x
	<i>Valonia</i> sp. 1	x					
	<i>Chaetomorpha linum</i> (O. F. Muller) Kützing	x	x		x	x	x
	<i>Chaetomorpha</i> cf. <i>antennina</i> (Bory de Saint Vincent) Kützing						x
	<i>Chaetomorpha</i> sp. 1				x		x
	<i>Cladophora fascicularis</i> (Mert.) Kützing	x	x	x	x	x	x
	<i>Cladophora</i> cf. <i>rupestris</i> (Linnaeus) Kützing						x
	<i>Cladophora</i> cf. <i>stimpsonii</i> Harvey				x		x
	<i>Cladophora</i> sp. 1(1987/88)	x		x			
	<i>Cladophora</i> sp. 2 (1987/88)		x				
	<i>Cladophora</i> sp. 1(1998)						x
	<i>Cladophora</i> sp. 2 (1998)						x
	<i>Cladophora</i> sp. 3 (1998)						x
	<i>Cladophora</i> sp. 4 (1998)						x
	<i>Cladophoropsis</i> sp.				x		
	<i>Rhizoclonium</i> sp.				x		
	<i>Caulerpa racemosa</i> (Forsskaal) J. Agardh		x		x	x	x
	<i>C. peltata</i> Lamouroux					x	
	<i>C. verticillata</i> J. Agardh	x			x		
	<i>C. serrulata</i> (Forsskaal) J. Agardh	x					
	<i>C. taxifolia</i> (Vahl) C. Agardh	x		x			
	<i>Udotea javensis</i> (Montagne) A & E. S. Gepp	x	x	x	x	x	x
	<i>Udotea flabellum</i> (Ellis & Sonder) Howe	x					
	<i>Avrainvillea erecta</i> (Berkeley) A & E. S. Gepp				x		
	<i>Acetabularia</i> sp. 1(1987/88)			x			
	<i>Acetabularia</i> sp. 1(1998)				x		x
	<i>Acetabularia</i> sp. 2 (1998)						x
<i>Bryopsis pennata</i> Lamouroux				x		x	
<b>Total</b>		<b>15</b>	<b>10</b>	<b>8</b>	<b>17</b>	<b>8</b>	<b>23</b>

**Appendix 9 [Continued]**

Seaweed Composition Checklist for Cape Rachado, A20 as recorded in the surveys of 1987/88 and 1998

Division	Species	1987/88			1998		
		Site A	Site B	Site C	Site A	Site B	Site C
Rhodophyta	<i>Achrochaetium</i> sp.					X	X
	<i>Gelidiella acerosa</i> (Forsskaal) Feldmann & Hamel	X	X	X		X	X
	<i>Gelidiella cf. pannosa</i> (Bornet) Feldmann & Hamel				X		
	<i>Gelidiella bornetti</i> (Weber van Bosse) Feldmann & Hamel		X				
	<i>Gelidiella</i> sp. 1			X			
	<i>Gelidium</i> sp. 1		X	X			
	<i>Gelidium</i> sp. 2	X	X	X			
	<i>Gelidium</i> sp. 3	X	X				
	<i>Pterocladia caerulescens</i> (Kutzing) Santelices	X	X	X	X	X	X
	<i>Amphiroa fragilissima</i> (Linnaeus) Lamouroux	X	X	X	X	X	X
	<i>Amphiroa cf. anceps</i> (Lamarck) Decaisne						X
	<i>Amphiroa foliaceae</i> (Linnaeus) Lamouroux		X				
	<i>Jania rubens</i> (Linnaeus) Lamouroux		X			X	X
	<i>Hypnea pannosa</i> J. Agardh				X	X	X
	<i>H. cf. esperi</i> Bory de Saint Vincent	X	X	X	X		X
	<i>H. cf. charoides</i> Lamouroux				X		X
	<i>Hypnea</i> sp. 1 (1987/88)	X	X	X			
	<i>Hypnea</i> sp. 2 (1987/88)	X	X				
	<i>Champia parvula</i> C. Agardh	X	X	X	X	X	X
	<i>Gracilaria salicornia</i> (C. Agardh) Dawson	X	X	X	X		
	<i>Gracilaria</i> sp. 1 (1987/88)	X	X	X			
	<i>Gelidiopsis</i> sp.				X		X
	<i>Ceratodictyon spongiosum</i> Zanardini	X	X	X	X		
	<i>Ceramium tenuissimum</i> (Roth) J. E. Areschoug				X	X	X
	<i>C. gracillimum</i> (Kutzing) Zanardini				X	X	X
	<i>Ceramium</i> sp. (1987/88)	X	X	X			
	<i>Centroceras</i> sp.	X	X		X		
	<i>Anotrichium tenue</i> (C. Agardh) Nageli					X	
	<i>Griffithsia</i> sp.				X		X
	<i>Callithamnion</i> sp.				X		X
	<i>Acanthophora spicifera</i> (Vahl.) Bergesen	X	X	X		X	X
	<i>Acanthophora orientalis</i> J. Agardh	X		X			
	<i>Chondria</i> sp. 1						X
	<i>Chondria</i> sp. 2						X
	<i>Laurencia parvipapillata</i> Tseng					X	X
	<i>Laurencia cf. corymbosa</i> J. Agardh				X	X	X
	<i>Laurencia cf. concreta</i> Cribb		X	X			
	<i>Laurencia</i> sp. 1 (1998)				X		X
	<i>Laurencia</i> sp. 2 (1998)						X
	<i>Laurencia</i> sp. 2 (1987/88)	X	X	X			
	<i>Laurencia</i> sp. 3 (1987/88)		X				
	<i>Leveillea junggermannioides</i> (Martens et Herling) Harvey	X	X		X	X	X
	<i>Polysiphonia cf. nigrescens</i> (Hudson) Greville				X	X	
	<i>Polysiphonia cf. violaceae</i> (Greville)						X
	<i>Polysiphonia</i> sp. 1 (1987/88)	X	X	X			
	<i>Polysiphonia</i> sp. 1 (1998)						X
	<i>Polysiphonia</i> sp. 2 (1998)				X	X	X
	<i>Tolyptocladia glomerulata</i> (C. Agardh) Schmitz	X			X		X
	<i>Galaxaura cf. filamentosa</i> Chou			X			
	<i>Herposiphonia</i> sp.	X	X		X	X	X
<i>Heterosiphonia</i> sp.	X						
<i>Erythrotrichia</i> sp.		X					
<i>Hildenbrandia</i> sp.				X		X	
<b>Total</b>		<b>22</b>	<b>26</b>	<b>19</b>	<b>23</b>	<b>17</b>	<b>31</b>

**Appendix 9 [Continued]**

Seaweed Composition Checklist for Cape Rachado, as recorded in the surveys of 1987/88 and 1998

Division	Species	1987/88			1998		
		Site A	Site B	Site C	Site A	Site B	Site C
Phaeophyta	<i>Ectocarpus</i> sp.		x	x		x	x
	<i>Padina commersonii</i> Domantay	x	x	x	x	x	x
	<i>P. tetrastromatica</i> Hauck	x	x	x		x	x
	<i>Lobophora variegata</i> (Lamouroux) Womersley	x	x	x	x	x	x
	<i>Dictyota dentata</i> Lamouroux				x		
	<i>D. dichotoma</i> (Hudson) Lamouroux		x	x	x		x
	<i>D. bartayresi</i> Lamouroux	x	x	x			
	<i>D. ceylanica</i> Kutzing	x	x	x			
	<i>D. cervicornis</i> Kutzing	x	x	x			
	<i>Dictyopteris delicatula</i> Lamouroux				x		
	<i>Sargassum baccularia</i> (Mert.) C. Agardh	x	x	x	x	x	x
	<i>S. oligocystum</i> Montagne	x	x	x		x	x
	<i>S. siliquosum</i> Agardh		x	x			x
	<i>Sargassum</i> sp. 1(1987/88)			x			
	<i>Sargassum</i> sp. 2 (1987/88)	x	x	x			
	<i>Sargassum</i> sp. 1(1998)						x
	<i>Turbinaria conoides</i> (J. Agardh) Kutzing	x	x	x	x	x	x
	<i>Turbinaria ornata</i> (Turner) J. Agardh	x	x	x			
	<i>Turbinaria</i> sp. 1	x		x			
	<i>Hormophysa triquetra</i> C. Agardh			x			
<i>Hydroclathrus clathratus</i> (Bory de Saint Vincent) Howe			x				
<i>Sphacelaria</i> sp.				x	x	x	
<b>Total</b>		12	14	18	9	8	11
<b>Total number of species at each site</b>		51	50	45	54	35	69
<b>Total number of species at Cape Rachado</b>				69			81

## **Appendix 10**

**Results of the Newman-Keuls post-hoc test for two-way ANOVA, performed on the water quality and biotic parameters of 1987/88 and 1998**

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_PH (2wayanov.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		(1) .9316571	(2) .9700856	(3) .9663006	(4) .9668885	(5) .9653460
A	98	(1)		.000010*	.000020*	.000026*	.000017*
A	875	(2)	.000010*		.425656	.404471	.315719
A	878	(3)	.000020*	.425656		.813411	.701561
A	883	(4)	.000026*	.404471	.813411		.809557
A	8711	(5)	.000017*	.315719	.701561	.809557	
B	98	(6)	.000022*	.311595	.915242	.901801	.961245
B	875	(7)	.000032*	.990921	.287017	.203327	.231658
B	878	(8)	.000017*	.785244	.175635	.228412	.085548
B	883	(9)	.000008*	.383474	.911239	.916736	.978122
B	8711	(10)	.000012*	.863047	.438804	.464338	.300238
C	98	(11)	.000011*	.000032*	.000017*	.000020*	.000008*
C	875	(12)	.000023*	.294978	.009496*	.017266*	.002694*
C	878	(13)	.000026*	.027223*	.000139*	.000342*	.000036*
C	883	(14)	.000015*	.932930	.417963	.472663	.264866
C	8711	(15)	.000020*	.319480	.014738*	.024854*	.004606*

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_PH (2wayanov.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		(6) .9646790	(7) .9700572	(8) .9724181	(9) .9652777	(10) .9705153
A	98	(1)	.000022*	.000032*	.000017*	.000008*	.000012*
A	875	(2)	.311595	.990921	.785244	.383474	.863047
A	878	(3)	.915242	.287017	.175635	.911239	.438804
A	883	(4)	.901801	.203327	.228412	.916736	.464338
A	8711	(5)	.961245	.231658	.085548	.978122	.300238
B	98	(6)		.257118	.059194	.810091	.270115
B	875	(7)	.257118		.878108	.307152	.981537
B	878	(8)	.059194	.878108		.096866	.725186
B	883	(9)	.810091	.307152	.096866		.350860
B	8711	(10)	.270115	.981537	.725186	.350860	
C	98	(11)	.000009*	.000026*	.000015*	.000022*	.000010*
C	875	(12)	.001281*	.354585	.484069	.002912*	.310461
C	878	(13)	.000027*	.033682*	.119378	.000040*	.034479*
C	883	(14)	.219529	.983214	.560901	.301673	.855288
C	8711	(15)	.002367*	.397571	.337348	.005098*	.311444

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_PH (2wayanov.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		(11) .9434561	(12) .9752795	(13) .9779330	(14) .9709697	(15) .9748079
A	98	(1)	.000011*	.000023*	.000026*	.000015*	.000020*
A	875	(2)	.000032*	.294978	.027223*	.932930	.319480
A	878	(3)	.000017*	.009496*	.000139*	.417963	.014738*
A	883	(4)	.000020*	.017266*	.000342*	.472663	.024854*
A	8711	(5)	.000008*	.002694*	.000036*	.264866	.004606*
B	98	(6)	.000009*	.001281*	.000027*	.219529	.002367*
B	875	(7)	.000026*	.354585	.033682*	.983214	.397571
B	878	(8)	.000015*	.484069	.119378	.560901	.337348
B	883	(9)	.000022*	.002912*	.000040*	.301673	.005098*
B	8711	(10)	.000010*	.310461	.034479*	.855288	.311444
C	98	(11)		.000020*	.000023*	.000012*	.000017*
C	875	(12)	.000020*		.286731	.307882	.849862
C	878	(13)	.000023*	.286731		.041405*	.421045
C	883	(14)	.000012*	.307882	.041405*		.271788
C	8711	(15)	.000017*	.849862	.421045	.271788	



STAT. GENERAL MANOVA			Newman-Keuls test; LOG_SALT (2wayanov.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{1} 1.495666	{2} 1.503771	{3} 1.494444	{4} 1.521542	{5} 1.529303
A	98	{1}		.039163*	.713146	.000010*	.000015*
A	875	{2}	.039163*		.025831*	.000027*	.000010*
A	878	{3}	.713146	.025831*		.000012*	.000017*
A	883	{4}	.000010*	.000027*	.000012*		.051168
A	8711	{5}	.000015*	.000010*	.000017*	.051168	
B	98	{6}	.000028*	.037859*	.000032*	.031565*	.000028*
B	875	{7}	.022472*	.678286	.011203*	.000031*	.000032*
B	878	{8}	.419554	.000640*	.527430	.000020*	.000026*
B	883	{9}	.000483*	.236744	.000154*	.001832*	.000026*
B	8711	{10}	.000032*	.000022*	.000010*	.967220	.081776
C	98	{11}	.874943	.027941*	.900743	.000015*	.000020*
C	875	{12}	.019564*	.917415	.018886*	.000033*	.000012*
C	878	{13}	.795403	.010273*	.846457	.000017*	.000023*
C	883	{14}	.000026*	.046513*	.000027*	.030359*	.000025*
C	8711	{15}	.000012*	.000032*	.000015*	.063208	.633468

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_SALT (2wayanov.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{6} 1.513168	{7} 1.505150	{8} 1.489938	{9} 1.509159	{10} 1.521406
A	98	{1}	.000028*	.022472*	.419554	.000483*	.000032*
A	875	{2}	.037859*	.678286	.000640*	.236744	.000022*
A	878	{3}	.000032*	.011203*	.527430	.000154*	.000010*
A	883	{4}	.031565*	.000031*	.000020*	.001832*	.967220
A	8711	{5}	.000028*	.000032*	.000026*	.000026*	.081776
B	98	{6}		.074760	.000015*	.449506	.013217*
B	875	{7}	.074760		.000152*	.227774	.000026*
B	878	{8}	.000015*	.000152*		.000010*	.000017*
B	883	{9}	.449506	.227774	.000010*		.001327*
B	8711	{10}	.013217*	.000026*	.000017*	.001327*	
C	98	{11}	.000010*	.010649*	.434926	.000128*	.000012*
C	875	{12}	.039638*	.862312	.000714*	.310772	.000027*
C	878	{13}	.000012*	.003096*	.420537	.000048*	.000015*
C	883	{14}	.817410	.074382	.000012*	.329452	.018526*
C	8711	{15}	.000077*	.000026*	.000023*	.000020*	.138988

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_SALT (2wayanov.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{11} 1.494029	{12} 1.503426	{13} 1.492615	{14} 1.512401	{15} 1.527718
A	98	{1}	.874943	.019564*	.795403	.000026*	.000012*
A	875	{2}	.027941*	.917415	.010273*	.046513*	.000032*
A	878	{3}	.900743	.018886*	.846457	.000027*	.000015*
A	883	{4}	.000015*	.000033*	.000017*	.030359*	.063208
A	8711	{5}	.000020*	.000012*	.000023*	.000025*	.633468
B	98	{6}	.000010*	.039638*	.000012*	.817410	.000077*
B	875	{7}	.010649*	.862312	.003096*	.074382	.000026*
B	878	{8}	.434926	.000714*	.420537	.000012*	.000023*
B	883	{9}	.000128*	.310772	.000048*	.329452	.000020*
B	8711	{10}	.000012*	.000027*	.000015*	.018526*	.138988
C	98	{11}		.024288*	.670613	.000033*	.000017*
C	875	{12}	.024288*		.010086*	.053970	.000010*
C	878	{13}	.670613	.010086*		.000010*	.000020*
C	883	{14}	.000033*	.053970	.000010*		.000055*
C	8711	{15}	.000017*	.000010*	.000020*	.000055*	

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_TSS (2wayanov.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		(1) 2.044294	(2) 1.971501	(3) 2.600069	(4) 1.866103	(5) 2.008487
A	98	{1}		.996945	.029846*	.979116	.984395
A	875	{2}	.996945		.063063	.872384	.983359
A	878	{3}	.029846*	.063063		.026886*	.053576
A	883	{4}	.979116	.872384	.026886*		.961307
A	8711	{5}	.984395	.983359	.053576	.961307	
B	98	{6}	.017555*	.022515*	.000138*	.049669*	.020569*
B	875	{7}	.506487	.369499	.001143*	.261860	.469756
B	878	{8}	.978318	.706742	.030208*	.904994	.945275
B	883	{9}	.037587*	.037790*	.000129*	.060859	.039989*
B	8711	{10}	.925996	.994462	.042480*	.974581	.939868
C	98	{11}	.001215*	.001799*	.000151*	.005048*	.001535*
C	875	{12}	.191009	.159001	.000282*	.176725	.187006
C	878	{13}	.317169	.753310	.111680	.588068	.642349
C	883	{14}	.041274*	.046545*	.000136*	.085781	.045803*
C	8711	{15}	.996125	.903725	.065042	.925211	.957826

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_TSS (2wayanov.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		(6) 1.239936	(7) 1.626253	(8) 1.891488	(9) 1.322424	(10) 2.024535
A	98	{1}	.017555*	.506487	.978318	.037587*	.925996
A	875	{2}	.022515*	.369499	.706742	.037790*	.994462
A	878	{3}	.000138*	.001143*	.030208*	.000129*	.042480*
A	883	{4}	.049669*	.261860	.904994	.060859	.974581
A	8711	{5}	.020569*	.469756	.945275	.039989*	.939868
B	98	{6}		.369400	.048180*	.919719	.019656*
B	875	{7}	.369400		.427099	.329685	.499841
B	878	{8}	.048180*	.427099		.069013	.969695
B	883	{9}	.919719	.329685	.069013		.040181*
B	8711	{10}	.019656*	.499841	.969695	.040181*	
C	98	{11}	.355788	.080809	.004520*	.553339	.001395*
C	875	{12}	.658918	.503094	.228730	.449264	.194933
C	878	{13}	.000965*	.091890	.597721	.002365*	.516201
C	883	{14}	.716866	.467182	.089026	.979944	.044931*
C	8711	{15}	.019828*	.410611	.871616	.036356*	.990927

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_TSS (2wayanov.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		(11) 1.042910	(12) 1.483652	(13) 2.257867	(14) 1.317057	(15) 1.997227
A	98	{1}	.001215*	.191009	.317169	.041274*	.996125
A	875	{2}	.001799*	.159001	.753310	.046545*	.903725
A	878	{3}	.000151*	.000282*	.111680	.000136*	.065042
A	883	{4}	.005048*	.176725	.588068	.085781	.925211
A	8711	{5}	.001535*	.187006	.642349	.045803*	.957826
B	98	{6}	.355788	.658918	.000965*	.716866	.019828*
B	875	{7}	.080809	.503094	.091890	.467182	.410611
B	878	{8}	.004520*	.228730	.597721	.089026	.871616
B	883	{9}	.553339	.449264	.002365*	.979944	.036356*
B	8711	{10}	.001395*	.194933	.516201	.044931*	.990927
C	98	{11}		.242186	.000166*	.403466	.001514*
C	875	{12}	.242186		.019080*	.712070	.165313
C	878	{13}	.000166*	.019080*		.002564*	.732433
C	883	{14}	.403466	.712070	.002564*		.042979*
C	8711	{15}	.001514*	.165313	.732433	.042979*	

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_SP (hjbanova.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{1}	{2}	{3}	{4}	{5}
			.9479923	.6346822	.4135936	.6385682	.6033112
A	98	{1}		.001278*	.000017*	.001282*	.000243*
A	875	{2}	.001278*		.009136*	.958758	.676298
A	878	{3}	.000017*	.009136*		.014622*	.011581*
A	8711	{4}	.001282*	.958758	.014622*		.885735
A	8803	{5}	.000243*	.676298	.011581*	.885735	
B	98	{6}	.600577	.000151*	.000020*	.000157*	.000036*
B	875	{7}	.043462*	.519209	.000536*	.298349	.431879
B	878	{8}	.281075	.237334	.000039*	.199543	.120434
B	8711	{9}	.270032	.248412	.000037*	.193613	.138017
B	8803	{10}	.691121	.000053*	.000023*	.000054*	.000024*
C	98	{11}	.001789*	.000020*	.000026*	.000017*	.000023*
C	875	{12}	.310999	.179662	.000012*	.158202	.080200
C	878	{13}	.038849*	.667463	.000657*	.524089	.527758
C	8711	{14}	.467579	.055668	.000012*	.050887	.018678*
C	8803	{15}	.738042	.003996*	.000015*	.003859*	.000902*

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_SP (hjbanova.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{6}	{7}	{8}	{9}	{10}
			.9873315	.7167085	.8000242	.7877082	1.009542
A	98	{1}	.600577	.043462*	.281075	.270032	.691121
A	875	{2}	.000151*	.519209	.237334	.248412	.000053*
A	878	{3}	.000020*	.000536*	.000039*	.000037*	.000023*
A	8711	{4}	.000157*	.298349	.199543	.193613	.000054*
A	8803	{5}	.000036*	.431879	.120434	.138017	.000024*
B	98	{6}		.009598*	.126076	.109461	.767545
B	875	{7}	.009598*		.684065	.611789	.003863*
B	878	{8}	.126076	.684065		.869810	.077913
B	8711	{9}	.109461	.611789	.869810		.062673
B	8803	{10}	.767545	.003863*	.077913	.062673	
C	98	{11}	.005850*	.000015*	.000033*	.000010*	.005358*
C	875	{12}	.162295	.657205	.806358	.911902	.111735
C	878	{13}	.008954*	.965004	.535886	.367562	.003725*
C	8711	{14}	.323880	.400629	.707090	.773674	.268284
C	8803	{15}	.666797	.087688	.358919	.374194	.656153

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_SP (hjbanova.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{11}	{12}	{13}	{14}	{15}
			1.218847	.8184438	.7200054	.8596377	.9228621
A	98	{1}	.001789*	.310999	.038849*	.467579	.738042
A	875	{2}	.000020*	.179662	.667463	.055668	.003996*
A	878	{3}	.000026*	.000012*	.000657*	.000012*	.000015*
A	8711	{4}	.000017*	.158202	.524089	.050887	.003859*
A	8803	{5}	.000023*	.080200	.527758	.018678*	.000902*
B	98	{6}	.005850*	.162295	.008954*	.323880	.666797
B	875	{7}	.000015*	.657205	.965004	.400629	.087688
B	878	{8}	.000033*	.806358	.535886	.707090	.358919
B	8711	{9}	.000010*	.911902	.367562	.773674	.374194

STAT. GENERAL MANOVA			Newman-Keuls test; LOG SP (hjbanova.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{11} 1.218847	{12} .8184438	{13} .7200054	{14} .8596377	{15} .9228621
B	8803	{10}	.005358*	.111735	.003725*	.268284	.656153
C	98	{11}		.000027*	.000012*	.000044*	.000788*
C	875	{12}	.000027*		.556414	.583518	.346269
C	878	{13}	.000012*	.556414		.340077	.075218
C	8711	{14}	.000044*	.583518	.340077		.400096
C	8803	{15}	.000788*	.346269	.075218	.400096	

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_BIO (hjbanova.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{1}	{2}	{3}	{4}	{5}
			1.140461	.8026197	.7819315	.8628028	.9222190
A	98	{1}		.228548	.234010	.318499	.370844
A	875	{2}	.228548		.898640	.710954	.741795
A	878	{3}	.234010	.898640		.872276	.823500
A	8711	{4}	.318499	.710954	.872276		.714475
A	8803	{5}	.370844	.741795	.823500	.714475	
B	98	{6}	.068618	.000059*	.000044*	.000242*	.000994*
B	875	{7}	.888397	.211325	.234327	.258962	.228774
B	878	{8}	.028531*	.000026*	.000026*	.000053*	.000204*
B	8711	{9}	.506524	.012258*	.009791*	.032007*	.069001
B	8803	{10}	.003276*	.000023*	.000026*	.000021*	.000025*
C	98	{11}	.070189	.000076*	.000055*	.000340*	.001232*
C	875	{12}	.253702	.016103*	.014261*	.035323*	.062202
C	878	{13}	.050936	.000063*	.000047*	.000273*	.000956*
C	8711	{14}	.392824	.012649*	.010544*	.030957*	.062013
C	8803	{15}	.043150*	.000032*	.000029*	.000099*	.000439*

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_BIO (hjbanova.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{6}	{7}	{8}	{9}	{10}
			1.600946	1.117669	1.673607	1.365956	1.780151
A	98	{1}	.068618	.888397	.028531*	.506524	.003276*
A	875	{2}	.000059*	.211325	.000026*	.012258*	.000023*
A	878	{3}	.000044*	.234327	.000026*	.009791*	.000026*
A	8711	{4}	.000242*	.258962	.000053*	.032007*	.000021*
A	8803	{5}	.000994*	.228774	.000204*	.069001	.000025*
B	98	{6}	.058632	.058632	.895547	.469926	.687378
B	875	{7}	.895547	.021848*	.021848*	.543566	.002245*
B	878	{8}	.895547	.021848*	.021848*	.405492	.511782
B	8711	{9}	.469926	.543566	.405492		.141587
B	8803	{10}	.687378	.002245*	.511782	.141587	
C	98	{11}	.912790	.063039	.944688	.374343	.743836
C	875	{12}	.535643	.405478	.387970	.966907	.115902
C	878	{13}	.992807	.048379*	.980480	.182685	.828277
C	8711	{14}	.541924	.471484	.428195	.932757	.143421
C	8803	{15}	.806428	.034738*	.839648	.438818	.666545

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_BIO (hjbanova.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{11}	{12}	{13}	{14}	{15}
			1.583159	1.325823	1.582358	1.352252	1.640743
A	98	{1}	.070189	.253702	.050936	.392824	.043150*
A	875	{2}	.000076*	.016103*	.000063*	.012649*	.000032*
A	878	{3}	.000055*	.014261*	.000047*	.010544*	.000029*
A	8711	{4}	.000340*	.035323*	.000273*	.030957*	.000099*
A	8803	{5}	.001232*	.062202	.000956*	.062013	.000439*
B	98	{6}	.912790	.535643	.992807	.541924	.806428
B	875	{7}	.063039	.405478	.048379*	.471484	.034738*
B	878	{8}	.944688	.387970	.980480	.428195	.839648
B	8711	{9}	.374343	.966907	.182685	.932757	.438818

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_BIO (hjbanova.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{11} 1.583159	{12} 1.325823	{13} 1.582358	{14} 1.352252	{15} 1.640743
B	8803	{10}	.743836	.115902	.828277	.143421	.666545
C	98	{11}		.507284	.996070	.485580	.933045
C	875	{12}	.507284		.390250	.870736	.454135
C	878	{13}	.996070	.390250		.332106	.984099
C	8711	{14}	.485580	.870736	.332106		.481064
C	8803	{15}	.933045	.454135	.984099	.481064	

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_H (hjbanova.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{1}	{2}	{3}	{4}	{5}
			.2153919	.2206327	.1872593	.1796189	.2190918
A	98	{1}		.991158	.511165	.409845	.976528
A	875	{2}	.991158		.569836	.387875	.931105
A	878	{3}	.511165	.569836		.903665	.557407
A	8711	{4}	.409845	.387875	.903665		.395696
A	8803	{5}	.976528	.931105	.557407	.395696	
B	98	{6}	.465025	.362372	.019994*	.005657*	.424508
B	875	{7}	.564282	.581882	.942066	.721819	.579406
B	878	{8}	.890789	.986550	.521107	.385997	.943963
B	8711	{9}	.940733	.989061	.426241	.449879	.983100
B	8803	{10}	.088062	.083173	.000650*	.000132*	.092243
C	98	{11}	.000010*	.000020*	.000020*	.000026*	.000026*
C	875	{12}	.965679	.997225	.416106	.363039	.994477
C	878	{13}	.877212	.944035	.406654	.589453	.931494
C	8711	{14}	.926602	.603486	.288093	.149972	.816918
C	8803	{15}	.428210	.261284	.020873*	.006234*	.350200

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_H (hjbanova.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{6}	{7}	{8}	{9}	{10}
			.2496576	.1859640	.2178391	.2094587	.2657962
A	98	{1}	.465025	.564282	.890789	.940733	.088062
A	875	{2}	.362372	.581882	.986550	.989061	.083173
A	878	{3}	.019994*	.942066	.521107	.426241	.000650*
A	8711	{4}	.005657*	.721819	.385997	.449879	.000132*
A	8803	{5}	.424508	.579406	.943963	.983100	.092243
B	98	{6}		.018258*	.475174	.369488	.365164
B	875	{7}	.018258*		.555779	.551153	.000552*
B	878	{8}	.475174	.555779		.965602	.100434
B	8711	{9}	.369488	.551153	.965602		.050362
B	8803	{10}	.365164	.000552*	.100434	.050362	
C	98	{11}	.000221*	.000023*	.000032*	.000015*	.002123*
C	875	{12}	.505252	.492001	.982236	.771901	.095663
C	878	{13}	.185228	.638726	.902144	.677507	.015365*
C	8711	{14}	.508304	.287685	.906165	.913635	.182402
C	8803	{15}	.948575	.019441*	.420941	.356948	.595824

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_H (hjbanova.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{11}	{12}	{13}	{14}	{15}
			.3205605	.2146250	.2020474	.2298890	.2485081
A	98	{1}	.000010*	.965679	.877212	.926602	.428210
A	875	{2}	.000020*	.997225	.944035	.603486	.261284
A	878	{3}	.000020*	.416106	.406654	.288093	.020873*
A	8711	{4}	.000026*	.363039	.589453	.149972	.006234*
A	8803	{5}	.000026*	.994477	.931494	.816918	.350200
B	98	{6}	.000221*	.505252	.185228	.508304	.948575
B	875	{7}	.000023*	.492001	.638726	.287685	.019441*
B	878	{8}	.000032*	.982236	.902144	.906165	.420941
B	8711	{9}	.000015*	.771901	.677507	.913635	.356948

STAT. GENERAL MANOVA		Newman-Keuls test; LOG_H (hjbanova.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION	{11} .3205605	{12} .2146250	{13} .2020474	{14} .2298890	{15} .2485081
B	8803 {10}	.002123*	.095663	.015365*	.182402	.595824
C	98 {11}		.000012*	.000017*	.000020*	.000312*
C	875 {12}	.000012*		.760079	.956614	.479355
C	878 {13}	.000017*	.760079		.772934	.183412
C	8711 {14}	.000020*	.956614	.772934		.296126
C	8803 {15}	.000312*	.479355	.183412	.296126	



STAT. GENERAL MANOVA			Newman-Keuls test; LOG_J (hjbanova.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{1} .2218657	{2} .2486083	{3} .2237438	{4} .2310587	{5} .2698632
A	98	{1}		.385443	.900545	.813679	.037758*
A	875	{2}	.385443		.347967	.472421	.618269
A	878	{3}	.900545	.347967		.626400	.044644*
A	8711	{4}	.813679	.472421	.626400		.131274
A	8803	{5}	.037758*	.618269	.044644*	.131274	
B	98	{6}	.313098	.952501	.247702	.267719	.680943
B	875	{7}	.358600	.948356	.362706	.576808	.511365
B	878	{8}	.052172	.788462	.064101	.189373	.999940
B	8711	{9}	.323493	.800115	.312971	.486036	.651345
B	8803	{10}	.027773*	.705857	.035727*	.123354	.994053
C	98	{11}	.014420*	.641355	.019671*	.080370	.995176
C	875	{12}	.013813*	.658388	.019110*	.080293	.997568
C	878	{13}	.087182	.718458	.096390	.228230	.717314
C	8711	{14}	.031455*	.760664	.040931*	.141167	.998954
C	8803	{15}	.044320*	.712635	.053593	.158825	.992719

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_J (hjbanova.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{6} .2477131	{7} .2532689	{8} .2700205	{9} .2524133	{10} .2737254
A	98	{1}	.313098	.358600	.052172	.323493	.027773*
A	875	{2}	.952501	.948356	.788462	.800115	.705857
A	878	{3}	.247702	.362706	.064101	.312971	.035727*
A	8711	{4}	.267719	.576808	.189373	.486036	.123354
A	8803	{5}	.680943	.511365	.999940	.651345	.994053
B	98	{6}		.982753	.816023	.947498	.727324
B	875	{7}	.982753		.798826	.954601	.750216
B	878	{8}	.816023	.798826		.850429	.805273
B	8711	{9}	.947498	.954601	.850429		.792151
B	8803	{10}	.727324	.750216	.805273	.792151	
C	98	{11}	.649265	.732169	.955510	.754851	.961014
C	875	{12}	.661326	.761115	.980498	.776429	.989166
C	878	{13}	.800313	.457928	.982371	.703507	.972102
C	8711	{14}	.774419	.819414	.965523	.846386	.995392
C	8803	{15}	.755034	.681215	.998934	.768350	.966706

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_J (hjbanova.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{11} .2777613	{12} .2784616	{13} .2644227	{14} .2738122	{15} .2700005
A	98	{1}	.014420*	.013813*	.087182	.031455*	.044320*
A	875	{2}	.641355	.658388	.718458	.760664	.712635
A	878	{3}	.019671*	.019110*	.096390	.040931*	.053593
A	8711	{4}	.080370	.080293	.228230	.141167	.158825
A	8803	{5}	.995176	.997568	.717314	.998954	.992719
B	98	{6}	.649265	.661326	.800313	.774419	.755034
B	875	{7}	.732169	.761115	.457928	.819414	.681215
B	878	{8}	.955510	.980498	.982371	.965523	.998934
B	8711	{9}	.754851	.776429	.703507	.846386	.768350

STAT.  
GENERAL  
MANOVA

Newman-Keuls test; LOG\_J (hjbanova.sta)  
Probabilities for Post-Hoc Tests  
INTERACTION: 1 x 2

SITE OCCASION			{11} .2777613	{12} .2784616	{13} .2644227	{14} .2738122	{15} .2700005
B	8803	{10}	.961014	.989166	.972102	.995392	.966706
C	98	{11}		.962836	.974506	.792715	.985750
C	875	{12}	.962836		.982768	.948598	.993342
C	878	{13}	.974506	.982768		.989231	.926869
C	8711	{14}	.792715	.948598	.989231		.994279
C	8803	{15}	.985750	.993342	.926869	.994279	

## **Appendix 11**

**Results of the Newman-Keuls post-hoc test for one-way ANOVA, performed on the pooled data of 1987/88 versus the data of 1998**

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_PH (2wayanv2.sta) Probabilities for Post_Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{1}	{2}	{3}	{4}	{5}
			.9316571	.9669102	.9646790	.9695172	.9434561
A	98	{1}		.000008*	.000022*	.000017*	.000036*
A	87/88	{2}	.000008*		.428526	.354929	.000022*
B	98	{3}	.000022*	.428526		.198837	.000009*
B	87/88	{4}	.000017*	.354929	.198837		.000008*
C	98	{5}	.000036*	.000022*	.000009*	.000008*	
C	87/88	{6}	.000020*	.013240*	.001729*	.057820	.000017*

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_PH (2wayanv2.sta) Probabilities for Post_Hoc Tests INTERACTION: 1 x 2	
SITE	OCCASION		{6}	
			.9748633	
A	98	{1}	.000020*	
A	87/88	{2}	.013240*	
B	98	{3}	.001729*	
B	87/88	{4}	.057820	
C	98	{5}	.000017*	
C	87/88	{6}		

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_SALT (2wayanv2.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{1}	{2}	{3}	{4}	{5}
			1.495666	1.514023	1.513168	1.505912	1.494029
A	98	{1}		.075086	.067213	.150719	.818456
A	87/88	{2}	.075086		.904651	.666211	.056784
B	98	{3}	.067213	.904651		.565594	.056277
B	87/88	{4}	.150719	.666211	.565594		.218142
C	98	{5}	.818456	.056784	.056277	.218142	
C	87/88	{6}	.153703	.748571	.544950	.680022	.159946

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_SALT (2wayanv2.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
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SITE	OCCASION		{6}
			1.508852
A	98	{1}	.153703
A	87/88	{2}	.748571
B	98	{3}	.544950
B	87/88	{4}	.680022
C	98	{5}	.159946
C	87/88	{6}	

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_TSS (2wayanv2.sta) Probabilities for Post-Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{1}	{2}	{3}	{4}	{5}
			2.044294	2.111540	1.239936	1.695618	1.042910
A	98	{1}		.763501	.003414*	.267281	.000400*
A	87/88	{2}	.763501		.002178*	.251555	.000264*
B	98	{3}	.003414*	.002178*		.044837*	.379216
B	87/88	{4}	.267281	.251555	.044837*		.012947*
C	98	{5}	.000400*	.000264*	.379216	.012947*	
C	87/88	{6}	.164088	.209348	.077849	.874118	.015446*

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_TSS (2wayanv2.sta) Probabilities for Post-Hoc Tests INTERACTION: 1 x 2	
SITE	OCCASION		{6}	
			1.731923	
A	98	{1}	.164088	
A	87/88	{2}	.209348	
B	98	{3}	.077849	
B	87/88	{4}	.874118	
C	98	{5}	.015446*	
C	87/88	{6}		

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_SP (hjbanov2.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{1} .9479923	{2} .5474777	{3} .9873315	{4} .8332209	{5} 1.218847
A	98	{1}		.000008*	.520737	.060982	.000049*
A	87/88	{2}	.000008*		.000017*	.000030*	.000020*
B	98	{3}	.520737	.000017*		.031856*	.000164*
B	87/88	{4}	.060982	.000030*	.031856*		.000008*
C	98	{5}	.000049*	.000020*	.000164*	.000008*	
C	87/88	{6}	.123514	.000013*	.046352*	.934993	.000017*

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_SP (hjbanov2.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2	
SITE	OCCASION		{6} .8282241	
A	98	{1}	.123514	
A	87/88	{2}	.000013*	
B	98	{3}	.046352*	
B	87/88	{4}	.934993	
C	98	{5}	.000017*	
C	87/88	{6}		

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_BIO (hjbanov2.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{1}	{2}	{3}	{4}	{5}
			1.140461	.8364168	1.600946	1.496081	1.583159
A	98	{1}		.020055*	.003938*	.017949*	.003965*
A	87/88	{2}	.020055*		.000020*	.000010*	.000017*
B	98	{3}	.003938*	.000020*		.701751	.891795
B	87/88	{4}	.017949*	.000010*	.701751		.505414
C	98	{5}	.003965*	.000017*	.891795	.505414	
C	87/88	{6}	.010682*	.000024*	.767305	.867684	.682734

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_BIO (hjbanov2.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
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SITE	OCCASION		{6}
			1.474297
A	98	{1}	.010682*
A	87/88	{2}	.000024*
B	98	{3}	.767305
B	87/88	{4}	.867684
C	98	{5}	.682734
C	87/88	{6}	



STAT. GENERAL MANOVA			Newman-Keuls test; LOG_H (hjbanov2.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{1}	{2}	{3}	{4}	{5}
			.2153919	.2032851	.2496576	.2217164	.3205605
A	98	{1}		.387004	.068305	.651335	.000017*
A	87/88	{2}	.387004	.387004	.008205*	.385652	.000020*
B	98	{3}	.068305	.008205*		.113047	.000009*
B	87/88	{4}	.651335	.385652	.113047		.000008*
C	98	{5}	.000017*	.000020*	.000009*	.000008*	
C	87/88	{6}	.839419	.481285	.059466	.911031	.000022*

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_H (hjbanov2.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{6}				
			.2232803				
A	98	{1}	.839419				
A	87/88	{2}	.481285				
B	98	{3}	.059466				
B	87/88	{4}	.911031				
C	98	{5}	.000022*				
C	87/88	{6}					

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_J (hjbanov2.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
SITE	OCCASION		{1} .2218657	{2} .2444446	{3} .2477131	{4} .2634402	{5} .2777613
A	98	{1}		.047712*	.060594	.001535*	.000033*
A	87/88	{2}	.047712*		.774418	.218477	.028725*
B	98	{3}	.060594	.774418		.167865	.041877*
B	87/88	{4}	.001535*	.218477	.167865		.420359
C	98	{5}	.000033*	.028725*	.041877*	.420359	
C	87/88	{6}	.000125*	.075246	.085213	.456944	.608707

STAT. GENERAL MANOVA			Newman-Keuls test; LOG_J (hjbanov2.sta) Probabilities for Post Hoc Tests INTERACTION: 1 x 2				
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SITE	OCCASION		{6} .2719235
A	98	{1}	.000125*
A	87/88	{2}	.075246
B	98	{3}	.085213
B	87/88	{4}	.456944
C	98	{5}	.608707
C	87/88	{6}	