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To the Graduate Council:

I am submitting herewith a dissertation written by David Hothersall entitled "Operant Conditioning of Heart Rate Changes in Curarized Rats with Brain Stimulation Reinforcement." I have examined the final electronic copy of this dissertation for form and content and recommend that it be accepted in partial fulfillment of the requirements for the degree of Doctor of Philosophy, with a major in Psychology.

Jasper Brener, Major Professor

We have read this dissertation and recommend its acceptance:

William H. Calhoun, James N. Liles, Harry M. B. Hurwitz

Accepted for the Council:

Carolyn R. Hodges

Vice Provost and Dean of the Graduate School

(Original signatures are on file with official student records.)

August 12, 1968

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VCalhoin

Accepted for the Council:

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Vice Chancellor for Graduate Studies and Research

OPERANT CONDITIONING OF HEART RATE CHANGES IN CURARIZED RATS WITH BRAIN STIMULATION REINFORCEMENT

A Dissertation Presented to the Graduate Council of The University of Tennessee

In Partial Fulfillment of the Requirements for the Degree Doctor of Philosophy

by

David Hothersall

August 1968

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I wish to express my sincere thanks to the chairman of my committee, Dr. Jasper Brener, for his advice and encouragement. My thanks also go to the other members of my committee, Doctors William H. Calhoun, James N. Liles, and particularly to Dr. Harry M. B. Hurwitz.

This investigation was carried out while the author was the recipient of a Predoctoral Fellowship awarded by the National Institutes of Health.

Finally I would like to thank my wife, Lesley.

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ABSTRACT

Many learning theorists have distinguished between autonomic responses which can be conditioned using classical conditioning procedures, and skeletal responses which can be conditioned using operant or instrumental conditioning procedures. In recent years this distinction has been challenged since such autonomic responses as changes in heart rate, GSRs, vasoconstriction and vasodilation, and changes in blood pressure have all been conditioned as operants.

The aim of this study was to investigate the possibility that both increases and decreases in heart rate could be conditioned in groups of curarized rats using an operant conditioning procedure with brain stimulation as the reinforcement for the heart rate changes. Curarized rats were used in order to markedly reduce the possibility of skeletal mediation of the heart rate changes which were conditioned. Brain stimulation was used since it has been found to have powerful reinforcing effects for more typical operant responses. A feedback stimulus was presented to the rats during the curare conditioning sessions whenever they were emitting criterion heart rates, i.e. heart rates which if sustained for a fixed number of beats would be followed by the brain stimulation. Two groups of six rats were used, one group being reinforced for heart rate increases, and one for decreases. All rats were given four curare conditioning sessions during which heart rate changes were reinforced.

During the first curare conditioning session no evidence of operant conditioning of heart rate changes was found. On the three

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later conditioning sessions significant differences were found in the heart rates of the two groups, with the group reinforced for heart rate increases showing the higher heart rate. There was also some evidence of an extinction effect during the later curare sessions in that the heart rates of the two groups which diverged during conditioning, tended to converge during extinction. The results of this experiment show that changes in heart rate can be conditioned in curarized rats using an operant conditioning procedure with brain stimulation as the reinforcement for the heart rate changes.

In a second experiment three rats were used in an attempt to first condition a heart rate change in one direction, and then to reinforce heart rate changes in the opposite direction. Although the three rats showed good acquisition of the initial heart rate change none of them showed any evidence of acquisition of the heart rate change in the opposite direction. The results from these three rats suggest that heart rate reversals will only be conditioned with great difficulty.

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INTRODUCTION

Traditionally many learning theorists have held that instrumental or operant conditioning procedures are effective only in the modification of skeletal responses under the control of the somatic nervous system, and not in generating control over the smooth muscles, cardiac muscles and glands. In short responses under the control of the autonomic nervous system are thought not to obey the law of effect.¹ Konorski and Miller (1937), Schlosberg (1937) and Skinner (1938) all distinguished between skeletal responses which can be conditioned as operants, and autonomic responses, which can only be conditioned using classical conditioning procedures. For over twenty years this distinction was widely accepted (Mowrer, 1950; Keller and Schoenfeld, 1950; Solomon and Wynne, 1954; Morgan, 1965). Bugelski (1956), in discussing the experiments of Pavlov on classical conditioning, and of Skinner and Mowrer on operant and instrumental conditioning, wrote:

For Mowrer and Skinner, Pavlov might account satisfactorily for the learning of autonomic nervous system responses...but for any behavior involving essentially the whole organism, some kind of reward learning is presumed (Bugelski, 1956, p. 68).

The clearest statement of this distinction was made by Kimble (1961)

¹The first clear statement of the law of effect is to be found in Thorndike (1911, p. 24). He proposed that responses which have satisfying consequences are strengthened and those followed by discomfort or annoyance are weakened. More recent statements of this law differ from Thorndike's original one in that other terms (reward, reinforcement, punishment) have been substituted for satisfaction and annoyance.

in his text on Conditioning and Learning. Kimble stated:

Thus for autonomically mediated behavior, the evidence points unequivocally to the conclusion that such responses can be modified by classical, but not instrumental, training methods (Kimble, 1961, p. 100).

The validity of this long-standing distinction has been challenged, especially in recent years, by the results of a number of experiments which apparently demonstrate operant conditioning of autonomic responses.² The general procedure used in these experiments has been to place a reinforcing stimulus under the control of an autonomic response such as heart rate, vasodilation or vasoconstriction, the galvanic skin response, or intestinal motility. Using this operant conditioning procedure systematic changes in the frequency of occurrence of these autonomic responses have been observed. However, some other investigators have reported negative results from attempts to condition autonomic responses as operants.³ Still other investigators have challenged the 'positive' results claiming that they do not in fact demonstrate operant conditioning of autonomic responses, but conditioning of some other aspect of the organism's behavior which in turn elicits the autonomic changes which have been observed. Such critics believe that the original distinction between classical and operant conditioning in terms of

²The results of these experiments will be presented and discussed in Chapters I and II of this dissertation.

⁵The results of these experiments will be presented and discussed in Chapters I and II of this dissertation.

the responses to which these procedures can be applied retains its validity, and that all of the results seemingly showing operant conditioning of autonomic responses can be accounted for in terms of the conditioning of some form of mediating response. The foremost proponent of this point of view has been Kendon Smith. In 1954 he formulated his mediation or artifact hypothesis which states that only skeletal responses can be conditioned and:

. . . every 'conditioned visceral response' is in reality an artifact, an innate accompaniment of the skeletal responses inculcated by the conditioning process (Smith, 1954, p. 217).

Smith recently made this point even more strongly in his supplementary comments on his paper 'Conditioning as an Artifact'. He stated:

The broad idea that autonomic conditioning does not really exist and that it is merely counterfeited by somatic learning is suggested so strongly . . . that there seems little distinction in being the one to formalize it (Smith in Kimble, 1967, p. 100).

Briefly stated the aim of the experiments reported in this dissertation was to demonstrate operant conditioning of an autonomic response under conditions where the possibility that the autonomic changes were 'counterfeited by somatic learning' was minimized.

There are at least two positions which might be adopted in replying to the proponents of the mediation or artifact hypothesis.

1. Results showing, for example, operant conditioning of heart rate changes might be regarded as of interest and significance without any consideration of the manner in which the changes in heart rate were actually produced. The important thing, it might be claimed, is that such results show that changes in heart rate can be conditioned in accordance with the principles of operant conditioning. How such conditioning is mediated might be regarded as of little or no interest. An analogy might be drawn between this and the situation in a Skinner box in which the experimenter is rarely interested in, or even aware of, the manner in which the rat actually presses the lever. What is of interest is how the rate of lever pressing can be controlled, not how the response is made. For the present investigator such a position, though methodologically defensible, does not seem satisfactory. The analogy between a discrete and arbitrary response such as lever pressing and a continuous response such as heart rate is tentative. In addition due to the controversial nature of results showing operant conditioning of changes in heart rate, it seems that the onus should rest on the investigator to consider the possibility of skeletal mediation of the autonomic changes which he has obtained, before claiming that operant conditioning of an autonomic response has been demonstrated.

2. The position which was adopted in planning the experiments reported in this dissertation was to give close consideration to the arguments proposed by mediation theorists, and to specifically consider the behaviors which could possibly mediate changes in the response being studied. Such a consideration led to an attempt to eliminate, or at least to markedly reduce, the possibility of such mediation and thus, at least in the ideal case, to the study of operant conditioning of an autonomic response isolated from possible mediating behaviors. However, in adopting this position the investigator is faced with the unhappy task of attempting to establish a negative proposition, i.e., that operant

conditioning of an autonomic response has occurred in the absence of skeletal mediation. Logically such negative propositions have a dubious status and just how impregnable they can be is shown by the exchange of papers between Black and Lang (1964) and Smith (1964). In spite of this difficulty in the experiments reported in this dissertation an attempt was made to at least reduce the possibility of skeletal mediation of the autonomic changes which were conditioned by using curarized Ss.

In the experiments reported in this dissertation both increases and decreases in heart rate have been conditioned in curarized rats using an operant conditioning procedure. Black (1965) termed the heart a 'promiscuous organ' since it is subject to so many different influences. Heart rate is influenced by⁴:

- (a) respiration heart rate increases during inspiration and decreases during expiration (sinus arrythmia);
- (b) skeletal tension;
- (c) blood pressure level under resting conditions there is normally an inverse relationship between blood pressure level and heart rate. This inverse relationship is sometimes described as Marey's law of the heart;
- (d) oxygen deficiencies anoxia leads to acceleration of heart rate (anoxic tachycardia);

⁴This information is taken from pages 123 and 124 of 'Samson Wright's Applied Physiology', 11th edition by Cyril A. Keele and Eric Neil.

- (e) endocrine factors;
- (f) temperature a rise in body temperature leads to an increase in heart rate.

In curarized rats the possibility of changes occurring in respiration and skeletal tension is at least considerably reduced.

Having decided to attempt to operantly condition changes in heart rate in curarized rats one further decision which had to be made concerned the reinforcer to be used in these experiments. Due to the nature of the curarized preparation the number of potential reinforcers is extremely limited. It was decided to use positively reinforcing brain stimulation since this reinforcer had a number of definite advantages. It could be made contingent on heart rate changes in a reliable and controlled manner, thus eliminating any delay between the change in heart rate and the delivery of the reinforcement. Secondly it seems desirable that powerful reinforcers be used in attempts to condition autonomic responses as operants. Brain stimulation has been found to yield powerful reinforcing effects in a number of experiments in which typical operant responses have been conditioned (Olds, 1958; Valenstein and Beer, 1964). All of the rats used in the experiments reported in this dissertation were first trained to lever press for brain stimulation, and this same intensity of stimulation was subsequently used to reinforce heart rate changes during the curare conditioning sessions.

A second important feature of the experiments reported in this dissertation was that a feedback stimulus was presented during the

conditioning phase of each curare conditioning session. This stimulus was contingent on criterion heart rates, i.e., heart rates which if sustained for a fixed number of beats would produce the brain stimulation reinforcement. This feedback stimulus was used since the results of a number of previous experiments (Lisina, 1961; Hnatiow and Lang, 1965; Lang, Sroufe and Hastings, 1967; Brener and Hothersall, 1966, 1967) have shown the importance of feedback for the modification and control of autonomic responses.

A third feature of this experiment was that the animals were given repeated curare conditioning sessions. This was done since it was felt that extent and length of training might be an important factor in the development of autonomic responses as operants. The alternative to using repeated curare conditioning sessions would have been either:

1. To use a large dose of curare which would have maintained the <u>Ss</u> in a completely paralyzed condition for a longer period of time, thus allowing the conditioning phase of the curare session to be prolonged. This procedure was not used in the experiments reported in this dissertation since Black (1967) has shown that large doses of d-tubocurarine have a direct effect on the neural innervation of the heart. In three deeply curarized dogs Black found that cardiac responses to direct vagal stimulation were markedly attenuated. At medium and light levels of curarization the cardiac response to this stimulation was recovered.

2. A second procedure which could have been used would have been to infuse curare at a slow rate throughout the curare conditioning session

by means of a venous cannula and an infusion pump. This technique would allow a constant level of curarization to be maintained throughout the curare session.

The repeated curare conditioning sessions which were used allowed a study of the development over sessions of the heart rate changes as operant responses. In addition this procedure allowed an assessment of the 'extinction' effect within each session, and the carry-over of the conditioned heart rate changes from one session to the next.

Finally I investigated the possibility that both increases and decreases in heart rate could be conditioned in the same rat under different stimulus conditions. A number of different procedures were used, and the results of the application of these procedures are reported.

CHAPTER I

OPERANT CONDITIONING OF CHANGES IN HEART RATE

Harwood (1962) published the first report of an experiment concerned with operant conditioning of changes in heart rate. He used human Ss and attempted to condition heart rate decreases. If a cardiac deceleration occurred in the presence of a blue light a nickel was given to the S. Periods of light and no light alternated with reinforcement only being given in the presence of the light. No evidence of operant conditioning of decreases in heart rate was found in comparisons of heart rates during periods when the light was and was not on. Harwood pointed out that the method which he had used to monitor and record heart rate had resulted in a variable delay between the occurrence of a cardiac deceleration - the operant response - and the delivery of the reinforcement. The average length of these delays was three seconds. It seems likely that with a continuous response such as heart rate delays of this magnitude could have resulted in the reinforcement of a heart rate response other than the one being conditioned. In view of this technical weakness the results of his experiment can not be regarded as in any way conclusive.

The first major attempt to condition heart rate as an operant was reported in 1960 in a doctoral dissertation by Shearn. A brief report of this experiment was published in 1962 (Shearn, 1962). Shearn used a Sidman avoidance procedure in which temporary heart rate accelerations avoided an electric shock. Yoked control Ss were paired with the

experimental <u>Ss</u>. The experimental <u>Ss</u> showed an increase in the number of temporary accelerations in heart rate, while the control <u>Ss</u> showed a decrease. Although this result might be regarded as evidence of operant conditioning of a change in heart rate, there was no decrease in the number of shocks received by the experimental <u>Ss</u> - one criterion of successful avoidance conditioning. In addition all of the <u>Ss</u>, both experimental and yoked control, showed an overall decrease in heart rate during the experimental session.

Brener (1966) used an escape avoidance procedure in an attempt to condition changes in heart rate as an operant. In his first experiment low heart rates, within the repertoire of the S, served to terminate or avoid an electric shock. In the second experiment the effective response was an increase in heart rate. Yoked control Ss were used. Significant heart rate differences were found between the experimental and yoked control Ss in both these experiments. These differences were consonant with predictions based on the principles of operant conditioning. Brener found it easier in these experiments to condition heart rate decreases. An important feature of the procedure of these experiments was the use of visual feedback presented to the Ss when criterion heart beats were emitted. Brener concluded that both increases and decreases in heart rate can be conditioned as operants. The results of these two experiments presented the first clear challenge to the widely held belief that autonomic responses can not be conditioned as operants.

Frazier (1966) used a discriminated avoidance conditioning technique with human Ss. During training ten twenty minute 'epochs', during which a light was present and punishment was available (s^D) periods, alternated with no light periods during which the contingencies were not in effect (S^{Δ}) . If during successive minutes of the 'epoch' S's heart rate did not increase, at the end of the period he was given an electric shock. Frazier attempted to achieve the goal of 'conditioning without awareness' by giving the S a monitoring task to perform during the conditioning period. This goal appears to have been met in that none of the Ss reported that they realized that a change in heart rate was the effective response. In acquisition a steady increase in heart rate was found. In addition there was a maintenance of a high heart rate during the 'epoch' and a return to a lower heart rate during the S $^{\Delta}$ period. During the last training period the light was presented alone without any shocks, but the Ss were not told that they would not receive shocks during this period. Sustained high heart rates were observed. Frazier claims that this demonstrates that the changes in heart rate during the S^{D} periods were due to the conditioning and were not due to some type of sensitizing effect of the electric shocks. Frazier reports that clear increases in heart rate were found during the S^D periods, but they were not accompanied by parallel changes in respiration rate or amplitude. Frazier was also able to demonstrate what he termed 'cardiovascular oscillation'. Following training repeated presentations of the \textbf{S}^{D} stimulus elicited clear heart rate increases, with the S's heart rate decreasing when the stimulus was withdrawn. These results represent an

impressive demonstration of stimulus control over changes in heart rate. It would be interesting to see whether similar results could be obtained in an experiment in which a similar procedure was used but with a decrease in heart rate as the effective response. As yet the results of such an experiment have not been published.

Engel and Hansen (1966) used human $\underline{S}s$ and reinforced them on a beat to beat basis for changes in heart rate. If the <u>S</u>'s heart rate decreased a light in the <u>S</u>'s room was switched on and a timer which the <u>S</u> could see began to run. The light served as a cue for the <u>S</u> and the sight of the timer running was assumed to be reinforcing since the <u>S</u> was later paid a sum of money proportional to the elapsed time shown on the timer. In this experiment five of the ten <u>S</u>s learned to slow their hearts under these conditions. Four of the five non-learners correctly inferred that a decrease in heart rate was the effective response and so it appears that, at least in this experiment, <u>S</u>s were better able to modify their heart rates when they were 'unaware' that heart rate changes were actually being conditioned.

In a second experiment Engel and Chism (1967) used a similar technique in an attempt to condition heart rate increases. In this experiment all five of the <u>Ss</u> learned to increase their heart rates. Since all of the <u>Ss</u> acquired the response in this second experiment, whereas only 50 percent of the <u>Ss</u> acquired the response in their earlier experiment Engel and his co-workers concluded that it is easier to condition heart rate increases as an operant than it is to condition heart rate decreases. This conclusion is in opposition to the results

reported by Brener (1966). Two of the <u>S</u>s in Engel and Chism's experiment were observed to increase their respiration rates, and so it seems that at least in these <u>S</u>s a somatic manoeuvre had been employed. It is interesting that when two of the <u>S</u>s who had successfully increased their heart rates were asked how they had achieved control over the light and the timer, they reported that they had tried to relax completely, a form of behavior which might have been expected on a priori grounds to have led to a decrease rather than to an increase in heart rate. Engel and his co-workers concluded on the basis of the results of these two experiments that human <u>S</u>s can learn to both increase and decrease their heart rates, and that the techniques which are used to regulate heart rate vary widely from one S to another.

Human <u>Ss</u> were used in all of the experiments which have been presented thus far. There have been six reports of experiments on operant conditioning of heart rate changes in which animal <u>Ss</u> have been used. Shearn and Clifford (1964) used rabbits and recorded frequency distributions of inter heart beat intervals (IBIs) under constant conditions and with no experimental contingencies in effect. Within sessions they found that the IBI distributions tended to drift towards the longer IBIs, i.e., there was a decrease in heart rate. During later sessions different <u>Ss</u> were shocked whenever they emitted either a long or a short IBI. The rabbits shocked for short IBIs became 'trapped' and continued to emit short IBIs throughout the session. Consequently there was no decrease in the number of shocks

they received. The <u>Ss</u> shocked for long IBIs oscillated between short and long IBIs. These results are contrary to what might have been expected on the basis of the principles of operant conditioning. The results of this experiment provide little evidence of operant conditioning of a change in heart rate.¹

Black (1967) reported the results of a series of experiments on operant conditioning of changes in heart rate in curarized dogs. In the first of these experiments two groups of dogs were used, one of which - the pretraining group - was operantly conditioned during the first phase of the experiment to press a pedal when a Conditioned Stimulus (CS) was presented in order to avoid a shock; the other group was simply given a series of presentations of the CS alone. In the second phase of the experiment all of the dogs were curarized and each group divided into two sub-groups one of which was reinforced for heart rate increases and one for heart rate decreases. A six second CS - UCS interval was used and if a criterion heart rate response occurred during this interval <u>S</u> avoided the electric shock. If such a response did not occur the shock was presented at the end of the interval. During the second phase of the experiment there was a significant increase in the number of

¹Miller and Carmona (1967) and Miller and DiCara (1967) suggest that the results of this experiment, together with the results of a number of others, seriously challenge the view that visceral responses can not be conditioned as operants. Inclusion of Shearn and Clifford's experiment in a list of experiments giving positive results from attempts to condition autonomic responses as operants does not seem justified.

successful avoidance responses made by both the increase and decrease groups, indicating that both increases and decreases in heart rate could be conditioned in curarized dogs. However increases in heart rate were found to be easier to condition than decreases. A nonsignificant difference favoring the dogs given pre-training in the pedal pressing situation was found. It is interesting that although the responses of the dogs in the increase and decrease groups during the CS - UCS interval were similar, an increase in heart rate followed by a decrease, they were very differnt in form: the duration of the increase in the dogs trained to increase their heart rates was significantly longer than the duration of the increase in dogs for whom the avoidance response was a decrease in heart rate. In later experiments Black used different levels of curarization, light, medium and deep. He found that operant conditioning of heart rate changes occurred more readily at the light level of curarization, than it did at either of the other two. In addition, conditioned cardiac responses which had been formed under light levels of curarization were found to be attenuated when they were transferred to medium or deep levels of curarization. Black suggests that this may have been due to the fact that with large doses of curare there may be a direct blocking of the vagus. Support for this suggestion came from results obtained by Black showing that in a deeply curarized dog the cardiac response to direct electrical stimulation of the vagus was weakened. In general the results of Black's experiments provide a convincing demonstration of operant conditioning of both increases and

decreases in heart rate, under conditions in which the possibility of skeletal mediation of the cardiac changes had been, at least, markedly reduced.

Recently Miller and his students and co-workers have reported the results of a number of experiments on operant conditioning of heart rate changes in curarized rats. In three of these experiments positively reinforcing brain stimulation has been used as the reinforcer for the heart rate changes. Trowill (1967) used rats which had previously been trained to lever press for brain stimulation. While completely paralyzed and artifically respirated these rats received brain stimulation on a fixed interval schedule of reinforcement for either relatively slow or relatively fast heart rates. Fifteen of 19 Ss reinforced for fast heart rates increased their rates, and the mean increase for all 19 Ss was 18 beats per minute (b.p.m.); fifteen of 17 Ss reinforced for slow heart rates decreased their rates, and the mean decrease was 19 b.p.m. The difference in the heart rate changes shown by the two groups was highly significant. Yoked control Ss were used in this experiment and a rather disconcerting feature of the results was that the yoked groups showed heart rate changes which were opposite to those of their experimental partners, and hence to each other. Trowill was unable to find any reason for this result and concluded that it may have been due to chance.

Since the heart rate changes conditioned in Trowill's experiment were relatively small, Miller and DiCara (1967) working in the same laboratory conducted a second experiment to see whether larger changes in heart rate could be conditioned. They attempted to 'shape' the heart

rate response by progressively shifting rats to a more difficult criterion heart rate after they had learned to meet an easier one. In addition they used a 20 second Time Out period to allow the rat's heart rate to recover from the effects of the brain stimulation. Two groups of rats were conditioned to either increase or decrease their heart rates while curarized in order to receive the positively reinforcing brain stimulation. Eleven of 12 rats reinforced for increases in heart rate progressively increased their heart rates from an initial rate of 422 to a final one of 510 b.p.m. Ten of the ll rats reinforced for decreases in heart rate decreased their heart rates from an initial rate of 400 to a final rate of 316 b.p.m. The initial difference in the mean heart rates of the two groups did not approach significance, but the difference at the end of conditioning was highly significant. Since the heart rate changes in the two groups were in different directions they could not be accounted for as due to the unconditioned effects of the brain stimulation. Throughout conditioning Miller and DiCara report that all skeletal responses were completely blocked by the curare.

In a second experiment a discrimination procedure was used with rats being reinforced for either increases or decreases in heart rate in the presence of one stimulus, but no reinforcement being delivered in the absence of the stimulus. The two groups of curarized rats acquired the discrimination of either increasing or decreasing their heart rates in response to the stimulus.

Miller and Banuazizi (1968) reinforced deeply curarized rats maintained on artificial respiration with brain stimulation for either increases or decreases in heart rate. The group reinforced for decreases progressively decreased its heart rate from an average of 425 b.p.m. at the beginning of training to an average of 373 b.p.m. at the end. The group reinforced for increases progressively increased its heart rate from an average of 440 b.p.m. at the beginning of training to an average of 468 b.p.m. at the end.

DiCara and Miller (1968) investigated the possibility of reinforcing changes in heart rate by escape from or avoidance of electric shock. Two groups of curarized rats were used. If on shock trials the criterion heart rate = either a decrease or an increase depending on the S's group membership - was achieved within 5 seconds of the onset of the shock signal the signal was terminated and no shock was delivered. If the S failed to meet the criterion heart rate the shock was presented. As soon as the criterion heart rate was achieved the shock and the signal were terminated. In addition an equal number of non-shock trials were given during which a second non-shock signal was presented for 5 seconds irrespective of the S's heart rate. This signal of course was never followed by an electric shock. Throughout training heart rate in the presence of the shock signal increased in the group reinforced for heart rate decreases. The differences in the average heart rates of the two groups in the presence of the shock signal were statistically significant. In addition there was some evidence that the no-shock stimulus elicited changes in heart rate opposite to those elicited by the shock stimulus.

In this experiment two extremely useful procedures were used by DiCara and Miller. The curare was infused at a slow but constant rate throughout the experimental session thus allowing much more precise control over the level of curarization which was actually achieved. Secondly electromyographic (EMG) recordings were taken from the lateral surface of the right gastrocnemius. Analysis of the EMG records for each \underline{S} at a sensitivity of 150 micro-volts/centimetre showed that within 15 minutes of the curare injection all skeletal muscle activity ceased. The first minute signs of muscle activity only appeared one hour after the end of training. DiCara and Miller concluded:

In the present experiment we have shown that changes in heart rate can be achieved by instrumental training techniques when the animals are so deeply curarized that muscle action potentials are completely abolished (DiCara and Miller, 1968, p. 11).

This series of experiments by Miller <u>et al.</u> has provided convincing evidence of operant conditioning of changes in heart rate in curarized rats using both brain stimulation and escape from, or avoidance of, electric shock as the reinforcement for the heart rate changes. However there is a great need for independent replications of these experiments.

Operant conditioning procedures were not specifically used in the final two sets of experiments I would like to present in this Chapter. However the procedures and results used in these experiments are relevant to the present discussion.

Hnatiow and Lang (1965) reported the results of an attempt to train human Ss to reduce the variability of their heart rates. No aversive stimuli were used in this experiment and S was simply instructed to try to keep his heart rate as steady as possible while observing a visual display which was synchronized with his heart rate. Ss were instructed to try to keep the pointer in this display in the target area in the center of a scale, and the only reinforcement used was the S's perception of his success or failure. Control Ss watched a display which was unrelated to their own heart rate variability and which merely simulated cardiac changes. In experimental Ss the total time spent within the target area was significantly greater during display than it was during non-display periods. In control Ss these differences were non-significant. In experimental Ss the amount of time spent on target increased progressively throughout training. Control Ss showed no such improvement. For the experimental Ss no significant correlations were obtained between measures of heart rate variability and any measure of respiration change.

Lang, Sroufe, and Hastings (1967) used a procedure similar to that of the earlier experiment. Five minute display (feedback) periods alternated with five minute non-display periods. Pairs of experimental and yoked control <u>S</u>s were run. Both <u>S</u>s were instructed to attempt to keep the pointer in the center of the dial. The position of the pointer was determined by the heart rate variability of the experimental <u>S</u>, and so the yoked control <u>S</u> received feedback which was determined by his partner's, rather than by his own, heart rate variability. The experimental $\underline{S}s$ learned to significantly reduce their heart rate variability, whereas the control $\underline{S}s$ did not show any significant reduction in heart rate variability. Lang, Sroufe, and Hastings found no evidence that the control of heart rate variability transferred to conditions of non-feedback. In the successful $\underline{S}s$ no differences were observed between display and non-display periods in respiration rate, variability, or the interaction of these two. The few $\underline{S}s$ who clearly seemed to be manipulating their respiration showed no greater improvement in the task than did $\underline{S}s$ showing very stable respiration during the display periods.

Brener and Hothersall (1966) gave \underline{S} s auditory feedback of their heart rates - a low frequency tone being associated with low heart rates and a high frequency tone with high heart rates. When one signal lamp was on \underline{S} was instructed to attempt to produce only the high tones and to inhibit the low ones; when the second signal lamp was on \underline{S} was instructed to attempt to produce only the low tones and to inhibit the high ones. No indication was given to the \underline{S} s that the tones were associated with their heart rates. The signal lamps alternated. The results showed that the \underline{S} s displayed high heart rates during one period, and low heart rates in the final two periods being statistically significant. The results of this experiment provided clear evidence that under conditions of augmented sensory feedback \underline{S} s can learn to control their heart rates. Changes in respiration were also observed during the two

signal periods, but although the changes in heart rate were very similar in different $\underline{S}s$, the changes in respiration were very dissimilar. The verbal reports of the $\underline{S}s$ as to how they had achieved control over the tones were also very variable.

In a second experiment Brener and Hothersall (1967) used two sessions. In the first session a procedure identical to that of the first experiment was used. The second session was divided into four phases, the first of which was the same as that of the first session. In the second phase a technique of respiration control similar to that of Wood and Obrist (1964) was used: S was instructed to pace his respiration by a signal lamp which flashed at a rate which he found comfortable. Ss were given training in pacing their respiration and after a little practice they were able to breathe very regularly under these conditions. In the third phase of the experiment the conditions of controlled respiration were maintained but S was instructed to attempt to control the tones as he had done before, but without changing his breathing. In the final phase of the experiment respiration control was removed. Once again good heart rate control was demonstrated both with and without respiration control. The results of these two experiments offer firm evidence of heart rate control under conditions of augmented feedback. The results of the second experiment strongly suggest that respiratory changes are not a prerequisite for such control, i.e., that heart rate can be controlled independently of changes in respiration.

The results of the independent and contemporaneous experiments of Lang <u>et al.</u> (1965, 1967) and Brener and Hothersall (1966, 1967) show that under conditions of augmented feedback human <u>Ss</u> can learn to exercise control over both heart rate variability and heart rate itself. These results also suggest that this control is independent of changes in respiration.

CHAPTER II

OPERANT CONDITIONING OF AUTONOMIC RESPONSES

OTHER THAN HEART RATE

In the late 1950s Kimmel and his co-workers began a series of experiments on operant conditioning of the galvanic skin response (GSR). The results of these experiments have been reviewed in some detail by Kimmel (1967) and so they will only be briefly presented in this chapter. Kimmel and Hill (1960) made either pleasant or unpleasant odors contingent on a criterion reduction in skin resistance in two experimental groups. Two groups of yoked control Ss were also used. No differences in rate of emission of the GSR were found between the groups reinforced with the different odors, or between the experimental and control groups. In extinction the experimental Ss showed an increase in response frequency and the yoked control Ss a decrease. Kimmel and Hill were unable to account for this result. This experiment had a number of unsatisfactory features. Due to difficulties in presenting and expelling the different odors there was a variable delay between the emission of the response and the presentation of the reinforcement. Secondly there was no independent demonstration of the reinforcing properties of the stimuli used as 'reinforcers' in this experiment. Without a demonstration that the pleasant and unpleasant odors could in fact function as positive and negative reinforcers, it could be claimed that the results reported by Kimmel and Hill (1960) demonstrate that the odors they used do not function as reinforcers, rather than the more obvious claim that the GSR can not be conditioned as an operant.

Fowler and Kimmel (1962) used the presentation of a dim light as a reinforcer - S being seated in a dark sound-proof room. It is important to note that the S was not instructed to attempt in any way to turn the light on as often as he could. Fowler and Kimmel simply assumed that for a S in a dark room the presentation of such a dim light would be reinforcing. In view of the difficulties which have been encountered in using light onset as a reinforcer in studies using animals (Hurwitz, 1956; Lockard, 1963; Kiernan, 1964), an independent demonstration of the reinforcing properties of the light would have been valuable. They found in this experiment that whereas the yoked control Ss showed a steady decrease in rate of emission of GSRs, the experimental Ss did not show a decrease. This resulted in a statistically significant difference between the two groups in the numbers of GSRs they emitted during the last two minutes of acquisition and the first two minutes of extinction. Kimmel and Kimmel (1963) replicated this study and according to Kimmel (1967) their results:

. . provided more convincing evidence of operant conditioning of an autonomically mediated response than had been obtained previously. Not only did the instrumental <u>Ss</u> make significantly more responses than their yoked controls during reinforcement and extinction, but the instrumental <u>Ss'</u> response frequency during acquisition and most of extinction was higher than it had been during the last 5 minutes of habituation prior to reinforcement (Kimmel, 1967, p. 338).

Johnson (1963) used verbal reinforcement (That's good! That's fine!) in an investigation of the effects of response contingent reinforcement upon the rate of emission of the GSR. Five experimental Ss read through a set of cards containing nonsense syllables and

whenever they emitted a GSR they received a verbal reinforcement. Three control <u>Ss</u> received the same number of reinforcements but they were randomly presented. The experimental group of <u>Ss</u> showed a significantly greater rate of emission of GSRs than did the control <u>Ss</u> on three of the four conditioning days. Johnson concluded that verbal reinforcement can modify the rate of emission of the GSR.

Greene (1966) used a light as the reinforcer for GSRs. During 'reinforcement' the yoked control <u>Ss</u> showed a decline in the rate of emission of GSRs, whereas in general the experimental <u>Ss</u> did not. This differential effect resulted in a statistically significant difference in the number of GSRs emitted by the two groups in acquisition. Rice (1966) presented visual reinforcement to his <u>Ss</u> only when a GSR occurred in the absence of observed EMG responses. Significant differences between experimental and control <u>Ss</u> in rate of emission of the GSR were found under these conditions.

Shapiro, Crider and Tursky (1964) presented experimental <u>Ss</u> with a tone whenever they emitted a criterion electrodermal response (EDR)¹. Control <u>Ss</u> received the same number of tones as their experimental partners but at times of no skin potential changes. The experimental Ss maintained a mean rate of responding at or above their resting level,

^LThe distinction between the galvanic skin response (GSR) and the electrodermal response (EDR) is based upon the technique of measurement which is used. A GSR is recorded by passing a small current between two electrodes on the skin and measuring the resistance between them with a sensitive galvanometer. The electrodermal response (EDR) is measured by determining the voltage potential across two electrodes on the skin. Electrical activity of the sweat glands underlies both the GSR and the EDR (Sidowski, 1966, p. 173).

while the rate of responding in the control Ss declined over time. This resulted in a statistically significant difference between the groups in numbers of EDRs. Crider, Shapiro and Tursky (1964) reported the results of a second experiment in which an intra-S control procedure was used. Each S received a period of contingent reinforcement of EDRs and a period of non-contingent reinforcement, within a single session. During the conditioning periods Ss emitted more EDRs than they did during the periods of non-contingent reinforcement. Shapiro and Crider (1967) used monetary reinforcement of EDRs. Under different stimulus conditions they obtained differences in response rate consonant with changes in the scheduling of reinforcement. Birk, Crider, Shapiro and Tursky (1966) administered an immobilizing dose of d-tubocurarine to a volunteer prior to a reinforcement session in which EDRs were reinforced. Although S was unable to lift his arms or move his body he was able to breathe without difficulty throughout the session. Under these conditions an increased rate of emission of the EDR was found when each response produced a tone. They concluded that curarization had no effect on the conditioning phenomenon which their previous experiments had demonstrated.

Gavalas (1967) investigated the possibility that operant reinforcement could increase the rate of emission of the GSR. Eight human <u>Ss</u> were used and each <u>S</u> was run for ten consecutive days - each daily session being ten minutes in length. The first two days were used to determine the <u>S</u>'s operant level for emission of the GSR; on the third day

the reinforcer was introduced and was presented randomly for ten minutes in order to measure the <u>S</u>'s reaction to it; reinforcement of GSRs began during the second half of the third day's session and continued for the next four days; the last three days were extinction sessions. Control <u>S</u>s followed the same procedure except that they received random, noncontingent reinforcement during the four and a half days of conditioning. Criterion GSRs were reinforced during conditioning by a flash of light and the comment 'That's good' or 'That's fine'. During the experiment all <u>S</u>s performed a distracting task - pronouncing nonsense syllables. Experimental and control <u>S</u>s differed significantly on three of the four days of conditioning and on the last day of extinction in rate of emission of GSRs. No significant differences were found during adaptation or during the period when the reinforcer was first introduced during the third day.

In a second experiment Gavalas matched $\underline{S}s$ on the basis of their rate of emission of GSRs during adaptation. One member of each pair was assigned to the experimental group, and one member to the yoked control group. Six of the nine experimental $\underline{S}s$ showed a clear increase in rate of emission of the GSR as compared with their yoked controls, while three of the experimental $\underline{S}s$ did not show any such increase. However the group differences between the experimental and yoked control $\underline{S}s$ in rate of emission of the GSR were significant on the last day of conditioning and the second day of extinction.

Higgins(1967) made either a positive reinforcer, a visual stimulus, or a negative reinforcer, an electric shock, contingent on GSRs in two groups of rats. Statistically significant differences were found in the

number of GSRs given by these two groups and Higgins concluded that the GSR can be modified using instrumental conditioning procedures.

There have been two recent reports of attempts to suppress GSRs using operant techniques. Senter and Hummel (1965) gave electric shocks to human <u>S</u>s whenever they emitted a GSR. A decreased rate of GSR emission was observed in these <u>S</u>s. Johnson and Schwartz (1967) confirmed this finding in an experiment in which a loud, and presumably aversive, tone was presented following spontaneous GSRs. A significant suppression effect was found and this effect persisted in extinction.

The results which have been presented thus far give the impression that a clear and unequivocal affirmative answer can be given to the question 'Can the GSR be conditioned as an operant?' In order to correct this impression some negative results will now be presented.

Mandler, Preven and Kuhlman (1962) made a visual stimulus contingent upon GSRs emitted during reinforcement periods. <u>Ss</u> subsequently received a sum of money proportional to the number of times the light had been switched on. They used an intra-<u>S</u> control procedure in which evidence of conditioning was sought in comparisons of response rates during stimulus periods when the contingencies were and were not in effect. Although more GSRs were emitted during the S^D periods, evidence for operant conditioning of the GSR, there was a general decrease in the rate of emission of this response throughout the session. Mandler <u>et al.</u> concluded that their results showed no evidence of operant conditioning of the GSR. This conclusion is directly opposed to the

conclusion of Kimmel <u>et al.</u>, and is somewhat disturbing since Mandler <u>et al</u>. used what would appear to be a stronger reinforcer than the one used by Kimmel and Kimmel (1963). A light signalling money would be expected on a priori grounds to be a stronger reinforcer than a light alone. In support of Mandler <u>et al.</u> Mednick (1964) reported negative results from an attempt to condition the GSR as an operant.

Stern, Boles and Dionis (1966) gave <u>Ss</u> nickels whenever they emitted a spontaneous GSR, and failed to obtain operant conditioning of this response. Stern (1967) reported negative results in a study in which an attempt was made to condition spontaneous GSRs as operants. Four groups of sixteen human <u>Ss</u> were used. For the <u>Ss</u> in Group 1 a light was switched on as soon as they emitted a GSR; the <u>Ss</u> in Group 2 were yoked directly to those in Group 1 and received identical and simultaneous reinforcements. The <u>Ss</u> in Group 3 were indirectly yoked to Group 1 receiving the same number of reinforcements per minute, but only when <u>S</u> was not giving a GSR. Group 4 received no reinforcement. The GSR data showed no separation of these four groups in terms of number of GSRs during either conditioning or extinction, and there was no systematic change in rate of responding over time.

Kimmel and Kimmel (1968) reported the failure of an attempt to condition the GSR as an operant. They used their standard procedure for reinforcing spontaneous GSRs, except that a bright, red light rather than the previously used dim, white one was used as the reinforcer. The experimental Ss reinforced for spontaneous GSRs showed no tendency

towards increased responding during conditioning. In attempting to account for these unexpected results they could only suggest that possibly the red light was an aversive stimulus for the S. Support for this suggestion came from results reported by Kimmel and Kimmel (1968) for a group of Ss for whom the reinforcement for spontaneous GSRs was switching off the red light. In conditioning this group showed a systematic tendency toward increased responding, and persistence of the responding in extinction. Although these results do support the suggestion that the red light was aversive to the S it must be pointed out that initially they felt that changing from the dim white light used in their earlier experiments to the bright red light used in this one would be insignificant, and so the red light was not obviously aversive. It will be most interesting to see whether Kimmel is able to replicate his earlier findings of operant conditioning of the GSR, using the dim light as the reinforcer, in later studies. This latest finding of Kimmel and Kimmel is regarded as strongly supporting the suggestion, which was made earlier in this chapter, that an independent demonstration of the reinforcing properties of the stimuli used as 'reinforcers' in a number of Kimmel et al.'s experiments would have been very valuable. Post hoc designation of the stimuli as either positive or aversive, this designation being made on the basis of the results of attempts to use the stimuli to reinforce GSRs, seems completely unsatisfactory.

Why certain investigators have consistently reported negative results from attempts to operantly condition the GSR, while certain other investigators have reported positive results from such attempts,

is not clear at the present time. However it can safely be said that the question of whether or not the GSR can be conditioned as an operant is far from being completely answered.

Avoidance conditioning procedures have also been used in a number of attempts to operantly condition the GSR. The first of these was by Mowrer (1938) who reported his failure to condition the GSR in an experiment in which shock avoidance was contingent on a change in skin resistance. Kimmel and Baxter (1964) used a CS - UCS interval of five seconds, and if S made a criterion GSR during this period he avoided the electric shock. Yoked control Ss were used. The experimental Ss were found to emit significantly larger GSRs than did the controls, but the actual number of criterion GSRs did not increase, nor did the number of successful avoidances. Two replications of this experiment were reported by Kimmel, Sternthal and Strub (1966) in which a more objective criterion for the GSR was used. The differences between the experimental and yoked control groups were not significant in either of these replications, although the direction of these differences was the same as that of the earlier study, i.e., the experimental Ss made more and larger, but not significantly so, GSRs than did the control Ss. Grings (1965) found that avoidance Ss made slightly, but non-significantly, more GSRs than did control Ss. Grings and Carlin (1966) used both a punishment and an avoidance paradigm in an attempt to condition the GSR. The avoidance Ss were found to increase the number of GSRs they emitted, although the average magnitude of these responses was reduced. The punished Ss emitted fewer GSRs and the responses which they did emit were reduced in size.

Van Twywer and Kimmel (1966) attempted to condition the GSR as an avoidance response. An important feature of this experiment was that only GSRs which did not occur within five seconds of an EMG response, or a respiration irregularity, were reinforced. Although there may be difficulties in formulating a universally acceptable definition of exactly what constitutes an 'EMG response' or a 'respiration irregularity' the use of these measures does constitute an improvement of their experimental procedures. Significant differences in the numbers of GSRs emitted by the experimental and yoked control <u>Ss</u> were found. Similar results have been obtained by Kimmel and Sternthal (1967) who were able to successfully condition the GSR as an avoidance response. They also reported that the GSRs were not accompanied by any systematic changes in EMG frequency, respiration rate or respiration irregularities.

Since in these experiments both positive and negative results have been obtained it is difficult to formulate a conclusion as to whether or not the GSR can be conditioned as an operant. In addition it is difficult to generalize from the GSR to autonomic responses in general since the GSR can not be regarded as a representative autonomic response. The post-ganglionic fibers which innervate the sweat glands are cholinergic, unlike most post-ganglionic fibers of the sympathetic division of the autonomic nervous system (A.N.S.) which are adrenergic (Thompson, 1967, p. 198). Keele and Neil (1961) have pointed out that the GSR, unlike other autonomic responses, does not have any clear biological utility. In addition the GSR seems to function independently of the rest of the A.N.S. Shapiro, Crider and Tursky (1964), for example,

report operant conditioning of the GSR without any correlated changes in other autonomic activities - heart rate and skin potentials. So there is a need both for further experiments on the GSR as well as experiments on operant conditioning of other autonomic responses.

Very few experiments have been performed on operant conditioning of autonomic responses other than heart rate and the GSR. The first attempt to condition vasoconstriction as an operant was made by Skinner and Delabarre. In describing this experiment Skinner (1938) writes:

In collaboration with Dr. E. B. Delabarre I have attempted to condition vasoconstriction of the arm in human subjects by making a positive reinforcer dependent upon constriction. The experiments have so far yielded no conclusive results, but there are many clinical observations that seem to indicate conditioning of this sort² (Skinner, 1938, p. 112).

An experiment by the Russian worker Lisina, as quoted by Razran (1961) demonstrates operant conditioning of a vasomotor response. Lisina observed that while the unconditioned vasomotor reaction to electric shock was constriction, occasionally <u>S</u>s showed a vasodilatory response prior to the onset of the shock. An experimental

 $^{^{2}}$ The results of this experiment, together with those on operant conditioning of the GSR reported by Mowrer (1938), are the 'evidence' to which Kimble (1961) refers in his statement:

Thus, for autonomically mediated behavior, the evidence points unequivocally to the conclusion that such responses can be modified by classical, but not instrumental, training methods (Kimble, 1961, p. 100).

The evidence available to Kimble hardly seems to have justified such an unequivocal conclusion. Subsequent evidence has of course cast considerable doubt upon the validity of Kimble's conclusion.

procedure was used in which if a vasodilation occurred prior to shock onset, the shock was not presented, i.e., an avoidance conditioning paradigm was used with vasodilation as the effective response. Lisina ran this procedure for some time and found no increase in the frequency of dilatory responses. Visual feedback of vasomotor activity was then introduced by allowing the <u>Ss</u> to watch their plethysmographic records. Under the feedback conditions it was observed that vasodilation increased rapidly in frequency with a concomitant decrease in the number of shocks which the <u>Ss</u> received. The results of this experiment are of importance in indicating that vasodilation can be conditioned as an operant, and secondly in indicating the crucial role played by feedback in operant conditioning of an autonomic response.

Fromer (1963) conducted a study of instrumental conditioning of vasomotor responses in the rabbit. The results of this experiment failed to provide strong evidence of conditioning of the response. Snyder and Noble (1968) presented a light to human <u>Ss</u> on the peak amplitude of each vasoconstriction in the right index finger which was not preceded by bodily movement. <u>Ss</u> were told that the light would indicate correct responses. Two experimental groups were run as were two control groups, which received an equal number of presentations of the lights, but these presentations were contingent on vasomotor stability. Experimental <u>Ss</u> showed a significant increase in the number of vasoconstrictions during conditioning, while the control <u>Ss</u> did not show any such increase. In the experimental Ss conditioned vaso-

constrictions were independent of gross bodily movement, muscle tension in the finger and forearm, heart rate and respiratory irregularities.

DiCara and Miller (1968) reported a dramatic demonstration of operant conditioning of a vasomotor response. Rats were curarized and given a series of training trials signaled by the onset of a tone. During the tone Ss could obtain positively reinforcing brain stimulation by making specified vasomotor responses. Six rats were reinforced for relatively greater vasodilation in the right ear, and six for relatively greater dilation in the left ear. Ss reinforced for right ear vasodilation showed significant increases in dilation of the right ear, and significant decreases in vasodilation of the left ear, with the difference between the two ears in number of dilations being highly significant at the end of training. Ss reinforced for left ear dilation showed a significant increase in number of dilations of the left ear, but did not show a significant change in vasomotor activity in the right ear. Analyses of heart rate, vasomotor activity of the tail, and temperature did not show any differences between the two groups, or within the groups during training. So the conditioned response was highly specific.³

Plumlee (1968) implanted rhesus monkeys with chronic intraarterial catheters. For four monkeys a discriminated avoidance procedure was used with a ten second tone terminating with a ten ma. shock, unless

⁵DiCara and Miller have also been able to modify vasomotor responses in the tail of the rat using an operant conditioning procedure. A report of this experiment will appear in Commun. Behav. Biol. later this year.

the monkey's diastolic blood pressure rose to high levels and remained high for one second, in which case the tone terminated and the shock was not presented. All four monkeys acquired the discriminated avoidance response, showing diastolic elevations in blood pressure of up to 60 mm. of mercury. No changes in blood pressure were observed in response to a neutral tone which had never been paired with shock. For four other monkeys a similar procedure was used except that the avoidance response was a decrease in systolic blood pressure. All four monkeys showed drops in blood pressure, but the stimulus control over this response was not as good as was the case with the blood pressure increases.

Results such as these are extremely dramatic and if replicated they must surely have important implications for the medical treatment of a number of disorders of the cardiovascular system, for psychology in general, and particularly for the treatment of psychosomatic disorders.

In his 1961 address to the New York Academy of Sciences Neal Miller stated:

I believe it is important to direct research towards this neglected problem in order to find out whether the two branches of the nervous system obey different laws. If they obey the same laws a dog rewarded whenever he salivates strongly should secrete more saliva than one given the same average number of rewards at times when he is salivating least (Miller, 1961, p. 834-835).

Miller and Carmona (1967) reported the results of an experiment in which the procedure suggested in this quotation was used. They attempted to change the rate of spontaneous salivation of mildly thirsty dogs by using water to reinforce some of them when they increased their rate of salivation and others when they decreased their rate. The dogs increased their rate of salivation when reinforced for doing so, and the <u>Ss</u> reinforced for decreases in rate of salivation actually decreased their rates. The differences between the two groups were statistically significant. Miller and Carmona concluded that:

The results clearly show that the autonomically innervated visceral response of salivation can be modified by an instrumental training procedure (Miller and Carmona, 1967, p. 4).

Brown and Katz (1967) reported a similar set of results from an experiment on operant conditioning of the human parotid salivary response. One group of <u>S</u>s received ten cents each time a spontaneous increase in salivary rate occurred. A second group received ten cents in the absence of salivation or for minimal salivation. The number of salivations increased in the group reinforced for salivating but the increase did not attain statistical significance due to the marked inter-<u>S</u> variability. No significant changes in salivation were observed in the second group. They concluded that:

This study indicates that salivary secretion in the human subject may be brought under stimulus control by means of operant reward techniques without evidence of mediation by swallowing (Brown and Katz, 1967, p. 160).

The results of these two experiments indicate that another autonomic response - salivation - is subject to modification and control through the use of operant conditioning techniques.

The final experiment which will be described in this chapter is that of Miller and Banuazizi (1968) who conducted a study to determine whether curarized rats could learn yet another type of autonomically mediated response - the contraction or relaxation of the large intestine. While deeply curarized three $\underline{S}s$ were reinforced with brain stimulation for intestinal relaxation, and four $\underline{S}s$ were reinforced for contraction. Both groups of rats showed progressive changes in intestinal motility in the appropriate direction. Although the brain stimulation initially had an effect on intestinal motility this effect soon disappeared. In addition since the brain stimulation was used to reinforce different intestinal responses the results of this experiment can not be explained as solely due to the unconditioned effects of the brain stimulation.

CHAPTER III

METHOD

Subjects

The <u>Ss</u> used in these experiments were male, albino rats of the Sprague-Dawley strain purchased from Sprague-Dawley, Inc., of Madison, Wisconsin. The rats weighed from 200 to 250 grams at the time of delivery and were housed, six per cage, in large wire mesh cages in the colony room of the Department of Psychology of the University of Tennessee. Purina chow pellets and tap water were available in the home cages throughout the experiment. Supplementary cabbage and meat were given once a week. The colony room was maintained at a temperature of between 70° and 76° F., and was kept on a twelve hour light dark cycle changing at 6:30 A.M. and 6:30 P.M.

Implantation Procedure

For seven days before the operation the rats were housed in individual cages. Although the rats were not found to be particularly wild in order to further tame them and to accustom them to being handled by \underline{E} each rat was handled and petted briefly on each of the seven days prior to the operation. In general the handling technique developed by Weininger, McClelland and Arima (1954) was used.

Twenty four hours before the operation all food was removed from the rat's living cage, although water continued to be available on an <u>ad lib</u>. basis. This deprivation procedure was adopted in

accordance with the suggestion of Routtenberg (1968) that food deprivation for a period of 24 hours before the administration of pento-barbital anesthesia increases the number of rats reaching surgical levels of anesthetisation, decreases the latency of induction of anesthesia, and lengthens the duration of the anesthesia. My own observations of the reactions of rats to pentobarbital anesthesia with and without food deprivation have confirmed Routtenberg's observations.

On the day of the implantation operation the rat was given an intraperitoneal (IP) injection of sodium pentobarbital (Nembutal)¹ at a dosage of 50/mg/kg. Once a surgical level of anesthetisation had been attained, the rat was mounted in a David Kopff stereotaxic instrument, and the implantation operation was begun. In general the procedure described by Miller, Coons, Lewis and Jensen (1961) was employed except that a commercially manufactured bipolar electrode² was used, and, following a suggestion by Goldstein (1967), the screws used to anchor the electrode to the skull were located on the lateral surface of the skull. The bipolar electrode was aimed at the medial forebrain bundle in the posterior portion of the lateral hypothalamus. (Krieg stereotaxic coordinates 1.5 mm. posterior to Bregma, 8.5 mm.

¹The sodium pentobarbital used in these operations was manufactured and supplied by Abbott Laboratories, North Chicago, Illinois.

²The bipolar electrodes were manufactured by Plastic Products Company, Roanoke, Virginia.

ventral to the point of entry, and 1.7 mm. left of the midline. After the operation the rat was returned to his individual cage, and for the next ten days a careful record was kept of his weight. Most rats were found to lose weight following the operation, usually from ten to twenty grams, and then to slowly return to their pre-operative weights. If a rat continued to lose weight he was given supplementary meat.

Testing the Effect of the Brain Stimulation

Ten to fourteen days after the implantation operation each rat was tested in a modified Skinner box in order to assess the effects of the brain stimulation. The dimensions of this box were six by eight by twelve inches, and it was constructed of Plexiglass. A lever was mounted on one wall of this box three inches above the level of the floor. Each lever pressing response made by the rat produced half a second of 60 cycles per second, ac stimulation, which was delivered from a Grass SD5 Stimulator. Fluctuations in brain resistance were minimized by a 100K potentiometer in series with the rat, and the current intensity was measured with an ac microammeter. Initial shaping was used and once the rat was pressing the lever consistently the intensity of the current was adjusted, between the limits of 20 to 90 microamps, so as to find the current intensity which yielded the fastest rate of responding without any gross motor side effects. Once this current intensity had been found it was used on all subsequent Skinner box sessions and was also the intensity of brain stimulation used to reinforce changes in heart rate

during the curare conditioning sessions.³ Prior to the first curare session each rat was given two self-stimulation sessions in the Skinner box, with 500 responses being followed, on a continuous schedule of reinforcement (CRF), by the brain stimulation during each session. Only those rats which responded consistently during both these sessions were given curare conditioning sessions. Between each curare session each rat was tested in the Skinner box with at least 200 responses being reinforced with CRF brain stimulation. This procedure was adopted in order to ensure that curarization had not had any effect on the positively reinforcing properties of the brain stimulation. All of the rats which self-stimulated on the initial Skinner box sessions also self-stimulated on all of the later Skinner box sessions and so it seems safe to conclude that the brain stimulation continued to act as a positive reinforcer throughout the experiment. In two of the rats a small decrease of some ten microamps in the intensity of the brain stimulation which yielded the best self-stimulation rates was found between the first and last Skinner box sessions. In these rats the intensity of the brain stimulation used to reinforce heart rate changes during the curare conditioning sessions was adjusted to match the intensity used during the last Skinner box session. It is interesting that Olds (1958) has also reported that the intensity threshold for rewarding brain stimulation may decline during the course of an experiment.

 $^{^{3}}$ The two exceptions to this general rule are discussed later on this page.

Curare Conditioning Sessions

At the beginning of each curare conditioning session the rat was given an IP injection of 12 units/kg., of d-tubocurarine chloride. This dosage was found to be sufficient to maintain the rats used in this experiment in a completely immobile state for over two hours. During the curare conditioning session the rat lay on a small hammock in a sound shielded box which in turn was inside an electrically shielded enclosure. The hammock was tilted so that the rat's head was slightly lower than the rest of his body. This was done to allow saliva to run out of the rat's mouth. Once paralysis began, usually within two minutes of the injection, the rat was fitted with a mask made of moulded nylon which fitted over the animal's nose and respiration was begun. The mask was connected via plastic tubing to a small animal respirator (E and M Instrument Co., Model V5KG). Respirator settings of a 1:1 Inspiration/ Expiration ratio, and a respiration rate of 70 breaths per minute were used. During the first five minutes following curarization the rat's heart rate was monitored and adjustments made in respiration rate and volume so as to ensure a stable and regular heart rate. Once such a heart rate had been found respiration rate and volume were fixed and no further adjustments were made during the remainder of the experimental session.

⁴ The d-tubocurarine chloride used in this experiment was manufactured and supplied by Eli Lilly and Company, Indianapolis.

The electrocardiogram (EKG) electrodes were Grass EKG silver disc electrodes. They were clipped to shaved areas of the skin lateral to the chest, and to the rear of the stomach. The shaved area of the skin was liberally coated with EKG Sol electrode paste. The EKG was recorded by means of a Grass Model 7 Polygraph. The amplified EKG signal was taken from one of the polygraph's J6 output jacks and fed through a pulse shaping circuit which converted the EKG wave form to a square wave pulse of fixed duration. This pulse provided the input to the BRS solid state control and analysis circuits. A second output from the polygraph was used to drive a cardiotachometer. A third channel on the polygraph was used for a marker pen which recorded the onset and offset of the feedback stimulus, of the brain stimulation, and of the Time Out.

An electrode lead was connected to the bipolar electrode to allow delivery of the brain stimulation. This lead was connected through a relay to a SD5 Stimulator. Signal lamps for the delivery of visual stimuli, and a speaker for auditory stimuli were located respectively in the roof and the door of the sound-shielded box.

Pairs of rats were operated upon and tested at one time. This was done since only two rats could be given their series of curare conditioning sessions at one time. One rat in each pair was assigned on the basis of a coin toss to one of the experimental groups, and the second rat was assigned to the second group. This assignment of the rats to the different groups was done before the first curare conditioning session in order to avoid the possibility of experimenter bias.

The curare conditioning session began with an adaptation phase which lasted for approximately thirty minutes. During this time the rat's heart rate was monitored and recorded and an inter heart-beat interval (IBI) selected so that approximately 50 percent of the IBIs emitted by the rat would be longer than this criterion IBI, and approximately 50 percent would be shorter. Following this adaptation phase the conditioning phase of the session was begun. The conditioning also lasted for approximately thirty minutes. During conditioning the rats in Group 1 were reinforced for increases in heart rate, and the rats in Group 2 for decreases. The procedure in conditioning for a rat being reinforced for heart rate decreases has been notated in Figure 1.

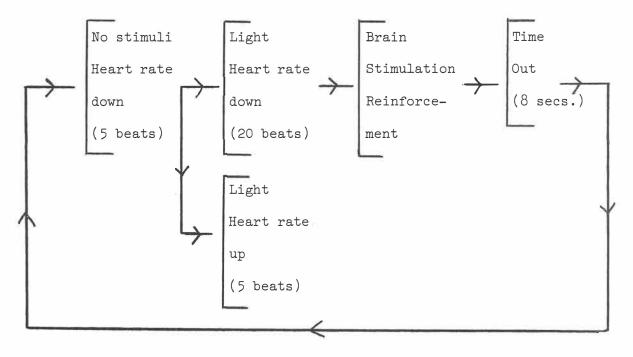


Figure 1. The procedure used in conditioning for a rat being reinforced for heart rate decreases.

At the beginning of the conditioning trial no stimuli were present but the rat's heart rate was monitored and recorded. If the first five IBIs emitted by the rat met the criterion which had been selected on the basis of the rat's adaptation heart rate, a light in the experimental chamber was switched on. If the rat's heart rate did not meet the criterion no stimuli were presented, and this condition continued until a sample of five IBIs was emitted which did meet the criterion. Once the light was switched on, provided that the rat's IBIs continued to meet the criterion, it remained on. When twenty, successive, additional, criterion IBIs occurred, the light terminated with the delivery to the rat of half a second of brain stimulation at the same current intensity as was used in the Skinner box. The brain stimulation was followed by an eight second Time Out period during which none of the stimuli were presented and none of the contingencies were in effect. This Time Out period was introduced to allow the rat's heart rate to recover from the effects of the brain stimulation, since this stimulation was found to elicit a clear decrease in heart rate. Malmo (1961) and Holdstock (1967) have reported a similar slowing of heart rate in rats following septal selfstimulation. At the end of the Time Out period the contingencies were reinstated and the procedure repeated until the end of the conditioning phase of the session. If, in the presence of the light, the rat's heart rate increased the light was switched off and the procedure recycled. If it was found that the rat was not meeting the criterion, and so was not receiving any brain stimulation reinforcements, the criterion was

made less stringent and was kept at this value until it was felt that a change to a more stringent criterion value could be made. If, on the other hand, it was found that the rat was easily meeting the criterion, the criterion was shifted to a more stringent value thus making it more difficult for the rat to obtain the brain stimulation reinforcements. Changes in the criterion were accomplished by shifting it in the appropriate direction by .01 seconds. The procedure for a rat being reinforced for heart rate increases would, of course, be the opposite to the procedure which has been described, i.e., an increase in heart rate for five beats would initially switch the light on and if maintained for an additional twenty beats would lead to the delivery of the brain stimulation. An extinction phase, during which neither the feedback stimulus or the brain stimulation were presented, followed the conditioning phase of the session. This extinction phase lasted for approximately thirty minutes. During the adaptation, conditioning and extinction phases of the curare session the numbers of heart beats in successive one minute periods were recorded. During conditioning the numbers of brain stimulation reinforcements and the criterion changes were also recorded.

Each rat was given four curare conditioning sessions separated by at least three days.

CHAPTER IV

RESULTS AND DISCUSSION

The numbers of heart beats for each of the twelve rats used in the first experiment during successive one minute periods of the adaptation, conditioning, and extinction phases of the four curare conditioning sessions are shown in Tables I to VIII of Appendix A. In addition criterion changes and numbers of brain stimulation reinforcements during conditioning periods are shown in these tables.

In order to present the changes in heart rate which occurred in the two experimental groups within each of the four curare conditioning sessions mean heart rates were calculated for each of the twelve rats during four blocks of adaptation periods, four blocks of conditioning periods, and four blocks of extinction periods. Since there was some variation¹ in the lengths of the three phases of the different curare sessions these means were not based upon equal numbers of one minute periods. Consequently the following procedure was adopted in calculating these means. The number of one minute periods in any phase of the curare session (N) was divided by four, and means calculated for four blocks of N/4 periods. These means for the first and second curare sessions are shown in Tables 1 and 2 and are presented graphically in Figure 2.

¹The minimum and maximum lengths of the adaptation, conditioning and extinction phases of the curare conditioning sessions were 20 and 31 minutes for adaptation, 20 and 31 minutes for conditioning, and 20 and 33 minutes for extinction. The average lengths of the adaptation, conditioning and extinction phases were respectively 25, 27, and 26 minutes.

Table l

Mean heart rates in beats per minute during four blocks of adaptation, conditioning and extinction periods

Session l Rat No. Adap. A A A Cond. C C C E E E Group l: Reinforced for heart rate increases 2 C 487 488 487 487 486 481 474 472 463 453 458 458 6 B 441 425 428 412 428 431 437 439 420 412 404 414 7 C 503 492 482 472 481 483 487 488 491 496 494 489 10 C 524 479 474 464 453 455 448 462 442 453 5 D 392 388 379 370 350 347 348 357 361 353 346 342 Means 472 456 454 449 448 450 449 440 442 Keans 472 456 454 449 448 450 449 447 448
Group 1: Reinforced for heart rate increases 2 C 487 488 487 486 481 474 472 463 453 458 458 6 B 441 425 428 412 428 431 437 439 420 412 404 414 7 C 503 492 482 472 481 483 487 488 491 496 494 489 10 C 524 479 474 486 471 493 497 487 482 490 495 493 4 D 486 465 471 464 453 455 448 462 442 442 453 5 D 392 388 379 370 350 347 348 357 361 353 346 342 Totals 2833 2737 2721 2694 2680 2688 2691 2679 2646 2639 2649 Means 472 456 454 449 447
2 C 487 488 487 487 486 481 474 472 463 453 458 458 6 B 441 425 428 412 428 431 437 439 420 412 404 414 7 C 503 492 482 472 481 483 487 488 491 496 494 489 10 C 524 479 474 486 471 493 497 487 482 490 495 493 4 D 486 465 471 467 464 453 455 448 462 442 442 453 5 D 392 388 379 370 350 347 348 357 361 353 346 342 Totals 2833 2737 2721 2694 2680 2688 2691 2679 2646 2639 2649 Means 472 456 454 449 447 448 450 449
6 B 441 425 428 412 428 431 437 439 420 412 404 414 7 C 503 492 482 472 481 483 487 488 491 496 494 489 10 C 524 479 474 486 471 493 497 487 482 490 495 493 4 D 486 465 471 467 464 453 455 448 462 442 442 453 5 D 392 388 379 370 350 347 348 357 361 353 346 342 Totals 2833 2737 2721 2694 2680 2688 2698 2691 2679 2646 2639 2649 Means 472 456 454 449 447 448 450 449 447 440 442
Means 472 456 454 449 447 448 450 449 447 441 440 442
Group 2: Reinforced for heart rate decreases
1 B5385325325295105054995014884784714896 C4834784824804654534524574494524584538 C3243464014203863823713703633683713749 C56255654153351951951852853453554054910 D482478479479474464456466457459467470
Totals 2839 2837 2876 2880 2795 2760 2734 2738 2717 2718 2735 2755
Means 473 473 479 480 466 460 456 456 453 453 456 459

Ta	ble	2
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Mean heart rates in beats per minute during four blocks of adaptation, conditioning and extinction periods

								-	-		-		
					Se	essio	n 2						
Rat No	. Adaj	p. A	A	А	Cond	. C	С	С	Ext	. E	Έ	Ε	
		Groi	up l:	Rein	force	l for	heart	; rate	incre	eases			
2 C 6 B 7 C 10 C 4 D 5 D	494 457 414 512 474 383	498 476 415 483 455 377	497 465 405 475 454 375	498 464 409 471 453 364	488 479 413 474 464 369	485 497 420 481 481 378	487 490 427 480 490 396	486 488 437 486 485 408	479 482 438 464 478 398	478 478 442 441 479 385	475 479 443 431 478 380	471 470 445 432 475 381	
Totals	2734	2704	2671	2659	2687	2742	2770	2790	2739	2703	2686	2674	
Means	456	451	445	443	448	457	462	465	457	451	448	446	
		Grou	ıp 2:	Rein	forced	l for	heart	rate	decre	eases			
1 B 6 C 8 C 9 C 9 D 10 D	473 411 391 534 455 426	459 414 384 528 439 407	442 413 382 523 411 414	456 410 379 518 366 410	449 405 363 516 356 408	432 401 354 510 342 402	430 395 351 496 335 398	433 397 337 481 331 395	444 404 340 485 329 398	453 408 354 490 330 405	459 409 359 492 331 408	462 416 372 491 331 415	
Totals	2690	2631	2585	2539	2497	2441	2405	2374	2400	2440	2458	2487	
Means	448	439	431	423	416	407	401	396	400	407	410	415	

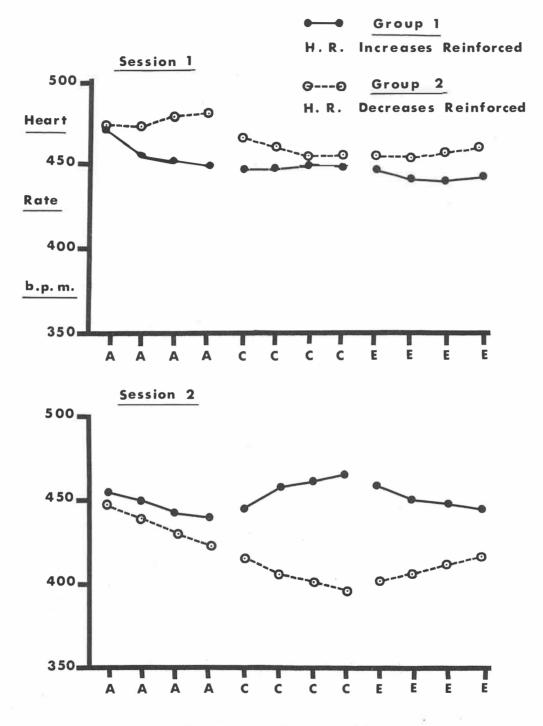


Figure 2. Group mean heart rates for four blocks of adaptation, conditioning and extinction periods during the first and second curare sessions.

During the conditioning phase of the first curare session the group reinforced for decreases in heart rate - Group 2 - showed a small decrease in heart rate resulting in a difference of 24 beats per minute (b.p.m.) between the means for the last block of adaptation periods and the mean for the last block of conditioning periods; and a difference of 10 beats per minute between the means for the first and last blocks of conditioning periods. The small decrease in heart rate shown by this group might be regarded as evidence of operant conditioning of a decrease in heart rate in curarized rats. However the group reinforced for heart rate increases - Group 1 - showed no difference in heart rate between the means for the last block of adaptation periods and the last block of conditioning periods, and a difference of only 2 b.p.m. between the means for the first and last blocks of conditioning periods. This result appears contrary to what might have been expected on the basis of the principles of operant conditioning. On the other hand in the present research project we have consistently found that within a curare session, in the absence of any experimental contingencies, there is an over-all decrease in heart rate, and so the fact that the heart rate of the group reinforced for heart rate increases did not in fact decrease might be regarded as evidence, although admittedly weak evidence, of an operant conditioning effect. However this argument could equally well be used to account for the small decrease in heart rate shown by Group 2 during the first curare conditioning session and so it seems most realistic to conclude that the results of this experiment show little if any evidence of operant conditioning of changes in heart rate during the first curare conditioning session.

Trowill (1967) reported a mean increase, during a curare conditioning session, of 18 b.p.m. in a group of 19 rats reinforced with brain stimulation for increases in heart rate, and a mean decrease of 19 b.p.m. in a group of 17 rats reinforced for decreases in heart rate. Trowill's results for the decrease group are very similar to those of the experiment reported in this dissertation - a decrease of 19 b.p.m. during conditioning as compared with the decrease of 24 b.p.m. in the present experiment. However he was much more successful in conditioning increases in heart rate. Trowill used a training period of ten minutes for nine of the rats, and a period of thirty minutes for the remaining Ss, whereas the average length of the conditioning phase of the first session in my experiment was 29 minutes. So at least for three quarters of Trowill's Ss the length of the training period was close to that of the conditioning phase of the first session used in the present experiment. The most important difference between the two experiments is that in my experiment the group to be reinforced for increases in heart rate had a much higher mean heart rate during adaptation (456 b.p.m.), than was the case in Trowill's experiment. His increase group had a pretraining mean heart rate of 401 b.p.m. Wilder's (1956) Law of Initial Values states that the magnitude of the response to an experimental stimulus or condition is related to the pre-stimulus level of the response system. Since there was a large difference in the pre-conditioning heart rates of the two increase groups used in these experiments this law provides an explanation of the difference in the effectiveness of the conditioning procedures in producing increases in heart rate.

The results of the present experiment also contrast with those of Miller and DiCara (1967). Miller and DiCara were able to condition both large increases and decreases in heart rate in two groups of curarized rats reinforced with brain stimulation for changes in heart rate. The result of this differential effect was a large difference of 194 b.p.m. in the mean heart rates of their two groups at the end of one curare conditioning session.

One possible explanation of the difference in the results of the two experiments is that Miller and DiCara used a ninety minute training period, whereas in the present experiment the conditioning phase of the first curare session only lasted for approximately thirty minutes. However even after thirty minutes Miller and DiCara were able to condition a difference of approximately 100 b.p.m.² in the mean heart rates of their two groups. Although it seems possible that if the conditioning phase of the first curare session had been increased in length in the present experiment a larger difference in the heart rates of the two groups would have been conditioned, since similar procedures were used in the two experiments, an explanation of the clear difference in their results must be sought.

At the beginning of conditioning in the Miller and DiCara experiment, there was a difference of 22 b.p.m. in the mean heart rates of their two groups, with the group which was to be reinforced for

 $^{^{2}}$ This difference has been taken from Figure 1 on page 14 in Miller and DiCara (1967).

increases in heart rate showing the higher average heart rate; in the present experiment, on the other hand, there was a difference of 34 b.p.m. in the mean heart rates of the two groups at the beginning of conditioning, with the group which was to be reinforced for increases in heart rate showing the lower average heart rate. Since rats were randomly assigned to the experimental groups in both experiments this difference was due to chance. In view of this difference in the initial heart rates of the groups used in these experiments it seems that an assessment of the conditioning effects in terms of the changes from these initial levels which were conditioned, is more satisfactory than a comparison of the four groups in terms of heart rates at the end of conditioning. Table 3 shows the changes in heart rate which were actually conditioned in the two experiments. Table 3 clearly shows how much more successful Miller and DiCara were in conditioning changes in heart rate than was the present experimenter. The results reported by Miller and DiCara represent a dramatic demonstration of operant conditioning of both increases and decreases in heart rate in curarized rats with brain stimulation reinforcement. Although there was some evidence of operant conditioning of a decrease in heart rate during the first curare session in the present experiment there was no evidence of operant conditioning of an increase in heart rate. I have not been able to find a satisfactory explanation of the large difference in the results of these two experiments in which similar procedures were used.

A similar position obtains with respect to the results of experiments in our laboratory on avoidance conditioning of changes in

	brain stimulation reinforcement					
Experiment	Group	H.R. at the beginning of conditioning	H.R. after 30 mins. of conditioning	Change in Heart rate		
Miller and DiCara	Increase	422 b.p.m.	460 b.p.m.	(460 - 422) = 38 b.p.m.		
(1967)	Decrease	400 b.p.m.	360 b.p.m.	(400 - 360) = 40 b.p.m.		
Present Experiment	Increase	449 b.p.m.	449 b.p.m.	(449 - 449) = 0 b.p.m.		
	Decrease	480 b.p.m.	456 b.p.m.	(480 - 456) + 24 b.p.m.		

Table 3

A comparison of the conditioned changes in heart rate in two experiments using curarized rats and

heart rate.³ In these experiments much smaller heart rate changes were conditioned than was the case in an experiment reported by DiCara and Miller (1968) in which changes in heart rate were conditioned using an avoidance conditioning procedure. Why Miller and his co-workers have been consistently more successful in operantly conditioning changes in heart rate than we have been is, at the present time, not clear.

Tables 2 (p. 51), 4 and 5, and Figures 2 (p. 52) and 3 show that on the second, third and fourth curare conditioning sessions of the present experiment, there were clear differences in the heart rates of the two groups, with the group reinforced for increases in heart rate showing a higher heart rate than did the group reinforced for heart rate decreases. Figures 2 and 3 clearly show the difference between the results for the three later sessions and the first session. Although there was at best little evidence of operant conditioning of changes in heart rate during the first curare session, there was clear evidence of such a result during the later sessions. Table 6 presents the changes in heart rate which were conditioned in the two experimental groups during the later curare conditioning sessions. Table 6 shows that during the later curare sessions the group reinforced for increases in heart rate showed a clear heart rate increase, and the group reinforced for decreases in heart rate showed a clear heart rate decrease. Figures 2 and 3 show that the difference in the heart rates of the two groups tended to increase during conditioning on the later curare sessions.

³These unpublished experiments were performed during 1967 by Brener and Goesling.

Mean heart rates in beats per minute during four blocks of adaptation, conditioning and extinction periods

				-					in and set			
					Se	ssior	13					
Rat No. A	dap.	А	А	A	Cond.	С	С	С	Ext.	Ε	E	Ε
		Grou	ıp l:	Rein	forced	for	heart	rate	incre	ases		
6 B 5 7 C 5 10 C 4 4 D 4	01 06 79 72	491 465 504 450 443 371	488 464 480 435 453 377	487 463 470 428 455 376	482 484 480 411 471 377	490 499 481 428 481 386	495 513 496 464 488 401	497 537 506 473 480 413	500 519 507 477 474 408	501 486 506 467 454 402	500 473 498 456 467 393	495 469 488 452 456 384
Totals 28	23 2	2724	2697	2679	2705	2765	2857	2906	2885	2816	2787	2744
Means 4	71	454	449	446	451	461	476	484	481	469	464	457
		Grou	ıp 2:	Rein	forced	for	heart	rate	decre	ases		
6 C 4 8 C 4 9 C 5 9 D 4	48 81 29 32	478 459 451 528 429 447	472 454 420 529 427 458	464 458 412 524 426 449	418 452 408 508 415 449	423 438 409 503 403 435	418 443 407 499 398 426	404 434 390 508 394 416	385 440 413 504 389 414	379 449 415 500 392 420	375 455 431 508 392 435	383 468 432 521 389 456
Totals 28	38 2	2792	2760	2733	2650	2611	2591	2546	2545	2555	2596	2649
Means 4	73	465	460	455	442	435	432	424	424	426	433	441

Mean heart rates in beats per minute during four blocks of adaptation, conditioning and extinction periods

					S	Sessi	on 4						
Rat No.	. Adaj	р. А	A	A	Cond	. C	С	С	Ext	• E	Ε	Ε	
		Groi	up 1:	Reir	forced	for	heart	. rate	incre	ases			
2 C 6 B 7 C 10 C 4 D 5 D	482 477 415 470 456 411	473 473 437 463 449 397	440 466 462 469 433 403	463 466 469 468	465 477 463 465 410 413	472 487 472 469 417 416	481 476 476 473 428 416	479 485 484 482 434 415	474 458 473 485 426 402	472 439 472 489 417 390	466 443 468 477 417 393	460 437 462 469 423 393	
Totals	2711	2692	2673	2703	2693	2733	2750	2779	2718	2679	2664	2644	
Means	450	449	445	450	449	456	458	463	453	447	444	441	
		Grou	up 2:	Rein	forced	l for	heart	: rate	decre	eases			
1 B 6 C 8 C 9 C 9 D 10 D	518 469 420 519 369 474	515 481 415 515 358 476	509 493 410 515 344 475	493 493 414 511 331 476	479 467 395 503 323 464	451 486 387 494 323 458	419 487 379 483 318 458	420 484 374 481 313 450	466 506 381 485 316 452	472 510 388 490 312 450	476 501 386 491 308 456	474 501 386 497 307 461	
Totals	2769	2760	2746	2718	2631	2599	2544	2522	2606	2622	2618	2626	
Means	461	460	458	453	438	433	424	420	434	439	436	438	

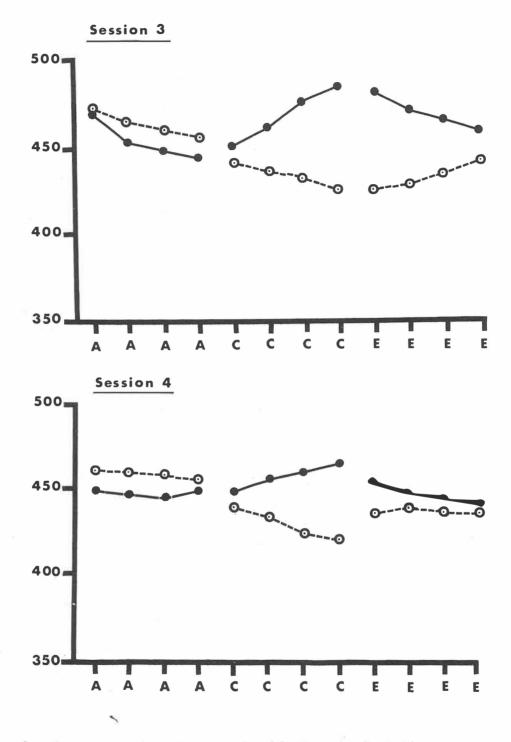


Figure 3. Group mean heart rates for blocks of adaptation, conditioning and extinction periods during the third and fourth curare sessions.

Group mean changes in heart rate conditioned during the second, third, and fourth curare sessions

ī

Session	Group	Mean heart rate for last block of adaptation periods	Mean heart rate for last block of conditioning periods	Change in heart rate (Cond. mean- Adap. mean)
	l			
2	H.R. Incr.	443 b.p.m.	465 b.p.m.	+ 22 b.p.m.
	2			
	H.R. Decr.	423 b.p.m.	396 b.p.m.	- 27 b.p.m.
3	l H.R. Incr.	446 b.p.m.	484 b.p.m.	+ 38 b.p.m.
5	2			
	H.R. Decr.	455 b.p.m.	424 b.p.m.	- 31 b.p.m.
	l			
4	H.R. Incr.	450 b.p.m.	463 b.p.m.	+ 13 b.p.m,
T	2			
	H.R. Decr.	453 b.p.m.	420 b.p.m.	- 33 b.p.m.

1.01

In the statistical analysis of the results of this experiment differences were computed between each rat's heart rate during the last block of adaptation periods and the last block of conditioning periods for each session. The results of an analysis of variance performed on this data are shown in Table 7. The treatments effect was found to be highly significant, F (1, 10) = 47.86, p < .001, indicating that in terms of this measure the two experimental groups showed significantly different changes in heart rate. The sessions effect was found to be non-significant, F (3, 30) = 2.89, p > .05, but the interaction between treatments and sessions was found to be significant, F (3, 30) = 5.14, p < .01. This significant interaction appears to have been due to the fact that the group reinforced for heart rate increases only showed clear evidence of operant conditioning of an increase in heart rate during the later curare conditioning sessions.

Figures 2 (p. 52) and 3 (p. 61) show evidence of an extinction effect in the results of this experiment in that during the second, third and fourth curare sessions the curves of the two experimental groups which tended to diverge during conditioning converged during extinction. In order to illustrate this extinction effect differences were computed for each session between the mean heart rates of the two groups during the final block of conditioning periods, and the final block of extinction periods. Table 8 shows these differences. Table 8 shows that although there were differences on the second, third and fourth curare sessions between the mean heart rates of the two groups during the last block of conditioning

betwee	en the	last	block	of	adaj	ptatic	on and	the	last	block	of	
(condit	ioning	g perio	ods	for	four	curare	e ses	ssions	3		
							1					

Summary of the Analysis of Variance on the difference in heart rate

Source	df	Sum of squares	Mean Square	F	Significance level (p)
Between <u>S</u> s	11	31,645	. <u>-</u> .	-	-
Treatments (A)	l	26,180	26,180	47.86	p < .001
Ss within groups	10	5,465	546.5	-	-
Error A					
Within Ss	36	11,146	: 8	-	-
Sessions (B)	3	1,784	594.7	2.89	ns
АхВ	3	3,180	1,060	5.14	p < .01
B x <u>S</u> s within	30	6,182	206.1	-	-
groups. Error B					

Table 7

Differences between the group mean heart rates for the last block of conditioning periods and the last block of extinction periods

Difference between the (449-456) (465-396) (484-424) (463-420)group means for the = -7 b.p.m. = +69 b.p.m. = +60 b.p.m. = +43 b.p.m. ditioning periods. Difference between the (442-459) (446-415) (457-441) (441-438)group means for the =-17 b.p.m. = +31 b.p.m. =+16 b.p.m. = +3 b.p.m. last block of extinction periods. periods, these differences were not maintained in extinction. The difference in the mean heart rates of the two groups became progressively smaller over the last three curare sessions. So in this experiment a heart rate change in the direction being conditioned occurred while reinforcement was being delivered, but the changed heart rate was not sustained once the reinforcement was no longer contingent on the change in heart rate. This result is important since it shows that one other phenomenon of operant conditioning can be demonstrated when the operant response is a heart rate change.

The results of this experiment show that both increases and decreases in heart rate can be conditioned when brain stimulation is used as the reinforcement for the heart rate change. Since these heart rate changes were conditioned in completely paralyzed rats the possibility that the heart rate changes were mediated by some type of skeletal response has been at least markedly reduced.⁴ The results of this experiment extend those reported by Trowill (1967) and Miller and DiCara (1967) in showing the importance of extent of conditioning in experiments on operant conditioning of changes in heart rate, and they also provide evidence of an extinction effect.

Since the results of this first experiment provided a clear demonstration that both increases and decreases in heart rate can be

⁴The implications of these results will be discussed in detail in Chapter V of this dissertation.

conditioned when brain stimulation is used as the reinforcer with curarized rats, it was decided to perform a second experiment in which an attempt was made to condition both increases and decreases in heart rate in individual rats. Since this experiment was a pilot study its procedures and results will be presented and discussed in Appendix B of this dissertation.

CHAPTER V

SUMMARY AND CONCLUSIONS

The aim of the first experiment reported in this dissertation was to investigate the possibility that both increases and decreases in heart rate can be conditioned in curarized rats when brain stimulation is used as the reinforcement for the heart rate changes. Curarized rats were used in an attempt to reduce the possibility of skeletal mediation of the conditioned heart rate changes. Brain stimulation was used as the reinforcer because such stimulation has been found to yield powerful reinforcing effects in many experiments in which more typical operant responses have been used. This stimulation could also be made contingent on the heart rate changes in a reliable and controlled manner. Two groups of six rats each were used in this experiment one of which was reinforced during the four curare conditioning sessions for increases in heart rate, and one for heart rate decreases. Little if any evidence of operant conditioning of either an increase or a decrease in heart rate was found in these groups during the first curare conditioning session. This result contrasts with the reports of Trowill (1967) and Miller and DiCara (1967). A number of possible explanations of the difference in the results of these experiments were considered.

During the second, third and fourth curare conditioning sessions clear differences in heart rate were found between the two groups, with the group reinforced for heart rate increases showing a clear increase in heart rate during conditioning, and the group reinforced for heart

rate decreases showing a clear decrease in heart rate during conditioning. An analysis of variance showed a statistically significant difference between the two groups during the later curare conditioning sessions, and a statistically significant interaction between treatments and sessions. The results for the last three curare sessions confirm and extend the findings of Trowill (1967) and Miller and DiCara (1967) and clearly show that both increases and decreases in heart rate can be conditioned in curarized rats when brain stimulation is used as the reinforcement for the heart rate changes. In addition there was some evidence of an extinction effect during the later curare sessions in that the heart rates of the two groups which tended to diverge during the conditioning phase of the session tended to converge during the extinction phase.

Since the results of this first experiment provided a clear demonstration of operant conditioning of both heart rate increases and heart rate decreases in different groups an attempt was made to condition both increases and decreases in heart rate in the same rat. Two different procedures for this reversal heart rate conditioning were used with three rats. All three rats showed good acquisition of the initial heart rate change, thus replicating the results of the first experiment, but no evidence of acquisition of the heart rate reversal. Since \underline{Ss} differ considerably in their cardiac behavior and in their reactions to stimuli a demonstration of operant conditioning of both increases and decreases in heart rate in the same \underline{S} would represent a convincing

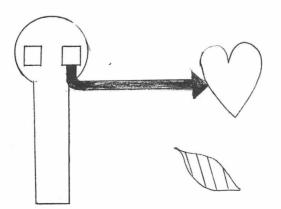
demonstration of the phenomenon of operant conditioning of changes in heart rate. However such a demonstration is <u>not</u> provided by the results for the three rats given a series of reversal conditioning trials. It must be remembered however that many investigators have reported failures to establish reversal conditioning with more conventional responses, and so difficulty in establishing heart rate reversals may be due to the nature of the reversal conditioning procedure, rather than due to the characteristics of heart rate as an operant response.

Since one of the most important features of this research was the use of curarized rats as experimental $\underline{S}s$, I would like to end this dissertation with some notes on the limitations inherent in the use of such $\underline{S}s$. It must be pointed out that when these experiments were planned I was not aware of a number of these limitations, and I only became aware of them as a result of my experience in conducting this research.

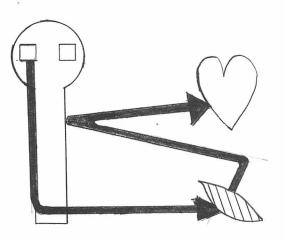
The most obvious starting point for research on conditioning of changes in heart rate is the question originally posed by Shearn (1961) 'Does the heart learn?' Since operant conditioning procedures have been found to lead to the modification and control of a variety of responses, one widely used procedure has been to make some type of reinforcement contingent on a change in heart rate, in a typical operant conditioning paradigm:

Response_(Heart rate change) Reinforcement Under such conditions let us assume that a clear increase in the frequency of occurrence of the heart rate change is observed. It might be concluded

that such results demonstrate that changes in heart rate can be operantly conditioned, and that a clear and unequivocal affirmative answer can be given to the question 'Does the heart learn?' However such a conclusion would certainly be challenged by proponents of the mediation or artifact hypothesis who would argue that in such an experiment some type of skeletal response could actually have been conditioned, and that it was these skeletal changes which mediated the heart rate changes which were observed. It might be further argued, as Smith (1954) suggested, that only skeletal responses can be conditioned and that 'conditioned' heart rate responses are no more than artifacts elicited by feedback from the skeletal responses. These different positions can be made clearer with reference to a series of models which were independently proposed by Black (1967) and Brener (1967). The claim that heart rate changes can be conditioned using an operant conditioning procedure is presumably based on the assumption that activity in some neural center directly controls heart rate and produces the heart rate change which has been conditioned.



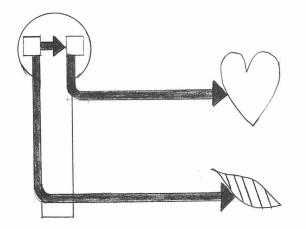
A mediation theorist however would challenge this type of explanation of the results and would propose that neural centers controlling some type of skeletal response had been activated, and that feedback from these skeletal responses produced the heart rate change. The following diagram illustrates this type of argument:



This model shows that if for example respiratory and heart rate changes were conditioned simultaneously, then the heart rate changes which were observed could have been the result of feedback from the respiratory changes.

Curarized rats were used in the experiments reported in this dissertation in an attempt to reduce, if not completely eliminate, the possibility of skeletal changes occurring during conditioning, and so to study operant conditioning of heart rate changes uncontaminated

by skeletal changes. With reference to the above model curarization might be regarded as a procedure which breaks the link between the neural control center and the skeletal response. If this model is accepted curarization is a useful and valuable procedure to adopt in experiments on operant conditioning of autonomic responses. However a third model might be proposed. This model is shown in the following diagram:



According to this model in order for a heart rate change to be mediated by a skeletal response it is not necessary for the skeletal response to actually occur, but only for what Black (1967) has termed the 'central movement process' underlying the skeletal response to occur. So even in a completely paralyzed \underline{S} , where there is no possibility of skeletal movement, it is still possible for a heart rate response to be mediated by a 'central movement process'. Black (1967) writes: When an operantly conditioned heart rate response occurs the result of the conditioning process is to activate neural centers controlling movements, and to produce a change in heart rate (Black, 1967, p. 10).

Smith (1967) has proposed a similar position:

Thus, it (Smith's 1954 paper 'Conditioning as an Artifact') insists that the somatic response and the autonomic response must be pictured, in the general sense, as arising coordinately from a common source (Smith, 1967, p. 101).

Obrist and Webb (1967) have adopted a similar position. They state:

However, these studies (studies in which curare has been used) also appear to be inconclusive because the technique evaluates only the extent to which cardiac events are influenced by the actual occurrence of peripheral somatic-motor events. What is not evaluated is the possibility that both responses have a common origin and means of control with the central nervous system . . .In this case, neither response 'causes' the other; rather they might be regarded as concomitants, or as simply different aspects of the same response process (Brackets mine. Obrist and Webb, 1967, p. 8).

Curare blocks only the peripheral manifestations of the 'central movement processes' or activity in some neural 'common source', but has no effect on the processes or neural sources themselves. So results from curarized <u>Ss</u> have little relevance to this type of mediation argument. Until suggestions such as those of Black and Smith are made more specific, for example what exactly is a 'central movement process', and what is the specific anatomical location of the 'common source', it is difficult to see how this mediation argument can ever be either supported or refuted. The study of mediation becomes extremely difficult when central processes are proposed as the basis of the mediation.

In the history of Psychology there are a number of examples of the misdirection of research due to the posing of questions which lack any possibility of empirical answer. The history of such questions as:

Is intelligence the result of heredity or environment?

and

What is learned?

shows that initially such questions appear simple and direct, but as more and more 'crucial' experiments designed to answer such questions are performed, the unanswerable nature of the question becomes apparent. After all of the research which has been done on the development of intelligence who, for example, would be prepared to state that intelligence is solely the result of heredity or environment? I suspect that the question, Can the heart be conditioned in the absence of skeletal responding?, is of a similar genre. Originally the use of curarized Ss seemed to offer a way of answering this question but difficulties of interpretation and analysis of the experimental results soon became apparent. In addition it is felt that too great a concern with problems of mediation may be unwise. The heart normally interacts with a number of other systems and so it must be remembered that the curarized S is a highly abnormal preparation, and so the generality of the results which are obtained using curarized Ss may not be very great. In the future it would seem more profitable, rather than continuing a seemingly never ending attempt to eliminate or reduce possible mediating factors, to direct research to the development of procedures which would facilitate the development of conditioned cardiac responses. A second promising approach would be to record skeletal concomitants - for example, respiration - of cardiac conditioning. This respiration data

could then be used in transfer equations similar to those published by Clynes (1960) which allow the prediction of heart rate from a knowledge of respiration. The difference between the heart rate predicted on the basis of the respiration change, and the actual change in heart rate, would allow an assessment of the effectiveness of the operant conditioning procedure.

In conclusion it is felt that curarization is a useful tool, and that the experiments reported in this dissertation represent a successful use of this procedure. But it must be remembered that it is only a tool and is not an end in itself. BIBLIOGRAPHY

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APPENDIXES

APPENDIX A

Tables I to VIII showing heart rates for the two experimental groups during successive one minute periods of adaptation, conditioning, and extinction for the four curare conditioning sessions. Criterion changes and number of brain stimulation reinforcements during conditioning periods.

		Group 1		te increases	reinforced	
			Ses	sion l		
lat	Number 2 C	6 в	7 C	10 C	4 D	5 D
	2 0	6.0		10 0	4 D	
	491	442	505	528	492	394
	491	442	505	531	490 484	394
	481 481	441 441	505 504	525 523	483	394 394
	487	439	501	513	489	389
	489	439	500	494	487	389
	489	430	497	496	474	389
	488	430	497	484	482	393
	486	417	491	460	482	393
	488 487	417 428	490 489	463 462	468 452	393 389
	487	428	489	402	454	388
	488	427	486	479	462	383
	488	430	480	484	457	379
	486	429	483	474	455	378
	486	427	482	478	467	380
	488 488	427 426	480 481	484 491	480 475	380 377
	488	426	478	489	473	381
	488	415	473	487	472	379
	487	411	471	.67 480 0	475	381
	487	408	468	.68 466 0	471	377
	487 487	405 411	472 473	.69 460 0 .70 466 3	471 471	378 378
	487	411	472	.69 485 6	471 471	378
	488 0	.74 417 0	.71 480 1	.68 486 6	466	378
	486 2	424 l	480 l	.67 491 6	465	371
	486 2	426 2	481 1	.65 496 4	466	358
	486 2 486 2	431 3 430 2	481 1 482 1	496 4 496 5	462 467	359 361
	487 1	429 5	482 l	.64 502 0	.62 475 0	.87 354 0
	484 3	437 6	481 1	501 0	473 0	.88 348 0
	482 2	432 5	483 l	.65 494 2	.63 467 0	.89 356 1
	480 3	432 l	483 l	.66 494 5	.64 453 0	355 l

Heart rate during successive one minute periods of adaptation, conditioning and extinction. Criterion changes and number of brain stimulation reinforcements during conditioning periods

TABLE I (continued)

Rat Number 2 C	6 в	7 0		4 D	5 D
$\begin{array}{c} 2 \\ 480 \\ 3 \\ 480 \\ 3 \\ 478 \\ 5 \\ 478 \\ 5 \\ 473 \\ 6 \\ 473 \\ 6 \\ 473 \\ 6 \\ 473 \\ 472 \\ 0 \\ 473 \\ 4 \\ 459 \\ 455 \\ 459 \\ 450$	$\begin{array}{c} 4 & 27 & 1 \\ 4 & 28 & 2 \\ 4 & 31 & 3 \\ 4 & 35 & 5 \\ 4 & 30 & 1 \\ 7 & 4 & 23 & 1 \\ 4 & 32 & 5 \\ 7 & 3 & 4 & 35 & 4 \\ 4 & 4 & 2 & 2 \\ 4 & 4 & 2 & 3 \\ 4 & 4 & 2 & 3 \\ 4 & 1 & 1 \\ 4 & 1 & 4 \\ 4 & 1 & 1 \\ 4 & 1 & 1 \\ 4 & 1 & 1 \\ 4 & 1 & 1 \\ 4 & 1 & 1 \\ 4 & 1 & 1 \\ 4 & 1 & 1 \\ 4 & 1 & 1 \\ 4 & 1 & 1 \\ 4 & 1 & 1 \\ 4 & 1 & 1 \\ 4 & 1 & 1 \\ 4 & 1 & 1 \\ 4 & 1 & 1 \\ 4 & 1 & 1 \\ 4 & 1 & 1 \\ 4 & 1 & 1 \\ 4 & 1 & 1 \\ 4 & 1 &$	$\begin{array}{c} 7 \\ 68 \\ 482 \\ 483 \\ 483 \\ 483 \\ 483 \\ 483 \\ 485 \\ 487 \\ 488 \\ 489 \\ 487 \\ 488 \\ 489 \\ 487 \\ 488 \\ 487 \\ 488 \\ 487 \\ 488 \\ 487 \\ 488 \\ 487 \\ 488 \\ 487 \\ 488 \\ 487 \\ 488 \\ 487 \\ 488 \\ 487 \\ 493 \\ 493 \\ 493 \\ 493 \\ 493 \\ 491 \\ 493 \\ 491 \\ 493 \\ 491 \\ 495 \\ 502 \\ 495 \\ 495 \\ 495 \\ 495 \\ 495 \\ 495 \\ 495 \\ 495 \\ 495 \\ 495 \\ 495 \\ 495 \\ 495 \\ 495 \\ 495 \\ 495 \\ 495 \\ 495 \\ 495 \\ 496 \\ 496 \\ 49$	$\begin{array}{c} 10 \ C \\ 496 \ 2 \\ 496 \ 2 \\ 495 \ 2 \\ 66 \ 482 \ 0 \\ 67 \ 482 \ 3 \\ 66 \ 484 \ 3 \\ 484 \ 1 \\ 459 \\ 457 \\ 489 \\ 494 \\ 495 \\ 497 \\ 493 \\ 488 \\ 486 \\ 487 \\ 491 \\ 495 \\ 497 \\ 497 \\ 497 \\ 497 \\ 497 \\ 497 \\ 497 \\ 495 \\ 494 \\ 494 \\ 494 \\ 494 \\ 493 \\ 490 \\ 490 \\ \end{array}$	$\begin{array}{c} 4 \\ 4 \\ 4 \\ 4 \\ 5 \\ 4 \\ 5 \\ 2 \\ 4 \\ 5 \\ 2 \\ 4 \\ 5 \\ 2 \\ 4 \\ 5 \\ 2 \\ 4 \\ 5 \\ 5$	5 D .90 343 4 .91 342 4 352 4 350 3 353 5 354 5 345 1 344 1 341 0 .92 341 0 .92 341 0 .92 341 0 .93 339 1 343 3 351 6 354 6 354 6 354 6 354 3 351 1 .94 351 6 .93 361 1 .94 351 6 .93 361 1 .94 351 6 .93 361 1 .94 356 4 357 2 373 370 366 354 3 357 2 373 370 366 354 3 359 351 349 348 350 349 353 366 352 349 349 353 366 352 349

Rat Number <u> </u>	6 в	7 C	10 C	4 D	5 D
462 458 457 455	415 412 417 421 423 423 409 407 405 405	488 <u>487</u>		442 439 442 447 450 450 452 460 458	345 347 342 339 347 349 344 338 340 341 338

TABLE I (continued)

Heart rate during successive one minute periods of adaptation, conditioning and extinction. Criterion changes and number of brain stimulation reinforcements during conditioning periods

TABLE II

	Group :		te decreases	reinforced	
Rat Numb		Ses	sion l		
l B	6 C	8 C	9 C	9 D	10 D
			48		
540	479	311	567	455	480
540	481	318	563	457	478
539	481	329	560	450	481
535	485	330	562	444	484
536	485	333	561	449	484
532	485	339	561	451	484
533	482	341	562	447	482
533	482	348	561	449	482
533	477	351	555	452	478
529	477	353	551	450	477
533	475	387	550	442	478
533	475	392	559	444	472
531	479	404	559	446	479
533	482	409	554	444	481
529	482	414	554	442	482
531	483	419	545	446	483
531	483	418	537	441	482
527	484	420	549	437	477
529	484	423	526	441	479
527	481	422	538	439	478
.61 512		.95 402 0	536	438	476
512		,90 381 0	536	441	475
510		.85 379 1	529	442	473
508		382 2	533	447	475
508		386 4	533	440	479
510		386 2	538	437	481
510 ¹		386 2	536	439	482
510		387 3	529	438	484
507		386 5	.60 526 2	435	480
508		.86 380 2	520 2	437	477
.64 506 (383 2	520 4	.70 444 6	.69 480 0
500 2		380 3	512 4	442 6	.68 478 3
510		382 3	517 5	.71 448 4	472 6
.63 496 1	458 5	.87 378 1	519 0	447 5	.69 478 0

TABLE II (continued)

Rat Number 1 B	6 C	8 C	9 C	9 D	10 D
502 2 498 4 498 4 498 3 500 2 500 4 501 3 501 3 497 491 487 485 485 485 485 484 479 476 471 476 477 478 476 477 478 476 473 469 466 478 498 498 494	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{c} 520 & 0 \\ 518 & 1 \\ 519 & 0 \\ 523 & 0 \\ 521 & 0 \\ 518 & 1 \\ 508 & 0 \\ 5225 & 0 \\ 528 & 0 \\ 532 & 532 \\ 5332 & 5332 \\ 5332 & 5332 \\ 5336 & 5338 \\ 538 & 539 \\ 540 & 540 \\ 540 & 540 \\ 54$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{c} 475 \\ 469 \\ 469 \\ 2 \\ 69 \\ 469 \\ 2 \\ 69 \\ 470 \\ 3 \\ 70 \\ 465 \\ 4 \\ 463 \\ 3 \\ 455 \\ 6 \\ 71 \\ 465 \\ 4 \\ 457 \\ 3 \\ 462 \\ 1 \\ 458 \\ 5 \\ 71 \\ 455 \\ 2 \\ 459 \\ 459 \\ 459 \\ 461 \\ 458 \\ 457 \\ 459 \\ 459 \\ 458 \\ 457 \\ 455 \\ 457 \\ 459 \\ 460 \\ 460 \\ 460 \\ 460 \end{array}$

Rat Number 1 B	6 C	8 C	9 C	9 D	10 D
482	461 457 455 452 453 451 452		540	426 428 429 427 425 427 431 432 432 428 428 428 428 427 428 430 432 432 430	461 462 462 463 467 472 474 473 471 469 471 469 473 469 466 470

TABLE II (continued)

Heart	rate	during	succes	sive	one	minute	period	s of	adaptat	ion,
	condi	tioning	g and e	xtind	ction	n. Cri	terion	chang	ges and	
	r	number d	of brain	n sti	imula	ation r	einforc	emen	ts	
			during	cond	litic	oning p	eriods			

	Group 1:		e increases ion 2	reinforced	
Rat Number 2 C	бв	7 C	10 C	4 D	5 D
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	465 469 4 9 477 3 8 483 5 7 488 2 6 481 0 7 491 3 6 493 3 494 2	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	526 521 518 516 510 479 489 494 486 475 476 476 477 461 467 476 476 476 472 462 468 471 476 477 2 476 6 479 1 479 3	485 486 480 473 463 461 458 450 455 455 455 455 455 455 455 455 455	382 387 389 379 376 376 376 378 382 375 375 375 375 379 376 381 380 374 376 373 372 372 365 363 363 363 363 363 363 363

TABLE III (continued)

Rat Number 2 C	бВ	7 C	10 C	4 D	5 D
$\begin{array}{c} 484 & 0 \\ 488 & 3 \\ 488 & 3 \\ 488 & 3 \\ 488 & 2 \\ 487 & 2 \\ 487 & 2 \\ 487 & 2 \\ 487 & 2 \\ 487 & 2 \\ 487 & 2 \\ 487 & 2 \\ 487 & 2 \\ 487 & 2 \\ 487 & 2 \\ 487 & 2 \\ 487 & 3 \\ 3 \\ 1 \\ 480 & 4 \\ 4$	500 0 498 1 497 0 499 5 486 5 487 5 487 5 487 5 487 5 487 5 489 4 493 2 493 2 65 499 5 66 499 5 66 499 5 66 500 4 68 475 0 66 500 4 68 475 0 68 494 5 486 5 486 5 486 484 483 481 480 475 478 475 478 479 480 476 478 478 479 480 478	.79 423 5 426 5 .76 427 4 423 5 430 3 431 3 430 1 434 1 433 2 435 5 .75 436 2 441 4 438 2 439 4 433 442 437 438 437 438 437 438 437 438 437 445 445 445 445 445 445 445 441 445 445 445 441 445 445 445 441 445 445 445 441 445 445 445 441 445 445 445 445 442 445 455	$\begin{array}{c} 481 & 3 \\ 481 & 3 \\ 481 & 3 \\ 483 & 4 \\ 482 & 2 \\ 479 & 2 \\ 481 & 3 \\ 484 & 2 \\ 466 & 3 \\ 484 & 2 \\ 466 & 3 \\ 488 & 3 \\ 486 & 3 \\ 486 & 3 \\ 486 & 3 \\ 486 & 3 \\ 485 & 3 \\ 486 & 3 \\ 485 & 3 \\ 485 & 3 \\ 485 & 3 \\ 485 & 3 \\ 486 & 4 \\ 486 & 3 \\ 485 & 3 \\ 485 & 3 \\ 485 & 3 \\ 486 & 4 \\ 486 & 3 \\ 485 & 3 \\ 486 & 4 \\ 486 & 3 \\ 485 & 3 \\ 486 & 4 \\ 486 & 3 \\ 485 & 3 \\ 486 & 4 \\ 486 & 3 \\ 485 & 3 \\ 486 & 4 \\ 486 & 3 \\ 485 & 3 \\ 486 & 4 \\ 486 & 3 \\ 485 & 3 \\ 486 & 4 \\ 486 & 3 \\ 485 & 3 \\ 486 & 4 \\ 486 & 3 \\ 485 & 3 \\ 486 & 4 \\ 486 & 3 \\ 485 & 3 \\ 485 & 3 \\ 485 & 3 \\ 486 & 4 \\ 486 & 3 \\ 485 & 3 \\ 485 & 3 \\ 485 & 3 \\ 486 & 4 \\ 486 & 3 \\ 485 & 3 \\ 485 & 3 \\ 486 & 4 \\ 486 & 3 \\ 485 & 3 \\ 485 & 3 \\ 485 & 3 \\ 485 & 3 \\ 486 & 4 \\ 486 & 3 \\ 485 & 3 \\ 485 & 3 \\ 485 & 3 \\ 485 & 3 \\ 486 & 4 \\ 486 & 3 \\ 485 & 3 \\$	$\begin{array}{c} 466 & 2 \\ 464 & 2 \\ 460 & 0 \\ 468 & 4 \\ 469 & 3 \\ 483 & 5 \\ 481 & 5 \\ 478 & 1 \\ 476 & 2 \\ 486 & 5 \\ 491 & 6 \\ 492 & 6 \\ 488 & 0 \\ 488 & 0 \\ 488 & 4 \\ 486 & 2 \\ 488 & 4 \\$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$

Rat Number 2 C	6 в	7 C	10 C	ЦD	5 D
471 470 470 471 470	480 478 476 472 470 468 464 461		434 433 431 432 432 432 432	482 478 473 486 477 475 480 478 475 476 482 472 473 470 477	387 382 383 383 381 378 380 379 379 379 379 378 383 384 383 384 382 375 378 378 381

TABLE III (continued)

TABLE IV

Heart rate during successive one minute periods of adaptation, conditioning and extinction. Criterion changes and number of brain stimulation reinforcements during conditioning periods

		Group 2:		ce decreases	reinforced	
Rat	Number 1 B	6 C	8 C	sion 2 9 C	9 D	l0 D
.71 .70	$\begin{array}{c} 1 \\ 490 \\ 478 \\ 468 \\ 466 \\ 463 \\ 466 \\ 466 \\ 466 \\ 466 \\ 466 \\ 466 \\ 466 \\ 466 \\ 466 \\ 437 \\ 439 \\ 4429 \\ 445 \\ 455 \\ 555 \\ 447 \\ 456 \\ 1 \\ 1 \\ 1 \\ 243 \\ 432 \\ 22 \\ 26 \\ 432 \\ 432 \\ 232 \\ 26 \\ 6 \\ 32 \\ 432 \\ 20 \\ 6 \\ 32 \\ 20 \\ 20 \\ 20 \\ 20 \\ 20 \\ 20 \\ 20$	$\begin{array}{c} 402 \\ 412 \\ 413 \\ 413 \\ 413 \\ 413 \\ 413 \\ 413 \\ 415 \\ 414 \\ 414 \\ 413 \\ 415 \\ 414 \\ 413 \\ 415 \\ 413 \\ 415 \\ 413 \\ 415 \\ 413 \\ 415 \\ 413 \\ 410 \\ 409 \\ 409 \\ 409 \\ 409 \\ 408 \\ 410 \\ 410 \\ 410 \\ 410 \\ 410 \\ 412 \\ 411 \\ .83 \\ 411 \\ 1 \\ 409 \\ 2 \\ .84 \\ 404 \\ 4 \\ .85 \\ 402 \\ 0 \\ .83 \\ 403 \\ 398 \\ 2 \\ .82 \\ 403 \\ 1 \\ 400 \\ 5 \\ 398 \\ 5 \\ .83 \\ 399 \\ 0 \\ 402 \\ 3 \\ 396 \\ 5 \\ .83 \\ .81$	395 399 393 387 383 385 385 385 382 384 382 380 384 382 382 382 382 382 382 382 382 381 377 381 377 381 377 381 377 381 377 381 377 381 377 381 377 381 377 381 377 381 377 381 377 381 377 379 .90 367 1 365 2 365 2 365 2 365 4 356 4 .87 356 4 .87 354 3 354 3 354 3 355 4 .86 356 5	$\begin{array}{c} 536\\ 534\\ 535\\ 534\\ 533\\ 532\\ 531\\ 527\\ 525\\ 526\\ 525\\ 526\\ 525\\ 523\\ 522\\ 521\\ 522\\ 521\\ 522\\ 521\\ 522\\ 519\\ 519\\ 519\\ 519\\ 519\\ 519\\ 519\\ 519$	463 457 457 458 455 448 447 445 451 445 451 446 437 437 431 427 431 427 431 427 431 427 431 427 431 427 431 427 431 427 431 409 412 409 412 409 412 409 412 409 412 409 412 409 412 409 403 400 399 400 386 350 350 346 348 347 .89 352 0 .90 364 0 363 2 .91 359 5	440 428 420 425 431 417 413 410 408 404 408 404 405 408 404 405 402 412 412 412 412 412 412 412 412 412 41

TABLE IV (continued)

Rat	Number 1 B	6 C	8 C	9 C	9 D	10 D
.74	430 2 430 2 430 2 430 2 430 2 430 2 430 2 433 3 428 3 433 2 433 3 433 2 433 3 433 2 433 3 433 2 433 3 433 2 447 5 556 6 66 7 89 0 21 4 460	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$.64 492 4 494 2 486 3 .65 481 5 479 5 .66 479 1 482 2 488 487 483 487 483 487 490 491 491 491 493 493 493 493 490 492 492 492 492 491 491 491 491 491	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$

Rat Number	6 с	8 C	9 C	9 D	10 D
461 464 464				327 331 328 330 333 332 330 328 330 330 330 334 333 333 328 328 328 328 328 328	402 407 409 411 414 415 417 411 414 413 415 418

TABLE IV (continued)

TABLE V

Heart rate during successive one minute periods of adaptation, conditioning and extinction. Criterion changes and number of brain stimulation reinforcements during conditioning periods

	Group 1:		e increases ion 3	reinforced	
Rat Number 2 C	6 в	7 C	10n 3 10 C	4 D	5 D
487 487 497 493 490 491 489 493 494 488 492 488 486 486 489 492 488 486 489 492 487 484 484 484 66 489 0 .68 .68 481 0 482 0	526 516 498 484 483 467 466 467 466 465 465 465 465 465 466 469 466 469 466 469 462 462 462 462 462 462 462 463 462 462 463 462 462 463 3 493 3 493 3 493 5 494 0 499 2 502 3 497 0 502 3 501 2 501 2 501 2 501 2 506 3	$\begin{array}{c} 502 \\ 505 \\ 509 \\ 508 \\ 505 \\ 504 \\ 503 \\ 504 \\ 503 \\ 504 \\ 506 \\ 503 \\ 501 \\ 500 \\ 472 \\ 469 \\ 459 \\ 459 \\ 459 \\ 459 \\ 459 \\ 459 \\ 467 \\ 475 \\ 478 \\ 473 \\ 69 \\ 485 \\ 2 \\ 68 \\ 487 \\ 475 \\ 478 \\ 476 \\ 0 \\ 69 \\ 481 \\ 3 \\ 70 \\ 475 \\ 0 \\ 481 \\ 3 \\ 0 \\ 491 \\ 1 \\ 495 \\ 1 \\ 497 \\ 5 \end{array}$	$\begin{array}{c} 482 \\ 477 \\ 472 \\ 483 \\ 474 \\ 460 \\ 460 \\ 448 \\ 442 \\ 441 \\ 439 \\ 436 \\ 435 \\ 435 \\ 432 \\ 426 \\ 429 \\ 430 \\ 428 \\ \hline \\ 78 \\ 411 \\ 0 \\ 79 \\ 408 \\ 0 \\ 428 \\ \hline \\ 77 \\ 420 \\ 418 \\ 406 \\ 0 \\ 418 \\ 405 \\ 77 \\ 420 \\ 418 \\ 406 \\ 77 \\ 420 \\ 418 \\ 406 \\ 77 \\ 420 \\ 418 \\ 406 \\ 71 \\ 420 \\ 418 \\ 406 \\ 71 \\ 420 \\ 418 \\ 406 \\ 71 \\ 420 \\ 418 \\ 418 \\ 2 \\ 430 \\ 5 \\ 76 \\ 423 \\ 3 \\ 434 \\ 445 \\ 6 \\ 74 \\ 465 \\ 7 \\ 465 $	$\begin{array}{c} 4 \\ 486 \\ 476 \\ 475 \\ 475 \\ 474 \\ 465 \\ 465 \\ 460 \\ 444 \\ 451 \\ 451 \\ 451 \\ 433 \\ 424 \\ 439 \\ 448 \\ 460 \\ 464 \\ 451 \\ 455 \\ 455 \\ 455 \\ 455 \\ 455 \\ 455 \\ 455 \\ 455 \\ 455 \\ 455 \\ 455 \\ 455 \\ 455 \\ 455 \\ 455 \\ 455 \\ 568 \\ 458 \\ 0 \\ .68 \\ 482 \\ 5 \\ .69 \\ 475 \\ 0 \end{array}$	380 376 373 375 373 371 368 367 365 369 375 376 374 373 376 376 376 376 377 379 380 377 379 380 377 379 380 378 377 379 380 376 377 379 380 375 376 377 379 380 377 379 380 378 377 379 380 378 379 380 375 376 377 379 380 378 379 380 378 379 380 375 376 377 379 380 378 379 380 375 376 377 379 380 376 378 377 379 380 376 378 377 379 380 376 378 377 379 380 376 377 379 380 376 377 379 380 376 377 379 380 376 377 379 380 376 377 379 380 376 376 377 379 380 376 377 379 380 376 377 379 380 376 377 379 380 376 377 379 380 376 377 379 380 376 377 379 380 376 377 379 380 377 379 380 376 377 379 380 377 379 380 377 379 380 377 379 380 377 379 380 377 379 380 375 371 369 375 371 369 375 371 369 375 6 376 376 377 379 375 6 376 377 379 375 6 376 375 371 369 375 6 376 376 377 376 376 377 379 375 6 376 376 377 379 375 6 376 377 379 375 6 376 377 377 377 377 377 377

TABLE V (continued)

Rat Number 2 C	6 в	7 C	10 C	4 D	5 D
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	513 4 $518 3$ $508 0$ $517 4$ $.63 521 5$ $527 6$ $531 6$ $.62 536 4$ $539 3$ $541 5$ $540 3$ $546 2$ 537 527 521 505 505 505 500 490 482 482 474 473 471 473 471 475 470 468 469 469	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$.69 468 2 473 5 .68 478 3 .67 481 5 .66 482 0 .67 483 4 .82 0 480 0 .67 483 4 .66 489 0 .67 483 4 .66 489 0 .65 493 4 .65 489 0 .65 488 0 .65 488 0 .65 488 3 .66 480 0 .65 488 3 .66 480 0 .65 488 3 .66 480 0 .65 488 3 .66 480 0 .65 488 3 .66 480 0 .65 488 4 .65 489 3 .66 480 0 .65 488 4 .65 488 4 .66 480 0 .65 488 4 .65 488 4 .66 480 0 .66 488 4 .65 488 3 .66 480 0 .66 488 4 .65 488 4 .66 480 0 .66 488 4 .65 488 4 .66 480 0 .66 488 4 .65 488 3 .66 480 0 .66 488 4 .65 488 4 .66 480 0 .66 488 4 .65 488 4 .66 480 0 .66 488 4 .65 488 4 .66 480 0 .66 488 4 .65 488 4 .66 489 3 .66 480 0 .66 488 4 .65 488 4 .66 480 0 .66 488 4 .65 488 4 .66 480 0 .66 400 400 0 .67 400 400 0 .68 477 3 .479 5 .476 476 476 476 476 468 469 452 438 463 468 469 455 468 469 455 468 469 455 468 469 455 468 469 455 468 469 455 468 469 455 468 469 455 468 469 455 468 469 455 468 469 455 468 469 455 468 469 455 468 469 455 468 469 455 66 465 465 465 465 465 465 465 465	$\begin{array}{cccccccccccccccccccccccccccccccccccc$

2 C	6 В	<u>7</u> C	10 C	4 D	5 D
				455	395
				458	394
				454	395
				453	390
				461	390
					388
					385
					381
					389
					384
					382
					384
					386
					381

TABLE V (continued)

Heart	rate	during	succes	ssive	one	minut	e perio	ls of	adapt	ation	ı,
	condi	itionin	g and e	extino	ction	n. Cr	iterion	chan	ges ar	nd	
	r	number	of bra:	in sti	imula	ation	reinfor	cemen	ts		
			during	g con	litio	oning	periods				

	Group 2:		e decreases	reinforced	
Rat Number l B	6 C	8 C	ion 3 9 C	9 D	10 D
$\begin{array}{c} 500 \\ 498 \\ 494 \\ 490 \\ 486 \\ 482 \\ 477 \\ 473 \\ 485 \\ 481 \\ 478 \\ 474 \\ 470 \\ 477 \\ 473 \\ 469 \\ 467 \\ 465 \\ 465 \\ 465 \\ 465 \\ 465 \\ 465 \\ 465 \\ 464 \\$	447 450 442 444 456 438 454 465 465 465 465 465 465 465 465 465 465 459 448 461 463 455 462 455 462 462 462 462 462 462 462 462 462 462 462 455 462 455 462 455 462 455 462 455 462 455 462 455 462 455 462 455 457 462 57 462 57 462 57 458 436 5 439 1 436 5 439 1 356 5 439 5 5 436 5 5 439 5 5 436 5 5 5 5 5 5 5 5	485 479 480 478 480 482 484 489 481 431 407 413 429 426 421 419 412 415 417 408 417 409 412 415 417 409 413 31.411 0 409 0 407	$\begin{array}{c} 529\\ 525\\ 528\\ 533\\ 532\\ 523\\ 521\\ 530\\ 533\\ 531\\ 533\\ 531\\ 533\\ 533\\ 528\\ 523\\ 529\\ 523\\ 529\\ 523\\ 529\\ 523\\ 529\\ 523\\ 529\\ 523\\ 529\\ 523\\ 529\\ 523\\ 529\\ 523\\ 529\\ 523\\ 521\\ 526\\ 524\\ 524\\ 524\\ 524\\ 524\\ 524\\ 524\\ 524$	$\begin{array}{c} 9 \\ 9 \\ 442 \\ 439 \\ 435 \\ 429 \\ 425 \\ 429 \\ 426 \\ 426 \\ 426 \\ 427 \\ 428 \\ 431 \\ 429 \\ 431 \\ 429 \\ 431 \\ 430 \\ 430 \\ 430 \\ 430 \\ 430 \\ 430 \\ 428 \\ 428 \\ 428 \\ 428 \\ 428 \\ 428 \\ 428 \\ 426 \\ 425 \\ 427 \\ 427 \\ 427 \\ 427 \\ 428 \\ 423 \\ 427 \\ 427 \\ 428 \\ 423 \\ 427 \\ 427 \\ 428 \\ 427 \\ 427 \\ 428 \\ 427 \\ 427 \\ 428 \\ 427 \\ 426 \\ 427 \\ 421 \\ 427 \\ 426 \\ 420 \\ 6 \end{array}$	$\begin{array}{c} 10 \ \text{D} \\ 462 \\ 462 \\ 457 \\ 453 \\ 454 \\ 453 \\ 451 \\ 445 \\ 445 \\ 445 \\ 445 \\ 445 \\ 445 \\ 445 \\ 445 \\ 445 \\ 445 \\ 455 \\ 456 \\ 468 \\ 472 \\ 464 \\ 455 \\ 455 \\ 455 \\ 455 \\ 455 \\ 455 \\ 455 \\ 455 \\ 457 \\ 447 \\ 448 \\ 447 \\ 448 \\ 447 \\ 448 \\ 447 \\ 448 \\ 447 \\ 448 \\ 447 \\ 448 \\ 447 \\ 448 \\ 447 \\ 448 \\ 447 \\ 448 \\ 447 \\ 448 \\ 447 \\ 448 \\ 447 \\ 448 \\ 447 \\ 448 \\ 447 \\ 448 \\ 447 \\ 445 \\ 445 \\ 455 \\$

TABLE VI

Rat Number 1 B	6 C	8 C	9 C	9D	10 D
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	404 4 406 2 408 1 407 1 407 1 407 2 407 4 407 4 407 4 407 4 407 4 415 4 425 4 430 4 35 4 37 4 430 4 30 4 30 4 430	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$.82 \begin{array}{c} 413 \\ 414 \\ 2 \\ 410 \\ 6 \\ .83 \\ 411 \\ 412 \\ 0 \\ .82 \\ 406 \\ 6 \\ .84 \\ 402 \\ 0 \\ .83 \\ 402 \\ 6 \\ .84 \\ 398 \\ 5 \\ .396 \\ 6 \\ .84 \\ 398 \\ 5 \\ .398 \\ 5 \\ .398 \\ 5 \\ .398 \\ 5 \\ .398 \\ 0 \\ .83 \\ 399 \\ 5 \\ .84 \\ 398 \\ 0 \\ .83 \\ 399 \\ 5 \\ .84 \\ 398 \\ 0 \\ .83 \\ 399 \\ 0 \\ .82 \\ 393 \\ 5 \\ .84 \\ 398 \\ 0 \\ .83 \\ 399 \\ 0 \\ .82 \\ 393 \\ 5 \\ .84 \\ 398 \\ 0 \\ .83 \\ 399 \\ 0 \\ .83 \\ 394 \\ 1 \\ 2 \\ .84 \\ 391 \\ 2 \\ .84 \\ 391 \\ 2 \\ .84 \\ 391 \\ 2 \\ .84 \\ 391 \\ 2 \\ .84 \\ 391 \\ 2 \\ .84 \\ 398 \\ 0 \\ .83 \\ 399 \\ .88 \\ 389 \\ 393 \\ 389 \\ 393 \\ $	$\begin{array}{cccccccccccccccccccccccccccccccccccc$

Ra	: Number 1 B	6 C	8 C	9 C	9 D	10 D
	376 381 382 384 387 385	468 471 469 469	430 432 430	520 526 531 531	393 393 393 393 393 390 391 388 393 394 393 394 393 392 394 393 393 393 393 393 393 393 393 392 390 385 383 380	420 422 428 428 432 431 435 435 435 435 435 435 435 435 440 444 447 448 455 459 455 460 460 461 461

TABLE VI (continued)

Heart rate during successive one minute periods of adaptation, conditioning and extinction. Criterion changes and number of brain stimulation reinforcements during conditioning periods

TABLE VII

	Group 1:		e increases ion 4	reinforced	
Rat Number 2 C	бв	7 C	10 C	4 D	5 D
456 .69 462 .68	471 3 479 5 481 1 475 1 482 3 480 0 482 0 487 1 490 3 491 3 494 5 484 1	$\begin{array}{c} 399\\ 417\\ 415\\ 424\\ 419\\ 423\\ 438\\ 436\\ 440\\ 446\\ 452\\ 460\\ 466\\ 466\\ 466\\ 466\\ 466\\ 466\\ 466$	$\begin{array}{c} 481\\ 481\\ 470\\ 450\\ 469\\ 461\\ 452\\ 458\\ 473\\ 473\\ 473\\ 475\\ 473\\ 465\\ 461\\ 462\\ 467\\ 470\\ 472\\ 469\\ 4661\\ 462\\ 1\\ 462\\ 1\\ 468\\ 4\\ 69\\ 469\\ 2\\ 1\\ 468\\ 4\\ 69\\ 469\\ 2\\ 1\\ 468\\ 4\\ 69\\ 469\\ 2\\ 70\\ 456\\ 0\\ 455\\ 5\\ 475\\ 3\\ 479\\ 472\\ 1\\ 72\\ 1\\ 2\\ 2\\ 1\\ 2\\ 1\\ 2\\ 1\\ 2\\ 2\\ 1\\ 2\\ 2\\ 1\\ 2\\ 2\\ 2\\ 2\\ 2\\ 2\\ 2\\ 2\\ 2\\ 2\\ 2\\ 2\\ 2\\$	$\begin{array}{c} 453\\ 457\\ 451\\ 451\\ 463\\ 464\\ 463\\ 464\\ 449\\ 447\\ 4439\\ 435\\ 428\\ 430\\ 435\\ 428\\ 430\\ 435\\ 428\\ 431\\ 435\\ 428\\ 424\\ 431\\ 435\\ 426\\ 424\\ 431\\ 435\\ 426\\ 426\\ 426\\ 424\\ 431\\ 435\\ 426\\ 426\\ 426\\ 426\\ 426\\ 426\\ 426\\ 426$	431 423 412 404 402 395 385 386 397 394 397 407 412 407 393 397 407 412 407 393 397 407 412 407 408 412 407 408 412 409 409 409 409 409 409 409 412 409 409 412 409 409 412 409 401 401 401 401 401 401 401 401 401 401

Rat	Number 2 C	6 в	7 C	10 C	4 D	5 D
.69 .68 .70	483 0 481 1 481 0 479 0 478 4	472 3 477 2 484 488 4 486 6 488 4 486 4 486 4 486 4 486 4 486 4 486 4 486 4 486 4 486 4 49 49 49 49 49 49 49 49 49 49 49 49 49 4	$\begin{array}{c} 476 & 0 \\ 476 & 3 \\ 473 & 2 \\ 473 & 2 \\ 473 & 2 \\ 473 & 2 \\ 1478 & 4 \\ 488 & 6 \\ 508 & 6 \\ 498 & 6 \\ 508 & 6 \\ 498 & 6 \\ 498 & 6 \\ 498 & 6 \\ 498 & 6 \\ 498 & 6 \\ 498 & 6 \\ 498 & 6 \\ 498 & 6 \\ 498 & 6 \\ 498 & 6 \\ 474 & 475 \\ 471 & 476 \\ 466 & 476 \\ 474 & 475 \\ 471 & 476 \\ 469 & 472 \\ 471 & 476 \\ 469 & 472 \\ 471 & 476 \\ 469 & 472 \\ 471 & 470 \\ 469 & 466 \\ 466 & 466 \\ 466 & 466 \\ 466 & 461 \\ 461 & 461 \\ 461 & 461 \\ 460 & 460 \\ 460 & 460 \\ \end{array}$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{c} 415 & 4 \\ 416 & 6 \\ 416 & 6 \\ 418 & 1 \\ 416 & 0 \\ 418 & 1 \\ 419 & 2 \\ 421 & 3 \\ 427 & 6 \\ 427 & 6 \\ 431 & 3 \\ 431 & 3 \\ 435 & 2 \\ 435 & 2 \\ 435 & 0 \\ 437 & 6 \\ 431 & 0 \\ 437 & 6 \\ 431 & 0 \\ 437 & 6 \\ 435 & 3 \\ 431 & 0 \\ 437 & 6 \\ 435 & 3 \\ 431 & 0 \\ 437 & 6 \\ 435 & 3 \\ 431 & 0 \\ 437 & 6 \\ 435 & 3 \\ 431 & 0 \\ 437 & 6 \\ 435 & 3 \\ 431 & 0 \\ 437 & 6 \\ 435 & 3 \\ 431 & 0 \\ 437 & 6 \\ 435 & 3 \\ 431 & 0 \\ 437 & 6 \\ 435 & 3 \\ 431 & 0 \\ 437 & 6 \\ 435 & 3 \\ 431 & 0 \\ 437 & 6 \\ 435 & 3 \\ 431 & 0 \\ 437 & 6 \\ 435 & 3 \\ 431 & 0 \\ 437 & 6 \\ 435 & 3 \\ 431 & 0 \\ 437 & 6 \\ 435 & 3 \\ 431 & 0 \\ 437 & 6 \\ 435 & 3 \\ 431 & 0 \\ 437 & 6 \\ 435 & 3 \\ 431 & 0 \\ 437 & 6 \\ 435 & 3 \\ 431 & 0 \\ 437 & 6 \\ 435 & 3 \\ 431 & 0 \\ 435 & 1 \\ 417 & 1 \\ 413 & 1 \\ 417 \end{array}$	414 6 412 2 411 0 414 2 416 4 421 5 425 4 421 3 419 2 413 1 413 1 413 1 415 1 412 1 412 1 412 1 412 1 412 1 412 0 415 3 416 4 418 4 418 4 418 4 418 4 418 4 415 411 407 403 395 391 394 391 394 391 391 392 393 393 393 393 393 396 388

TABLE VII (continued)

Rat Number 2 C	6в	7 C	10 C	4 D	5 D
458 <u>457</u>			468 467	419 417 421 421 420 420 422 424 424 424 419 425 424	392 393 394 396 400 401 393 389 391 388 390

TABLE VII (continued)

TABLE VIII

Heart rate during successive one minute periods of adaptation, conditioning and extinction. Criterion changes and number of brain stimulation reinforcements during conditioning periods

		Group 2		te decreases sion 4	reinforced	
Rat	Number 1 B	6 C	8 C	9 C	9 D	l0 D
.68 .69 .70	519 519 518 518 516 517 514 515 510 501 509 510 501 509 510 501 490 25 472 472 472 472 472 472 453 452 445 456 456 456 456 456 456 456 456 456 4	$\begin{array}{c} 455\\ 462\\ 475\\ 476\\ 470\\ 475\\ 466\\ 474\\ 481\\ 491\\ 488\\ 492\\ 494\\ 495\\ 497\\ 495\\ 497\\ 491\\ 488\\ 492\\ 494\\ 495\\ 494\\ 491\\ 489\\ 500\\ \hline \begin{array}{c} .68\\ 475\\ 5\\ .72\\ 454\\ 3\\ .73\\ 455\\ 4\\ .72\\ 478\\ 1\\ .69\\ 485\\ 3\\ 481\\ 6\\ \end{array}$	421 419 418 416 418 421 412 407 412 406 409 413 408 397 2 392 6 .87 390 4 .86 386 3 .85 390 0 384 1 383 2 .86 382 3 381 4 379 5 .87 376 3	$\begin{array}{c} 521\\ 520\\ 519\\ 518\\ 517\\ 517\\ 519\\ 516\\ 517\\ 513\\ 512\\ 513\\ 512\\ 514\\ 521\\ 519\\ 515\\ 511\\ 511\\ 510\\ 515\\ 511\\ 510\\ 514\\ 512\\ 516\\ 511\\ 507\\ 507\\ .62\\ 511\\ 507\\ 507\\ .62\\ 511\\ 507\\ 507\\ .63\\ 504\\ 6\\ .64\\ 501\\ 6\\ .64\\ 504\\ 2\\ .63\\ 504\\ 3\\ 498\\ 6\\ .64\\ 497\\ 5\\ 495\\ 5\\ .65\\ 494\\ 5\end{array}$	373 371 368 367 366 363 359 358 356 375 354 352 350 347 351 347 352 350 347 352 350 347 352 341 340 337 333 331 329 326 4 320 326 4 324 323 4	$\begin{array}{c} 468 \\ 462 \\ 468 \\ 477 \\ 482 \\ 480 \\ 481 \\ 478 \\ 475 \\ 473 \\ 469 \\ 475 \\ 481 \\ 475 \\ 469 \\ 475 \\ 482 \\ 477 \\ 477 \\ 482 \\ 477 \\ 475 \\ 466 \\ 468 \\ 471 \\ 475 \\ 480 \\ 474 \\ 477 \\ 473 \\ 475 \\ 475 \\ 477 \\ 70 \\ 475 \\ 0 \\ 471 \\ 0 \\ 71 \\ 468 \\ 4 \end{array}$

TABLE VIII (continued)

Rat Number 1 B	6 C	8 C	9 C	9 D	10 D
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	376 1 375 2 373 3 373 3 375 1 372 3 378 380 382 382 382 384 384 389 387 386 383 384 386 383 384 386 387 386 387 386 387 388 391 387 389 387 387 387 389 387 397 397 397 397 397 397 397 397 397 397 397 397 397 397 39	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	470 2 70 458 4 454 1 73 460 2 454 1 73 460 2 459 3 71 454 2 459 3 71 454 3 459 3 71 454 3 459 5 72 456 5 459 5 2466 5 74 452 2 460 2 459 5 72 456 0 458 2 459 3 459 5 74 452 2 450 6 75 454 0 454 0 454 0 454 0 454 3 449 3

t Number	6 C	8 C	9 C	9 D	10 D
	503 504		499 498 497	307 306 308 308 308	454 455 445 454 462 464 462 464 457 456 462

TABLE VIII (continued)

APPENDIX B

OPERANT CONDITIONING OF BOTH INCREASES AND DECREASES

IN HEART RATE IN THE SAME RAT

Since the results of the first experiment reported in this dissertation provided clear evidence of operant conditioning of both increases and decreases in heart rate in groups of curarized rats, a second experiment was performed. In this experiment an attempt was made to condition both increases and decreases in heart rate in the same rat. Two rats were used. The following changes were made in the procedure employed in this second experiment.

1. The rats were trained in the morning, with a curare session never starting earlier than 8:00 A.M. or later than 11:00 A.M. Since rats have clear diurnal activity cycles (Munn, 1950) controlling the time of the curare sessions represented a procedural improvement.

2. Curare sessions were separated by a fixed interval of four days. In contrast to this fixed interval, in the first experiment curare sessions were separated by varying intervals of from three to six days.

3. Following the sessions in the Skinner box in which the reinforcing properties of the brain stimulation were assessed, the rats were given a curare adaptation session during which they were curarized and their heart rates recorded, but the contingencies were not in effect and so neither the feedback stimulus or the brain stimulation was presented. The introduction of this adaptation curare

session provided baseline data against which the conditioned changes in heart rate could be evaluated.

4. The adaptation and extinction phases of the curare conditioning sessions lasted for thirty minutes and the conditioning phase lasted for forty minutes. This procedure eliminated the small variation in the lengths of the different phases of the curare sessions which was present in the first experiment.

Subjects

Two, male Sprague-Dawley rats were used in this experiment. They were housed and maintained under conditions identical to those of the first experiment.

Implantation Procedure

The implantation operation and the procedure used to assess the reinforcing properties of the brain stimulation were identical to those of the first experiment.

Curare Sessions

The procedure used during the curare sessions was similar to that of the first experiment but with the additional controls which have been mentioned. The first curare session was an adaptation session during which the contingencies were not in effect and no stimuli were presented. During this period the rats' heart rates were recorded each minute for one hundred minutes. Four days later the rats were given their second curare session, during which the reinforcement contingency was in effect. One of the rats was reinforced for increases in heart rate, and one for heart rate decreases. The decision as to the direction of the heart rate changes to be conditioned first in each rat was made on the basis of the toss of a coin. The first thirty minutes of the curare conditioning session was an adaptation phase during which no stimuli were presented. During the next forty minutes a conditioning procedure identical to that of the first experiment was employed. This conditioning phase was followed by a thirty minute extinction phase. During all three phases of the session the rats' heart rates were recorded during successive one minute periods.

The conditioning procedure was used for three curare sessions each of which had thirty minute adaptation and extinction phases, and a forty minute conditioning phase. All curare sessions were separated by four days. The numbers of heart beats for the two rats used in this experiment during successive minutes of the first four curare sessions are shown in Tables XIV to XX of Appendix C. In addition criterion changes and number of brain stimulation reinforcements during conditioning are also shown in these tables. In order to present the changes in heart rate which occurred in the two rats during each of the first four curare sessions mean heart rates were calculated for each of the rats during ten blocks of adaptation periods in the case of the curare adaptation session, and three blocks of adaptation periods, four blocks of conditioning periods, and three blocks of extinction periods during the three curare conditioning sessions. Unlike the first

experiment in this experiment each of these blocks of periods represented ten minutes of the curare session. These mean heart rates are shown in Tables IX and X and are presented graphically in Figures 4 and 5.

TABLE	IX
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Session 1 Adaptation				Session 2 Conditioning			_
Rat No. H.R. I Reinfo	ncr.	<u>3 E</u> H.R. Decr. Reinforced		H.R. I Reinfo	ncr.	<u>3</u> E H.R. Decr. Reinforced	
Adap.	511	517		Adap.	521	542	
Adap.	524	514		Adap.	515	552	
Adap.	522	514		Adap.	510	534	
Adap.	520	516		Cond.	490	539	
Adap.	520	512		Cond.	496	549	
Adap.	515	508		Cond.	495	546	
Adap.	509	504		Cond.	498	540	
Adap.	507	506		Ext.	500	551	
Adap.	505	512		Ext.	495	540	
Adap.	505	508	14	Ext.	494	534	

	TABLE	Х
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-							
Session 3				Session 4			
Conditioning				Conditioning			
Rat No.	<u>2 E</u>	3 E	2 E		<u>3 E</u>		
H.R. I	ncr.	H.R. Decr.	H.R. In	cr.	H.R. Decr.		
Reinfo	orced	Reinforced	Reinfor	ced	Reinforced		
-							
Adap.	510	517	Adap.	486	474		
Adap.	515	512	Adap.	486	465		
Adap.	524	520	Adap.	491	459		
Cond.	547	519	Cond.	490	453		
Cond.	550	522	Cond.	509	450		
Cond.	555	518	Cond.	504	446		
Cond.	554	514	Cond.	522	436		
Ext.	542	513	Ext.	533	439		
Ext.	530	515	Ext.	517	446		
Ext.	527	518	Ext.	506	448		

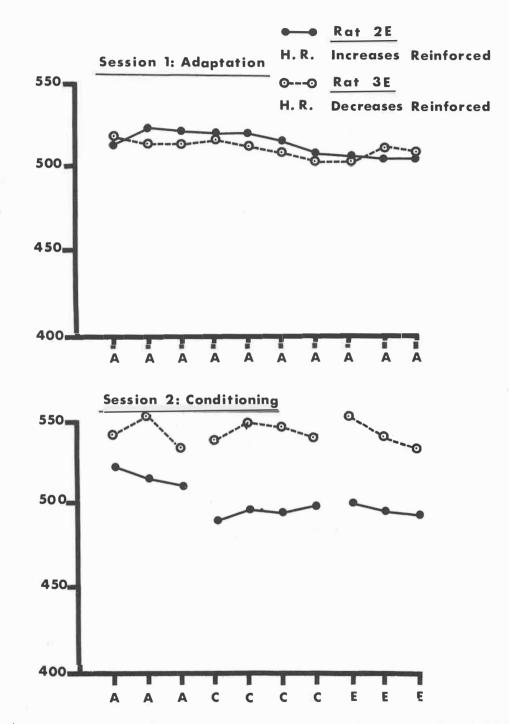


Figure 4. Mean heart rates during ten 10 minute blocks of one minute periods in Session 1, and mean heart rates during three blocks of adaptation periods, four blocks of conditioning periods, and three blocks of extinction periods in Session 2.

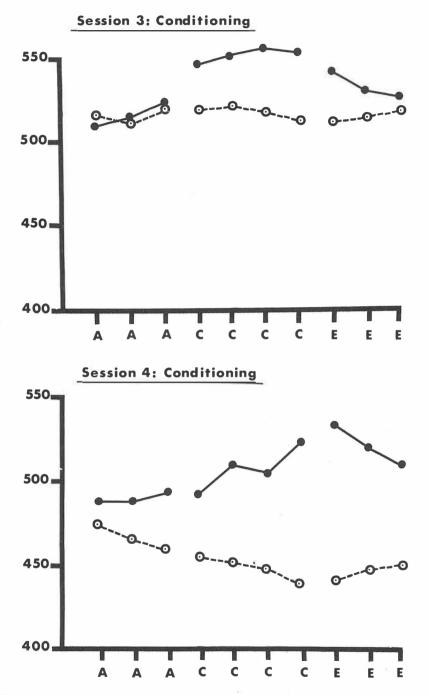


Figure 5. Mean heart rates during three blocks of adaptation periods, four blocks of conditioning periods, and three blocks of extinction periods in Sessions 3 and 4.

During the adaptation curare session both groups showed a small over-all decrease in heart rate. The two rats showed little if any evidence of operant conditioning of either an increase or a decrease in heart rate during the first curare conditioning session. On the third curare session there was some evidence of operant conditioning of changes in heart rate, and on the fourth session clear heart rate changes were conditioned. During the fourth curare session rat number 2 E showed an increase in heart rate of 31 b.p.m. during conditioning, and rat number 3 E a decrease of 23 b.p.m. So the results for these two rats in this second experiment provide additional evidence of operant conditioning of both increases and decreases in heart rate in curarized rats.

The fifth curare session was a reversal conditioning session during which the following procedure was adopted. As usual the session began with a thirty minute adaptation session. This was followed by a twenty minute conditioning phase during which the two rats were reinforced for heart rate changes in the direction which had previously been conditioned. During these conditioning periods rat $2 \ge 100$ showed a clear increase in heart rate, and rat $3 \ge 100$ a decrease. A ten minute extinction period, during which the contingencies were not in effect, followed these conditioning periods. A thirty minute reversal conditioning phase followed these extinction periods. During this reversal phase of the curare session the rat which had previously been reinforced for heart rate increases $-2 \ge -1000$ was reinforced for heart rate decreases, and the rat which had previously been reinforced for heart rate decreases - $3 \ge$ - was reinforced for heart rate increases. During the reversal conditioning periods the feedback stimulus was changed so that criterion IBIs - long IBIs in the case of rat 2 E, and short IBIs in the case of rat $3 \ge$ - were followed by a high frequency tone, and non-criterion IBIs were followed by a low frequency tone. It was hypothesized that changing the modality of the feedback stimulus and making it more informative would facilitate acquisition of the heart rate reversal. The reversal conditioning phase of this session lasted for thirty minutes and was followed by a twenty minute extinction phase. The results for the reversal conditioning session are shown in Table XI and are presented graphically in Figure 6.

Although both rats showed heart rate changes in the direction being conditioned during the first set of conditioning periods of this session, they showed no evidence of acquisition of the reverse heart rate change during the reversal conditioning periods. In fact rat $2 \ge$ which was being reinforced for heart rate decreases during the reversal periods, showed an increase in heart rate, while rat $3 \ge$ which was being reinforced for heart rate increases during the reversal periods, showed a heart rate decrease. So during the reversal phase of this session both rats continued to show heart rate changes in the direction which had been conditioned during the earlier curare conditioning sessions, in spite of the change in the direction of the heart rate change which produced the brain stimulation reinforcement. It seemed possible that

TABLE XI

	Sess	ion 5	
2 E	ersal C	Conditioning 3 E	
Adaptation	456	Adaptation	487
Adaptation	458	Adaptation	482
Adaptation	455	Adaptation	464
Cond. H.R. Incr. Reinforced	470	Cond. H.R. Decr. Reinforced	467
Cond. H.R. Incr. Reinforced	486	Cond. H.R. Decr. Reinforced	447
Extinction	467	Extinction	435
Cond. H.R. Decr. Reinforced	479	Cond. H.R. Incr. Reinforced	443
Cond. H.R. Decr. Reinforced	476	Cond. H.R. Incr. Reinforced	443
Cond. H.R. Decr. Reinforced	479	Cond. H.R. Incr. Reinforced	439
Extinction	487	Extinction	452
Extinction	481	Extinction	452

Mean heart rates in beats per minute during blocks of adaptation, conditioning and extinction periods

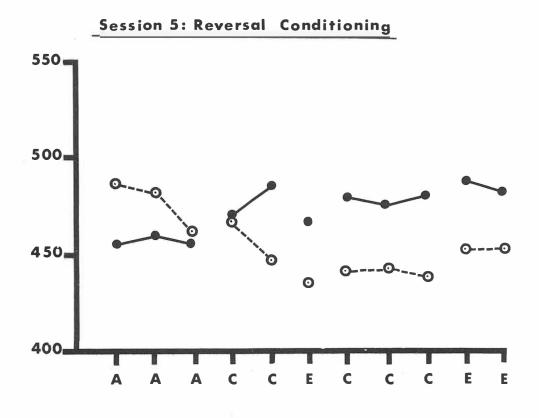


Figure 6. Mean heart rates during three blocks of adaptation periods, two blocks of conditioning periods, one block of extinction periods, three blocks of reversal conditioning periods, and two blocks of extinction periods in Session 5.

further conditioning might lead to acquisition of the heart rate reversal and so two further curare sessions were given during which the reversal conditioning procedure was employed. The results for these two curare reversal conditioning sessions are shown in Table XII and are presented graphically in Figure 7.

The only suggestion of any acquisition of the reversal response was the performance of rat $2 \ge$ during the sixth curare session. During the conditioning phase of this session rat $2 \ge$, which was being reinforced for decreases in heart rate showed a heart rate decrease of 14 b.p.m. However during the seventh curare session the rat showed an increase in heart rate of 18 b.p.m. during the reversal conditioning phase. On both of the last two curare sessions rat $3 \ge$ showed no evidence of acquisition of the heart rate reversal - an increase in heart rate. On the sixth curare session rat $3 \ge$ showed a heart rate decrease of 9 b.p.m., and on the seventh curare session a heart rate conditioning curare sessions the heart rate changes which had initially been conditioned.

Although the results for these two rats show good initial acquisition of both a heart rate increase and a heart rate decrease, thus replicating the results of the first experiment, they provide no evidence of any acquisition of the heart rate reversal. However the reversal response which was required in this experiment was a change in heart rate opposite to that which had originally been

TABLE	XII

Potto	Sessi		Derror	on 7	
		nditioning			litioning 3 E
Rat No.		$\frac{3 E}{H.R. Incr.}$	Territoria de la constante de la const	E	
H.R. I			H.R.		H.R. Incr.
Reinfo	breed	Reinforced	Reini	orced	Reinforced
Adap.	504	508	Adap.	480	515
Adap.	485	494	Adap.	478	499
Adap.	487	472	Adap.	467	487
Cond.	488	469	Cond.	474	474
Cond.	480	468	Cond.	476	463
Cond.	477	448	Cond.	479	464
Cond.	474	460	Cond.	482	469
Ext.	507	447	Ext.	481	479
Ext.	485	24 24 24	Ext.	478	480
Ext.	485	436	Ext.	474	479

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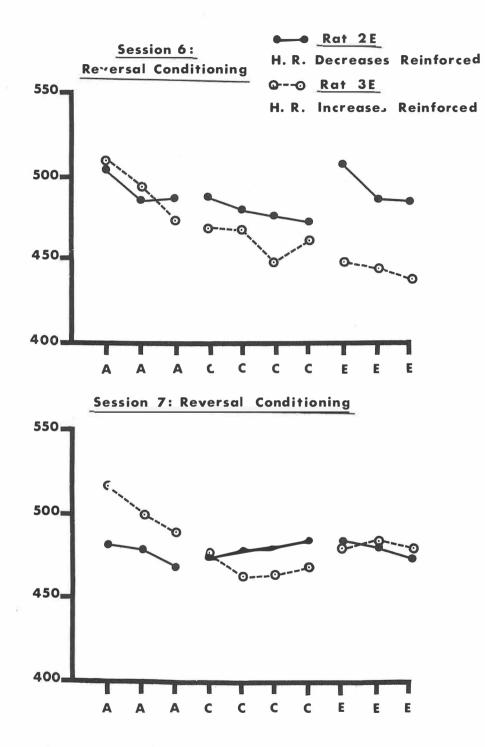


Figure 7. Mean heart rates during three blocks of adaptation periods, four blocks of conditioning periods, and three blocks of extinction periods in Sessions 6 and 7.

conditioned. Difficulty in conditioning such a heart rate reversal was not unexpected since even with a conventional operant response such as lever pressing, it is anticipated that considerable difficulty would be experienced in training a rat which had previously been reinforced for high rates of responding to respond at a lower rate. The results from these two rats do however strongly suggest that considerable difficulty will be experienced in conditioning both increases and decreases in heart rate in the same \underline{S} .

This suggestion is confirmed by the results from one additional rat in which an attempt was made to condition a heart rate reversal. It was hypothesized that a stimulus which was present during the initial conditioning, but not during the reversal conditioning, would facilitate acquisition of the reversal heart rate response. For this rat - number 5 E - a procedure similar to that of rats 2 E and 3 E was used except that:

1. A light was present during the initial conditioning periods, but not during the reversal conditioning periods. It was hypothesized that making the two sets of conditioning periods more discriminable would facilitate acquisition of the heart rate reversal.

2. During the initial conditioning periods the rat was reinforced for heart rate increases but no feedback stimuli were presented.

3. The fifth and sixth curare sessions were reversal conditioning sessions during which heart rate decreases were reinforced. During these two sessions criterion IBIs were followed by a high frequency tone, and non-criterion IBIs by a low frequency tone. The results for the adaptation curare session, the second, third and fourth conditioning sessions, and the fifth and sixth reversal conditioning sessions are shown in Tables XXI to XXIII of Appendix D, and are summarized in Table XIII and are presented graphically in Figures 8, 9 and 10.

On the third and fourth curare conditioning sessions this rat showed a clear increase in heart rate during conditioning - an increase of 33 b.p.m. during the conditioning phase of the third session, and an increase of 45 b.p.m. during the conditioning phase of the fourth session. So once again these results show operant conditioning of a heart rate change in a curarized rat. However on the fifth and sixth curare reversal conditioning sessions, during which heart rate decreases were reinforced, this rat continued to show an increase in heart rate. So with this rat, as was the case with rats 2 E and 3 E, there was no evidence of acquisition of the heart rate reversal. Although with extended conditioning it may be possible to condition heart rate reversals the results from these three rats strongly suggest that such heart rate reversals will only be conditioned with great difficulty.

Sessions l			2	3	4	5	6
Adaptation		H. R.	Increases	Reinfo	prced H.R	. Decr.	Reinforced
Adap.	468	Adap.	452	477	461	455	464
Adap.	460	Adap.	462	459	459	453	458
Adap.	463	Adap.	440	445	455	450	456
Adap.	452	Cond.	448	422	442	471	47ı
Adap.	452	Cond.	447	447	456	466	473
Adap.	456	Cond.	440	453	470	469	477
Adap.	462	Cond.	447	455	487	469	479
Adap.	459	Ext.	453	460	487	467	474
Adap.	455	Ext.	459	463	491	458	465
Adap.	460	Ext.	466	457	482	451	458

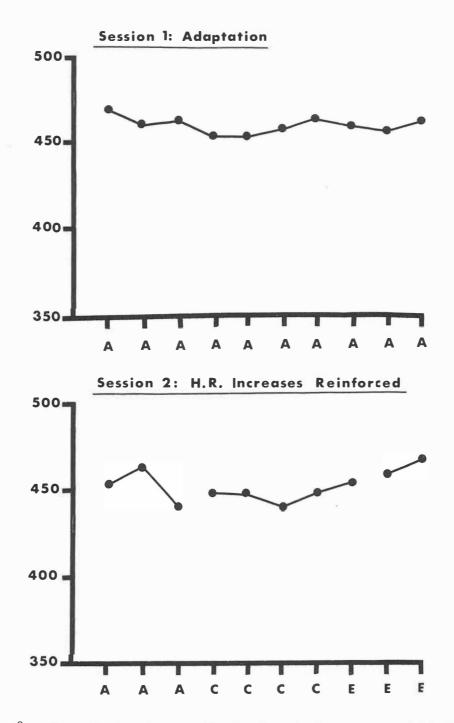


Figure 8. Heart rates during blocks of adaptation, conditioning and extinction periods. Adaptation and Session 2.

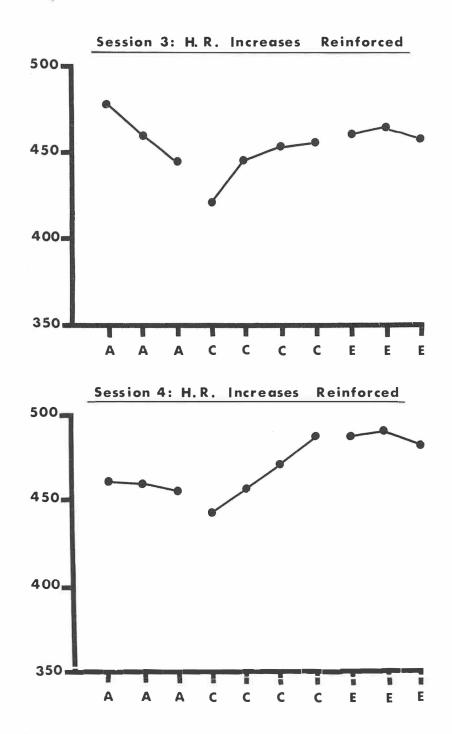


Figure 9. Heart rate during blocks of adaptation, conditioning and extinction periods. Sessions 3 and $4\,.$

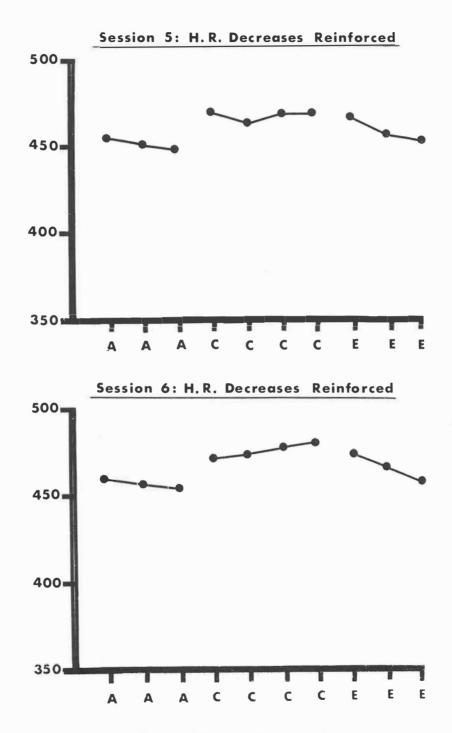


Figure 10. Heart rate during blocks of adaptation, conditioning and extinction periods. Sessions 5 and 6.

APPENDIX C

Tables XIV to XX showing heart rates for rat numbers $2 \ge and$ $3 \ge during$ successive one minute periods of one adaptation session, three conditioning sessions and three reversal conditioning sessions. Criterion changes and number of brain stimulation reinforcements during conditioning periods.

	Dot Number O I		D	ot Number 2 I	
	Rat Number <u>2 E</u> Adaptation		R	at Number <u>3 I</u> Adaptation	<u>-</u>
$\begin{array}{c} 558\\ 556\\ 553\\ 541\\ 523\\ 489\\ 473\\ 481\\ 503\\ 523\\ 523\\ 523\\ 522\\ 522\\ 522\\ 522\\ 52$	523 523 521 520 518 521 520 518 519 519 519 519 519 519 517 516 517 517 516 517 517 514 515 511 511 512 510 509 509 509 509 507 507 507 507 507 507 508 508 509 507 507 507	505 507 506 504 505 504 506 506 505 505 505 505 505 505 507 503 505 505 505 505	524 527 523 529 507 518 513 511 512 507 511 517 523 519 511 509 505 511 513 517 513 517 513 517 513 517 513 517 513 517 513 517 513 517 515 514 508 513 517 516 522 520 512 508 513 517 516 522 508 514 517 516 522 508 514 517 516 522 508 514 517 516 522 508 514 517 516 522 508 514 517 516 522 508 514 517 518 517 516 522 508 514 517 518 517 516 522 508 514 517 518 517 516 527 518 517 516 527 518 517 516 527 518 517 518 517 516 527 518 517 516 527 518 517 518 517 518 517 518 517 518 517 518 517 515 518 517 515 517 518 517 515 518 517 515 518 517 515 518 517 515 517 518 517 515 517 518 517 515 517 515 517 515 517 515 517 515 517 515 517 515 517 515 517 515 517 515 517 515 515 517 515	515 514 511 512 510 512 510 512 509 508 507 508 5112 508 505 504 502 505 504 505 504 505 504 505 504 505 504 505 504 505 504 505 504 505 504 505 504 505 504 505 504 505 504 502 506 510 502 506 510 502 506 510 502 506 510 502 506 500 502 505 504 505 504 505 504 505 506 510 502 505 504 505 504 505 506 510 505 507 509 510	513 512 511 510 509 513 512 512 512 512 512 512 512 512 510 508 506 508 506 508 506 508 509 507 508

TABLE	XV
-------	----

TABLE XVI

	Rat Number 2 E			Rat Number <u>3 E</u>	
Conditioning <u>Heart</u> rate		Conditioning Heart rate			
1nc	reases reinior	cea	deci	reases reinford	ea
Cond			Cond		
519 511 500 507 512 525 527 539 531 531 531 531 .62 538 4 .61 550 3 551 1 548 0 .62 549 4 .61 547 0	.60 557 1 .61 554 1 .62 549 3 .61 555 5 553 1 554 1 552 0 555 4 554 2 554 2 554 2 554 2 554 2 554 2 555 3 555 3 555 3 555 3 555 0 551 0	531 526 526 526 525 526 525 527 527	510 519 520 520 514 516 514 521 525 523 523 523 526 .70 522 0 .68 521 0 .66 518 1 519 0 .65 517 4 519 4	$\begin{array}{c} 510 & 6\\ .65 & 518 & 1\\ 521 & 0\\ 518 & 2\\ 514 & 4\\ .66 & 514 & 0\\ .65 & 515 & 2\\ 513 & 3\\ .66 & 516 & 0\\ .65 & 514 & 3\\ 513 & 4\\ 515 & 3\\ 515 & 3\\ 512 & 4\\ 512 & 4\\ 510 \\ 510 \\ 510 \\ \end{array}$	517 515 516 517 519 521 521 522 521

	Rat Number <u>2 E</u>			Rat Number <u>3 E</u>	
	Conditioning Heart rate		Conditioning Heart rate		
incr	eases reinfor	ced	decreases reinforced		ed
	$\begin{array}{c} \mbox{tensor} \begin{tabular}{lllllllllllllllllllllllllllllllllll$			0	

TABLE XVIII

Rat Number <u>2 E</u> Reversal Conditionin	g	at Number <u>3</u> sal Conditio	
	g 473 1 473 0 479 0 479 0 479 0 479 0 474 0 480 0 476 0 480 0 482 1 488 0 482 1 488 0 482 488 489 489 489 489 488 489 489 489 488 481 480 481 485 486 481 485 486 481 485 486 481 485 486 481 485 486 481 485 486 481 481 485 486 481 481 485 486 481 481 485 486 481 481 485 486 481 481 485 486 481 481 485 486 481 481 485 486 481 485 486 481 485 486 481 485 486 481 485 486 481 485 486 481 485 486 483 478 478 478 478 478 478 478 478 478 478		

TABLE XIX

the second se						
Rat Numb				t Number <u>3 E</u>		
Conditioning Heart rate		е	Conditioning Heart rate			
decreases r	decreases reinforced			ases reinforced		
513 49 513 49 505 48 506 48 503 48 500 48 500 48 500 48 500 48 500 48 500 48 500 48 503 47 499 47 495 47 491 48 486 47 483 48 477 47 483 47 483 47 483 47 483 47 483 47 480 47 480 47 480 47 486 47 486 47 486 47 486 47 486 47 486 47 486 47	98 0 93 0 93 0 93 0 93 0 93 0 93 0 93 0 93 0 93 0 93 0 93 0 93 0 93 0 93 1 93 1 93 1 93 1 93 1 93 1 93 1 93 1 93 1 93 1 93 1 93 1 93 1 93 1 93 1 93 1 93 1 93 1 93 1 93 2 93 2 93 3 93 3 93 <td< td=""><td></td><td>$\begin{array}{r} \text{incre} \\ 516 \\ 512 \\ 512 \\ 510 \\ 509 \\ 509 \\ 509 \\ 506 \\ 505 \\ 501 \\ 500 \\ 502 \\ 498 \\ 497 \\ 494 \\ 492 \\ 488 \\ 482 \\ 492 \\ 488 \\ 482 \\ 492 \\ 488 \\ 482 \\ 471 \\ 472 \\ 471 \\ 472 \\ 471 \\ 472 \\ 471 \\ 472 \\ 471 \\ 472 \\ 471 \\ 472 \\ 471 \\ 472 \\ 471 \\ 472 \\ 471 \\ 472 \\ 2 \end{array}$</td><td>$\begin{array}{r} 471 & 3 \\ 467 & 0 \\ .72 & 464 & 2 \\ 465 & 5 \\ 468 & 6 \\ .71 & 471 & 5 \\ 471 & 5 \\ 472 & 6 \\ .70 & 474 & 0 \\ 474 & 0 \\ .71 & 472 & 0 \\ 467 & 0 \\ .71 & 472 & 0 \\ 467 & 0 \\ .71 & 472 & 0 \\ 467 & 0 \\ .71 & 453 & 0 \\ .73 & 454 & 0 \\ .74 & 453 & 0 \\ .74 & 453 & 0 \\ .75 & 450 & 3 \\ 448 & 0 \\ 441 & 0 \\ 435 & 0 \\ .74 & 450 & 5 \\ .73 & 454 & 6 \\ 462 & 5 \\ .72 & 462 & 2 \\ 466 & 3 \\ 462 & 5 \\ .72 & 462 & 2 \\ 466 & 3 \\ 462 & 1 \\ 458 & 0 \\ .73 & 462 & 6 \\ 458 & 3 \\ 460 & 5 \\ 457 & 3 \\ 450 & 0 \\ 447 \end{array}$</td><td>446 443 446 4443 4446 4445 4448 4450 44522 4446 44522 4446 44522 4446 44522 4446 44522 4446 44522 4446 44522 4446 44522 4446 44522 4446 44522 4446 44522 4446 445227 44526 445227 44526 445227 44526 445227 44527777777777</td></td<>		$\begin{array}{r} \text{incre} \\ 516 \\ 512 \\ 512 \\ 510 \\ 509 \\ 509 \\ 509 \\ 506 \\ 505 \\ 501 \\ 500 \\ 502 \\ 498 \\ 497 \\ 494 \\ 492 \\ 488 \\ 482 \\ 492 \\ 488 \\ 482 \\ 492 \\ 488 \\ 482 \\ 471 \\ 472 \\ 471 \\ 472 \\ 471 \\ 472 \\ 471 \\ 472 \\ 471 \\ 472 \\ 471 \\ 472 \\ 471 \\ 472 \\ 471 \\ 472 \\ 471 \\ 472 \\ 2 \end{array}$	$\begin{array}{r} 471 & 3 \\ 467 & 0 \\ .72 & 464 & 2 \\ 465 & 5 \\ 468 & 6 \\ .71 & 471 & 5 \\ 471 & 5 \\ 472 & 6 \\ .70 & 474 & 0 \\ 474 & 0 \\ .71 & 472 & 0 \\ 467 & 0 \\ .71 & 472 & 0 \\ 467 & 0 \\ .71 & 472 & 0 \\ 467 & 0 \\ .71 & 453 & 0 \\ .71 & 453 & 0 \\ .71 & 453 & 0 \\ .71 & 453 & 0 \\ .73 & 454 & 0 \\ .74 & 453 & 0 \\ .74 & 453 & 0 \\ .75 & 450 & 3 \\ 448 & 0 \\ 441 & 0 \\ 435 & 0 \\ .74 & 450 & 5 \\ .73 & 454 & 6 \\ 462 & 5 \\ .72 & 462 & 2 \\ 466 & 3 \\ 462 & 5 \\ .72 & 462 & 2 \\ 466 & 3 \\ 462 & 1 \\ 458 & 0 \\ .73 & 462 & 6 \\ 458 & 3 \\ 460 & 5 \\ 457 & 3 \\ 450 & 0 \\ 447 \end{array}$	446 443 446 4443 4446 4445 4448 4450 44522 4446 44522 4446 44522 4446 44522 4446 44522 4446 44522 4446 44522 4446 44522 4446 44522 4446 44522 4446 44522 4446 445227 44526 445227 44526 445227 44526 445227 44527777777777	

Heart rate during successive one minute periods of Session $\boldsymbol{6}$

	umber <u>2 E</u>			at Number <u>3 E</u>	
Conditioning Heart rate		Conditioning Heart rate			
decrease	s reinfor	ced	incr	eases reinford	ed
472 478 482 490 486 .69	475 0 475 0 475 0 474 1 474 2	486 484 483 479 478	524 524 520 516 515	467 1 .72 474 4 467 3 461 0 454 0	479 481 476 476 479
481 480 478 479 474 470 474	474 2 476 0 476 0 477 0 474 2 474 2 475 2	478 479 478 481 478 478 479 478 481	514 512 511 508 508 505 504	463 4 474 6 .71 469 1 470 2 468 0 .72 458 0 .73 455 0 460 2	478 478 480 480 478 478 478 480
483 482 485	479 0 481 0 480 0 479 1 479 1 478 2 480 0 480 1	481 480 478 476 476 476 477 477 475	502 501 496 499 497 495 494	460 2 458 1 455 1 460 2 462 5 461 4 464 6 .72 468 3	480 482 480 480 481 481 480 478
477 474 467 464 461 463 464	478 3 478 3 481 0 481 0 481 0 481 0 484 0 484 0 484 0 483 1	477 474 474 471 471 473 471 471 473	492 490 490 487 485 485 488 488	467 3 466 3 470 6 .71 470 4 470 4 470 4 470 4 470 4 470 4	484 482 483 479 478 476 478 478 476
466 470 .70 473 3 473 3 472 4 474 1 474 1 474 1	482 1 481 2 481 2 483 1 483 1 482 2 484 485		486 484 .70 484 5 .69 483 0 .70 480 2 477 .71 474 4 472 2	470 4 469 2 469 2 468 1 468 1 468 1 468 1 478 482	

APPENDIX D

Tables XXI to XXIII showing heart rates for rat number 5 Eduring successive one minute periods of one adaptation session, three conditioning sessions, and two reversal conditioning sessions. Criterion changes and number of brain stimulation reinforcements during conditioning periods.

TABLE XXI

Session l			Session 2	
Adaptation	Conditioning Heart rate increases reinforced			
Adaptation 484 457 487 459 483 460 475 458 471 454 463 455 465 452 463 451 466 453 466 456 466 452 465 450 466 452 465 450 466 452 465 450 464 449 452 457 458 461 454 459 458 456 458 456 462 456 462 456 462 456 462 456 464 461 466 462 441 462 438 461 444 464 449 467 455 464 441 464 449 467 455 464	460 460 458 457 455 455 455 455 455 455 455 455 455			

Rat Number <u>5 E</u>. Heart rate during successive one minute periods of Sessions 1 and 2

TABLE XXII

Rat Number <u>5 E</u>. Heart rate during successive one minute periods of Sessions 3 and 4

TABLE XXIII

Deee	Session 5 Revenuel Conditioning Meant			Session 6 Reversal Conditioning Heart		
	Reversal Conditioning Heart rate decreases reinforced			rate decreases reinforced		
rate decreases reinforced rate decreases reinfo					lordeu	
459	468 3	468	465	474 l	475	
459	469 1	469	462	474 l	475	
457	468 3	468	467	475 0	474	
457	470 2	468	465	475 0	474	
454	468 3	464	464	.69 474 2	470	
456	468 3	463	464	474 2	471	
451	471 0	465	465	470 4	473	
453	469 3	466	461	470 4	471	
454	466 3	465	460	476 0	469	
451	.74 462 2	463	462	476 0	468	
453	464 0	460	459	476 0	467	
454	464 O	459	459	472 1	468	
453	465 0	458	458	472 3	464	
454	465 0	456	457	474 2	464	
453	.72 466 0	455	457	.68 479 1	465	
454	466 0	457	458	479 l	464	
451	466 0	454	458	479 l	461	
454	468 O	453	458	476 3	460	
453	.70 471 0	451	456	476 3	459	
454	471 l	454	460	475 4	458	
451	472 l	451	454	478 l	458	
450	472 l	450	456	478 l	457	
449	469 3	451	457	476 3	458	
450	471 l	451	455	478 O	458	
450	.69 468 0	452	458	476 3	458	
450	468 O	451	456	481 O	458	
452	471 O	451	456	481 O	460	
448	467 l	450	456	481 O	456	
450	467 l		455	.67 478 3		
450	471 O		457	478 3		
.70 475	0 466 3		.71 461 0	480 l		
475			463 0	478 3		
471			465 0	480 l		
473			.70 473 3	478 3		
471			473 3	476		
.71 468	4 469		472 4	476		

Rat Number $5 \pm 10^{\circ}$ Heart rate during successive one minute periods of Sessions 5 and 6

David Hothersall was born in Longridge, England on August 25, 1940. He received his early education at Bishop's Court in Freshfield, Lancashire. In 1952 he emigrated with his parents to Johannesburg in the Republic of South Africa and continued his schooling at Christian Brothers' College in Boksburg, matriculating in 1957. In 1959 he entered the University of the Witwatersrand in Johannesburg and graduated in 1961 with a Bachelor of Arts degree with a first class pass in Psychology. He was accepted into the Department of Psychology's honors program and in 1963 received an Honors degree with a first class pass in Psychology. In 1964 he was awarded a Master's degree with a distinction in Psychology.

VITA

A post-graduate scholarship awarded by the Council of the University of the Witwatersrand allowed him to return to England and to work with Dr. Jasper Brener in the laboratory of Dr. Harry Hurwitz at Birkbeck College in the University of London. Together with Doctors Brener and Hurwitz he emigrated to the United States of America in 1965 and entered the University of Tennessee as a graduate student in the Department of Psychology in September 1965. In 1966 he was awarded a pre-doctoral research fellowship by the National Institutes of Health.

David Hothersall is married and has one daughter born in Knoxville on September 4, 1967.

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