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Polynucleotide encoding a gene conferring resistance to *Bacillus thuringiensis* toxins

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Heckel et al.

(10) **Patent No.:** **US 7,029,851 B2**

(45) **Date of Patent:** **Apr. 18, 2006**

(54) **POLYNUCLEOTIDE ENCODING A GENE CONFERRING RESISTANCE TO *BACILLUS THURINGIENSIS* TOXINS**

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(73) Assignee: **Clemson University**, Clemson, SC (US)

(*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 528 days.

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Related U.S. Application Data

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(51) **Int. Cl.**
C12Q 1/68 (2006.01)
C07H 21/02 (2006.01)
C07H 21/04 (2006.01)

(52) **U.S. Cl.** **435/6; 536/23.1; 536/24.3**

(58) **Field of Classification Search** **435/6; 530/350; 536/23.1, 24.3**

See application file for complete search history.

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(57) **ABSTRACT**

Nucleic acid (DNA) probes are provided which will specifically identify a gene for resistance of Bt in insect populations. Sequences are identified associated with the onset of resistance to *Bacillus thuringiensis* toxins. The sequences are used as probes to monitor the presence of acquired insect resistance associated with transgenic crops.

7 Claims, 5 Drawing Sheets

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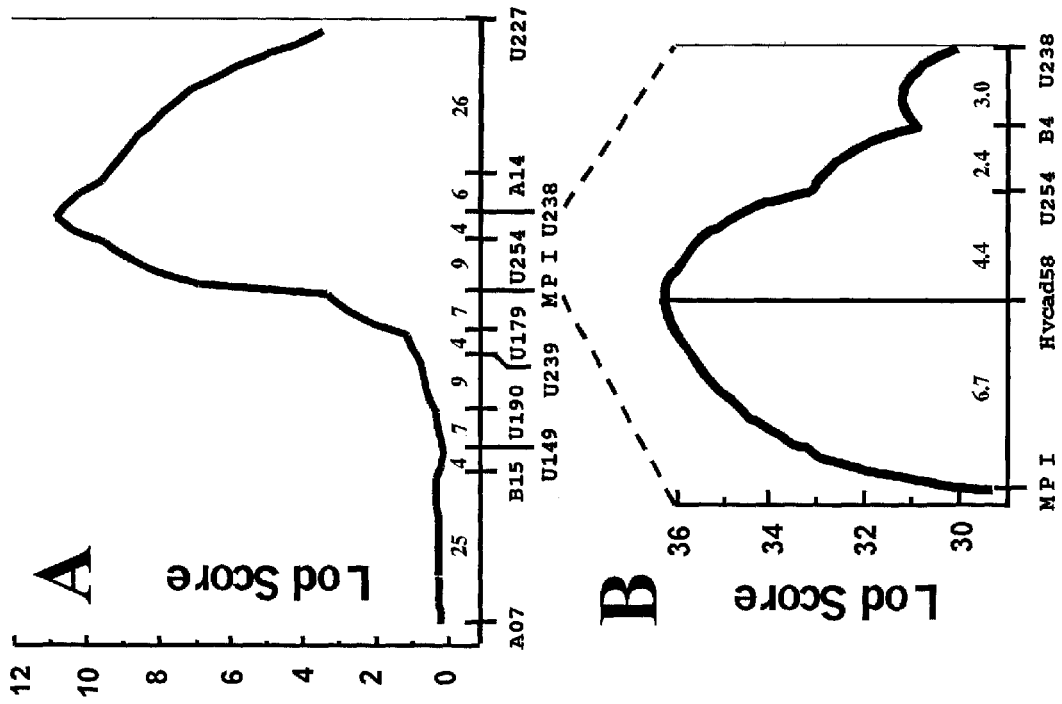


Fig. 1

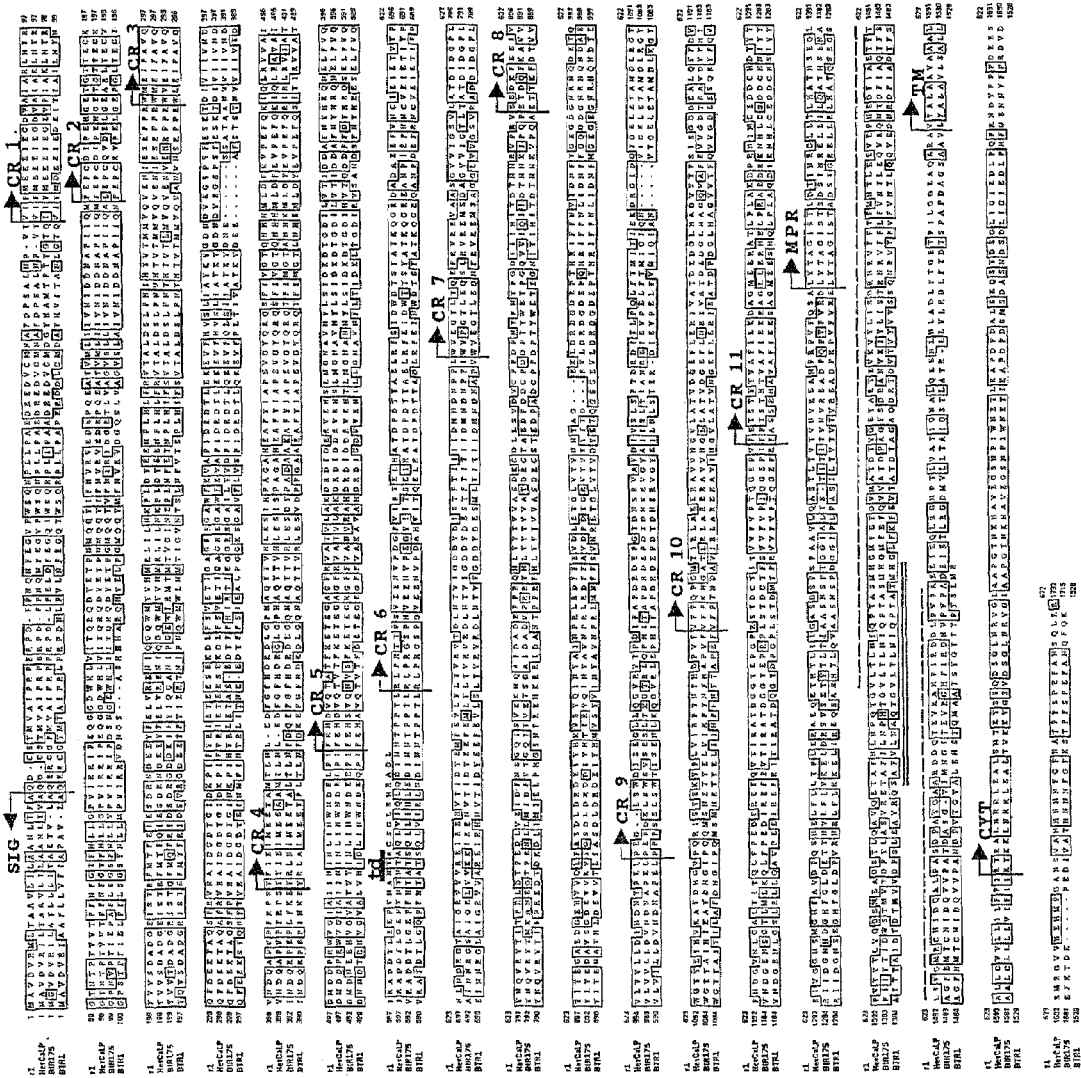


Fig. 2

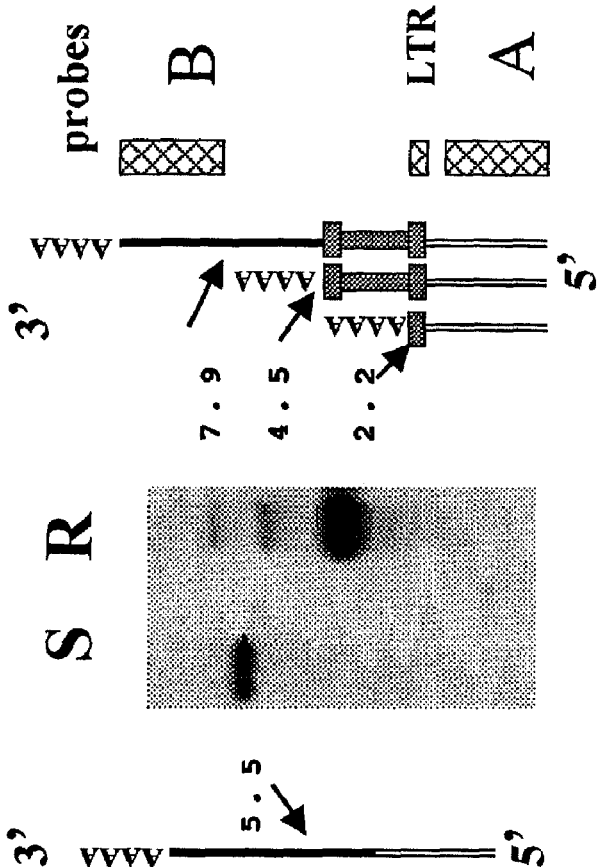
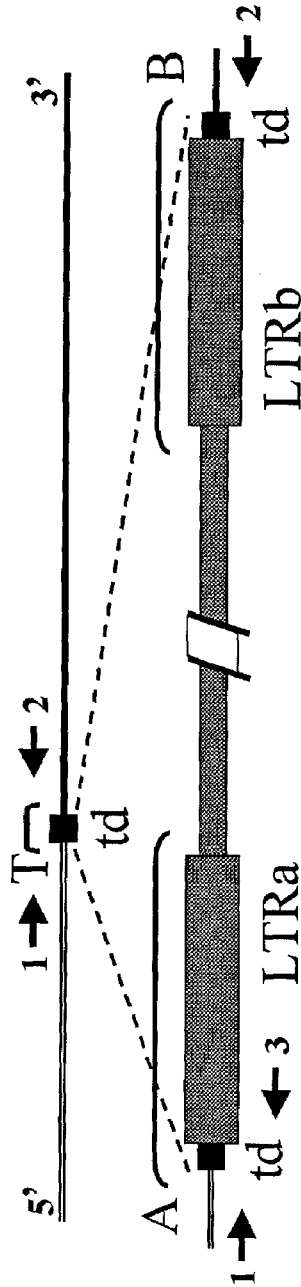


Fig. 3



T Y N T N T A Q L V
 TACAAACACCAACTGCCCCAACTGGTG
 |||||
A TACRACACCAACTGCCCTGTCCGGACTGTACATCGCGCGCTATGAGTGGCGGACACACGTCATCGTGGCCCCCACCTAA
 GCTGGGC
 |||||
B gcttcaaccgggggataTGTCGGGACTGTACATCGCGCGCTATGAGTGGCGGACACACGTCATCGTGGCCCCCACCTAAGGTG
 GGC
 |||||
A CCTCACCATACGGCCGGACCCCGGACACTCGCTCAGGGACCCCGGTGCGGCATACACGCCGCGGCAACCGCGGATTTTCTCTTG
 T-ACATA-CTT
 |||||
B CCTCACCATACGGCCGGACCCCGGACACTCGCTCATGACCCCGGTCCGCATACACGCCGCGGCAACCGCGGATCTACTCTTG
 TCACCTATCTA
 |||||
T CAAATACAGTCTTCT-
 TTGCAATCGAAGTTTCATGTGAACCGCCGAGACGATCATCTCCTACATCTGGACCTTGGCGCTCAAGcatggccccctggcaa
 |||||
T TAATACAGTCTTCTACTTTGAAATGGAATGTTTATTGAAACCGCGGAGACCACTACACCTGCACTCGCGCCCTCAAACTGCCCC
 AACTGGTG
 |||||
 TACAAACCAACTGCCAACTGGTG
 Y N T N T A Q L V

Fig. 4

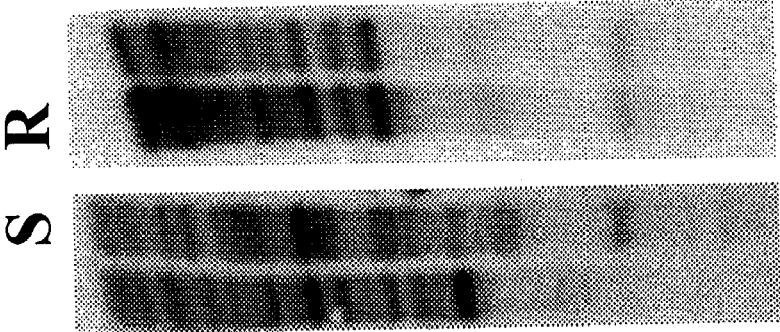


Fig. 5

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**POLYNUCLEOTIDE ENCODING A GENE
CONFERRING RESISTANCE TO *BACILLUS
THURINGIENSIS* TOXINS**

RELATED APPLICATIONS

This application claims the benefit of U.S. provisional application having Ser. No. 60/276,180 filed on Mar. 15, 2001, and which is incorporated herein by reference.

STATEMENT AS TO RIGHTS TO INVENTIONS
MADE UNDER FEDERALLY SPONSORED
RESEARCH AND DEVELOPMENT

The United States Government may have rights to this invention under the terms of a sponsored research agreement by the National Science Foundation, grant number MCB-9816056.

FIELD OF THE INVENTION

This invention is directed towards the occurrence and identification of pesticide tolerance of certain insects. The invention makes use of specific polynucleotide sequences associated with the onset of resistance to *Bacillus thuringiensis* toxins which are used as probes to monitor the presence of acquired insect resistance associated with transgenic crops. The specific polynucleotide sequences are also used to monitor changes in the frequencies of alleles which confer the resistance to the toxins.

BACKGROUND OF THE INVENTION

The bacterium *Bacillus thuringiensis* (Bt) contains genes encoding insecticidal proteins. Bt proteins are toxic when ingested by susceptible insect larvae. The protein attacks the insect's midgut, causes cessation of feeding, and eventually kills the insect. Bt toxins have been produced as fermentation products of Bt cultures and used in spray formulations for crop protection. Bt genes have also been used commercially to transform crop plants; these transgenic crop plants' cells then produce the insecticidal protein which attacks susceptible insects that attempt to feed on the plant.

The general mode of action of Bt toxins is well known in the art and is described for example by Rajamohan F, Lee M K, Dean D H (1998) *Progress in Nucleic Acid Research and Molecular Biology* 60: 1-27. The protein produced by the bacterium is usually a protoxin, which itself is not toxic until it is proteolytically cleaved by the insect's own proteases. The smaller protein resulting from proteolysis is the active toxin. This toxin diffuses through the peritrophic membrane to the midgut epithelium, where it binds to one or more sites in the membrane. This initial binding step may be reversible, but eventually the toxin becomes irreversibly bound to the membrane. A conformational change occurs in the toxin, whereby membrane-spanning alpha helices are inserted into the membrane, where they aggregate and form pores. These pores disrupt the normal osmotic balance of the epithelial cells. The cells swell and lyse, leading to destruction of the midgut epithelial cell layer and eventual death of the insect.

The initial binding step is believed to be necessary for toxin action; consequently there have been many studies on binding interactions of Bt toxins and components of the midgut, described for example by Pietrantonio P V and Gill S S (1996) in *Biology of the Insect Midgut*, Chapman & Hall, London, pp 345-372. Techniques used to study binding often start with the isolation of a brush border membrane

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vesicles (BBMVs) from the microvillar portion of columnar epithelial cells. Binding to BBMVs in suspension can be measured using labeled toxin. Alternatively, proteins can be isolated from BBMVs, separated by denaturing electrophoresis conditions, transferred to membranes, and probed with toxin. In addition, histological sections of insect midguts can be prepared and binding of labeled toxin can be visualized using microscopy.

Binding of Bt toxins to specific insect proteins can also be measured. Several proteins that interact with Bt toxins are well known in the art. Aminopeptidases exist in many different forms in insect midguts, and many of them have been shown to bind Bt toxins (Knight P J K, Knowles B H, Ellar D J (1995) *Journal of Biological Chemistry* 270 (30): 17765-17770; Gill S S, Cowles E A, Francis V (1995) *Journal of Biological Chemistry* 270 (45): 27277-27282; Luo K, Sangadala S, Masson L, Mazza A, Brousseau R, Adang M J (1997) *Insect Biochemistry and Molecular Biology* 27 (8-9): 735-743). Members of the cadherin superfamily have also been shown to bind Bt toxins (Vadlamudi R K, Weber E, Ji I H, Ji T H, and Bulla L A (1995) *Journal of Biological Chemistry* 270: 5490-5494; and Nagamatsu Y, Koike T, Sasaki K, Yoshimoto A, Furukawa Y, (1999) *FEBS Letters* 460: 385-390). Phosphatase enzymes have also been implicated in Bt toxin binding (Sangadala S, Walters F S, English L H, Adang M J, (1994) *Journal of Biological Chemistry* 269 (13): 10088-10092). TPP-75, an elastase-like serine protease, binds to certain Bt toxins and causes them to precipitate (Milne R E, Pang A S D, Kaplan H (1995) *Insect Biochemistry and Molecular Biology* 25 (10): 1101-1114). BTR-270, a peptidoglycan, binds CryIA toxins with high affinity (Valaitis A P, Jenkins J L, Lee M K, Dean D H, Garner K J (2001) *Archives of Insect Biochemistry and Physiology* 46 (4): 186-200). Bt toxins have also been shown to bind to nonprotein components of midgut epithelial membranes. Glycolipids from *Manduca sexta* have been shown to bind CryIA toxins using an overlay technique (Garczynski S F and Adang M J (2000) in *Entomopathogenic Bacteria: From Laboratory to Field Application*, Kluwer Academic Publishers, pp 181-197). Neutral lipids are involved in Bt toxin binding to *Manduca sexta* brush border membranes (Sangadala S, Azadi P, Carlson R, Adang M J (2001) *Insect Biochemistry and Molecular Biology* 32 (1): 97-107). Neutral glycolipids, especially hexa- and trisaccharylceramides, are implicated in CryIA toxin binding in diamondback moth (Kumaraswami N S, Maruyama T, Kurabe S, Kishimoto T, Mitsui T, Hori H, (2001) *Comparative Biochemistry and Physiology B-Biochemistry & Molecular Biology* 129 (1): 173-183).

The relationship between binding targets for Bt-toxins and susceptibility or resistance to Bt is very complicated and not completely understood at the present time. Several hundred strains of *Bacillus thuringiensis* exist, with considerable specificity toward various groups of insects. Co-evolution between the insects and Bt has resulted in specificity of the interaction between Bt-toxin and the membranes of insect gut cells. The Bt-toxin of a particular strain of *Bacillus thuringiensis* may bind to the gut of some insect larvae but not to others. Thus, the Bt-toxins may have a high specificity for a small number of insect pest species while having no significant activity against beneficial insects, wildlife, or humans.

Plants transformed to carry Bt genes and express insecticidal proteins are known in the art and include potato, cotton, tomato, corn, tobacco, lettuce, and canola. Transformed plants are known in the art as reflected in U.S. Pat. Nos. 5,608,142; 5,495,071; 5,349,124; and 5,254,799, the

specifications of which are incorporated in their entirety herein by reference. The use of genetically engineered plants is designed to reduce the use of broad spectrum insecticides.

There is concern that resistance may evolve to Bt toxins, whether they are applied to plants in spray formulations or the plants are genetically engineered to express them. The development of resistance to Bt-toxin expressing crops may also result in resistance to commercial formulations of fermented strains of Bt such as DIPEL® (Abbott Laboratories).

Rapid, reliable methods for broad screening to distinguish and detect the development of Bt resistance in populations of insects are needed. Heretofore, all methods require living or fresh-frozen insect larvae or preparations derived from them. The simplest methods employ bioassays on living insects, in which survivorship or larval metabolic rates are determined over time following a diet containing a specified concentration of a Bt-toxin. One such bioassay based on reduced metabolic rates after exposure to low doses of toxin mixed into artificial diet is discussed in U.S. Pat. No. 6,060,039 to Roe et al. which is incorporated herein by reference. Other bioassays are based on survival after exposure to a single, high diagnostic dose of toxin (for example, Diaz-Gomez O, Rodriguez J C, Shelton A M, Lagunes-T A, Bujanos-M R, (2000) *Journal of Economic Entomology* 93 (3): 963-970).

In principle, these bioassay methods can detect resistance no matter what its biochemical or physiological mechanism is. However, they require living, healthy larvae for their use, which are not always available. A more severe limitation on these methods is that, depending on the frequency of resistance genes in the populations, millions of individuals may need to be tested to detect a single resistant larva. High-level resistance to Bt is usually recessive, which means that an insect must have two copies of the resistance gene to be resistant. To a very good approximation, the frequency of such homozygous individuals is given by the square of the frequency of the resistance allele. For example, if the resistance allele frequency is one in a thousand, the frequency of homozygous resistant individuals is one in a million. In this example, more than a million larvae would need to be screened to detect resistance.

One solution to this problem is to develop methods for detecting the resistance genes directly. In the example just given, the frequency of heterozygous carriers of one copy of the resistance allele is $2 \times 0.001 \times 0.999$ or approximately 2 in a thousand. When resistance is recessive, these individuals would not be identified by bioassay because the one resistance allele they carry is not enough to make them fully resistant. But a direct, DNA-based method for detecting the resistance allele would identify these individuals, and sample sizes on the order of a thousand, rather than a million, would suffice.

The main limitation to developing DNA-based methods for detecting resistance alleles is that, up to now, the identity of resistance-causing genes has been unknown. In spite of much work on Bt toxin mode of action, prior to the invention described herein there has not been a demonstration of which genes, when mutated, actually cause resistance. Accordingly, there is room for variation and improvement in the art of screening assays useful in detecting the presence of genes conferring Bt resistance in natural populations.

SUMMARY OF THE INVENTION

It is one aspect of one of the present inventions to provide a genetic probe to identify and monitor resistance for the

Bt-toxin in target insect populations. One such insect pest is the tobacco budworm (*Heliothis virescens*) which is a major economic pest of cotton.

It is yet another aspect of one of the present inventions to develop a DNA probe and assay protocol which distinguishes between the conditions of homozygotes and heterozygotes with respect to resistance to Bt in populations of *Heliothis virescens* and other insects.

It is yet another aspect of one of the present inventions to provide a process and useful sequences in which nucleotide probes are used to monitor the presence of acquired insect resistance associated with a transgenic crop.

It is yet another aspect of one of the present inventions to provide a process and useful nucleotide sequences which are used to monitor population changes in the frequency of alleles which are associated with the resistance to Bt toxin.

These and other features, aspects, and advantages of the present invention will become better understood with reference to the following description and appended claims.

BRIEF DESCRIPTION OF THE DRAWINGS

A full and enabling disclosure of the present invention, including the best mode thereof, to one of ordinary skill in the art, is set forth more particularly in the remainder of the specification, including reference to the accompanying drawings.

FIG. 1 is a QTL map of the Cry1Ac resistance trait on linkage group 9 of *Heliothis virescens*.

FIG. 2 is a conceptual translation of HevCaLP (s1 allele and r1 allele) in alignment with BmBtR175 of *Bombyx mori* and BtR1 of *Manduca sexta*.

FIG. 3 is a northern analysis of mRNA isolated from susceptible and resistant strains following probing with the gene sequences set forth herein.

FIG. 4 sets forth the insertion point of the Hel-1 element in the r1 allele of HevCaLP.

FIG. 5 shows the multi-copy occurrence of Hel-1 in genomic DNA of resistant and susceptible strains of *Heliothis virescens*.

BRIEF DESCRIPTION OF THE SEQUENCE LISTINGS

The accompanying sequence ID listings are identified below. The sequence listings appear following the claims and are incorporated herein by reference.

The first sequence 1 identifies SEQ ID NO: 1 which is the DNA sequence of the susceptible allele s1 of HevCaLP.

Sequence 2 is the protein sequence SEQ ID NO: 2 of a conceptual translation of allele s1 as used in the protein alignment to *Bombyx* and *Manduca*.

Sequence 3 is the DNA sequence of SEQ ID NO: 3 which is the resistant allele r1 of HevCaLP, including the Hel-1 insert and the duplicated target sequences.

Sequence 4 is the DNA insert identified as SEQ ID NO: 4 for the Hel-1 insert which does not include duplicated target sequences.

Sequence 5, having SEQ ID NO: 5, is a DNA sequence corresponding to the left LTR of the Hel-1 insert.

Sequence 6, having SEQ ID NO: 6, is a DNA sequence corresponding to the right LTR of the Hel-1 insert.

Sequence 7, having SEQ ID NO: 7, is a DNA sequence of primer F1 corresponding to bases 1982 to 2001 of SEQ ID NO: 3.

Sequence 8, having SEQ ID NO: 8, is a DNA sequence corresponding to primer R2 consisting of the reverse complement of bases 4322 to 4351 of SEQ ID NO: 3.

Sequence 9, having SEQ ID NO: 9, is a DNA sequence corresponding to primer R3 consisting of the reverse complement of bases 2029 to 2052 of SEQ ID NO: 3.

DESCRIPTION OF THE PREFERRED EMBODIMENT

Reference now will be made in detail to the embodiments of the invention, one or more examples of which are set forth below. Each example is provided by way of explanation of the invention, not limitation of the invention. In fact, it will be apparent to those skilled in the art that various modifications and variations can be made in the present invention without departing from the scope or spirit of the invention. For instance, features illustrated or described as part of one embodiment, can be used on another embodiment to yield a still further embodiment. Thus, it is intended that the present invention cover such modifications and variations as come within the scope of the appended claims and their equivalents. Other objects, features, and aspects of the present invention are disclosed in the following detailed description. It is to be understood by one of ordinary skill in the art that the present discussion is a description of exemplary embodiments only and is not intended as limiting the broader aspects of the present invention, which broader aspects are embodied in the exemplary constructions.

In describing the various figures herein, the same reference numbers are used throughout to describe the same material, apparatus or process pathway. To avoid redundancy, detailed descriptions of much of the apparatus once described in relation to a figure is not repeated in the descriptions of subsequent figures, although such apparatus or process is labeled with the same reference numbers.

Applicants' protocols and procedures may be found in reference to "Identification of a Gene Associated with Bt resistance in *Heliothis virescens*" which was published in Science, volume 293, pp 857-860, on Aug. 3, 2001; and which is incorporated herein by reference.

A resistant strain of *Heliothis virescens* was previously developed in the laboratory by selection using artificial diet containing various concentrations of Bt toxin (Gould F, Anderson A, Reynolds A, Bumgarner L, Moar W (1995) *Journal of Economic Entomology* 88 (6): 1545-1559). The strain, named YHD2, is 10,000 fold more resistant to the toxin Cry1Ac and is conditioned in a large part by a single recessive gene named BtR-4 which is located in linkage group 9 of *H. virescens*. The initial localization of the resistance gene BtR4 has been reported in the Applicants' prior publication (Heckel D G, Gahan L C, Gould F, Anderson A (1997) *Journal of Economic Entomology* 90: 75-86) and which is incorporated herein by reference.

Further localization of BtR-4 to a particular region of linkage group 9 was carried out using a total of 11 polymorphic markers spanning a length of 105 cM. The markers were scored on a segregating backcross family derived from YHD2 females crossed with susceptible males. The linkage group was scanned for quantitative trait loci (QTLs) conferring Bt resistance following the methods of Lander, E S and Botstein D (1989) *Genetics* 121: 185-193. A single, highly significant peak of the log-likelihood function indicated that the BtR-4 resistance gene is located between A14 and MPI as set forth in FIG. 1.

The cadherin superfamily was chosen as a candidate for BtR-4. Partially degenerate oligonucleotide primers Btmp5

and Btmp8 as shown in Table 1 were designed based on published sequence of the BtR175 gene from *Bombyx mori* (GenBank Accession No AB026260, described by Nagamatsu Y, Toda S, Koike T, Miyoshi Y, Shigematsu S, Kogure M (1998) *Bioscience, Biotechnology and Biochemistry* 62 (4): 727-734). These primers were used in the polymerase chain reaction (PCR) with cDNA prepared from midgut mRNA of larval *Heliothis virescens*. A PCR product of 334 basepairs designated Hvcad58 was amplified, cloned and sequenced using conventional methodology well-known to those skilled in the art. The sequence of Hvcad58 corresponds to bases 4279 to 4612 of SEQ ID NO: 1.

Radiolabeled Hvcad58 was used to probe Southern filters made from additional segregating backcross families for further mapping on linkage group 9. Finer scale QTL mapping in this region using 268 backcross progeny yielded a single peak of the log-likelihood function directly above the map location of Hvcad58 (FIG. 1). The data clearly indicates that the gene containing Hvcad58 is a strong candidate for the BTR-4 resistance gene.

The Hvcad58 probe was used to screen midgut cDNA libraries made from resistant (YHD2) and susceptible strains of *Heliothis virescens*. Clones recovered from these libraries were sequenced and used to design additional primers to amplify the full-length coding sequence from susceptible cDNA. In addition to the cDNA methods, a five-prime RACE (rapid amplification of cDNA ends) technique was used to complete the full sequence.

The sequencing yielded one transcript (s1) cloned from a susceptible strain as given in SEQ ID NO: 1. Conceptual translation of this transcript produced a protein product (that we have named HevCaLP, *Heliothis virescens* cadherin-like protein) of 1732 amino acids as given in SEQ ID NO: 2. HevCaLP is 70% identical to the BtR175 protein, sharing a signal sequence at the amino terminus, 11 extra-cellular cadherin-type repeats, a non-cadherin proximal membrane region, a transmembrane region, and a highly conserved cytoplasmic domain at the carboxy terminus as shown in FIG. 2. It shows somewhat less similarity to the BT-R1 protein from *Manduca sexta*, as given in GenBank Accession No. AAB33758 and reported by Vadlamudi R K, Weber E, Ji I H, Ji T H, and Bulla L A (1995) *Journal of Biological Chemistry* 270: 5490-5494. The transmembrane and cytoplasmic domains are absent from that sequence of BT-R1.

Expression of the mRNA encoding HevCaLP in susceptible and resistant larval midguts was studied using northern analysis and sequencing of clones from the resistant library. As shown in FIG. 3, susceptible larvae show a single transcript of 5.5 kb. YHD2 larvae show three transcripts. The sequence of the rarest (7.9 kb) is denoted as the r1 allele, and given as set forth in SEQ ID NO: 3. It is similar to the susceptible transcript except for a 2.3 kb insert denoted as Hel-1 as given in the accompanying SEQ ID NO: 4. Hel-1 shows several hallmarks of the LTR-type retrotransposons. Hel-1 has an approximately 255 nucleotide long terminal repeat (LTR) sequence at both ends and an unrelated sequence in the middle. The left LTR sequence, LTRa, is given in SEQ ID NO: 5 and the right LTR sequence, LTRb, is given in SEQ ID NO: 6. Hel-1 is flanked by an 8-nt duplication of the host sequence AACTGACC, as shown in FIG. 4. The transcript of intermediate abundance (4.4 kb) is an abbreviated form, truncated at the second LTR of Hel-1 by a poly-A tail. The third, highly abundant transcript (2.1 kb), is truncated at the first LTR of Hel-1 by a poly-A tail.

Because of an in-frame stop codon 30 bases into the first LTR of Hel-1, conceptual translation of the three different YHD2 transcripts produces the same truncated 622-aa pro-

tein (as shown in the translation of the r1 allele in FIG. 2). Multiple stop codons in all three reading frames of the LTR follow the initial stop codon, preventing translation of a larger protein containing the carboxy-terminus of HevCaLP. Thus, the predicted protein product of the YHD2 r1 allele (if one is produced) would possess the same signal sequence as HevCaLP (possibly directing its secretion into the midgut lumen) but no predicted transmembrane domain or toxin-binding region.

Genomic Southern blots probed with the LTR region of Hel-1 show that it occurs with a copy number of 10-15 in both YHD2 and susceptible insects (FIG. 5). Insertion of this Hel-1 element into the gene encoding HevCaLP has created the novel, knockout r1 allele which confers resistance when homozygous (present in two copies in an individual insect). This insertion event could have occurred in the laboratory during the Bt-resistance selection protocol that produced YHD2, or may already have been present in the field-collected founders of the selection line. Thus it is now evident that a DNA-based method for detecting Bt resistance in *Heliothis virescens* may be devised, based on detection of the specific insertion of the Hel-1 element into the gene encoding HevCaLP, producing the r1 allele.

To illustrate detection of the r1 allele, a PCR assay was designed using two primers flanking the insertion point (F1 and R2) and a third (R3) internal to the left LTR (FIG. 4). Primer F1 consists of bases 1982 to 2001 of SEQ ID NO: 3, 5' ATA CGA GCT GAC GAC ACG CTG GGA GA 3', primer R2 consists of the reverse complement of bases 4322 to 4351 of SEQ ID NO: 3, 5' TCT GAG CGT AGG AGG TGT GTT GAT GTC 3', and primer R3 consists of the reverse complement of bases 2029 to 2052 of SEQ ID NO: 3, 5' GCG CGA TGT GAC AGT CCG GM CAG 3'. Primers F1 and R3 produce a 71-bp band from the r1 allele. Primers F1 and R2 amplify a 99-bp band from s1 or other susceptible alleles lacking the Hel-1 insert. Heterozygotes produce both bands. This is a marked improvement on a conventional bioassay, which would not distinguish heterozygotes from homozygous susceptibles because the resistant allele is recessive. It also confirms that the resistant strain is fixed for the r1 allele, as all YHD2 individuals examined to date have the 71-bp band only. It will be evident to those skilled in the art that the detection method for the r1 allele is not limited to PCR with these specific primers, and that there are many other molecular methods of detecting the specific insertion of the Hel-1 element into the HevCaLP gene, based on the sequence information disclosed herein.

It is believed that the gene encoding HevCaLP is identical to BtR-4, the major resistance gene in YHD2. Recessivity of the resistant allele at BtR-4 is explained by Hel-1 inactivation of HevCaLP. HevCaLP functions as a "lethal target" of Bt-toxin, since two copies of the disrupted allele are required for 10,000-fold resistance. Heterozygotes still present a "lethal target" since they have one copy of the susceptible allele.

The normal physiological function of HevCaLP is unknown, although other members of the cadherin superfamily are involved in cell adhesion and signalling (T. Uemura (1998) *Cell* 93 (7): 1095-1098). Whatever its function, it is not essential for life, as YHD2 is viable and fertile under laboratory conditions, despite being a "natural knockout" strain for HevCaLP. Whether its absence confers a fitness disadvantage in the field has important implications for resistance management, and this question can now be addressed with the information developed here. Target-site resistance to other insecticides usually involves modification but not knockout of the target, which is generally essential

for life (e.g., acetylcholinesterase for organophosphates, sodium channel for pyrethroids, GABA receptor for cyclo-dienes) (French-Constant R H, Pittendrigh B, Vaughan A, Anthony N (1998) *Philosophical Transactions of the Royal Society of London Series B-Biological Sciences* 353 (1376): 1685-1693.). However, methoprene resistance in *Drosophila melanogaster* provides another example of resistance by gene inactivation (Wilson T G & Ashok M, (1998) *Proceedings of the National Academy of Sciences of the USA* 95 (24): 14040-14044).

The present invention now makes possible the application of molecular methods to Bt-resistance monitoring. We previously estimated the frequency of YHD2-type resistant alleles in field populations of *Heliothis virescens* prior to widespread planting of transgenic Bt-cotton to be 0.002 (Gould F, Anderson A, Jones A, Sumerford D, Heckel D G, Lopez J, Micinski S, Leonard R, Laster M (1997) *Proceedings of the National Academy of Sciences of the USA* 94 (8): 3519-3523). This labor-intensive, bioassay-based estimate was derived by testing progeny of more than 1,000 field-caught males mated to YHD2 females, for alleles which would confer resistance when heterozygous with r1. Our results now suggest that this estimate covers the entire class of HevCaLP knockouts regardless of the nature of the molecular lesion, as well as other mutants preventing any expressed HevCaLP from functioning as a toxic target. Development of efficient DNA-based methods to detect these other types of mutants at BtR4 should be a high priority and is now possible with the methods described herein.

Only by monitoring allele frequencies at resistance genes like BtR-4 will it be possible to verify that the high-dose/refuge resistance management strategy for Bt-cotton mandated by the US Environmental Protection Agency (EPA) is actually working to keep resistance allele levels low. The present invention affords a new method of complying with EPA regulations which require monitoring resistance levels in *Heliothis virescens*. The present invention provides a nucleic acid probe that will specifically identify genes for resistance in field populations. Further, the probes and protocols set forth herein provide for a method of monitoring the population of homozygous and heterozygous resistant individuals in field populations.

Bt resistance in *Heliothis virescens* caused by other types of mutations that inactivate the HevCaLP gene product may also be screened for using the information provided herein. Such methods may include obtaining portions of the gene or its homologues by cDNA cloning or the polymerase chain reaction, determining the DNA sequence by standard methods, and examining the sequence for the occurrence mutations that may include nucleotide substitution, insertions, or deletions. Such mutations may affect protein sequences encoded by the gene by causing amino acid substitutions, insertions, or deletions as well as incorrect intron splicing, premature chain termination due to nonsense mutations, or errors in the normal initiation or termination of the transcription or translation.

By way of example, DNA or RNA isolated from individual *Heliothis virescens* is used as the template for PCR using primers specifically designed from SEQ ID NO: 1. The PCR products are directly sequenced, or cloned and sequenced, using standard methods. The sequences are examined using commercially available computer programs well known in the art, such as the Wisconsin Genetics Computer Group package. Mutations, such as individual nucleotide substitutions, insertions, or deletions; or insertions or deletions of several nucleotides, are detected by

comparison to SEQ ID NO: 1. Such mutations may alter the amino acid in the protein sequence, leading to reduced binding of Bt toxins to the HevCaLP gene product and thereby conferring resistance. Or such mutations may cause frameshifts or premature occurrence of stop codons, resulting in a truncated or absent protein that fails to bind to Bt toxins and thereby confers resistance.

In the course of this invention, an isolated nucleic acid molecule of the present invention includes a nucleic acid that is at least about 85%, preferably at least about 90%, and still more preferably at least about 95%, and even more preferably at least about 99% identical to the sequence of the susceptible allele s1 of HevCaLP. Additionally, any isolated polynucleotide or naturally occurring polynucleotide that hybridizes to the sequence set forth in SEQ ID NO: 1 at 60° C. in 1×SSC will have properties useful in carrying out the present invention.

Other embodiments of the present invention include isolated nucleic acid molecules that are at least about 85%, preferably at least about 90%, still more preferably at least about 95%, and even more preferably at least about 99%, identical to the sequences set forth in SEQ ID NO: 3 and SEQ ID NO: 4.

Bt resistance in other insect species may also be screened for using the same approach. These species may contain one or more genes homologous to the *Heliothis virescens* HevCaLP gene, whose products interact with Bt toxins. Resistance in these other species can be detected by obtaining the sequence of those genes, designing PCR primers, and amplifying and sequencing DNA from individual insects collected from the field or reared in the laboratory. Examination of the sequence for inactivating mutations as described herein will detect Bt resistance in those species. Representative sequences of HevCaLP homologues in other species and which may be used in the screening process described herein include the following:

- 1) *Manduca sexta* BT-R1, GenBank Accession No. I77078, U.S. Pat. No. 5,693,491 (SEQ ID NO: 1) and U.S. Pat. No. 6,007,981 (SEQ ID NO: 1);
- 2) *Bombyx mori* BtR175, GenBank Accession No. AB026260, described by Nagamatsu Y, Toda S, Koike T, Miyoshi Y, Shigematsu S, Kogure M (1998) *Bioscience, Biotechnology and Biochemistry* 62 (4): 727-734;
- 3) *Pectinophora gossypiella* BT-R2, GenBank Accession No. AX150183, Patent Application, International Publication No. WO01/34807 (SEQ ID NO: 1);
- 4) *Ostrinia nubilalis*, GenBank Accession No. AX147201, Patent application, International Publication No. WO 01/36639 (SEQ ID NO: 1);
- 5) *Helicoverpa zea*, GenBank Accession No. AX147203, Patent application, International Publication No. WO01/36639 (SEQ ID NO: 3);
- 6) *Spodoptera frugiperda*, GenBank Accession No. AX147205, Patent application, International Publication No. WO01/36639 (SEQ ID NO: 5); and
- 7) *Lymantria dispar* BTR-CAD, GenBank Accession No. AF317621.

The above identified sequences and the referenced publications are all incorporated herein by reference as is set forth in their entirety.

The current methodology includes detecting resistance to *Bacillus thuringiensis* endotoxin in insect populations by screening for mutations that alter the structure or function of a protein as set forth in SEQ ID NO: 2. For the purposes of screening protocols, it is believed that using the sequence set forth in SEQ ID NO: 2 may include homologues and other species which would display at least 60% similarity to the sequence set forth in SEQ ID NO: 2. More preferably, the sequence similarity is at least about 75%, preferably at least about 80%, more preferably at least about 85%, even more preferably at least about 90%, still more preferably at least about 95%, and even more preferably at least about 99% identical to the amino acid sequence set forth in SEQ. ID. NO: 2.

Several of the mutations in other species detected by this approach may not have an obvious effect of activating the HevCaLP homologue. In that case, evidence that the mutation confers resistance may be obtained by conducting a linkage analysis and mapping the gene as described herein for *Heliothis virescens*. For that purpose, a strain of the species of interest with the mutation is crossed with a wild-type strain, and the F1 hybrids are intercrossed or backcrossed to one of the parental strains. The F2 or backcross progeny are tested for resistance by any of the bioassay methods described previously and well known in the art, and DNA is isolated from each individual progeny. The DNA is analyzed for the presence of the mutation, using restriction fragment polymorphism analysis, allele-specific PCR, denaturing gradient gel electrophoresis, single-stranded conformation polymorphism, denaturing high-performance liquid chromatography, or any other mutation detection system well known in the art. Evidence that the mutation confers resistance is obtained from the correlation across progeny between presence of the mutation and presence of resistance.

A straightforward extension of this method of detecting Bt- resistance is to examine the DNA sequence of genes encoding other proteins that interact with Bt toxins, including but not limited to aminopeptidases, alkaline phosphatases, elastin-like serine proteases, and peptidoglycans.

All cited references, publications, and sequence listings set forth herein are incorporated by reference in their entirety.

These and other modifications and variations to the present invention may be practiced by those of ordinary skill in the art, without departing from the spirit and scope of the present invention. In addition, it should be understood that aspects of the various embodiments may be interchanged both in whole or in part. Furthermore, those of ordinary skill in the art will appreciate that the foregoing description is by way of example only, and is not intended to limit the invention.

TABLE 1

Primers Used in Determining the Structure of BtR4, the Cadherin-like Polynucleotide in *Heliothis virescens*

Bmtp 5	5'-GTR CTG ACK GTT AAY ATC GAG CCC ACK GC-3'
Smtp 8	5'-TAG GGG YAC RTT RTC SCG KAT GAA GTG KCC-3'
Hvtp05	5'-AGC CCA CTG CAT CTA TGC ACG GCA TGT TTG A-3'
Hvtp08	5'-CCT GAG TTG GGT CTG GTG GTC CCT GGC-3'

TABLE 1-continued

Primers Used in Determining the Structure of Btr4, the Cadherin-like Polynucleotide in <i>Heliothis virescens</i>	
GGp1	5'-TGT GGA GTC AGC TTC CAT AGA GTC TTG TAT GAG CGT GTA-3'
CGnotp2	5'-GAT ACG CGG CCG CAG GTC AGC AGA GCT CTG TTG ATG GTG TCG AGG GTG GAG A-3'
T7p1	5'-TAA GTT GGG TAA CGC GAG GGT TTT CCC AGT GAC-3'
T7p2	5'-GGC CAG TGA ATT GTA ATA CGA CTC ACT ATA GGG CG-3'
T3p1	5'-GAT AAC AAT TTC ACA CAG GAA ACA GCT ATG ACC ATG-3'
T3p2	5'-GAA ATT AAC CAC CCT TAA AGG GAA CAA AAG CTG GAG-3'
CGp3	5'-GGC ACG TTT TTT TCC ACT GAC GGG GTC GTG CG-3'
Cgnotp4	5'-GAT ACG CGG CCG CGG GCA GTC TGA GCG TAG GAG GTG TGT TGT TGA T-3'
RC36T4	5'-GAC GTG TGT TCG CCT GAT CCT AAC TAC T-3'
RC36cg5	5'-AGC CTC TTA AAT CCA TAG GGG TCT CCA G-3'
RC36cg5+	5'-CTG GAG ACC GCT ATG GAT TTA AGA-3'
SC3T6	5'-ATG TTC GAG GTG CTG TAC CTC ACC G-3'
SC3cg7	5'-ACA CGA ACA CAG GAT CGT GGA AGT T-3'
CGp5	5'-TGT ATC TTC TGG AAC TCC GGC ACT TCG AAG TC-3'
CGnotp6	5'-GAT ACG CGG CCG CAT GTG ATG GTT CTG CGT GCC GAC GAT GAA GGA CTG-3'
Sint1	5'-GCT AAG GAC CGG GAT ATT GAT GAT AGA GT-3'
Sint2	5'-CGT GCG GGG CAG TCT GAG AGT AG-3'
RUNI1	5'-CAT ACA CGA CCG CAC GCG CAA CG-3'
RUNI2	5'-TGA GCG CCG AGG TGC AGG TGT AGG-3'
Hvtp13	5'-CTG TAC ACA GCC GGC ATC TCC AC-3'
Hvtp14	5'-CTG GAA GTT GAG GGT CAG CAC TCC AGT-3'
Hvtp15	5'-AAC CGT CGT CTG GAA GCT CT-3'
Hvtp16	5'-TCT TCG ATG CCG ATC AGA TCC GAG TC-3'
Hvtp17	5'-GCG GCG CCG GGC ACC AAC AAG CA-3'
HvA11-RT	5'-AAT AGA TGC TCT TAC ATA ATA CGA GTA TCT TAC-3'
5'R5A4/8	5'-GAT ACG CGG CCG CGA GAA CTA TGA GAT GGC AGT CGA CGT GAG AAT A-3'
HvA11F1	5'-GAA CTA TGA GAT GGC AGT CGA CGT GAG AAT-3'
HvA11F3	5'-TTA ACT TTC GCG CAA GAT TGT TCC TAT ATG-3'
HvA11R2	5'-GAA CTC TGG GCT GAA GGG GGT AGC-3'
HvA11R4	5'-CCC GAA GTT RTT GTT ATG GTT TGC TAC TGA-3'
USTP01	5'-ATG GGC AAC GCA GTT AAC TAC CTG-3'
USTP02	5'-CAT CCT CGT GAC AAT CGA CGA TGC-3'
USTP03	5'-CAG ACA GAA CGA GCT CTT TGT GCA-3'
F771-5Ksp1	5'-GCC GTG CAG CAG TTC GAT GAG AAG-3'
F771-5Ksp2	5'-CTC CCA CTG TAT CAG TAG CCA TCA-3'
738-3.4Ksp1	5'-ACA ATC CTT CAG GGT TCG AGC CAT C-3'
738-3.4Ksp2	5'-GTA CAA GAG AAA ATC GCG CGT TGC GT-3'
738-3.4Ksp3	5'-CCT GAT CAA CTG GAA CGA TGA GCT G-3'
738-3.4Ksp4	5'-CCA AAG TCC ACG GGC GGT TGC GCA C-3'
738-3.8sp6	5'-GTG TAA CGT AGT GTG CTC GTG TAA TGC-3'
738-C10sp8-	5'-CCG TCT GAA ACA TGT CGA AGT CAT-3'
TBR01	5'-GAG ACT AGC ACC TAC ACG GTC GCT-3'
TBR02	5'-TCC AAC GAG CTG TTC CTG CTG ACG-3'
CR9TBR	5'-CAC TGT TAC TGT CAA TGT TCG AGA-3'
LTR-Pr1	5'-CAC ACG TCA TCG TGC GCC CCA CCT AAG CTG-3'
LTR-Pr2	5'-CTG GCG CGA CCT CAT AGG CCG GCG CGA TGT-3'
LTR-1.9Ksp1	5'-CGA ATC AGC TGA TTC ATT GTC GCT-3'
LTR-1.9Ksp2	5'-GTA GTG TGT GAT GTG ATC CAG-3'
Rint-Fwd1	5'-ATA CGA GCT GAC GAC ACG CTG GGA GAG CC-3'
Rint-Rev2	5'-TCT GAG CGT AGG AGG TGT GTT GAT GTC-3'
C36RESEQ-F	5'-CCC GGC ACC GAC AAC TCC-3'
C36RESEQ-R	5'-CTC CAT GGT CGT ATG CCT TGA CAT GTA-3'
pc11Fa	5'-GAG ATG GCA GTC GAC GTG AGA ATA CTG A-3'
pc12Fa	5'-CCC GTT TCG CCG TGT TCA GGA ATG TC-3'
pc12Ra	5'-TGG TAC CTC GGT AGT TAA GCC TGG CAA T-3'
pc13Fa	5'-GAA CAC GGC GAA ACG GGC ACC ACA GA-3'
pc13Ra	5'-TGC CAG GCT TAA CTA CCG AGG TAC CA-3'
pc14Fa	5'-AAC CCG CTG CAT TTG TTT AGA GTT ACA G-3'
pc14Ra	5'-CGA ACT GCT GCA CGG CGA AGA TCT CCA T-3'
pc15Ra	5'-TTC CTT CCA CGT CAT TGT CGC CAT ATT T-3'
RintS-F1	5'-ATA CGA GCT GAC GAC ACG CTG GGA GA-3'
RintS-R2	5'-TCT GAG CGT AGG AGG TGT GTT GAT GTC-3'
RintR-R3	5'-GCG CGA TGT GAC AGT CCG GAA CAG-3'
RintR-F4	5'-ACG CGC AAC GCG CGA TCT ACT CTT-3'

SEQUENCE LISTING

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<221> NAME/KEY: CDS

<222> LOCATION: (10)..(5205)

<400> SEQUENCE: 1

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gat ctc aac acg gtc att gga gac gac tat gat ata tca aca ttc acg Asp Leu Asn Thr Val Ile Gly Asp Asp Tyr Asp Ile Ser Thr Phe Thr 735 740 745 750	2259
atc att ata ata gac atg aac gac aac cct ccg ctg tgg gtg gaa ggc Ile Ile Ile Ile Asp Met Asn Asp Asn Pro Pro Leu Trp Val Glu Gly 755 760 765	2307
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gtt ata gga tcc gta ctc gcc act gat atc gac gga cct ctt tat aat Val Ile Gly Ser Val Leu Ala Thr Asp Ile Asp Gly Pro Leu Tyr Asn 785 790 795	2403
caa gtg ccg tat acc atc act cct aga tta gat act cca gaa gac cta Gln Val Arg Tyr Thr Ile Thr Pro Arg Leu Asp Thr Pro Glu Asp Leu 800 805 810	2451
gtg gag atc gac ttc aat tcg ggt cag atc tca gtg aag aag cac cag Val Glu Ile Asp Phe Asn Ser Gly Gln Ile Ser Val Lys Lys His Gln 815 820 825 830	2499
gcc atc gac gcg gac gag ccg ccg cgc cag cac ctc tac tac acc gtg Ala Ile Asp Ala Asp Glu Pro Pro Arg Gln His Leu Tyr Tyr Thr Val 835 840 845	2547
gtc gcc agc gac aag tgc gac ctg ctc tct gtc gac gtg tgt ccg cct Val Ala Ser Asp Lys Cys Asp Leu Leu Ser Val Asp Val Cys Pro Pro 850 855 860	2595
gat cct aac tac ttc aac aca ccg gga gac ata acg atc cac ata aca Asp Pro Asn Tyr Phe Asn Thr Pro Gly Asp Ile Thr Ile His Ile Thr 865 870 875	2643
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ctc ttc gcc agc gat ctg gat aga gat gaa atc tac cac aaa gtg agc Leu Phe Ala Ser Asp Leu Asp Arg Asp Glu Ile Tyr His Lys Val Ser 915 920 925			2787
tac cag atc aac tac gcg atc aac cct cgt ctc cgc gac ttc ttc gag Tyr Gln Ile Asn Tyr Ala Ile Asn Pro Arg Leu Arg Asp Phe Phe Glu 930 935 940			2835
gta gac ctg gag acc ggc ctg gtg tac gtc aac aac acg gcc ggg gag Val Asp Leu Glu Thr Gly Leu Val Tyr Val Asn Asn Thr Ala Gly Glu 945 950 955			2883
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gtt atc gat aac ttc tat ggg gaa gga gat ggc aac cgg aac cag gac Val Ile Asp Asn Phe Tyr Gly Glu Gly Asp Gly Asn Arg Asn Gln Asp 975 980 985 990			2979
gag aca caa gtg tta gtg gtg ctg ttg gac atc aac gac aac tat ccg Glu Thr Gln Val Leu Val Val Leu Leu Asp Ile Asn Asp Asn Tyr Pro 995 1000 1005			3027
gag ctg cct gag ggt ctc tca tgg gat atc tct gag gga ttg cta cag Glu Leu Pro Glu Gly Leu Ser Trp Asp Ile Ser Glu Gly Leu Leu Gln 1010 1015 1020			3075
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ggc acc gac aac tcc cgc gtg gcg tac gac atc gtc agc ctc tcg ccc Gly Thr Asp Asn Ser Arg Val Ala Tyr Asp Ile Val Ser Leu Ser Pro 1040 1045 1050			3171
acc gac agg gac atc aca ctt cct caa ctc ttc acc atg atc acc ata Thr Asp Arg Asp Ile Thr Leu Pro Gln Leu Phe Thr Met Ile Thr Ile 1055 1060 1065 1070			3219
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gat tta aga ggc tat tgg ggc act tat gaa ata cat gta aag gca tac Asp Leu Arg Gly Tyr Trp Gly Thr Tyr Glu Ile His Val Lys Ala Tyr 1090 1095 1100			3315
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gga atg act atc aga ctc gcg aag gag cga gca gta gtg aac gga gtg Gly Met Thr Ile Arg Leu Ala Lys Glu Arg Ala Val Val Asn Gly Val 1135 1140 1145 1150			3459
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gat gag gcg ttg cag tac ttc gac gtg ttt aac gac gga gtg aac ttg Asp Glu Ala Leu Gln Tyr Phe Asp Val Phe Asn Asp Gly Val Asn Leu 1185 1190 1195			3603
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Gly Ala Leu Thr Ile Thr Gln Leu Phe Pro Glu Asp Phe Arg Glu Phe 1200 1205 1210	
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agt acc gac tgc acg atc acc gta gtg ttt gtg cct acg cag gga gag Ser Thr Asp Cys Thr Ile Thr Val Val Phe Val Pro Thr Gln Gly Glu 1235 1240 1245	3747
cct gtg ttc gaa act agc acc tac acg gtc gct ttt atc gag aaa gat Pro Val Phe Glu Thr Ser Thr Tyr Thr Val Ala Phe Ile Glu Lys Asp 1250 1255 1260	3795
gct ggt atg gaa gag cgg gcc acg ctg cct ctc gcc aag gac ccg cgt Ala Gly Met Glu Glu Arg Ala Thr Leu Pro Leu Ala Lys Asp Pro Arg 1265 1270 1275	3843
aac ata atg tgt gaa gat gat tgt cac gac acc tat tac agc att gtt Asn Ile Met Cys Glu Asp Asp Cys His Asp Thr Tyr Tyr Ser Ile Val 1280 1285 1290	3891
gga ggc aac tcg atg ggc cac ttt gcg gtg gac cct cag tcc aac gag Gly Gly Asn Ser Met Gly His Phe Ala Val Asp Pro Gln Ser Asn Glu 1295 1300 1305 1310	3939
ctg ttc ctg ctg acg cca ctg gag cgc gcg gag cag gag acg cac acc Leu Phe Leu Leu Thr Pro Leu Glu Arg Ala Glu Gln Glu Thr His Thr 1315 1320 1325	3987
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aca ctg caa gct gtg cag gag aca gcc ttc aac ttg aac cct cag act Thr Leu Gln Ala Val Gln Glu Thr Ala Phe Asn Leu Asn Pro Gln Thr 1410 1415 1420	4275
gga gtg ctg acc ctc aac ttc cag ccc aca gca tct atg cac ggc atg Gly Val Leu Thr Leu Asn Phe Gln Pro Thr Ala Ser Met His Gly Met 1425 1430 1435	4323
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gcg gag aca ttt acc ctg ttc ttc ggc atg cgg tgc aac atc gac cag Ala Glu Thr Phe Thr Leu Phe Phe Gly Met Arg Cys Asn Ile Asp Gln 1490 1495 1500	4515
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Thr Glu Val Arg Ala His Phe Ile Arg Asp Asp Leu Pro Val Pro Ala
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gag gag atc gaa caa tta cgc ggc aac ccg acc cta gtg gcg acc atc 4659
Glu Glu Ile Glu Gln Leu Arg Gly Asn Pro Thr Leu Val Ala Thr Ile
1535                1540                1545                1550

cag aac gcc ctg cag gag gag aac ctg aac ctg gcc gac ctg ttc acg 4707
Gln Asn Ala Leu Gln Glu Glu Asn Leu Asn Leu Ala Asp Leu Phe Thr
                1555                1560                1565

ggc gag act ccc atc ctg ggc ggc gag gcg cag gcg cgg gcg gtc tat 4755
Gly Glu Thr Pro Ile Leu Gly Gly Glu Ala Gln Ala Arg Ala Val Tyr
                1570                1575                1580

gct ctc gcg gcg gtg gcg gct gcg ctc gcg ctg ctc tgc gtc gtg ctg 4803
Ala Leu Ala Ala Val Ala Ala Leu Ala Leu Leu Cys Val Val Leu
                1585                1590                1595

ctt ata ctc ttc ttc atc agg act agg gcc ctc aac cgt cgc ctg gaa 4851
Leu Ile Leu Phe Phe Ile Arg Thr Arg Ala Leu Asn Arg Arg Leu Glu
    1600                1605                1610

gcc cta tcc atg acc aag tac agt tcc caa gac tca gga ctc aac cgc 4899
Ala Leu Ser Met Thr Lys Tyr Ser Ser Gln Asp Ser Gly Leu Asn Arg
1615                1620                1625                1630

gtg ggt ctg gcg gcg ccg ggc acc aac aag cac gcg gtg gag ggc tcc 4947
Val Gly Leu Ala Ala Pro Gly Thr Asn Lys His Ala Val Glu Gly Ser
                1635                1640                1645

aac ccc atc tgg aac gaa act ctt aag gca ccg gac ttt gat gct ctt 4995
Asn Pro Ile Trp Asn Glu Thr Leu Lys Ala Pro Asp Phe Asp Ala Leu
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agc gag cag tcg tac gac tcg ggt ctg atc ggc atc gaa gac ttg ccg 5043
Ser Glu Gln Ser Tyr Asp Ser Gly Leu Ile Gly Ile Glu Asp Leu Pro
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cag ttc agg aac gac tac ttc ccg cct gac gag gag agc tcc atg cgg 5091
Gln Phe Arg Asn Asp Tyr Phe Pro Pro Asp Glu Glu Ser Ser Met Arg
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gga gtc gtc aat gaa cac atg cct gga gct aat tca gta gca aac cat 5139
Gly Val Val Asn Glu His Met Pro Gly Ala Asn Ser Val Ala Asn His
1695                1700                1705                1710

aac aat aac ttc ggg ttc aac gct acc ccc ttc agc cca gag ttc gcg 5187
Asn Asn Asn Phe Gly Phe Asn Ala Thr Pro Phe Ser Pro Glu Phe Ala
                1715                1720                1725

aac tcg cag ctc agg aga taaaacatta tagtattttt tatataatat 5235
Asn Ser Gln Leu Arg Arg
    1730

tataaagaag tgatataacg cactaaaatt tacctataag tatatattga agtgaagat 5295

actcgtatta tgtaagagca tctatttttt taccaccaga caataaaaac tttataaag 5355

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<210> SEQ ID NO 2

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<212> TYPE: PRT

<213> ORGANISM: Heliothis virescens

<400> SEQUENCE: 2

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                20                25                30

Arg Pro Glu Arg Pro Asp Phe Pro Asn Gln Asn Phe Glu Gly Val Pro
                35                40                45

Trp Ser Gln Asn Pro Leu Leu Pro Ala Glu Asp Arg Glu Asp Val Cys
    50                55                60

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Met Asn Ala Phe Asp Pro Ser Ala Leu Asn Pro Val Thr Val Ile Phe
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 Met Glu Glu Glu Ile Glu Gly Asp Val Ala Ile Ala Arg Leu Asn Tyr
 85 90 95
 Arg Gly Thr Asn Thr Pro Thr Val Val Thr Pro Phe Asn Phe Gly Thr
 100 105 110
 Phe His Leu Leu Gly Pro Val Ile Arg Arg Ile Pro Glu Gln Gly Gly
 115 120 125
 Asp Trp His Leu Val Ile Thr Gln Arg Gln Asp Tyr Glu Thr Pro Asn
 130 135 140
 Met Gln Gln Tyr Ile Phe Asn Val Arg Val Glu Asp Glu Pro Gln Glu
 145 150 155 160
 Ala Thr Val Met Leu Ile Ile Val Asn Ile Asp Asp Asn Ala Pro Ile
 165 170 175
 Ile Gln Met Phe Glu Pro Cys Asp Ile Pro Glu His Gly Glu Thr Gly
 180 185 190
 Thr Thr Glu Cys Lys Tyr Val Val Ser Asp Ala Asp Gly Glu Ile Ser
 195 200 205
 Thr Arg Phe Met Thr Phe Gln Ile Glu Ser Asp Arg Asn Asp Glu Glu
 210 215 220
 Tyr Phe Glu Leu Val Arg Glu Asn Ile Gln Gly Gln Trp Met Tyr Val
 225 230 235 240
 His Met Arg Leu Ile Leu Asn Lys Pro Leu Asp Tyr Glu Glu Asn Pro
 245 250 255
 Leu His Leu Phe Arg Val Thr Ala Leu Asp Ser Leu Pro Asn Val His
 260 265 270
 Thr Val Thr Met Met Val Gln Val Glu Asn Ile Glu Ser Arg Pro Pro
 275 280 285
 Arg Trp Met Glu Ile Phe Ala Val Gln Gln Phe Asp Glu Lys Thr Ala
 290 295 300
 Gln Ala Phe Arg Val Arg Ala Ile Asp Gly Asp Thr Gly Ile Asp Lys
 305 310 315 320
 Pro Ile Phe Tyr Arg Ile Glu Thr Glu Glu Ser Glu Lys Asp Leu Phe
 325 330 335
 Ser Val Glu Thr Ile Gly Ala Gly Arg Glu Gly Ala Trp Phe Lys Val
 340 345 350
 Ala Pro Ile Asp Arg Asp Thr Leu Glu Lys Glu Val Phe His Val Ser
 355 360 365
 Leu Ile Ala Tyr Lys Tyr Gly Asp Asn Asp Val Glu Gly Ser Pro Ser
 370 375 380
 Phe Glu Ser Lys Thr Asp Ile Val Ile Ile Val Asn Asp Val Asn Asp
 385 390 395 400
 Gln Ala Pro Val Pro Phe Arg Pro Ser Tyr Tyr Ile Glu Ile Met Glu
 405 410 415
 Glu Ala Ala Met Thr Leu Asn Leu Glu Asp Phe Gly Phe His Asp Arg
 420 425 430
 Gly Leu Gly Pro His Ala Gln Tyr Thr Val His Leu Glu Ser Ile Ser
 435 440 445
 Pro Ala Gly Ala His Glu Ala Phe Tyr Ile Ala Pro Glu Val Gly Tyr
 450 455 460
 Gln Arg Gln Ser Phe Ile Val Gly Thr Gln Asn His His Met Leu Asp
 465 470 475 480

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Phe	Glu	Val	Pro	Glu	Phe	Gln	Lys	Ile	Gln	Leu	Arg	Ala	Val	Ala	Ile
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Asp	Met	Asp	Asp	Pro	Arg	Trp	Val	Gly	Ile	Ala	Ile	Ile	Asn	Ile	Asn
			500					505					510		
Leu	Ile	Asn	Trp	Asn	Asp	Glu	Leu	Pro	Ile	Phe	Glu	His	Asp	Val	Gln
		515					520					525			
Thr	Val	Thr	Phe	Lys	Glu	Thr	Glu	Gly	Ala	Gly	Phe	Arg	Val	Ala	Thr
	530					535					540				
Val	Leu	Ala	Lys	Asp	Arg	Asp	Ile	Asp	Asp	Arg	Val	Glu	His	Ser	Leu
545					550					555					560
Met	Gly	Asn	Ala	Val	Asn	Tyr	Leu	Ser	Ile	Asp	Lys	Asp	Thr	Gly	Asp
				565					570					575	
Ile	Leu	Val	Thr	Ile	Asp	Asp	Ala	Phe	Asn	Tyr	His	Arg	Gln	Asn	Glu
			580					585					590		
Leu	Phe	Val	Gln	Ile	Arg	Ala	Asp	Asp	Thr	Leu	Gly	Glu	Pro	Tyr	Asn
		595					600					605			
Thr	Asn	Thr	Ala	Gln	Leu	Val	Ile	Gln	Leu	Gln	Asp	Ile	Asn	Asn	Thr
	610					615					620				
Pro	Pro	Thr	Leu	Arg	Leu	Pro	Arg	Thr	Thr	Pro	Ser	Val	Glu	Glu	Asn
625					630						635				640
Val	Pro	Asp	Gly	Phe	Val	Ile	Pro	Thr	Glu	Leu	His	Ala	Thr	Asp	Pro
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Asp	Thr	Thr	Ala	Glu	Leu	Arg	Phe	Ser	Ile	Asp	Trp	Asp	Thr	Ser	Tyr
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Ala	Thr	Lys	Gln	Gly	Arg	Asp	Ala	Asp	Ala	Glu	Glu	Phe	Val	Asn	Cys
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Ile	Glu	Ile	Glu	Thr	Val	Tyr	Pro	Asn	Leu	Asn	Asp	Arg	Gly	Thr	Ala
	690					695					700				
Ile	Gly	Arg	Val	Val	Val	Arg	Glu	Ile	Arg	Glu	His	Val	Thr	Ile	Asp
705					710					715					720
Tyr	Glu	Met	Phe	Glu	Val	Leu	Tyr	Leu	Thr	Val	Arg	Val	Thr	Asp	Leu
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Asn	Thr	Val	Ile	Gly	Asp	Asp	Tyr	Asp	Ile	Ser	Thr	Phe	Thr	Ile	Ile
			740					745					750		
Ile	Ile	Asp	Met	Asn	Asp	Asn	Pro	Pro	Leu	Trp	Val	Glu	Gly	Thr	Leu
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Thr	Gln	Glu	Phe	Arg	Val	Arg	Glu	Val	Ala	Ala	Ser	Gly	Val	Val	Ile
	770					775					780				
Gly	Ser	Val	Leu	Ala	Thr	Asp	Ile	Asp	Gly	Pro	Leu	Tyr	Asn	Gln	Val
785					790					795					800
Arg	Tyr	Thr	Ile	Thr	Pro	Arg	Leu	Asp	Thr	Pro	Glu	Asp	Leu	Val	Glu
			805						810					815	
Ile	Asp	Phe	Asn	Ser	Gly	Gln	Ile	Ser	Val	Lys	Lys	His	Gln	Ala	Ile
			820					825					830		
Asp	Ala	Asp	Glu	Pro	Pro	Arg	Gln	His	Leu	Tyr	Tyr	Thr	Val	Val	Ala
		835					840						845		
Ser	Asp	Lys	Cys	Asp	Leu	Leu	Ser	Val	Asp	Val	Cys	Pro	Pro	Asp	Pro
	850					855					860				
Asn	Tyr	Phe	Asn	Thr	Pro	Gly	Asp	Ile	Thr	Ile	His	Ile	Thr	Asp	Thr
865					870					875					880
Asn	Asn	Arg	Val	Pro	Arg	Val	Glu	Glu	Asp	Lys	Phe	Glu	Glu	Ile	Val
			885						890					895	
Tyr	Ile	Tyr	Glu	Gly	Ala	Glu	Asp	Gly	Glu	His	Val	Val	Gln	Leu	Phe

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900					905					910					
Ala	Ser	Asp	Leu	Asp	Arg	Asp	Glu	Ile	Tyr	His	Lys	Val	Ser	Tyr	Gln
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Ile	Asn	Tyr	Ala	Ile	Asn	Pro	Arg	Leu	Arg	Asp	Phe	Phe	Glu	Val	Asp
	930					935					940				
Leu	Glu	Thr	Gly	Leu	Val	Tyr	Val	Asn	Asn	Thr	Ala	Gly	Glu	Lys	Leu
945					950					955					960
Asp	Arg	Asp	Gly	Asp	Glu	Pro	Thr	His	Arg	Ile	Phe	Phe	Asn	Val	Ile
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Asp	Asn	Phe	Tyr	Gly	Glu	Gly	Asp	Gly	Asn	Arg	Asn	Gln	Asp	Glu	Thr
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Gln	Val	Leu	Val	Val	Leu	Leu	Asp	Ile	Asn	Asp	Asn	Tyr	Pro	Glu	Leu
		995					1000					1005			
Pro	Glu	Gly	Leu	Ser	Trp	Asp	Ile	Ser	Glu	Gly	Leu	Leu	Gln	Gly	Val
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Arg	Val	Thr	Pro	Asp	Ile	Phe	Ala	Pro	Asp	Arg	Asp	Glu	Pro	Gly	Thr
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Asp	Asn	Ser	Arg	Val	Ala	Tyr	Asp	Ile	Val	Ser	Leu	Ser	Pro	Thr	Asp
			1045						1050					1055	
Arg	Asp	Ile	Thr	Leu	Pro	Gln	Leu	Phe	Thr	Met	Ile	Thr	Ile	Glu	Lys
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Asp	Arg	Gly	Ile	Asp	Gln	Thr	Gly	Glu	Leu	Glu	Thr	Ala	Met	Asp	Leu
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Arg	Gly	Tyr	Trp	Gly	Thr	Tyr	Glu	Ile	His	Val	Lys	Ala	Tyr	Asp	His
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Gly	Val	Pro	Gln	Arg	Ile	Ser	Tyr	Glu	Lys	Tyr	Pro	Leu	Val	Ile	Arg
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Pro	Tyr	Asn	Phe	His	Asp	Pro	Val	Phe	Val	Phe	Pro	Gln	Pro	Gly	Met
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Thr	Ile	Arg	Leu	Ala	Lys	Glu	Arg	Ala	Val	Val	Asn	Gly	Val	Leu	Ala
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Thr	Val	Asp	Gly	Glu	Phe	Leu	Glu	Arg	Ile	Val	Ala	Thr	Asp	Glu	Asp
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Gly	Leu	His	Ala	Gly	Val	Val	Thr	Phe	Ser	Ile	Ser	Gly	Asp	Asp	Glu
	1170					1175					1180				
Ala	Leu	Gln	Tyr	Phe	Asp	Val	Phe	Asn	Asp	Gly	Val	Asn	Leu	Gly	Ala
1185					1190					1195					1200
Leu	Thr	Ile	Thr	Gln	Leu	Phe	Pro	Glu	Asp	Phe	Arg	Glu	Phe	Gln	Val
				1205					1210					1215	
Thr	Ile	Arg	Ala	Thr	Asp	Gly	Gly	Thr	Glu	Pro	Gly	Pro	Arg	Ser	Thr
			1220					1225					1230		
Asp	Cys	Thr	Ile	Thr	Val	Val	Phe	Val	Pro	Thr	Gln	Gly	Glu	Pro	Val
		1235					1240					1245			
Phe	Glu	Thr	Ser	Thr	Tyr	Thr	Val	Ala	Phe	Ile	Glu	Lys	Asp	Ala	Gly
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Met	Glu	Glu	Arg	Ala	Thr	Leu	Pro	Leu	Ala	Lys	Asp	Pro	Arg	Asn	Ile
1265					1270					1275					1280
Met	Cys	Glu	Asp	Asp	Cys	His	Asp	Thr	Tyr	Tyr	Ser	Ile	Val	Gly	Gly
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Asn	Ser	Met	Gly	His	Phe	Ala	Val	Asp	Pro	Gln	Ser	Asn	Glu	Leu	Phe
			1300					1305					1310		
Leu	Leu	Thr	Pro	Leu	Glu	Arg	Ala	Glu	Gln	Glu	Thr	His	Thr	Leu	Ile
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Ile Gly Ala Ser Asp Ser Pro Ser Pro Ala Ala Val Leu Gln Ala Ser
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 Thr Leu Thr Val Thr Val Asn Val Arg Glu Ala Asn Pro Arg Pro Val
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 1365 1370 1375
 Asn Arg Gly Leu Leu Thr Leu His Ala Thr His Ser Glu Gly Leu Pro
 1380 1385 1390
 Val Thr Tyr Thr Leu Val Gln Asp Ser Met Glu Ala Asp Ser Thr Leu
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 Gln Ala Val Gln Glu Thr Ala Phe Asn Leu Asn Pro Gln Thr Gly Val
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 1425 1430 1435 1440
 Phe Asp Val Met Ala Thr Asp Thr Val Gly Glu Thr Ala Arg Thr Glu
 1445 1450 1455
 Val Lys Val Tyr Leu Ile Ser Asp Arg Asn Arg Val Phe Phe Thr Phe
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 Met Asn Thr Leu Glu Glu Val Glu Pro Asn Glu Asp Phe Ile Ala Glu
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 1490 1495 1500
 Pro Ala Ser Asp Pro Ala Thr Gly Ala Ala Arg Asp Asp Gln Thr Glu
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 1620 1625 1630
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 Gln Ser Tyr Asp Ser Gly Leu Ile Gly Ile Glu Asp Leu Pro Gln Phe
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 1685 1690 1695
 Val Asn Glu His Met Pro Gly Ala Asn Ser Val Ala Asn His Asn Asn
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<212> TYPE: DNA
<213> ORGANISM: Heliothis virescens

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<211> LENGTH: 2263

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<213> ORGANISM: *Heliothis virescens*

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<212> TYPE: DNA

<213> ORGANISM: Heliothis virescens

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<211> LENGTH: 257

<212> TYPE: DNA

<213> ORGANISM: Heliothis virescens

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<212> TYPE: DNA

<213> ORGANISM: Heliothis virescens

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<212> TYPE: DNA
<213> ORGANISM: Heliothis virescens

<400> SEQUENCE: 9

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<210> SEQ ID NO 10
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<213> ORGANISM: Heliothis virescens

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35 40 45
Trp Ser Gln Asn Pro Leu Leu Pro Ala Glu Asp Arg Glu Asp Val Cys
50 55 60
Met Asn Ala Phe Asp Pro Ser Ala Leu Asn Pro Val Thr Val Ile Phe
65 70 75 80
Met Glu Glu Glu Ile Glu Gly Asp Val Ala Ile Ala Arg Leu Asn Tyr
85 90 95
Arg Gly Thr Asn Thr Pro Thr Val Val Thr Pro Phe Asn Phe Gly Thr
100 105 110
Phe His Leu Leu Gly Pro Val Ile Arg Arg Ile Pro Glu Gln Gly Gly
115 120 125
Asp Trp His Leu Val Ile Thr Gln Arg Gln Asp Tyr Glu Thr Pro Asn
130 135 140
Met Gln Gln Tyr Ile Phe Asn Val Arg Val Glu Asp Glu Pro Gln Glu
145 150 155 160
Ala Thr Val Met Leu Ile Ile Val Asn Ile Asp Asp Asn Ala Pro Ile
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Ile Gln Met Phe Glu Pro Cys Asp Ile Pro Glu His Gly Glu Thr Gly
180 185 190
Thr Thr Glu Cys Lys Tyr Val Val Ser Asp Ala Asp Gly Glu Ile Ser
195 200 205
Thr Arg Phe Met Thr Phe Glu Ile Glu Ser Asp Arg Asn Asp Glu Glu
210 215 220
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His Met Arg Leu Ile Leu Asn Lys Pro Leu Asp Tyr Glu Glu Asn Pro
245 250 255

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Arg Trp Met Glu Ile Phe Ala Val Gln Gln Phe Asp Glu Lys Thr Ala
      290                295                300

Gln Ser Phe Arg Val Arg Ala Ile Asp Gly Asp Thr Gly Ile Asp Lys
      305                310                315                320

Pro Ile Phe Tyr Arg Ile Glu Thr Glu Glu Ser Glu Lys Asp Leu Phe
      325                330                335

Ser Val Glu Thr Ile Gly Ala Gly Arg Glu Gly Ala Trp Phe Lys Val
      340                345                350

Ala Pro Ile Asp Arg Asp Thr Leu Glu Lys Glu Val Phe His Val Ser
      355                360                365

Leu Ile Ala Tyr Lys Tyr Gly Asp Asn Asp Val Glu Gly Ser Pro Ser
      370                375                380

Phe Glu Ser Lys Thr Asp Ile Val Ile Ile Val Asn Asp Val Asn Asp
      385                390                395                400

Gln Ala Pro Val Pro Phe Arg Pro Ser Tyr Phe Ile Glu Ile Met Glu
      405                410                415

Glu Thr Ala Met Thr Leu Asn Leu Glu Asp Phe Gly Phe His Asp Arg
      420                425                430

Asp Leu Gly Pro His Ala Gln Tyr Thr Val His Leu Glu Ser Ile His
      435                440                445

Pro Ala Gly Ala His Glu Ala Phe Tyr Ile Ala Pro Glu Val Gly Tyr
      450                455                460

Gln Arg Gln Ser Phe Ile Val Gly Thr Gln Asn His His Met Leu Asp
      465                470                475                480

Phe Glu Val Pro Glu Phe Gln Lys Ile Gln Leu Arg Val Val Ala Ile
      485                490                495

Asp Met Asp Asp Pro Arg Trp Val Gly Ile Ala Ile Ile Asn Ile Asn
      500                505                510

Leu Ile Asn Trp Asn Asp Glu Leu Pro Ile Phe Glu His Asp Val Gln
      515                520                525

Thr Ala Thr Phe Lys Glu Thr Glu Gly Ala Gly Phe Arg Val Ala Thr
      530                535                540

Val Leu Ala Lys Asp Arg Asp Ile Asp Glu Arg Val Glu His Ser Leu
      545                550                555                560

Met Gly Asn Ala Val Asn Tyr Leu Ser Ile Asp Lys Asp Thr Gly Asp
      565                570                575

Ile Leu Val Thr Ile Asp Asp Ala Phe Asn Tyr His Arg Gln Asn Glu
      580                585                590

Leu Phe Val Gln Ile Arg Ala Asp Asp Thr Leu Glu Glu Pro Tyr Asn
      595                600                605

Ala Asn Thr Ala Cys Ser Gly Leu Ser His Arg Ala Gly Leu
      610                615                620

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<210> SEQ ID NO 11

<211> LENGTH: 1714

<212> TYPE: PRT

<213> ORGANISM: Bombyx mori

<400> SEQUENCE: 11

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Met Gly Val Asp Val Arg Ile Leu Ala Thr Leu Leu Leu Ile Tyr Ala
  1           5           10           15

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Glu Thr Val Leu Ala Gln Glu Arg Cys Gly Phe Met Val Ala Ile Pro
 20 25 30

Arg Pro Pro Arg Pro Asp Leu Pro Glu Leu Asp Phe Glu Gly Gln Thr
 35 40 45

Trp Ser Gln Arg Pro Leu Ile Pro Ala Ala Asp Arg Glu Asp Val Cys
 50 55 60

Met Asp Gly Tyr His Ala Met Thr Pro Thr Tyr Gly Thr Gln Ile Ile
 65 70 75 80

Tyr Met Glu Glu Glu Ile Glu Gly Glu Val Pro Ile Ala Lys Leu Asn
 85 90 95

Tyr Arg Gly Pro Asn Val Pro Tyr Ile Glu Pro Ala Phe Leu Ser Gly
 100 105 110

Ser Phe Asn Leu Leu Val Pro Val Ile Arg Arg Ile Pro Asp Ser Asn
 115 120 125

Gly Glu Trp His Leu Ile Ile Thr Gln Arg Gln Asp Tyr Glu Thr Pro
 130 135 140

Gly Met Gln Gln Tyr Val Phe Asn Ile Arg Ile Asp Gly Glu Thr Leu
 145 150 155 160

Val Ala Gly Val Ser Leu Leu Ile Val Asn Ile Asp Asp Asn Ala Pro
 165 170 175

Ile Ile Gln Ala Leu Glu Pro Cys Gln Val Asp Glu Leu Gly Glu Ala
 180 185 190

Arg Leu Thr Glu Cys Val Tyr Val Val Thr Asp Ala Asp Gly Arg Ile
 195 200 205

Ser Thr Gln Phe Met Gln Phe Arg Ile Asp Ser Asp Arg Gly Asp Asp
 210 215 220

Lys Ile Phe Tyr Ile Gln Gly Ala Asn Ile Pro Gly Glu Trp Ile Arg
 225 230 235 240

Met Thr Met Thr Val Gly Ile Asn Glu Pro Leu Asn Phe Glu Thr Asn
 245 250 255

Pro Leu His Ile Phe Ser Val Thr Ala Leu Asp Ser Leu Pro Asn Thr
 260 265 270

His Thr Val Thr Leu Met Val Gln Val Glu Asn Val Glu His Arg Pro
 275 280 285

Pro Arg Trp Val Glu Ile Phe Ala Val Gln Gln Phe Asp Glu Lys Thr
 290 295 300

Ala Gln Ser Phe Pro Val Arg Ala Ile Asp Gly Asp Thr Gly Ile Asn
 305 310 315 320

Lys Pro Ile His Tyr Arg Leu Glu Thr Ala Glu Glu Asp Thr Phe Phe
 325 330 335

His Ile Arg Thr Ile Glu Gly Gly Arg Ser Gly Ala Ile Leu Tyr Val
 340 345 350

Asp Pro Ile Asp Arg Asp Thr Leu Gln Arg Glu Val Phe Gln Leu Ser
 355 360 365

Ile Ile Ala Tyr Lys Tyr Asp Asn Glu Ser Ser Ala Thr Ala Ala Asn
 370 375 380

Val Val Ile Ile Val Asn Asp Ile Asn Asp Gln Arg Pro Glu Pro Leu
 385 390 395 400

Phe Lys Glu Tyr Arg Leu Asn Ile Met Glu Glu Thr Ala Leu Thr Leu
 405 410 415

Asn Phe Asp Gln Glu Phe Gly Phe His Asp Arg Asp Leu Gly Gln Asn
 420 425 430

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Ala	Gln	Tyr	Thr	Val	Arg	Leu	Glu	Ser	Asp	Tyr	Pro	Ala	Asp	Ala	Ala
	435						440					445			
Lys	Ala	Phe	Tyr	Ile	Ala	Pro	Glu	Val	Gly	Tyr	Gln	Arg	Gln	Thr	Phe
	450					455					460				
Ile	Met	Gly	Thr	Ala	Asn	His	Lys	Met	Leu	Asp	Tyr	Glu	Val	Pro	Glu
465					470					475					480
Phe	Gln	Arg	Ile	Arg	Leu	Arg	Val	Ile	Ala	Thr	Asp	Met	Asp	Asn	Glu
				485					490					495	
Glu	His	Val	Gly	Val	Ala	Tyr	Val	Tyr	Ile	Asn	Leu	Ile	Asn	Trp	Asn
			500					505					510		
Asp	Glu	Glu	Pro	Ile	Phe	Glu	His	Ser	Val	Gln	Asn	Val	Ser	Phe	Lys
		515					520					525			
Glu	Thr	Glu	Gly	Lys	Gly	Phe	Phe	Val	Ala	Asn	Val	Arg	Ala	His	Asp
	530					535				540					
Arg	Asp	Ile	Asp	Asp	Arg	Val	Glu	His	Thr	Leu	Met	Gly	Asn	Ala	Asn
545					550					555					560
Asn	Tyr	Leu	Ser	Ile	Asp	Lys	Asp	Thr	Gly	Asp	Ile	His	Val	Thr	Gln
				565					570					575	
Asp	Asp	Phe	Phe	Asp	Tyr	His	Arg	Gln	Ser	Glu	Leu	Phe	Val	Gln	Val
			580					585					590		
Arg	Ala	Asp	Asp	Thr	Leu	Gly	Glu	Pro	Phe	His	Thr	Ala	Thr	Ser	Gln
		595					600					605			
Leu	Leu	Ile	His	Leu	Glu	Asp	Ile	Asn	Asn	Thr	Pro	Pro	Thr	Leu	Arg
	610					615					620				
Leu	Pro	Arg	Gly	Ser	Pro	Asn	Val	Glu	Glu	Asn	Val	Pro	Glu	Gly	Tyr
625					630					635					640
Ile	Ile	Thr	Ser	Glu	Ile	Arg	Ala	Thr	Asp	Pro	Asp	Thr	Thr	Ala	Glu
				645					650					655	
Leu	Arg	Phe	Glu	Ile	Asp	Trp	Thr	Thr	Ser	Tyr	Ala	Thr	Lys	Gln	Gly
			660					665					670		
Arg	Glu	Ala	Asn	Pro	Ile	Glu	Phe	His	Asn	Cys	Val	Glu	Ile	Glu	Thr
		675					680					685			
Ile	Tyr	Pro	Ala	Ile	Asn	Asn	Arg	Gly	Ser	Ala	Ile	Gly	Arg	Leu	Val
	690				695					700					
Val	Lys	Lys	Ile	Arg	Glu	Asn	Val	Thr	Ile	Asp	Tyr	Glu	Glu	Phe	Glu
705					710					715					720
Met	Leu	Tyr	Leu	Thr	Val	Arg	Val	Arg	Asp	Leu	Asn	Thr	Val	Ile	Gly
				725					730					735	
Asp	Asp	Tyr	Asp	Glu	Ser	Thr	Phe	Thr	Ile	Thr	Ile	Ile	Asp	Met	Asn
			740					745					750		
Asp	Asn	Pro	Pro	Ile	Trp	Val	Pro	Gly	Thr	Leu	Glu	Gln	Ser	Leu	Arg
		755					760					765			
Val	Arg	Glu	Met	Ser	Asp	Ala	Gly	Val	Val	Ile	Gly	Thr	Leu	Thr	Ala
	770					775					780				
Thr	Asp	Ile	Asp	Gly	Pro	Leu	Tyr	Asn	Gln	Val	Arg	Tyr	Thr	Met	Lys
785					790					795					800
Ala	Asn	Glu	Gly	Thr	Pro	Glu	Asn	Leu	Leu	Met	Ile	Asp	Phe	Tyr	Thr
				805					810					815	
Gly	Gln	Ile	Thr	Val	Lys	Thr	Ser	Gly	Ala	Ile	Asp	Ala	Asp	Val	Pro
			820					825					830		
Arg	Arg	Tyr	Asn	Leu	Tyr	Tyr	Thr	Val	Val	Ala	Thr	Asp	Arg	Cys	Tyr
		835					840					845			
Ala	Glu	Asp	Pro	Asp	Asp	Cys	Pro	Asp	Asp	Pro	Thr	Tyr	Trp	Glu	Thr

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850					855					860					
Pro	Gly	Gln	Val	Val	Ile	Gln	Ile	Ile	Asp	Thr	Asn	Asn	Lys	Ile	Pro
865					870					875					880
Gln	Pro	Glu	Thr	Asp	Gln	Phe	Lys	Ala	Val	Val	Tyr	Ile	Tyr	Glu	Asp
				885					890					895	
Ala	Val	Ser	Gly	Asp	Glu	Val	Val	Lys	Val	Ile	Gly	Ser	Asp	Leu	Asp
			900					905					910		
Arg	Asp	Asp	Ile	Tyr	His	Thr	Ile	Arg	Tyr	Gln	Ile	Asn	Tyr	Ala	Val
	915					920						925			
Asn	Pro	Arg	Leu	Arg	Asp	Phe	Phe	Ala	Val	Asp	Pro	Asp	Thr	Gly	Arg
	930				935					940					
Val	Tyr	Val	Tyr	Tyr	Thr	Thr	Asp	Glu	Val	Leu	Asp	Arg	Asp	Gly	Asp
945					950					955					960
Glu	Pro	Gln	His	Arg	Ile	Phe	Phe	Asn	Leu	Ile	Asp	Asn	Phe	Phe	Gln
			965						970					975	
Gln	Gly	Asp	Gly	Asn	Arg	Asn	Gln	Asn	Asp	Ala	Glu	Val	Leu	Val	Val
			980					985					990		
Leu	Leu	Asp	Val	Asn	Asp	Asn	Ala	Pro	Glu	Leu	Pro	Glu	Pro	Asp	Glu
		995					1000					1005			
Leu	Ser	Trp	Ser	Val	Ser	Glu	Ser	Leu	Thr	Lys	Gly	Thr	Arg	Leu	Gln
	1010					1015					1020				
Pro	His	Ile	Tyr	Ala	Pro	Asp	Arg	Asp	Glu	Pro	Asp	Thr	Asp	Asn	Ser
1025				1030					1035					1040	
Arg	Val	Gly	Tyr	Ala	Ile	Ile	Ser	Leu	Thr	Ile	Ala	Asn	Arg	Glu	Ile
			1045					1050						1055	
Glu	Val	Pro	Glu	Leu	Phe	Thr	Met	Ile	Gln	Ile	Gln	Asn	Val	Thr	Gly
			1060					1065					1070		
Glu	Leu	Glu	Thr	Ala	Met	Asp	Leu	Arg	Gly	Tyr	Trp	Gly	Thr	Tyr	Ala
		1075					1080					1085			
Ile	His	Ile	Lys	Ala	Tyr	Asp	His	Gly	Ile	Pro	Gln	Gln	Met	Ser	Asn
	1090					1095					1100				
Glu	Thr	Tyr	Glu	Leu	Val	Ile	Arg	Pro	Tyr	Asn	Phe	His	Ala	Pro	Val
1105				1110					1115					1120	
Phe	Val	Phe	Pro	Lys	His	Gly	Ala	Thr	Leu	Arg	Leu	Ala	Arg	Glu	Arg
			1125					1130						1135	
Ala	Val	Val	Asn	Gly	Leu	Ala	Thr	Val	Asp	Gly	Glu	Phe	Leu	Asn	Arg
			1140					1145					1150		
Ile	Val	Ala	Thr	Asp	Glu	Asp	Gly	Leu	His	Ala	Gly	Gln	Val	Ala	Phe
	1155						1160					1165			
Glu	Val	Val	Gly	Asp	Thr	Glu	Ala	Val	Asp	Tyr	Phe	His	Ile	Val	Asn
	1170					1175					1180				
Asp	Gly	Glu	Asn	Ser	Gly	Thr	Leu	Met	Leu	Lys	Gln	Leu	Phe	Pro	Glu
1185				1190					1195					1200	
Asp	Ile	Arg	Glu	Phe	Glu	Val	Thr	Ile	Arg	Ala	Thr	Asp	Gly	Gly	Thr
			1205					1210						1215	
Glu	Pro	Arg	Pro	Leu	Ser	Thr	Asp	Cys	Thr	Phe	Ser	Val	Val	Phe	Val
			1220					1225					1230		
Pro	Ile	Gln	Gly	Glu	Pro	Ile	Phe	Pro	Thr	Ser	Thr	His	Thr	Val	Ala
	1235						1240					1245			
Phe	Ile	Glu	Lys	Glu	Ala	Gly	Leu	Leu	Glu	Arg	His	Glu	Leu	Pro	Arg
	1250					1255						1260			
Ala	Glu	Asp	Arg	Lys	Asn	His	Leu	Cys	Ser	Asp	Asp	Cys	His	Asn	Ile
1265				1270							1275			1280	

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Tyr Tyr Arg Ile Ile Asp Gly Asn Asn Asp Gly His Phe Gly Leu Asp
 1285 1290 1295
 Glu Thr Thr Asn Val Leu Phe Leu Val Lys Glu Leu Asp Arg Ser Val
 1300 1305 1310
 Ser Glu Thr Tyr Thr Leu Thr Ile Ala Ala Ser Asn Ser Pro Thr Gly
 1315 1320 1325
 Gly Ile Ala Leu Thr Ser Thr Ile Thr Ile Thr Val Asn Val Arg Glu
 1330 1335 1340
 Ala Asp Pro Gln Pro Tyr Phe Val Arg Asp Leu Tyr Thr Ala Gly Ile
 1345 1350 1355 1360
 Ser Thr Ser Asp Ser Ile Asn Arg Glu Leu Leu Ile Leu Gln Ala Thr
 1365 1370 1375
 His Ser Glu Asn Ala Pro Ile Ile Tyr Thr Ile Asp Trp Ser Thr Met
 1380 1385 1390
 Val Thr Asp Pro Thr Leu Ala Ser Val Arg Glu Thr Ala Phe Ile Leu
 1395 1400 1405
 Asn Pro His Thr Gly Val Leu Thr Leu Asn Ile Gln Pro Thr Ala Ser
 1410 1415 1420
 Met His Gly Met Phe Glu Phe Gln Val Val Ala Thr Asp Pro Ala Gly
 1425 1430 1435 1440
 Tyr Ser Asp Arg Ala Asn Val Lys Ile Tyr Leu Ile Ser Thr Arg Asn
 1445 1450 1455
 Arg Val Phe Phe Leu Phe Val Asn Thr Leu Glu Gln Val Glu Gln Asn
 1460 1465 1470
 Thr Asp Phe Ile Ala Gln Thr Phe Ser Ala Gly Phe Glu Met Thr Cys
 1475 1480 1485
 Asn Ile Asp Gln Val Val Pro Ala Thr Asp Ala Ser Gly Val Ile Met
 1490 1495 1500
 Asn Gly Ile Thr Glu Val Arg Gly His Phe Ile Arg Asp Asn Val Pro
 1505 1510 1515 1520
 Val Pro Ala Asp Glu Thr Ile Glu Thr Leu Arg Gly Asp Met Val Leu Leu
 1525 1530 1535
 Thr Ala Ile Gln Ser Thr Leu Ala Thr Arg Leu Leu Val Leu Arg Asp
 1540 1545 1550
 Leu Phe Thr Asp Thr Ser Pro Ala Pro Asp Ala Gly Ser Ala Ala Val
 1555 1560 1565
 Leu Tyr Ala Leu Ala Val Leu Ser Ala Leu Leu Ala Ala Leu Cys Leu
 1570 1575 1580
 Leu Leu Leu Val Ile Phe Ile Ile Arg Thr Lys Lys Leu Asn Arg Arg
 1585 1590 1595 1600
 Leu Glu Ala Leu Thr Val Lys Lys Tyr Gly Ser Val Asp Ser Gly Leu
 1605 1610 1615
 Asn Arg Val Gly Ile Ala Ala Pro Gly Thr Asn Lys His Ala Val Glu
 1620 1625 1630
 Gly Ser Asn Pro Ile Trp Asn Glu Thr Ile Lys Ala Pro Asp Phe Asp
 1635 1640 1645
 Ser Met Ser Asp Ala Ser Asn Asp Ser Asp Leu Ile Gly Ile Glu Asp
 1650 1655 1660
 Leu Pro His Phe Gly Glu Asn Asn Tyr Phe Pro Arg Asp Val Asp Glu
 1665 1670 1675 1680
 Phe Lys Thr Asp Lys Pro Glu Asp Ile Val Ala Thr His Asn Asn Asn
 1685 1690 1695

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Phe Gly Phe Lys Ser Thr Pro Phe Ser Pro Glu Phe Ala Asn Gln Phe
 1700 1705 1710

Gln Lys

<210> SEQ ID NO 12
 <211> LENGTH: 1527
 <212> TYPE: PRT
 <213> ORGANISM: *Manduca sexta*

<400> SEQUENCE: 12

Met Ala Val Asp Val Arg Ile Ala Ala Phe Leu Leu Val Phe Ile Ala
 1 5 10 15

Pro Ala Val Leu Ala Gln Glu Arg Cys Gly Tyr Met Thr Ala Ile Pro
 20 25 30

Arg Leu Pro Arg Pro Asp Asn Leu Pro Val Leu Asn Phe Glu Gly Gln
 35 40 45

Thr Trp Ser Gln Arg Pro Leu Leu Pro Ala Pro Glu Arg Asp Asp Leu
 50 55 60

Cys Met Asp Ala Tyr His Val Ile Thr Ala Asn Leu Gly Thr Gln Val
 65 70 75 80

Ile Tyr Met Asp Glu Ile Glu Asp Glu Ile Thr Ile Ala Ile Leu
 85 90 95

Asn Tyr Asn Gly Pro Ser Thr Pro Phe Ile Glu Leu Pro Phe Leu Ser
 100 105 110

Gly Ser Tyr Asn Leu Leu Met Pro Val Ile Arg Arg Val Asp Asn Gly
 115 120 125

Ser Ala Ser His His His Ala Arg Gln His Tyr Glu Leu Pro Gly Met
 130 135 140

Gln Gln Tyr Met Phe Asn Val Arg Val Asp Gly Gln Ser Leu Val Ala
 145 150 155 160

Gly Val Ser Leu Ala Ile Val Asn Ile Asp Asp Asn Ala Pro Ile Ile
 165 170 175

Gln Asn Phe Glu Pro Cys Arg Val Pro Glu Leu Gly Glu Pro Gly Leu
 180 185 190

Thr Glu Cys Thr Tyr Gln Val Ser Asp Ala Asp Gly Arg Ile Ser Thr
 195 200 205

Glu Phe Met Thr Phe Arg Ile Asp Ser Val Arg Gly Asp Glu Glu Thr
 210 215 220

Phe Tyr Ile Glu Arg Thr Asn Ile Pro Asn Gln Trp Met Trp Leu Asn
 225 230 235 240

Met Thr Ile Gly Val Asn Thr Ser Leu Asn Phe Val Thr Ser Pro Leu
 245 250 255

His Ile Phe Ser Val Thr Ala Leu Asp Ser Leu Pro Asn Thr His Thr
 260 265 270

Val Thr Met Met Val Gln Val Ala Asn Val Asn Ser Arg Pro Pro Arg
 275 280 285

Trp Leu Glu Ile Phe Ala Val Gln Gln Phe Glu Glu Lys Ser Tyr Gln
 290 295 300

Asn Phe Thr Val Arg Ala Ile Asp Gly Asp Thr Glu Ile Asn Met Pro
 305 310 315 320

Ile Asn Tyr Arg Leu Ile Thr Asn Glu Glu Asp Thr Phe Ser Ile Glu
 325 330 335

Ala Leu Pro Gly Gly Lys Ser Gly Ala Val Phe Leu Val Ser Pro Ile
 340 345 350

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Asp	Arg	Asp	Thr	Leu	Gln	Arg	Glu	Val	Phe	Pro	Leu	Thr	Ile	Val	Ala
		355					360					365			
Tyr	Lys	Tyr	Asp	Glu	Glu	Ala	Phe	Ser	Thr	Ser	Thr	Asn	Val	Val	Ile
	370					375					380				
Ile	Val	Thr	Asp	Ile	Asn	Asp	Gln	Arg	Pro	Glu	Pro	Ile	His	Lys	Glu
385					390					395					400
Tyr	Arg	Leu	Ala	Ile	Met	Glu	Glu	Thr	Pro	Leu	Thr	Leu	Asn	Phe	Asp
				405					410					415	
Lys	Glu	Phe	Gly	Phe	His	Asp	Lys	Asp	Leu	Gly	Gln	Asn	Ala	Gln	Tyr
			420					425					430		
Thr	Val	Arg	Leu	Glu	Ser	Val	Asp	Pro	Pro	Gly	Ala	Ala	Glu	Ala	Phe
			435				440					445			
Tyr	Ile	Ala	Pro	Glu	Val	Gly	Tyr	Gln	Arg	Gln	Thr	Phe	Ile	Met	Gly
	450					455					460				
Thr	Leu	Asn	His	Ser	Met	Leu	Asp	Tyr	Glu	Val	Pro	Glu	Phe	Gln	Ser
465					470					475					480
Ile	Thr	Ile	Arg	Val	Val	Ala	Thr	Asp	Asn	Asn	Asp	Thr	Arg	His	Val
			485						490					495	
Gly	Val	Ala	Leu	Val	His	Ile	Asp	Leu	Ile	Asn	Trp	Asn	Asp	Glu	Gln
			500					505					510		
Pro	Ile	Phe	Glu	His	Ala	Val	Gln	Thr	Val	Thr	Phe	Asp	Glu	Thr	Glu
		515					520					525			
Gly	Glu	Gly	Phe	Phe	Val	Ala	Lys	Ala	Val	Ala	His	Asp	Arg	Asp	Ile
	530					535					540				
Gly	Asp	Val	Val	Glu	His	Thr	Leu	Leu	Gly	Asn	Ala	Val	Asn	Phe	Leu
545					550					555					560
Thr	Ile	Asp	Lys	Leu	Thr	Gly	Asp	Ile	Arg	Val	Ser	Ala	Asn	Asp	Ser
			565						570					575	
Phe	Asn	Tyr	His	Arg	Glu	Ser	Glu	Leu	Phe	Val	Gln	Val	Arg	Ala	Thr
			580					585					590		
Asp	Thr	Leu	Gly	Gln	Pro	Phe	His	Thr	Ala	Thr	Ser	Gln	Leu	Val	Ile
		595					600					605			
Arg	Leu	Asn	Asp	Ile	Asn	Asn	Thr	Pro	Pro	Thr	Leu	Arg	Leu	Pro	Arg
	610					615					620				
Gly	Ser	Pro	Gln	Val	Glu	Glu	Asn	Val	Pro	Asp	Ala	His	Val	Ile	Thr
625					630					635					640
Gln	Glu	Leu	Arg	Ala	Thr	Asp	Pro	Asp	Thr	Thr	Ala	Asp	Leu	Arg	Phe
				645					650					655	
Glu	Ile	Asn	Trp	Asp	Thr	Ser	Phe	Ala	Thr	Lys	Gln	Gly	Arg	Gln	Ala
			660					665					670		
Asn	Pro	Asp	Glu	Phe	Arg	Asn	Cys	Val	Glu	Ile	Glu	Thr	Ile	Phe	Pro
		675					680					685			
Glu	Ile	Asn	Asn	Arg	Gly	Leu	Ala	Ile	Gly	Arg	Val	Val	Ala	Arg	Glu
	690					695					700				
Ile	Arg	His	Asn	Val	Thr	Ile	Asp	Tyr	Glu	Glu	Phe	Glu	Val	Leu	Ser
705					710					715					720
Leu	Thr	Val	Arg	Val	Arg	Asp	Leu	Asn	Thr	Val	Tyr	Gly	Asp	Asp	Tyr
				725					730					735	
Asp	Glu	Ser	Met	Leu	Thr	Ile	Thr	Ile	Ile	Asp	Met	Asn	Asp	Asn	Ala
			740					745					750		
Pro	Val	Trp	Val	Glu	Gly	Thr	Leu	Glu	Gln	Asn	Phe	Arg	Val	Arg	Glu
		755					760					765			
Met	Ser	Ala	Gly	Gly	Leu	Val	Val	Gly	Ser	Val	Arg	Ala	Asp	Asp	Ile

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770					775					780					
Asp	Gly	Pro	Leu	Tyr	Asn	Gln	Val	Arg	Tyr	Thr	Ile	Phe	Pro	Arg	Glu
785					790					795					800
Asp	Thr	Asp	Lys	Asp	Leu	Ile	Met	Ile	Glu	Leu	Pro	His	Gly	Ser	Asn
				805					810					815	
Phe	Arg	Glu	His	Lys	Arg	Arg	Ile	Asp	Ala	Asn	Thr	Pro	Pro	Arg	Phe
			820					825					830		
His	Leu	Tyr	Tyr	Thr	Val	Val	Ala	Ser	Asp	Arg	Cys	Ser	Thr	Glu	Asp
		835					840					845			
Pro	Ala	Asp	Cys	Pro	Pro	Asp	Pro	Thr	Tyr	Trp	Glu	Thr	Glu	Gly	Asn
	850					855					860				
Ile	Thr	Ile	His	Ile	Thr	Asp	Thr	Asn	Asn	Lys	Val	Pro	Gln	Ala	Glu
865					870					875					880
Thr	Thr	Lys	Phe	Asp	Thr	Val	Val	Tyr	Ile	Tyr	Glu	Asn	Ala	Thr	His
				885					890					895	
Leu	Asp	Glu	Val	Val	Thr	Leu	Ile	Ala	Ser	Asp	Leu	Asp	Arg	Asp	Glu
			900					905					910		
Ile	Tyr	His	Met	Val	Ser	Tyr	Val	Ile	Asn	Tyr	Ala	Val	Asn	Pro	Arg
		915					920					925			
Leu	Met	Asn	Phe	Phe	Ser	Val	Asn	Arg	Glu	Thr	Gly	Leu	Val	Tyr	Val
	930					935					940				
Asp	Tyr	Glu	Thr	Gln	Gly	Ser	Gly	Glu	Val	Leu	Asp	Arg	Asp	Gly	Asp
945					950					955					960
Glu	Pro	Thr	His	Arg	Ile	Phe	Phe	Asn	Leu	Ile	Asp	Asn	Phe	Met	Gly
				965					970					975	
Glu	Gly	Glu	Gly	Asn	Arg	Asn	Gln	Asn	Asp	Thr	Glu	Val	Leu	Val	Ile
			980					985					990		
Leu	Leu	Asp	Val	Asn	Asp	Asn	Ala	Pro	Glu	Leu	Pro	Pro	Pro	Ser	Glu
		995					1000						1005		
Leu	Ser	Trp	Thr	Ile	Ser	Glu	Asn	Leu	Lys	Gln	Gly	Val	Arg	Leu	Glu
	1010					1015					1020				
Pro	His	Ile	Phe	Ala	Pro	Asp	Arg	Asp	Glu	Pro	Asp	Thr	Asp	Asn	Ser
1025				1030					1035					1040	
Arg	Val	Gly	Tyr	Glu	Ile	Leu	Asn	Leu	Ser	Thr	Glu	Arg	Asp	Ile	Glu
			1045					1050						1055	
Val	Pro	Glu	Leu	Phe	Val	Met	Ile	Gln	Ile	Ala	Asn	Val	Thr	Gly	Glu
			1060					1065					1070		
Leu	Glu	Thr	Ala	Met	Asp	Leu	Lys	Gly	Tyr	Trp	Gly	Thr	Tyr	Ala	Ile
	1075					1080						1085			
Tyr	Ile	Leu	Ala	Phe	Asp	His	Gly	Ile	Pro	Gln	Met	Ser	Met	Asn	Glu
1090					1095						1100				
Thr	Tyr	Glu	Leu	Ile	Ile	His	Pro	Phe	Asn	Tyr	Tyr	Ala	Pro	Glu	Phe
1105				1110					1115					1120	
Val	Phe	Pro	Thr	Asn	Asp	Ala	Val	Ile	Arg	Leu	Ala	Arg	Glu	Arg	Ala
			1125					1130					1135		
Val	Ile	Asn	Gly	Val	Leu	Ala	Thr	Val	Asn	Gly	Glu	Phe	Leu	Glu	Arg
		1140						1145				1150			
Ile	Ser	Ala	Thr	Asp	Pro	Asp	Gly	Leu	His	Ala	Gly	Val	Val	Thr	Phe
	1155					1160					1165				
Gln	Val	Val	Gly	Asp	Glu	Glu	Ser	Gln	Arg	Tyr	Phe	Gln	Val	Val	Asn
	1170					1175					1180				
Asp	Gly	Glu	Asn	Leu	Gly	Ser	Leu	Arg	Leu	Leu	Gln	Ala	Val	Pro	Glu
1185				1190					1195					1200	

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Glu Ile Arg Glu Phe Arg Ile Thr Ile Arg Ala Thr Asp Gln Gly Thr
 1205 1210 1215
 Asp Pro Gly Pro Leu Ser Thr Asp Met Thr Phe Arg Val Val Phe Val
 1220 1225 1230
 Pro Thr Gln Gly Glu Pro Arg Phe Ala Ser Ser Glu His Ala Val Ala
 1235 1240 1245
 Phe Ile Glu Lys Ser Ala Gly Met Glu Glu Ser His Gln Leu Pro Leu
 1250 1255 1260
 Ala Gln Asp Ile Lys Asn His Leu Cys Glu Asp Asp Cys His Ser Ile
 1265 1270 1275 1280
 Tyr Tyr Arg Ile Ile Asp Gly Asn Ser Glu Gly His Phe Gly Leu Asp
 1285 1290 1295
 Pro Val Arg Asn Arg Leu Phe Leu Lys Lys Glu Leu Ile Arg Glu Gln
 1300 1305 1310
 Ser Ala Ser His Thr Leu Gln Val Ala Ala Ser Asn Ser Pro Asp Gly
 1315 1320 1325
 Gly Ile Pro Leu Pro Ala Ser Ile Leu Thr Val Thr Val Thr Val Arg
 1330 1335 1340
 Glu Ala Asp Pro Arg Pro Val Phe Val Arg Glu Leu Tyr Thr Ala Gly
 1345 1350 1355 1360
 Ile Ser Thr Ala Asp Ser Ile Gly Arg Glu Leu Leu Arg Leu His Ala
 1365 1370 1375
 Thr Gln Ser Glu Gly Ser Ala Ile Thr Tyr Ala Ile Asp Tyr Asp Thr
 1380 1385 1390
 Met Val Val Asp Pro Ser Leu Glu Ala Val Arg Gln Ser Ala Phe Val
 1395 1400 1405
 Leu Asn Ala Gln Thr Gly Val Leu Thr Leu Asn Ile Gln Pro Thr Ala
 1410 1415 1420
 Thr Met His Gly Leu Phe Lys Phe Glu Val Thr Ala Thr Asp Thr Ala
 1425 1430 1435 1440
 Gly Ala Gln Asp Arg Thr Asp Val Thr Val Tyr Val Val Ser Ser Gln
 1445 1450 1455
 Asn Arg Val Tyr Phe Val Phe Val Asn Thr Leu Gln Gln Val Glu Asp
 1460 1465 1470
 Asn Arg Asp Phe Ile Ala Asp Thr Phe Ser Ala Gly Phe Asn Met Thr
 1475 1480 1485
 Cys Asn Ile Asp Gln Val Val Pro Ala Asn Asp Pro Val Thr Gly Val
 1490 1495 1500
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 1505 1510 1515 1520
 Tyr Pro Tyr Ser Leu Met Arg
 1525

<210> SEQ ID NO 13
 <211> LENGTH: 27
 <212> TYPE: DNA
 <213> ORGANISM: Heliothis virescens
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (1)..(27)

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 Tyr Asn Thr Asn Thr Ala Gln Leu Val
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27

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<210> SEQ ID NO 14
 <211> LENGTH: 9
 <212> TYPE: PRT
 <213> ORGANISM: *Heliothis virescens*

<400> SEQUENCE: 14

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<210> SEQ ID NO 15
 <211> LENGTH: 287
 <212> TYPE: DNA
 <213> ORGANISM: *Heliothis virescens*

<400> SEQUENCE: 15

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 cactcgcctca gcgaccccg tgcgcatac acgaccgcac gggcaacgcy cgattttctc 180
 ttgtacatac ttcaatacag tcttctttgc aaatcgaagt ttcattgaac cgccgagacc 240
 atcatcctac atctggacct cggcgcctca gatttggctc ctgcgaa 287

<210> SEQ ID NO 16
 <211> LENGTH: 291
 <212> TYPE: DNA
 <213> ORGANISM: *Heliothis virescens*

<400> SEQUENCE: 16

gcttcaaccc ggggaatatg ttccggactgt cacatcgcg cggcctatga ggtcgcgcc 60
 gcacacgcca tcgtgcgcc cacctaagct gggccctcac catacgccg acccccggac 120
 actcgcctat cgaccccggt cgcgcataca cgaccgcacg cgcaacgcgc gatctactct 180
 tgtcacctat ctataatata gtcttctact ttgaacatcg aagttttatt gaaacgccga 240
 gaccagcaac ctacacctgc acctcggcgc tcaaacactg cccaactggt g 291

That which is claimed is:

1. A method of detecting resistance to *Bacillus thuringiensis* endotoxin in *Heliothis virescens* populations by screening for the presence of mutations having a sequence selected from the group consisting of SEQ ID NO: 3 or SEQ ID NO: 4.

2. A method of detecting resistance to *Bacillus thuringiensis* endotoxin in insect populations by screening for mutations that alter the structure or function of any protein encoded by the nucleotide sequence set forth in SEQ ID NO: 1.

3. A method of detecting resistance to *Bacillus thuringiensis* endotoxin in insect populations by screening for mutations that alter the structure or function of SEQ ID NO: 2 or homologues of SEQ ID NO: 2, wherein SEQ ID NO: 2 and said homologues of SEQ ID NO: 2 bind *Bacillus thuringiensis* endotoxin.

4. A method for detecting mutations in genes from insect populations by screening for the presence of insertions of a DNA sequence that hybridizes to SEQ ID NO: 4 or the complement of SEQ ID NO: 4 at 60° C. in 1×SSC.

5. A process for monitoring Bt resistance associated with the presence of an r1 allele in an insect population associated with transgenic crops comprising the steps of:
 obtaining DNA from an individual insect;
 amplifying said DNA using primers having nucleotide sequences of SEQ ID NO: 7, SEQ ID NO: 8, and SEQ ID NO: 9.

measuring the molecular size of said amplified DNA, thereby determining whether said individual has zero, one, or two copies of said r1 allele.

6. A method of detecting mutations in purified nucleic acid sequences obtained from an insect population by screening for a sequence of at least 24 contiguous nucleotides, wherein the at least 24 contiguous nucleotides are on a sequence selected from the group consisting of SEQ ID NO: 4, SEQ ID NO: 5, and SEQ ID NO: 6.

7. A method of detecting resistance to *Bacillus thuringiensis* endotoxin in insect populations by:
 providing purified genomic DNA from an individual insect;
 performing PCR using oligonucleotide primers of 24 nucleotides or greater, identical in at least 16 positions

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of 24 to any sequence of 24 contiguous nucleotides of SEQ ID NO: 1 or the complement of SEQ ID NO: 1; determining the DNA sequences of the PCR products; computing the conceptual translations of the DNA sequences of the PCR products in all six reading frames; 5
comparing each of the predicted polypeptide sequences to SEQ ID NO: 2 or homologues thereof, wherein SEQ ID NO: 2 and said homologues of SEQ ID NO: 2 bind *Bacillus thuringiensis* endotoxin;

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whereupon the comparison, if indicating any change that would lead to the premature termination of the protein such that the last 12 amino acids or more of the carboxy-terminus of SEQ ID NO: 2 or homologues thereof would be predicted to be lacking in the mature protein, the insect will be at least heterozygous for resistance to *Bacillus thuringiensis* endotoxin.

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