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#### **Review** article

# The role of inflammation and potential pharmacological therapy in intracranial aneurysms



AND NEUROSURGERY

Wojciech Gruszka<sup>a,b,\*</sup>, Miłosz Zbroszczyk<sup>a</sup>, Jacek Komenda<sup>a</sup>, Katarzyna Gruszczyńska<sup>a</sup>, Jan Baron<sup>a</sup>

<sup>a</sup> Department of Radiology and Interventional Radiology, Medical University of Silesia, Katowice, Poland <sup>b</sup> Pathophysiology Unit, Department of Pathophysiology, Medical Faculty in Katowice, Medical University of Silesia, Katowice, Poland

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#### ABSTRACT

Intracranial aneurysms remain important clinical concern. There is relatively low risk of rupture of symptomless aneurysms incidentally found in MRA or CTA performed due to other indications. Not all of the intracranial aneurysms should or can be treated with neurosurgery intervention or endovascular embolization. Clinical strategy for small, symptomless, unruptured aneurysms is still questionable. Mechanisms underlying aneurysms formation, progression and rupture are poorly understood. Inflammation is one of the factors suspected to participate in these processes. Therefore the aim of this manuscript is to present current state of knowledge about the role of inflammation in the formation and progression of intracranial aneurysms and in their rupture process. Current knowledge about possible pharmacological treatment of intracranial aneurysms will also be presented. Macrophages infiltration seems to participate in the formation of intracranial aneurysms. Inhibition of signals sent by macrophages may prevent the aneurysms formation. Inflammation present in the wall of the aneurysm seems to be also related to the aneurysm's rupture risk. However it does not seem to be the only cause of the degeneration, but it can be a possible target of drug therapy. Some preliminary studies in humans indicate the potential role of aspirin as a factor that decrease the level of inflammation and lower the risk of rupture of intracranial aneurysms. However further research including a greater number of subjects and a prospective randomized design are necessary to assess the role of aspirin in preventing strategy for small, symptomless, unruptured intracranial aneurysms.

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E-mail address: wojtekgruszka@yahoo.pl (W. Gruszka).

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<sup>\*</sup> Corresponding author at: Pathophysiology Unit, Department of Pathophysiology, Medical University of Silesia, Medyków Street 18 20, 40-752 Katowice, Poland.

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#### 1. Introduction

Intracranial aneurysms remain important clinical issue due to its high prevalence in general population. According to American Heart Association Statistics Committee and Stroke Statistics Subcommittee [1] in every 1 000 000 adults of mean age of 50 years about 32 000 have intracranial aneurysm. Every year about 80 of them rupture causing subarachnoid hemorrhage that can be the cause of death of about 50% of patients. About 1/3 of patients who survive will likely suffer from a major neurological deficits. Only neurosurgery intervention or endovascular embolization is medical treatment strategy to prevent rupture or to treat ruptured aneurysms. Both procedures are not fully safe for patients and are associated with the risk of complications, including death. Up to now there is no recognized pharmacological strategy for aneurysms treatment.

The risk of rupture of symptomless aneurysms, which were incidentally found in MRA or CTA performed due to other indications, is relatively low [2]. There is no definite method that can predict this risk, as it is related to gender, age, size of aneurysms, its location and irregular shape, growth time, coexistence of other vascular lesions, family history of SAH and prior SAH. Smoking, excessive alcohol consumption and hypertension are the only modifiable risk factors [2]. In American Heart Association/American Stroke Association 2015'Guidelines [2] there is no indication which aneurysm size should be treated. Also in European Stroke Association 2013'Guidelines lacks such informations [3].

Taking above into account, not all of the intracranial aneurysms should or can be treated by surgery or endovascular embolization. One of the obstacles can be the lack of patients consent or technical problems including atherosclerosis in blood vessels, location of aneurysms, making surgical or endovascular access difficult or even impossible. This cause a question about the clinical strategy for small aneurysms, not included to the treatment. In these cases only regular MRA or CTA examinations are recommended [2]. However there is a lack of optimal time interval for such check-ups [2]. Another problem is the anxiety associated with potential rupture. Patients knowing about intracranial aneurysm in their heads ask for the treatment. Argumentation that the risk of rupture is low or lower that the risk associated with neurosurgery intervention or endovascular embolization can be difficult to accept by some patients. Recent study by Yamashiro et al. [4] showed that preoperative and postoperative quality of life of patients with intracranial aneurysms is significantly limited. Potential pharmacological therapy in above cases seems to be a reasonable solution.

Mechanisms regulating aneurysms formation, progression and rupture are poorly understood. Inflammation is one of the factors suspected to participate in these processes. Detailed analysis of particles taking part in this state seems to lead to identification of therapeutic targets and creation pharmacological strategy which should inhibit the ruptures. Therefore the aim of this manuscript is to present current state of knowledge about the role of inflammation in the formation and progression of intracranial aneurysms and in their rupture process. Also current knowledge about possible pharmacological treatment of intracranial aneurysms will be presented.

#### 2. Formation of intracranial aneurysms

In most of human intracranial aneurysms it is impossible to say when and how aneurysm occurred. That's way all of the studies investigating aneurysms formations are based on animals. In one of the first studies by Kanematsu et al. [5] mice aneurysms were induced using a combination of single injections of elastase into the cerebrospinal fluid and angiotensin II induced hypertension. Next in the half of mice the reduction in the number of macrophages was induced using clodronate liposome. In the group without blocked macrophage the occurrence of aneurysms was 6 out of 10; in the group with blocked macrophage - 1 out of 10, what suggest that reduction in the number of macrophages inhibits the formation of aneurysms. In the same study [5] also reduced incidence of aneurysms in mice lacking monocyte chemotactic protein-1 (MCP-1), which is critical chemotactic factor for proper macrophage function, compared with the incidence of aneurysms in wild type mice, were observed. Secretion of proteases destroying stromal tissue (matrix), triggering oxidative stress and secretion of inflammatory cytokines can link macrophages with the process of aneurysms formation. What is more, Aoki et al. [6], based on mice and rats with induced aneurysms studies, suggested that induction of macrophage activity is based on cyclooxygenase 2 (COX-2) pathway activation and prostaglandin E (PGE2) via prostaglandin E receptor (EP2) through an amplifying loop and activation of nuclear factor-кВ (NF-кВ) in the endothelium of the cerebral arteries. It was also suggested in latter study be Aoki et al. [7] that blocking tumor necrosis factor  $\alpha$  receptor (TNF $\alpha$ R1) for TNF reduced the formation of aneurysms. These results were confirmed by other authors [8]. In the study by Nuki et al. [9] metalloproteinase-9 (MMP-9) knockout mice, but not MMP-2 knockout mice, had a reduced incidence of intracranial aneurysms. Metalloproteinases, produced mainly by macrophages, are thought to destroy arterial wall extracellular matrix.

The role of the most important inflammation pathways in the formation, progression and rupture of intracranial aneurysms is illustrated in Scheme 1.

Later studies focused on pharmacological agents, which can inhibit above described processes. In the study by Nuki et al. [9] doxycycline, a broad-spectrum MMP inhibitor, reduced the incidence of aneurysm to 10%. Among others, treatment with simvastatin, pitavastatin, pravastatin, nifedipine, olmesartan, ibudilast and erythropoietin (EPO) [10–15] suppressed the development of intracranial aneurysms in different mechanisms. All above studies were also based on animals, however the drugs used in them should be a candidate for medical treatment strategy of intracranial aneurysms in humans. Moreover, COX-2 inhibitors like aspirin or indomethacin [16] should be taken into account.

Summarizing, the inflammation seems to participate in the formation of intracranial aneurysms. Pharmacological inhibition of macrophage infiltration and signaling (via the COX-2 or NF- $\kappa$ B or TNF $\alpha$  pathways inhibition) may inhibit the formation of aneurysms.



Scheme 1 – The role of the most important inflammation pathways in the formation, progression and rupture of intracranial aneurysms.

#### 3. Progression of intracranial aneurysms

The role of inflammation in the progression process of intracranial aneurysms were also studied before. However, most of the above cited authors did not distinguish clearly between aneurysm formation and progression. What is more, below-described studies on rupture of intracranial aneurysms involved data about intracranial aneurysms wall destruction, so that progression process. Studies on progression or growth of aneurysms in general seems to need longer follow-ups so that seems to be more difficult to design.

Starke et al. has shown that inhibition of  $TNF\alpha$  by 3,6' dithiothalidomide (DTH) reduce progression of intracranial aneurysms in rats [8]. It was also shown in rats that simvastatin suppresses intracranial aneurysms progression. Statins seem to inhibit the expression of several matrix metalloproteinases, including MMP-2 and MMP-9 involved in inflammation [10]. Furthermore, the similar suppressing effect was observed in the study with pitavastatin. In this case reduction in progression of intracranial aneurysms were explained by inhibition of NF-KB by pitavastatin [11]. Also nifedipine through inhibition of NF-KB suppressed intracranial aneurysms progression in the study by Aoki et al. [12]. Further studies tried to focus on destruction and remodeling of aneurysm wall. Marbacher et al. [17] induced in rats 24 saccular aneurysms of the abdominal aorta - in half of them number of mural cells were reduced. Aneurysms with the reduced number of mural cells were growing and 50% of growing aneurysms ruptured in comparison to not growing. As concluded, the loss of mural cells causes an increase in the size of aneurysm and its rupture. This observations were confirmed by further study in which a transplantation of smooth muscle cells into the luminal thrombus of intracranial aneurysms in rats reduced their destruction resulting in lower recurrence and lower growth of aneurysms [18]. On the other hand, there is no direct evidence that the complement activation affects directly smooth muscles cells [19] and there is no direct evidence on apoptosis induction by inflammatory cells (activation of Caspase 8) in the wall of aneurysms in humans. This indicates a potential role of other factors in the activation of apoptosis process. In this field, the role of Caspase 9 and oxidative stress response is suspected [20]. Also hemooxygenaze-1, secreted by certain inflammatory cells, accompanies the destruction of the walls of aneurysms and ruptures [20]. Recently Ollikainen et al. [21,22] have indicated the presence of mast cells and lipid accumulation in the aneurysms wall and its contribution to remodeling and destruction of this wall.

Another problem related to the progression of intracranial aneurysms is their morphology which seems to be linked to hemodynamic factors and inflammation. Irregular shape, particularly presence of daughter sac, seems to be related to higher risk of rupture. In the recent study by Abboud et al. [23] risk of rupture increased from single sac with regular margin, through aneurysms with irregular margin and with daughter sac, and was the highest in multilobulated intracranial aneurysm. Formation of daughter sac in intracranial aneurysms seems to be related to hemodynamic factors causing local elevation of pressure at the site of aneurysms without surrounding structure, which may cancel perpendicular wall tension [24]. However, it seems that there are no studies investigating the role of inflammation factors on the formation of daughter sac in intracranial aneurysms.

In spite of the small amount of studies focusing only on progression of intracranial aneurysms all cited studies indi-

cate that the inflammation may participate also in this process. Pharmacological inhibition of macrophage infiltration and signaling (via the COX-2 or NF- $\kappa$ B or TNF $\alpha$  pathways inhibition) may inhibit also the progression of intracranial aneurysms.

#### 4. Rupture of intracranial aneurysms

It is difficult to predict which aneurysm will rupture and cause hemorrhages. It is suggested that the risk of rupture of intracranial aneurysms is related among other to gender, size of the aneurysm, its location and irregular shape, coexistence of other vascular lesions, smoking and hypertension [2,25-28]. However this does not exclude that other factors especially on biochemical level take part in this process. Frösen et al. [29] in histological examination of the material obtained during neurosurgical clipping of human 24 unruptured and 42 ruptured intracranial aneurysms found an increased infiltration of macrophages and proliferation of smooth muscle cells in ruptured aneurysms. In addition, in ruptured aneurysms M1/M2 subsets of macrophages balance was lost in favor of M1 proinflammatory cells [30]. Level of metalloproteinases, produced by macrophages, seems to be higher in ruptured than in unruptured aneurysms in humans [31]. In histological examination of the material obtained during neurosurgical clipping of human intracranial aneurysms Tulamo et al. [19] found that in addition to the macrophages infiltration, an activation of complement (humoral response) in ruptured aneurysms is present. In the histological examination by Gounis et al. [32] of the material obtained during the clipping of 20 unruptured and 3 ruptured human intracranial aneurysms, all ruptured aneurysms had increased myeloperoxidase activity. In another study a 2.7-fold higher concentration of myeloperoxidase was found in blood drawn directly from the lumen of intracranial aneurysms in comparison to peripheral blood [33]. It can be concluded that myeloperoxidase, secreted mainly by neutrophils and macrophages, results in oxidative stress and is accompanied by an increased risk of aneurysms rupture.

As discussed, many studies confirmed that inflammation accompanies the rupture of aneurysms in humans. There are other factors like loss of smooth muscle cells that seems to contribute to destruction of aneurysms wall and to the ruptures of aneurysms, as well. Some inflammatory cells may cause oxidative stress and this in turn may affect the ruptures of aneurysms.

It is unknown how well-known risk factors like smoking or hypertension [2,25–28] can affect or trigger the inflammation and cause rupture. Recent study on humans by Cebral et al. [34] indicated that flow conditions in saccular intracranial aneurysms, assessed on the basis of patient-specific computational models of hemodynamic (computational fluid dynamics – CFD) created from preoperative CT angiographies were associated with wall remodeling. More diffuse inflow was associated with degenerated and decellularized saccular aneurysm walls, assessed in histological examinations of material obtained in neurosurgical clipping. Also low flow conditions were associated with wall changes. These observations suggest that endothelial injury may be a mechanism by which flow induces the inflammation in the intracranial aneurysms wall. Also Jamous et al. [35,36] indicated that endothelial cell injury is the earliest change in aneurysm wall. However there are many controversy about the role of hemodynamic factors on the rupture risk of intracranial aneurysms [34,37-39]. In the last review and meta analysis study Can et al. [37] showed that increase in wall shear stress (WSS), defined as tangiental frictional force exerted by flowing blood on the arterial endothelium, proportional to the blood viscosity and velocity gradients [38], may contribute to aneurysms formation, whereas low WSS seems to be associated with raptured aneurysms. It can be explained that high WSS implicates wall remodeling and potential destruction via endothelial injury. Low WSS, on the other hands, can cause localized stasis of blood flow near the aneurysms wall causing endothelial dysfunction and elevation of proinflammatory factors [37,39]. Interpretation of the results of studies on hemodynamic factors remains difficult, while among others except WSS other indexes have been proposed and used [37], there are differences induced by diverse configurations and geometries of vessels of the anterior and posterior circulations [40]. Furthermore, some studies investigating hemodynamics factors were performed for sidewall aneurysms, while others - for bifurcation aneurysms [37].

There is a group of other factors involved in rupture process. It cannot be excluded that they modify above described observations. As proved by Pena Silva et al. [41] angiotensin (1-7) via Mas receptor reduces the formation of intracranial aneurysms and reduces their rupture probability. Shimada et al. [42] indicated also the protective role of angiotensin (1-7) however via angiotensin II type 2 receptor (ATR2) pathway. What is more, angiotensin (1–7) in this study reduced the expression of  $TNF\alpha$ . In another study Shimada et al. [43] showed that activation of PPAR-y (Peroxisome Proliferator-Activated Receptor-y) on macrophages by pioglitazone (PGZ) reduced macrophages infiltration and level of inflammatory cytokines (interleukin-1 and interleukin-6) resulting in decreasing aneurysm rupture risk. Hasan et al. [44] confirmed this results, indicating the important role of smooth muscle cell PPAR-y. Pena Silva et al. [45] suggests that hepatocyte growth factor (HGF) reduces inflammation and prevents rupture of intracranial aneurysms. In another study by Tada et al. [46,47] estrogen (via  $\beta$  receptor for estrogen) seems to prevent the rupture of intracranial aneurysms. This seems to stay in line with clinical observations that the prevalence of hemorrhage caused by intracranial aneurysms rupture in postmenopausal women is higher than in premenopausal women [48]. Also hormonal replacement therapy (HRT) including estrogens seems to decrease rupture risk in postmenopausal women [49]. All above studies were based on animals.

Furthermore, comparison of gene expression by Kurki et al. [50] of 8 ruptured and 11 unruptured human aneurysms showed that the accumulation of lipids was strongly linked with rupture. Additionally, this seems to be linked to the fact that lipid accumulation in the wall of aneurysms is associated with loss of wall cells [51] and is associated with an activation of the complement system [52]. On the other hand, Pyysalo et al. [53,54] showed the presence of the bacterial DNA specific to the mouth in the intracranial aneurysms. This results raised

the question about potential role of bacteria in the formation and rupture of intracranial aneurysms.

Summarizing, inflammation in the wall of the aneurysm seems to be accompanied by destruction of the wall and the risk of rupture. This suggest a potential role of inflammation as a marker of rupture risk. If proper assessment tool is created, we will be able to recognize aneurysms with high risk of rupture and treat only them. This can be also useful in case of SAH and multiple intracranial aneurysms, when it is impossible to say which aneurysm caused hemorrhage. Assessing the inflammation state should help clinicians to indicate this aneurysm [55,56]. Secondly, inflammation does not seem to be the only cause of the destruction of the wall of the aneurysm, but it can be a possible target of drug therapy.

## 5. Perspectives of possible pharmacological treatment of intracranial aneurysms

There are some studies focusing on the role of aspirin as a well-known agent against inflammation. Except for blocking COX-2 and microsomal prostaglandin E2 synthase 1 (mPGES-1), aspirin was shown to inhibit MMP-2 and MMP-9 expression [57], TNF $\alpha$  in smooth muscle cells [58] and to reduce NF- $\kappa$ B activity [58,59].

In the study by Li et al. [60], firstly intracranial aneurysms in rats were induced. The group was divided into aspirin-treated and untreated control group. Then the aneurysms wall were histologically examined. As concluded, aspirin reduced the destruction of the wall of the aneurysm and lowered inflammatory markers (reduced expression of MMP-2 and MMP-9, reverse upregulation of NF- $\kappa$ B, MCP-1 and vascular cell adhesion molecule-1 – VCAM-1).

A retrospective evaluation by Hasan et al. [61] of 271 untreated aneurysms of patients from International Study of Unruptured Intracranial Aneurysms (ISUIA), in which patients were divided on the basis of interview into group using aspirin (≥3 times a week) and groups of nonusers of aspirin ("less than once o month" and "between once a month and twice a week"). In the group of patients taking aspirin (≥3 times a week) a fewer incidents of aneurysms ruptures were observed. Another retrospective assessment of 1797 patients with intracerebral haemorrhages and 1340 patients with SAH by Garcia-Rodriguez et al. [62] showed that aspirin was not associated with the risk of intracerebral bleeding, however chronic low-dose of aspirin (75-300 mg/day) reduced the risk of SAH. Results of the retrospective study by Gross et al. [63] of 747 patients with brain aneurysms indicated that the risk of SAH is lower in the group receiving aspirin compared to those not taking it (28% vs 40%). However in this study among patients with SAH there were no significant difference in presenting clinical and radiological grade, measured by Hunt-Hess and Fisher Scales, between patients taking and nottaking aspirin. What is more, aspirin use was not associated with 1-year poor outcome.

Another study design was proposed by Hasan et al. [64]. In this study 11 patients with intracranial aneurysms were divided into two groups: 6 people – treated with aspirin (81 mg/ day) and 5 people – control group. Initially inflammation in aneurysms wall was assessed in MR enhanced with Ferumoxytol. After three months second MR scanning was performed to assess the inflammation process. Then, surgical clipping of aneurysms was performed and obtained material was histologically examined. Both histological and MR results suggested that aspirin reduced inflammation in the wall of aneurysms. Ferumoxytol used in this study is a relatively new nanoparticle containing iron oxide coated by carbohydrate shell. It belongs to ultrasmall superparamagnetic iron oxide (USPIOs) and was initially developed for threating iron deficiency anemia in patients with chronic renal failure [65]. It can be used as enhancement medium and T2\*-weight gradient-echo imaging is usually applied for evaluation of macrophage infiltration within the aneurysms wall. Ferumoxytol is usually administered intravascular about 24-72 h before MR imaging. In this time it is absorbed by macrophages and they, as mentioned above, accumulate in the area of inflammation [66-68].

Objective comparison of above described studies is difficult because of their limitations like relatively small sample size. Presence of potential confounding variables should be also taken into account. It should be emphasized, that most of studies on humans had retrospective design and doses of aspirin used in particular studies were different. However, on the basis of all above preliminary studies in animals and humans, it seems that small doses of aspirin can reduce the risk of aneurysms rupture. Further research including a greater number of subjects and prospective randomized design to assess the role of aspirin as a potential preventing strategy for small, symptomless, unruptured intracranial are inevitable.

It is worth to mention that some of authors indicate the possible role of another pharmacological agents like tetracycline (minocycline and doxycycline) in prevention of intracranial aneurysms rupture [69]. However this study, like most of the above, was based on mice. Also previously mentioned statins and other medications [10–15], blocking formations of intracranial aneurysm in animal's models, should be taken into account as potential prevention strategy. However initial studies with statins performed on humans gave inconclusive results [70–72].

#### 6. Conclusion

Following above review of literature (however not a systematic review) it can be concluded that inflammation appears to mediate in the formation and progression of intracranial aneurysms and seems to be related to its rupture risk. This suggests a potential role of inflammation as a marker of aneurysm's rupture risk. Furthermore, inflammation can be a possible target for drug therapy. Preliminary studies on animals and humans indicate that small doses of aspirin can reduce the risk of aneurysms rupture. However further research including greater number of subjects and prospective randomized design are necessary to assess the role of aspirin as a potential preventing strategy for small, symptomless, unruptured intracranial aneurysms.

#### **Conflict of interest**

None declared.

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#### REFERENCES

- [1] Go AS, Mozaffarian D, Roger VL, Benjamin EJ, Berry JD, Blaha MJ, et al. Heart disease and stroke statistics – 2014 update: a report from the American Heart Association. Circulation 2014;129(3):e28–92.
- [2] Thompson BG, Brown Jr RD, Amin-Hanjani S, Broderick JP, Cockroft KM, Connolly Jr, et al. Guidelines for the management of patients with unruptured intracranial aneurysms: a guideline for healthcare professionals from the American Heart Association/American Stroke Association. Stroke 2015;46(8):2368–400.
- [3] Wiebers DO, Whisnant JP, Huston 3rd J, Meissner I, Brown Jr RD, Piepgras DG, et al. Unruptured intracranial aneurysms: natural history, clinical outcome, and risks of surgical and endovascular treatment. Lancet 2003;362(9378):103–10.
- [4] Yamashiro S, Nishi T, Koga K, Goto T, Kaji M, Muta D, et al. Improvement of quality of life in patients surgically treated for asymptomatic unruptured intracranial aneurysms. J Neurol Neurosurg Psychiatry 2007;78(5):497–500.
- [5] Kanematsu Y, Kanematsu M, Kurihara C, Tada Y, Tsou TL, van Rooijen N, et al. Critical roles of macrophages in the formation of intracranial aneurysm. Stroke 2011;42(1):173–8.
- [6] Aoki T, Nishimura M, Matsuoka T, Yamamoto K, Furuyashiki T, Kataoka H, et al. PGE2–EP2 signaling in endothelium is activated by haemodynamic stress and induced cerebral aneurysm through an amplifying loop via NF-KB. Br J Pharmacol 2011;163(6):1237–49.
- [7] Aoki T, Fukuda M, Nishimura M, Nozaki K, Narumiya S. Critical role of TNF-alpha-TNFR1 signaling in intracranial aneurysm formation. Acta Neuropathol Commun 2014;2:34.
- [8] Starke RM, Chalouhi N, Jabbour PM, Tjoumakaris SI, Gonzalez LF, Rosenwasser RH, et al. Critical role of  $TNF-\alpha$  in cerebral aneurysm formation and progression to rupture. J Neuroinflammation 2014;11:77.
- [9] Nuki Y, Tsou TL, Kurihara C, Kanematsu M, Kanematsu Y, Hashimoto T. Elastase-induced intracranial aneurysms in hypertensive mice. Hypertension 2009;54(6):1337–44.
- [10] Aoki T, Kataoka H, Ishibashi R, Nozaki K, Hashimoto N. Simvastatin suppresses the progression of experimentally induced cerebral aneurysms in rats. Stroke 2008;39(4):1276– 85.
- [11] Aoki T, Kataoka H, Ishibashi R, Nakagami H, Nozaki K, Morishita R, et al. Pitavastatin suppresses formation and progression of cerebral aneurysms through inhibition of the nuclear factor kappaB pathway. Neurosurgery 2009;64 (2):357–65.
- [12] Aoki T, Kataoka H, Ishibashi R, Nozaki K, Hashimoto N. Nifedipine inhibits the progression of an experimentally induced cerebral aneurysm in rats with associated downregulation of NF-kappa B transcriptional activity. Curr Neurovasc Res 2008;5(1):37–45.
- [13] Kimura N, Shimizu H, Eldawoody H, Nakayama T, Saito A, Tominaga T, et al. Effect of olmesartan and pravastatin on experimental cerebral aneurysms in rats. Brain Res 2010;1322:144–52.
- [14] Yagi K, Tada Y, Kitazato KT, Tamura T, Satomi J, Nagahiro S. Ibudilast inhibits cerebral aneurysms by down-regulating inflammation-related molecules in the vascular wall of rats. Neurosurgery 2010;66(3):551–9. discussion 559.

- [15] Xu Y, Tian Y, Wei HJ, Chen J, Dong JF, Zacharek A, et al. Erythropoietin increases circulating endothelial progenitor cells and reduces the formation and progression of cerebral aneurysm in rats. Neuroscience 2011;181:292–9.
- [16] Miralles M, Wester W, Sicard GA, Thompson R, Reilly JM. Indomethacin inhibits expansion of experimental aortic aneurysms via inhibition of the cox2 isoform of cyclooxygenase. Vasc Surg 1999;29(5):884–92. discussion 892–3.
- [17] Marbacher S, Marjamaa J, Bradacova K, von Gunten M, Honkanen P, Abo-Ramadan U, et al. Loss of mural cells leads to wall degeneration, aneurysm growth, and eventual rupture in rat aneurysm model. Stroke 2014;45(1):248–54.
- [18] Marbacher S, Frösén J, Marjamaa J, Anisimov A, Honkanen P, von Gunten M, et al. Intraluminal cell transplantation prevents growth and rupture in a model of rupture-prone saccular aneurysms. Stroke 2014;45(12):3684–90.
- [19] Tulamo R, Frösen J, Junnikkala S, Paetau A, Pitkäniemi J, Kangasniemi M, et al. Complement activation associates with saccular cerebral artery aneurysm wall degeneration and rupture. Neurosurgery 2006;59(5):1069–76.
- [20] Laaksamo E, Tulamo R, Liiman A, Baumann M, Friedlander RM, Hernesniemi J, et al. Oxidative stress is associated with cell death, wall degeneration, and increased risk of rupture of intracranial aneurysm wall. Neurosurgery 2013;72(1):109– 17.
- [21] Ollikainen E, Tulamo R, Frösen J, Lehti S, Honkanen P, Hernesniemi J, et al. Mast cells, neovascularization, and microhemorrhages are associated with saccular intracranial artery aneurysm wall remodeling. J Neuropathol Exp Neurol 2014;73(9):855–64.
- [22] Ollikainen E, Tulamo R, Lehti S, Lee-Rueckert M, Hernesniemi J, Niemelä M, et al. Smooth muscle cell foam cell formation, apolipoproteins, and ABCA1 in intracranial aneurysms: implications for lipid accumulation as a promoter of aneurysm wall rupture. J Neuropathol Exp Neurol 2016;75(7):689–99.
- [23] Abboud T, Rustom J, Bester M, Czorlich P, Vittorazzi E, Pinnschmidt HO, et al. Morphology of ruptured and unruptured intracranial aneurysms. World Neurosurg 2017;99:610–7.
- [24] Sugiyama SI, Endo H, Omodaka S, Endo T, Niizuma K, Rashad S, et al. Daughter sac formation related to blood inflow jet in an intracranial aneurysm. World Neurosurg 2016;96:396–402.
- [25] Caranci F, Briganti F, Cirillo L, Leonardi M, Muto M. Epidemiology and genetics of intracranial aneurysms. Eur J Radiol 2013;82(10):1598–605.
- [26] Rahman M, Smietana J, Hauck E, Hoh B, Hopkins N, Siddiqui A, et al. Size ratio correlates with intracranial aneurysm rupture status: a prospective study. Stroke 2010;41(5):916– 20.
- [27] Lindgren AE, Räisänen S, Björkman J, Tattari H, Huttunen J, Huttunen T, et al. De novo aneurysm formation in carriers of saccular intracranial aneurysm disease in Eastern Finland. Stroke 2016;47(5):1213–8.
- [28] Kleinloog R, de Mul N, Verweij BH, Post JA, Rinkel GJE, Ruigrok YM. Risk factors for intracranial aneurysm rupture: a systematic review. Neurosurgery 2018;82(4):431–40.
- [29] Frösen J, Pippo A, Paetau A, Kangasniemi M, Niemelä M, Hernesniemi J, et al. Remodeling of saccular cerebral artery aneurysms wall is associated with rapture: histological analysis of 24 unruptured and 42 ruptured cases. Stroke 2004;35(10):2287–93.
- [30] Hasan D, Chalouhi N, Jabbour P, Hashimoto T. Macrophage imbalance (M1 vs. M2) and upregulation of mast cells in wall of ruptured human cerebral aneurysms: preliminary results. J Neuroinflammation 2012;9:222.

- [31] Jin D, Sheng J, Yang X, Gao B. Matrix metalloproteinases and tissue inhibitors of metalloproteinases expression in human cerebral ruptured and unruptured aneurysm. Surg Neurol 2007;68(Suppl. 2):S11–6. discussion S16.
- [32] Gounis MJ, Vedantham S, Weaver JP, Puri AS, Brooks CS, Wakhloo AK, et al. Myeloperoxidase in human intracranial aneurysms: preliminary evidence. Stroke 2014;45(5):1474–7.
- [33] Chu Y, Wilson K, Gu H, Wegman-Points L, Dooley SA, Pierce GL, et al. Myeloperoxidase is increased in human cerebral aneurysms and increases formation and rupture of cerebral aneurysms in mice. Stroke 2015;46(6):1651–6.
- [34] Cebral J, Ollikainen E, Chung BJ, Mut F, Sippola V, Jahromi BR, et al. Flow conditions in the intracranial aneurysm lumen are associated with inflammation and degenerative changes of the aneurysm wall. Am J Neuroradiol 2017;38 (1):119–26.
- [35] Jamous MA, Nagahiro S, Kitazato KT, Satoh K, Satomi J. Vascular corrosion casts mirroring early morphological changes that lead to the formation of saccular cerebral aneurysm: an experimental study in rats. J Neurosurg 2005;102(3):532–5.
- [36] Jamous MA, Nagahiro S, Kitazato KT, Tamura T, Aziz HA, Shono M, et al. Endothelial injury and inflammatory response induced by hemodynamic changes preceding intracranial aneurysm formation: experimental study in rats. J Neurosurg 2007;107(2):405–11.
- [37] Can A, Du R. Association of hemodynamic factors with intracranial aneurysm formation and rupture: systematic review and meta-analysis. Neurosurgery 2016;78(4):510–20.
- [38] Meng H, Wang Z, Hoi Y, Gao L, Metaxa E, Swartz DD, et al. Complex hemodynamics at the apex of an arterial bifurcation induces vascular remodeling resembling cerebral aneurysm initiation. Stroke 2007;38(6):1924–31.
- [39] Meng H, Tutino VM, Xiang J, Siddiqui A. High WSS or low WSS? Complex interactions of hemodynamics with intracranial aneurysm initiation, growth, and rupture: toward a unifying hypothesis. Am J Neuroradiol 2014;35 (7):1254–62.
- [40] Doddasomayajula R, Chung B, Hamzei-Sichani F, Putman CM, Cebral JR. Differences in hemodynamics and rupture rate of aneurysms at the bifurcation of the basilar and internal carotid arteries. Am J Neuroradiol 2017;38(3):570–6.
- [41] Pena Silva RA, Kung DK, Mitchell IJ, Alenina N, Bader M, Santos RA, et al. Angiotensin 1–7 reduces mortality and rupture of intracranial aneurysms in mice. Hypertension 2014;64(2):362–8.
- [42] Shimada K, Furukawa H, Wada K, Wei Y, Tada Y, Kuwabara A, et al. Angiotensin-(1–7) protects against the development of aneurysmal subarachnoid hemorrhage in mice. J Cereb Blood Flow Metab 2015;35(7):1163–8.
- [43] Shimada K, Furukawa H, Wada K, Korai M, Wei Y, Tada Y, et al. protective role of peroxisome proliferator-activated receptor-y in the development of intracranial aneurysm rupture. Stroke 2015;46(6):1664–72.
- [44] Hasan DM, Starke RM, Gu H, Wilson K, Chu Y, Chalouhi N, et al. Smooth muscle peroxisome proliferator-activated receptor γ plays a critical role in formation and rupture of cerebral aneurysms in mice in vivo. Hypertension 2015;66 (1):211–20.
- [45] Pena Silva RA, Chalouhi N, Wegman-Points L, Ali M, Mitchell I, Pierce GL, et al. A novel role for endogenous HGF in the pathogenesis of intracranial aneurysms. Hypertension 2015;65(3):587–93.
- [46] Tada Y, Wada K, Shimada K, Makino H, Liang EI, Murakami S, et al. Estrogen protects against intracranial aneurysm rupture in ovariectomized mice. Hypertension 2014;63 (6):1339–44.
- [47] Tada Y, Makino H, Furukawa H, Shimada K, Wada K, Liang EI, et al. Roles of estrogen in the formation of intracranial

aneurysms in ovariectomized female mice. Neurosurgery 2014;75(6):690–5.

- [48] de Rooij NK, Linn FH, van der Plas JA, Algra A, Rinkel GJ. Incidence of subarachnoid haemorrhage: a systematic review with emphasis on region, age, gender and time trends. J Neurol Neurosurg Psychiatry 2007;78(12):1365–72.
- [49] Mhurchu CN, Anderson C, Jamrozik K, Hankey G, Dunbabin D, Australasian Cooperative Research on Subarachnoid Hemorrhage Study (ACROSS) Group. Hormonal factors and risk of aneurysmal subarachnoid hemorrhage: an international population-based, case–control study. Stroke 2001;32(3):606–12.
- [50] Kurki MI, Häkkinen SK, Frösen J, Tulamo R, von und zu Fraunberg M, Wong G, et al. Upregulated signaling pathways in ruptured human saccular intracranial aneurysm wall: an emerging regulative role of Toll-like receptor signaling and nuclear factor NF-KB, hypoxiainducible factor-1A, and ETS transcription factors. Neurosurgery 2011;68(6):1667–76.
- [51] Frösen J, Tulamo R, Heikura T, Sammalkorpi S, Niemelä M, Hernesniemi J, et al. Lipid accumulation, lipid oxidation, and low plasma levels of acquired antibodies against oxidized lipids associate with degeneration and rupture of the intracranial aneurysm wall. Acta Neuropathol Commun 2013;1:71.
- [52] Tulamo R, Frösen J, Junnikkala S, Paetau A, Kangasniemi M, Peláez J, et al. Complement system becomes activated by the classical pathway in intracranial aneurysm walls. Lab Investig 2010;90(2):168–79.
- [53] Pyysalo MJ, Pyysalo LM, Pessi T, Karhunen PJ, Öhman JE. The connection between ruptured cerebral aneurysms and odontogenic bacteria. J Neurol Neurosurg Psychiatry 2013;84(11):1214–8.
- [54] Pyysalo MJ, Pyysalo LM, Pessi T, Karhunen PJ, Lehtimäki T, Oksala N, et al. Bacterial DNA findings in ruptured and unruptured intracranial aneurysms. Acta Odontol Scand 2016;74(4):315–20.
- [55] Matouk CC, Mandell DM, Günel M, Bulsara KR, Malhotra A, Hebert R, et al. Vessel wall magnetic resonance imaging identifies the site of rupture in patients with multiple intracranial aneurysms: proof of principle. Neurosurgery 2013;72(3):492–6. discussion 496.
- [56] Kondo R, Yamaki T, Mouri W, Sato S, Saito S, Nagahata M, et al. Magnetic resonance vessel wall imaging reveals rupture site in subarachnoid hemorrhage with multiple cerebral aneurysms. No Shinkei Geka 2014;42(12):1147–50.
- [57] Yiqin Y, Meilin X, Jie X, Keping Z. Aspirin inhibits MMP-2 and MMP-9 expression and activity through PPARalpha/gamma and TIMP-1-mediated mechanisms in cultured mouse celiac macrophages. Inflammation 2009;32(4):233–41.
- [58] Sánchez de Miguel L, de Frutos T, González-Fernández F, del Pozo V, Lahoz C, Jiménez A, et al. Aspirin inhibits inducible nitric oxide synthase expression and tumour necrosis factor-alpha release by cultured smooth muscle cells. Eur J Clin Investig 1999;29(2):93–9.
- [59] Yotsui T, Yasuda O, Kawamoto H, Higuchi M, Chihara Y, Umemoto E, et al. Aspirin prevents adhesion of T lymphoblasts to vascular smooth muscle cells. FEBS Lett 2007;581(3):427–32.
- [60] Li S, Wang D, Tian Y, Wei H, Zhou Z, Liu L, et al. Aspirin inhibits degenerative changes of aneurysmal wall in a rat model. Neurochem Res 2015;40(7):1537–45.
- [61] Hasan DM, Mahaney KB, Brown Jr RD, Meissner I, Piepgras DG, Huston J, et al. Aspirin as a promising agent for decreasing incidence of cerebral aneurysm rupture. Stroke 2011;42(11):3156–62.
- [62] Garcia-Rodriguez LA, Gaist D, Morton J, Cookson C, González-Pérez A. Antithrombotic drugs and risk of hemorrhagic stroke in the general population. Neurology 2013;81(6):566–74.

- [63] Gross BA, Rosalind Lai PM, Frerichs KU, Du R. Aspirin and aneurysmal subarachnoid hemorrhage. World Neurosurg 2014;82(3):1127–30.
- [64] Hasan DM, Chalouhi N, Jabbour P, Dumont AS, Kung DK, Magnotta VA, et al. Evidence that acetylsalicylic acid attenuates inflammation in the walls of human cerebral aneurysms: preliminary results. J Am Heart Assoc 2013;2(1):e000019.
- [65] Lu M, Cohen MH, Rieves D, Pazdur R. FDA report: ferumoxytol for intravenous iron therapy in adult patients with chronic kidney disease. Am J Hematol 2010;85(5):315–9.
- [66] Hasan D, Chalouhi N, Jabbour P, Dumont AS, Kung DK, Magnotta VA, et al. Early change in ferumoxytol-enhanced magnetic resonance imaging signal suggests unstable human cerebral aneurysm: a pilot study. Stroke 2012;43 (12):3258–65.
- [67] Chalouhi N, Jabbour P, Magnotta V, Hasan D. The emerging role of ferumoxytol-enhanced MRI in the management of cerebrovascular lesions. Molecules 2013;18(8):9670–83.

- [68] Bashir MR, Bhatti L, Marin D, Nelson RC. Emerging applications for ferumoxytol as a contrast agent in MRI. J Magn Reson Imaging 2015;41(4):884–998.
- [69] Makino H, Tada Y, Wada K, Liang EI, Chang M, Mobashery S, et al. Pharmacological stabilization of intracranial aneurysms in mice: a feasibility study. Stroke 2012;43 (9):2450–6.
- [70] Marbacher S, Schläppi JA, Fung C, Hüsler J, Beck J, Raabe A. Do statins reduce the risk of aneurysm development? A case-control study. J Neurosurg 2012;116(3):638–42.
- [71] Yoshimura Y, Murakami Y, Saitoh M, Yokoi T, Aoki T, Miura K, et al. Statin use and risk of cerebral aneurysm rupture: a hospital-based case–control study in Japan. J Stroke Cerebrovasc Dis 2014;23(2):343–8.
- [72] Bekelis K, Smith J, Zhou W, MacKenzie TA, Roberts DW, Skinner J, et al. Statins and subarachnoid hemorrhage in Medicare patients with unruptured cerebral aneurysms. Int J Stroke 2015;10(Suppl. A100):38–45.