

# In vitro evaluation of electroporated gold nanoparticles and extremely-low frequency electromagnetic field anticancer activity against Hep-2 laryngeal cancer cells

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## Abstract

**Introduction.** The extremely-low frequency electromagnetic field (ELFEMF) has been proposed for use in cancer therapy since it was found that magnetic waves interfere with many biological processes. Gold nanoparticles (Au-NPs) have been widely used for drug delivery during cancer in vitro studies due to their low cytotoxicity and high biocompatibility. The electroporation of cancer cells in a presence of Au-NPs (EP Au-NPs) can induce cell apoptosis, alterations of cell cycle profile and morphological changes. The impact of ELFEMF and EP Au-NPs on morphology, cell cycle and activation of apoptosis-associated genes on Hep-2 laryngeal cancer cell line has not been studied yet.

**Materials and methods.** ELFEMF on Hep-2 cells were carried out using four different conditions: 25/50 mT at 15/30 min, while Au-NPs were used as direct contact (DC) or with electroporation (EP, 10 pulses at 200V, equal time intervals of 4 sec). MTT assay was used to check the toxicity of DC Au-NPs. Expression of *CASP3*, *P53*, *BAX* and *BCL2* genes was quantified using qPCR. Cell cycle was analyzed by flow cytometry. Hematoxylin and eosin (HE) staining was used to observe cell morphology.

**Results.** Calculated  $IC_{50}$  of DC Au-NPs  $24.36 \mu\text{M}$  ( $4.79 \mu\text{g/ml}$ ) and such concentration was used for further DC and EP AuNPs experiments. The up-regulation of pro-apoptotic genes (*CASP3*, *P53*, *BAX*) and decreased expression of *BCL2*, respectively, was observed for all analyzed conditions with the highest differences for EP AuNPs and ELFEMF 50 mT/30 min in comparison to control cells. The highest content of cells arrested in G2/M phase was observed in ELFEMF-treated cells for 30 min both at 25 or 50 mT, while the cells treated with EP AuNPs or ELFEMF 50 mT/15 min showed highest ratios of apoptotic cells. HE staining of electroporated cells and cells exposed to ELFEMF's low and higher frequencies for different times showed nuclear pleomorphic

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cells. Numerous apoptotic bodies were observed in the irregular cell membrane of neoplastic and necrotic cells with mixed euchromatin and heterochromatin.

**Conclusions.** Our observations indicate that treatment of Hep-2 laryngeal cancer cells with ELFEMF for 30 min at 25–50 mT and EP Au-NPs can cause cell damage inducing apoptosis and cell cycle arrest. (*Folia Histochemica et Cytobiologica* 2019, Vol. 57, No. 4, 159–167)

**Key words:** gold nanoparticles; electroporation; extremely-low frequency electromagnetic field; Hep-2 cells; apoptosis; qPCR

## Introduction

Laryngeal cancer most often refers to squamous cell carcinoma of the larynx. Other malignant tumors of the larynx (e.g., sarcoma, lymphoma, neuroendocrine tumors) are extremely rare in comparison. This disease consists of malignant tumors of mucosal origin that originate from the supra-glottis, glottis, and sub-glottis. Electromagnetic fields (EMFs) have been employed as useful tools in medical diagnosis. Recently, the use of EMFs has been expanded for therapeutic purposes because their interactions with living matter produce effects that initiate, accelerate or inhibit biological processes. Frequencies below 300 Hz are known as extremely low-frequency electromagnetic fields (ELFEMFs) which do not break molecular bonds due to low energy; therefore not causing direct DNA damage. Additionally, ELFEMFs are non-invasive and non-ionizing and even have non-thermal effects on cells and tissues. These properties have led to studies on the influence of ELFEMF on the development of various diseases, including cancer. While some researchers associate ELFEMF exposure with carcinogenesis [1, 2], other studies on experimental models and human cancers have shown that ELFEMF does not increase the risk of development of several tumor types, including liver cancer [3–5]. The ELFEMF treatment with tumor-specific frequencies is feasible and well-tolerated and may show positive biological efficacy in patients with advanced tumors [3–5]. Moreover, the exposure of female C3H/HeJ mice bearing human mammary adenocarcinoma to a frequency of 120 Hz at intensities of 4 and 5 mT resulted in a significant reduction in the growth of the tumors, which is a phenomenon associated with inhibition of angiogenesis [6]. Also, the exposure of female athymic nude mice with human breast cancer xenografts to a frequency of 120 Hz with an intensity of 15 mT, either alone or in combination with gamma radiation, resulted in decreased growth or reduced vascularization of the tumors [7]. Similarly, the effect of 50 Hz at 0.5  $\mu$ T and 0.5 mT on the development of chemically induced foci in rat livers showed a slight inhibition of foci formation [8]. Further studies on

ELFEMFs' impact on cancer cells are demanded since the current observations are insufficient as a proof of ELFEMF inhibition of carcinogenesis. The anticancer activity of gold nanoparticles (Au-NPs) has been studied against Dalton's lymphoma ascites (DLA) cell lines, human epithelial type 2 (Hep-2) cell lines and human leukemic monocyte lymphoma respectively [9, 10]. The anticancer effect of silver and gold nanoparticles against HepG-2 and lung cancer (A549) cell lines was performed [11, 12]. The results showed a good cytotoxic activity against the tested cancer cell lines. The concentration of Au-NPs plays an important role in their anticancer activity. Au-NPs showed expected effects against A549 cells in wide spectrum of concentrations: 100, 1 g, by 50  $\mu$ g, 25  $\mu$ g and 1  $\mu$ g. The lowest inhibitory effect on cells was observed for the concentration of 10  $\mu$ g. The cytotoxic effect of Au-NPs is the result of active physicochemical interaction of gold atoms with the functional groups of intracellular proteins, as well as with the nitrogen bases and phosphate groups in DNA. So, the aim of this study was to use electroporation in a presence of Au-NPs and ELFEMF effect on laryngeal carcinoma cell line (Hep-2) to evaluate genetic alteration under the effect of both electroporated Au-NPs and ELFEMF at different time and frequencies and related pathological changes [7].

## Materials and methods

**Cell culture.** Laryngeal adenocarcinoma cell line (Hep-2, ATCC: CCL-23) was kindly supplied from the Holding company, Tissue Culture Department, for production of vaccines, sera, and drugs (VACSERA, Giza, Egypt). Cells were maintained in MEM-E medium supplemented with 10% fetal bovine serum (GIBCO, Thermo Fisher Scientific, Waltham, MA, USA) in a humidified atmosphere of 5% CO<sub>2</sub> at 37°C (Jouan SA, Saint-herblain, Pays de la Loire, France). Cells were maintained according to manufacturing protocol where the growth medium was decanted and cells were washed with phosphate buffer saline (Adwia Pharmaceuticals, El Sharkeya, Egypt). Cells were treated with 0.25% trypsin enzyme and 0.05% (v/v) EDTA (GIBCO) for 5 min at 37°C. Detached cells were splatted according to need.

**Cell Viability and Au-NPs IC<sub>50</sub> evaluation (MTT Assay).**

Cytotoxicity was conducted according to the previous work [13] where human laryngeal cancer cells were propagated in 75 cm<sup>2</sup> cell culture (TPP-Swiss) as previously mentioned according to the previously reported work [13]. Cells were plated at a concentration of  $2 \times 10^5$  cell/ml in 96-well cell culture plates and incubated at 37°C for 24 h to achieve confluence. The growth medium was decanted and fresh medium containing 2 fold serially diluted Au-NPs dispensed to pre-cultured plate. 24 hrs later, dead cells were washed out using phosphate buffer saline (PBS, pH =  $7.2 \pm 0.2$  (Adwia) and 50  $\mu$ l of MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) stock solution (0.5 mg/ml) were added per well. After 4 h incubation period at 37°C, the supernatant was discarded and the formazan precipitate was solubilized by addition of 50  $\mu$ l/well of dimethyl sulfoxide (DMSO). Plates were incubated in the dark for 30 min at 37°C and absorbance was determined at a wavelength of 570 nm using microplate reader (ELx-800, Bio-Tek Instruments, Inc, Winooski, VT, USA). The cell viability percentage was calculated using the following formula: Viability percentage (%) = Mean OD of test dilution  $\times$  100/ Mean OD of control wells. The IC<sub>50</sub> value was determined using GraphPad Prism software (v.6, GraphPad Software, La Jolla, CA, USA). For cell cycle and apoptotic profile, Hep-2 cells were pre-cultured in 25 cm<sup>2</sup> surface area (SA) cell culture flasks, treated with the IC<sub>50</sub> of Au-NPs in MEM-E medium post decanting the growth medium. 24 h later the affected cells were collected and divided for cell cycle and genetic analysis, the cells were harvested and fixed gently with 70% (v/v) ethanol in PBS, maintained at a temperature of 4°C overnight, and re-suspended in PBS containing 40  $\mu$ g/ml PI (propidium iodide), 0.1 mg/ml RNase and 0.1% (v/v) Triton X-100 in a dark room. After 30 min at 37°C, the cells were analyzed using a flow cytometer (Becton-Dickinson, San Jose, CA, USA) equipped with an argon ion laser at a wavelength of 488 nm. The cell cycle and sub-G1 group were determined and analyzed, as described previously.

**Electroporation of laryngeal cancer cells.** Hep-2 cells were seeded in 75 cm<sup>2</sup> cell culture flasks and incubated at 37°C (5% CO<sub>2</sub>) until attachment. Cells were trypsinized and pelleted using cold centrifugation (Jouan-Ki21, ParisTech, France), the supernatant was decanted and Au-NPs (0.5  $\mu$ M/ mL) were added to 0.5 mL of Hep-2 cells and mixed well. Cells – Au-NPs (at the final concentration calculated as IC<sub>50</sub> for DC AuNPs) mix was placed in the sterile MicroPulser electroporation cuvette (Biorad, Hercules, CA, USA). Ten pulses were made at 200V and applied to the cells at equal time intervals of 4 s. Negative control was considered.

**Extremely-low frequency electromagnetic field treatment (ELFEMF).** Four sterile test tubes of detached Hep-2 cancer

cell line ( $2 \times 10^6$  cell/ml/tube) were categorized, where two of the tubes were subjected to the electromagnetic field as 25 mT for 15 and 30 mins in a cooling atmosphere. The other two tubes were exposed to the electromagnetic field at 50 mT for the same time intervals. The apparatus (Medical Physics, Swiss Tropical and Public Health Institute, Basel, Switzerland) was adjusted for each magnetic field frequency using a teslameter set for justification of a magnetic field. The test tubes were kept inside the center of the coil producing EMF. The test tubes were tested for apoptotic profile regarding the pro-and anti-apoptotic genes (P53, Bcl-2, Bax, caspase-3), cell cycle and histological profile were considered.

**Hematoxylin and eosin staining of Hep-2 cells.** Fifty microliters of Au-NPs electroporated and ELFEMF exposed cells were dispensed on clean slides (3 for each treatment). Slides were air-dried, methanol fixed and rehydrated in descending concentrations of alcohol (100%, 90%, 75% and 50%). Slides were washed in distilled water for 5 min. The slides were immersed in filtered hematoxylin (HE) stain for 3 min and washed with distilled water twice. Slides were immersed in filtered eosin stain for 5 seconds and washed with distilled water. Dried slides were immersed in xylene, mounted with Canada balsam then coverslips were placed and left to dry. Ten microscopic fields of each slide were photomicrographed using the power of 400 $\times$ . This was done using a digital camera (Canon, Tokyo, Japan), which was mounted on a light microscope. Images were transferred to the computer system for analysis. Field selection was based on the presence of the highest number of apoptotic cells. The photomicrographs were qualitatively evaluated for the presence of morphological criteria of apoptosis.

**Expression of apoptosis-related genes.** Total RNA was extracted from control, ELFEMF (25/50 mT for 15 and 30 min), 1 mM Au-NPs direct contact and electroporated exposed cells, respectively using RNeasy Mini Kit (Qiagen, Germantown, MD, USA) according to manufacturer's instructions. The concentration of extracted RNA was evaluated using a Beckman dual spectrophotometer (Beckman Instruments, Ramsey, MN, USA). The expression level of apoptosis-related genes; *P53* (F: 5'-TCAGATCCTAGC GTCGAGCCC-3' & R: 5'-GGGTGTGGAATCAACCCACAG-3'), *BAX* (F: 5'-ATGGACGGGTCCGGGGAGCA-3' & R: 5'-CCCAGTTGAAGTTGCCGTCA-3') and *BCL2* (F: 5'-GTGAACTGGGGGAGGATTGT-3' & R: 5'-GGAGAAATCAAACAGAGGCC-3') *CASP3* (F: 5'-CTCGGTCTGGTACAGATGTCTGA-3' & R: (5'-CATGGCTCAGAAGCACACAAAC-3) and housekeeping gene; *ACTB* (F: 5'-AGCGAGCATCCCCAAAGTT-3' & R: 5'-GGGCACGAAGGCTCATCATT-3) were determined using real-time PCR. 10 ng of the extracted total RNA from each sample was used for cDNA synthesis using high capacity cDNA reverse transcriptase kit (Applied

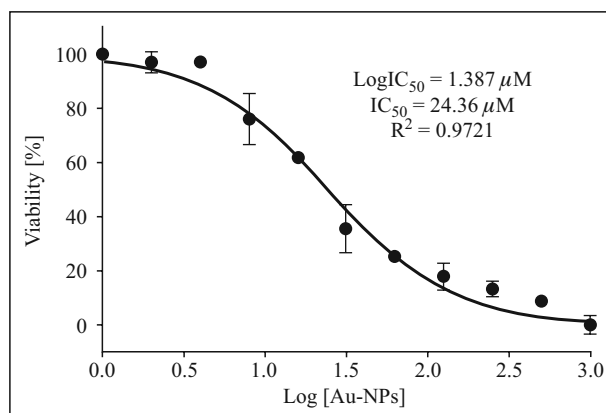
Biosystems-Thermo Fischer Scientific, USA). The obtained cDNA was subsequently amplified using Sybr Green I PCR master kit (Thermo Fisher Scientific Inc., Lithuania) using Step One apparatus (Applied Biosystems), as follows: 10 min at 95°C for enzyme activation followed by 40 cycles of 15 s at 95°C, 20 s at 55°C and 30 s at 72°C for the amplification step. Changes in the expression of each target gene were normalized relative to the mean critical threshold (CT) values of  $\beta$ -actin as housekeeping gene by the  $\Delta$ CT method.

**Cell cycle analysis.** Cell cycle distribution was examined by measuring the DNA content of nuclei labeled with propidium iodide (PI). Au-NPs electroporated and ELFEMF treated Hep-2 cells were pelleted by cold centrifugation (Jouan Ki-21-France), washed with 1 ml of cold PBS, centrifuged, and fixed in 70% cold ethanol at +4°C for 24 hrs. Subsequently, cells were washed twice and treated with RNase A (20 mg/ml) and PI (20 mg/ml), FITC conjugated Annexin-V according to the protocol described by the manufacturer for 30 min at 37°C in the dark. Finally, cell cycle distribution analysis was performed using flow cytometry and the percentages of cells at G1, S and G2/M phases were calculated by flow cytometry (Becton-Dickinson).

**Statistical analysis.** All experiments were carried out three independent times. Results were statistically analyzed using GraphPad Prism v. 6.07 (GraphPad Software, San Diego, CA, USA) with the following tests: t-test between 2 subgroups, one-way analysis of variance (ANOVA) between more than two subgroups, regression analysis and Spearman's correlation test. All results were presented as mean  $\pm$  SD. The difference was considered statistically significant at  $P < 0.05$ .

## Results

The toxicity test of Au-NPs was performed on Hep-2 cells and the results were shown in Figure 1. Based on that, the calculated  $IC_{50}$  concentration was  $24.36 \mu\text{M}$ /ml ( $4.79 \mu\text{g}/\text{ml}$ ). Such concentration of Au-NPs was used in further experiments with electroporation (EP Au-NPs) or by direct contact between Au-NPs and Hep-2 cells (DC Au-NPs). The influence of ELFEMF on Hep-2 cells was carried out using four different magnetic induction/time values: 25 mT/15 min, 25 mT/30 min, 50 mT/15 min and 50 mT/30 min. The expression profile of three pro-apoptotic genes (*CASP3*, *P53*, *BAX*) and one anti-apoptotic gene (*BCL2*) of Au-NPs and ELFEMF treated cells showed statistically different ratios (Fig. 2). We found the highest expression of *CASP3*, *P53* and *BAX* genes in EP Au-NPs and ELFEMFs cells at 50 mT/30 min (Fig. 2 A, B, C). Moreover, the tendency of increas-

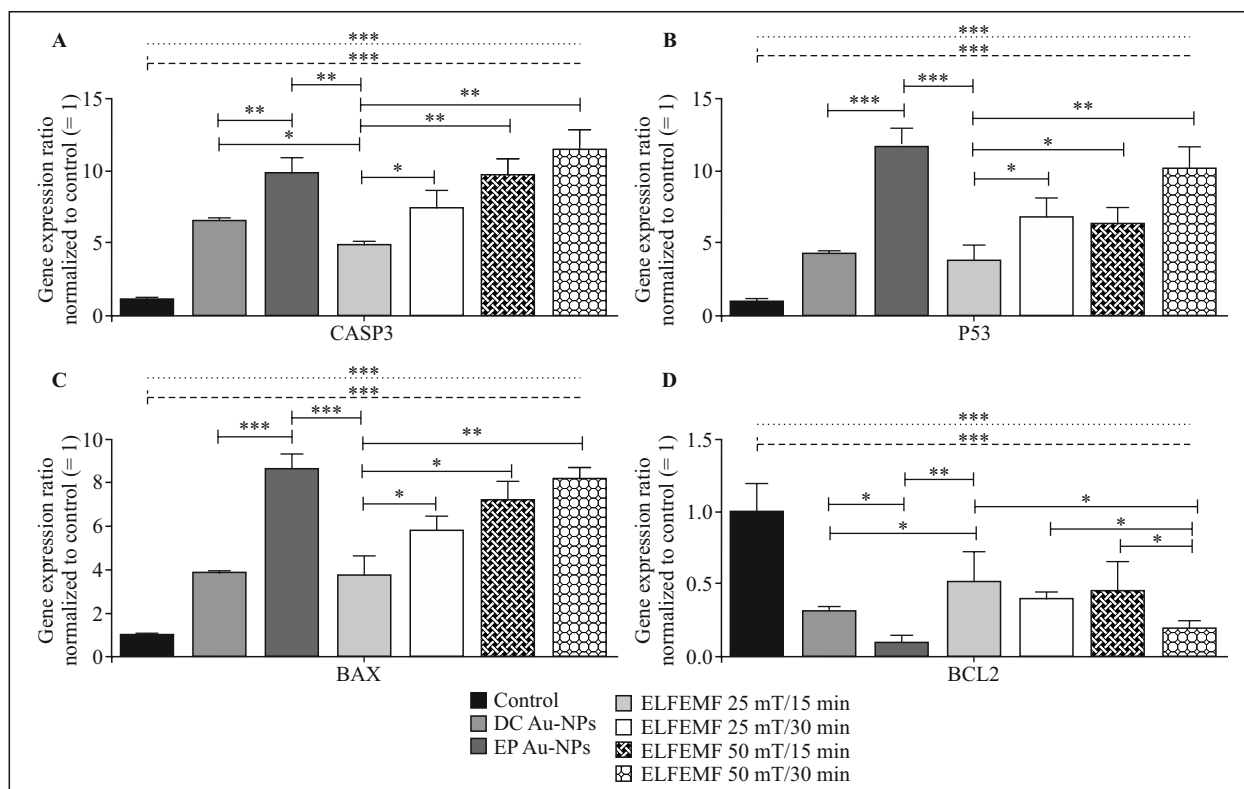


**Figure 1. Evaluation of cytotoxicity of gold nanoparticles on Hep-2 cells using MTT assay.** The calculated  $IC_{50}$  was  $24.26 \mu\text{M}$  using Graphpad Prism software. The values obtained from three separate experiments are shown (mean  $\pm$  SD) and the nonlinear regression curve was added to plot,  $R^2 = 0.9721$ .

ing expression of pro-apoptotic genes was observed in higher magnetic field/time values ( $P > 0.05$ , data not shown). We found that the electroporation with Au-NPs influenced the expression of *CASP3*, *P53* and *BAX* the similar way vs. direct contact with Au-NPs as increasing ELFEMF values. Regarding the *BCL2* gene, we did not observe the association between ELFEMF values and *BCL2* expression rate (except for the 25/15 vs. 50/30 mT/min values). Again, we found that EP Au-NPs-treated Hep-2 cells presented the lowest ratio of *BCL2* expression. The presented results showed that the electroporation with Au-NPs (at the concentration of  $24.36 \mu\text{M}$ ) as well as the usage of ELFEMF at 50 mT/30 min values activated the expression of analyzed pro-apoptotic genes and deactivated the expression of anti-apoptotic *BCL2* gene at the highest values of tested conditions.

Next, we checked the influence of Au-NPs and ELFEMF on the cell cycle of Hep-2 cells (Fig. 3). The significant G2/M phase arrest was observed for cells treated with all ELFEMFs (except for 50 mT/15 min) values (Fig. 3C). It seems that the G2/M arrest was associated with prolonged time of exposure rather than higher dose of electromagnetic field. The similar pattern of ELFEMF impact was found in apoptotic cell distribution (Fig. 3D). Although the distribution of apoptotic cells in EP Au-NPs treated cells was higher than in control cells (Fig. 3D), we did not observed cellular arrest in any of analyzed cell cycle phases when cells were treated with gold nanoparticles.

The microscopic observation of ELFEMF exposed and Au-NPs electroporated cells showed morphologi-



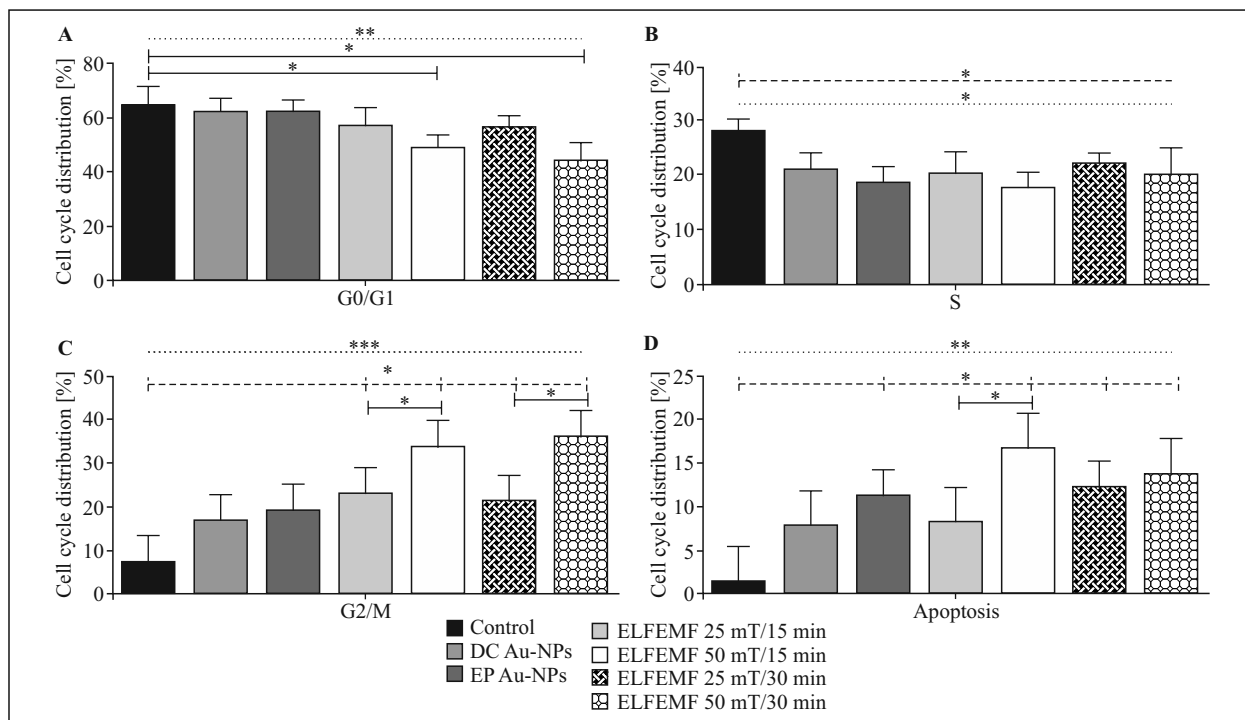
**Figure 2. Expression analysis of the *CASP3*, *P53*, *BAX* and *BCL2* genes in Hep-2 cells at the mRNA level.** Gene expression was assessed as described in the Materials and methods. (A–D) plots for *CASP3*, *P53*, *BAX* and *BCL2* genes' expression ratios in cells after treatment with different conditions, respectively. Bars and whiskers represent mean  $\pm$  standard deviation normalized to control samples. \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$ ; capped lines: t-test between subgroups; dotted line: one-way ANOVA between all subgroups; dashed line: t-test between control and tested cells, respectively. *CASP3*, caspase 3 gene; *P53*, tumor protein 53 gene; *BAX*, BCL2 associated X; *BCL2*, B-cell lymphoma 2 gene; DC Au-NPs, direct contact gold nanoparticles; EP Au-NPs, electroporation with gold nanoparticles; ELFEMF, extremely-low frequency electromagnetic field.

cal differences in comparison to control cells (Fig. 4). Hematoxylin and eosin staining of control, Hep-2 cells, showed regular cellular pleomorphic and nuclear pleomorphic cells (Fig. 4A), Au-NPs electroporated Hep-2 cells showed numerous apoptotic bodies, irregular cell membrane of neoplastic and necrotic cells with mixed euchromatin and heterochromatin (Fig. 4B). Also, Hep-2 cells showed numerous apoptotic bodies on post short term exposure to ELFEMF for 15 min at 25 mT and neoplastic cells with nuclear fragmentation and apoptotic body on post longer-term cell exposure (Fig. 4C); 30 min at 25 mT (Fig. 4D). Moreover, higher frequency at 50 mT for 15 min (Fig. 4E), exposed cells showed numerous apoptotic bodies and irregular neoplastic cell membrane and clear necrosis with mixed euchromatin and heterochromatin ruptured cell membrane of necrotic cells on post cell exposure to 50 mT for 30 min (Fig. 4F).

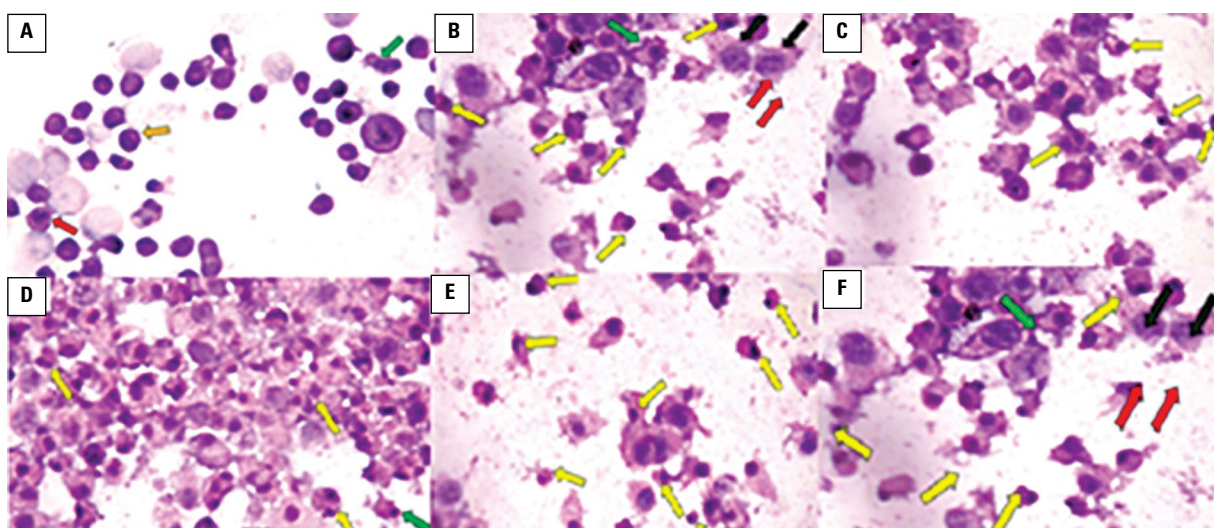
## Discussion

The present work aimed to use ELFEMF and electroporation with Au-NPs as alternative ways of promotion apoptosis of human laryngeal cancer cells. Also, we aimed to preview the genetic and cell cycle alterations in Hep-2 cells. The obtained data revealed that both electroporation in a presence of Au-NPs and cell exposure to ELFEMF could manage the death of cancer cells *via* apoptosis induction.

The present study indicated that intermittent exposure of laryngeal carcinoma cells (Hep-2) to a 25 and 50 mT of ELFEMF affected cell cycle profile and proved these *via* noticed elevated up and down-regulation of apoptosis-related genes. Thus, we showed that apoptotic profile and G2/M cellular arrest may play the crucial role in the biological effects of 25 and 50 mT ELFEMF and related induction of DNA



**Figure 3. Evaluation of cell cycle of Hep-2 cells using flow cytometry.** (A–D) G0/G1, S, G2/M phases and apoptotic cells’ proportions of Hep-2 cells. \*P < 0.05, \*\*P < 0.01, \*\*\*P < 0.001; capped lines: t-test between subgroups; dotted line: one-way ANOVA between all subgroups; dashed line: t-test between control and tested cells, respectively. DC Au-NPs, direct contact gold nanoparticles; EP Au-NPs, electroporation with gold nanoparticles; ELFEMF, extremely-low frequency electromagnetic field.



**Figure 4. Microphotographs of Hep-2 cells after Au-NPs or ELFEMF treatment.** A. Control, untreated Hep-2 cells present regular tumor cells (yellow arrow), cellular pleomorphism (green arrow) and nuclear pleomorphism (red arrow). B. Hep-2 Au-NPs electroperated cells show numerous apoptotic bodies (yellow arrows), irregular cell membrane of neoplastic cell (green arrow), necrotic cells with euchromatin and heterochromatin (red arrows) and ruptured cell membrane of necrotic cells (black arrows). C. Hep-2 cells showing numerous apoptotic bodies post short term exposure to ELFEMF for 15 min at 25 mT (yellow arrows). D. Hep-2 cells showing neoplastic cells with nuclear fragmentation (yellow arrows) and apoptotic bodies (green arrow) post cell exposure for 30 min at 25 mT. E. Hep-2 cells showing numerous apoptotic bodies (yellow arrows) after cell exposure to 50 mT for 15 min. F. Hep-2 cells showing numerous apoptotic bodies and irregular cell membrane of neoplastic cells (green arrow), necrotic cells (red arrows) and ruptured cell membrane of necrotic cells (black arrows) after cell exposure to 50 mT for 30 min. H&E staining, magnification ×40.



strand breaks. On the other hand, it was observed that subjecting cells continuously to a constant electromagnetic field may induce adaptive mechanisms, protecting the genome from harmful influences [14]. The point of view concerning this way and effect was based on the continuous exposure of people to such frequencies *via* different magnetic field sources such as radio and microwave in their homes. The potential of ELFEMF to exert harmful biological effects on human health is of increasing concern. It has been reported that DNA damage may be involved in cell proliferation, apoptosis as well as cell cycle [15, 16].

The presence of Au-NPs inside the cells allowed the introduction and progress of cell damage, especially cancerous cells. On the other hand, it has been noticed that the cells that were exposed to the ELFEMF have developed a high rate of cellular damage. This cellular damage was developed by the irreversible induction of pores in the cell membrane due to the overlapping between the externally applied ELFEMF and the magnetic properties of the cell membranes. In addition to the effectiveness of the magnetic field on the cancerous cells, Au-NPs activate the signaling pathways through acting on the membrane proteins that leads to the inhibition of the cancer cells proliferation. It was previously observed that the gold nanoparticles have the ability to enter the cells through the endocytosis, which produces mitochondrial dysfunction, formation of reactive free radicals and the destruction of nucleic acid and cellular proteins [17]. Hence, all the used effects allow the inhibition of cellular proliferation. Therefore, the utilizing of gold nanoparticles in the presence of electroporation and the effect of extremely-low electromagnetic field has shown remarkable effects. This was done by the activation of Au-NPs to start entering the tumor cells, especially through electroporation physics enhancers accordingly displaying a remarkable ability to treat the tumor cells. It was observed that the use of electroporation in the presence of gold nanoparticles can be effective to induce apoptosis in the Hep-2 cells. Moreover, the combination of electroporation and Au-NPs had moderate effect on the studied neoplastic cells. Hence, the effect on pro-apoptotic genes (*P53* and *BAX*) and anti-apoptotic genes (*BCL2*) was viewed to be significant in the presence of combined action of gold nanoparticles and electroporation and the ELFEMF treatment with 50 mT and 100 mT.

The influence of size, shape, and surface modification on cytotoxicity of gold nanoparticles to human Hep-2 and canine MDCK cells was investigated by Zhang et al. [11]. Au-NPs cell treatment revealed that cell death induced predominately within one hour through apoptosis induced gene expression, whereas

cell death by free cetyltrimethylammonium-bromide (CTAB) was a time- and dose-dependent. Both positively and negatively surface-charged polymer-coated gold nanorods GNRs showed similar levels of cytotoxicity, suggesting the significance of surface functionality rather than surface charge in this case [11].

As the interaction of gold nanoparticles with cells relies on properties of nanoparticles, the cytotoxicity is complex and still under debate. In a previously published work, they found that cetyltrimethylammonium-bromide (CTAB) encapsulated gold nanorods (GNRs) were relatively higher cytotoxic than gold nanorods (GNRs) undergone further polymer coating and citrate stabilized gold nanospheres (GNSs) [11].

The toxicity of CTAB-encapsulated GNRs was mainly caused by CTAB on GNRs' surface but not free CTAB in the solution. On the contrary to our data, Brisdelli and coworkers recorded that lower frequency ELF-MF (1 mT; 50 Hz) enhanced the apoptosis and that ULEMF (25/50 mT; 50 Hz) enhanced the apoptosis and cell arrest in human laryngeal carcinoma Hep-2 cells [18]. Four different compounds, namely vinblastine, etoposide, quercetin, and resveratrol, enhanced apoptosis in human K562 chronic myeloid leukemia cells. The exposure to ELFEMF did not affect growth and viability of untreated K562 cells and did not influence the anti-proliferative effects of resveratrol, vinblastine, and etoposide [18]. On the contrary, in quercetin treated cells, exposure to ELFEMF significantly reduced the percentage of apoptotic cells and the caspase-3 activity and modified the cell cycle profile especially after 48 hours of exposure. In addition, the simultaneous treatments for 24 hours with quercetin plus ELFEMF increased Bcl-2 protein expression and prevented quercetin-induced downregulation of Mcl-1 and Bcl-xL [18].

The ELFEMF-dependent modulation of the expression of anti-apoptotic Bcl-2 family and Hsp70 proteins could act as a pro-survival mechanism in K562 cells [18]. ELFEMF seems to produce several biological effects such as changes in cell proliferation, cell differentiation, cell cycle and enzyme activity [19–22]. Moreover, ELFEMF was found to affect cellular redox homeostasis, modulation of physiological functions such as mitochondrial membrane potential,  $\text{Ca}^{+2}$  signaling and homeostasis, and ATP synthesis [23–25]. Alterations in the apoptosis process are involved in the pathogenesis of many types of cancer [26]. There is evidence implicating ELFEMF in the development of this disease in humans [27], and many *in vitro* studies have also reported correlation between apoptotic process and ELFEMF. The latter has been involved in the reduction or delay of cell apoptosis process, as well as in its stimulation [28, 29].

Additionally, some studies reported evidence suggesting the combined effects of ELFEMF and various chemicals and physical agents in modulating the apoptotic cascade [30, 31]. However, current research is characterized by conflicting data, and synergistic and/or antagonistic effect induced by ELFEMF has been described in H<sub>2</sub>O<sub>2</sub>-treated erythroleukemia cell line [32]. Also, our data was on the contrary of the previous work recording that the effects of 50 Hz ELFEMF exposure on the growth of GC-2 and cell viability was detected with the CCK-8 kit following 50 Hz ELFEMF exposure at different magnetic intensities for 72 hours to explore the effects of the ELFEMF on the growth of GC-2 cells [33]. Data showed that the 50 Hz ELFEMF did not markedly affect the morphology or viability (CCK-8) of GC-2 cells. Also, the effect of 50 Hz ELFEMF exposure on the apoptosis and cell cycle of GC-2 cells result showed that 50 Hz (ELFEMF) did not induce apoptosis in GC-2 cells compared with the same group [33]. Additionally, the percentages of G1-, S-, and G2-M phase cells did not significantly differ between the exposed groups post-treatment with 50 Hz ELFEMF for 72 hours. These findings confirmed that 50 Hz ELFEMF did not induce apoptosis or cell cycle arrest in GC-2 cell [34–36].

Based on the presented data, it could be concluded that the treatment of Hep-2 cancer cell with ELFEMF and electroporation with Au-NPs can cause cell damage inducing cell cycle arrest and apoptosis. Apoptotic and cell cycle profiles were cell type, time and ELFEMF frequency dependent. Usage of either Au-NPs-electroporation or ELFEMF in anti-cancer therapy may be a promising cancer therapy tool, may substitute or enhance the radio-, chemotherapy and surgical therapy approaches with least adverse effects. More intensified trials are recommended on different cancer cell lines as well as in vivo studies and different formulation of different nanoparticles shapes and concentrations must be validated.

### Acknowledgement

The authors extend their appreciation to the Deanship of Scientific Research at King Khalid University, Abha, Saudi Arabia for funding this work through General Research Project under grant number G.R.P. 1/9/38-38.

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*Submitted: 24 April, 2019*

*Accepted after reviews: 12 November, 2019*

*Available as AoP: 20 November, 2019*