

The Relationship between Wilms' Tumor 1 (WT1) and Paired Box 8 (Pax-8) Protein Expressions in Papillary and Anaplastic Thyroid Carcinomas

Katsuhiro TANAKA, Masahiko IKEDA, Hiroshi SONOO, Mai HIRONO,
Tsunehisa NOMURA, Sumiko OHKUBO, Yutaka YAMAMOTO, Shigeo SHIIKI,
Kazutaka NAKAJIMA and Junichi KUREBAYASHI

*Department of Breast and Thyroid Surgery, Kawasaki Medical School : 577 Matsushima,
Kurashiki, Okayama, 701-0192 Japan*

Accepted for publication on September 7, 2006

ABSTRACT. Background : We elucidated the expression of WT1 and its correlation with Pax-8 and TTF-1 as a differentiation promoter in thyroid cancer. **Methods :** We investigated the expression of WT1, Pax-8, and TTF-1 immunohistochemically in 60 primary tumors (30 well-differentiated and 15 poorly differentiated papillary carcinomas and 15 anaplastic thyroid carcinomas). **Results :** Positive staining of WT1 was found in the cytoplasm and nucleus of the cancer cells. Each positive rate of cytoplasmic staining of WT1 was 100% in the 45 papillary carcinomas and 93.3% in the 15 anaplastic carcinomas. The immunohistochemical scores for WT1 in well differentiated, poorly differentiated papillary carcinomas and anaplastic carcinomas were 5.90 ± 1.7 , 6.53 ± 1.7 , and 3.60 ± 2.1 , respectively. The scores for the well and poorly differentiated papillary carcinomas were significantly higher than those for the anaplastic carcinomas. The expression level of WT1 showed a significant correlation with both Pax-8 and TTF-1. **Conclusions :** This is the first report to demonstrate a positive correlation of the expression of WT1 with Pax-8 and TTF-1 in clinical cases. In thyroid cancer, the decrease in expression of WT1 with anaplastic transformation is assumed to be closely related to Pax-8 and TTF-1.

Key words ① thyroid carcinoma ② WT1 ③ Pax-8 ④ differentiation
⑤ TTF-1

The Wilms' tumor 1 (WT1) gene was identified as a candidate gene among children harboring Wilms' tumor in 1990¹⁾. WT1 is a transcriptional repressor with zinc finger domains^{2), 3)}. Although it was demonstrated to be a suppressor gene at initially³⁾, overexpression of its wild type has been reported in several tumors subsequently. Expression of WT1 has been demonstrated in leukemia⁴⁾, malignant mesothelioma⁵⁾, bone and soft tissue sarcoma⁶⁾, and in breast⁷⁾, ovarian⁸⁾, lung⁹⁾, thyroid¹⁰⁾, and colorectal cancers¹¹⁾. Most reports have emphasized the overexpression of WT1 in cancer cells compared to the expression in normal tissues. Although mutated homozygous WT1 occurs in Wilms' tumor in the kidney, WT1 plays an opposite and transactivate role in several cancers without mutations. Little is known as yet

田中克浩, 池田雅彦, 園尾博司, 廣納麻衣, 野村長久, 大久保澄子, 山本 裕, 椎木滋雄, 中島一毅,
紅林淳一

e-mail:tanakaka@med.kawasaki-m.ac.jp

about these reversal mechanisms, but some investigators have noticed isoforms of WT1. WT1 has four isoforms that are formed by alternative RNA splicing¹²). In these isoforms, the presence or absence of three amino acids (lysine, threonine, and serine ; KTS) between the third and fourth zinc fingers in WT1 seems to be important, because, the insertion or omission of KTS affects DNA-binding affinity and the specificity of WT1¹³⁾⁻¹⁵). Some reports have noted reversal effects on targets and cell proliferation depending on the presence of KTS^{16), 17}).

Many target genes of WT1 have been identified, including insulin-like growth factor-2 (IGF-2)¹⁸), platelet-derived growth factor (PDGF) A-chain¹⁹), IGF1 receptor²⁰), epidermal growth factor receptor (EGFR)²¹), colony-stimulating factor-1 (CSF-1)²²), bcl-2²³), c-myc²³), E-cadherin²⁴), cyclin E²⁵), and mevalonate²⁶). Therefore, the candidate target genes of WT1 are representative and important for cell proliferation.

Although the regulatory factors for expression of WT1 are not as yet well known, paired-box (Pax)-2 and -8^{27), 28}), hypoxia-inducible factor 1²⁹), IGF-1³⁰) and Spl³¹) have been included. In these factors, Pax-8 is essential for all organs, and especially during development. Pax-8 is expressed in the developing and adult thyroid, the developing secretory system and, at lower levels, in the adult kidney³²). Therefore, the thyroid is the only adult organ in which Pax-8 is fully and persistently expressed. In the normal thyroid, Pax-8 is essential and critical for the expression of differentiation markers such as thyroglobulin (Tg), thyroid peroxidase (TPO), and thyrotropin receptor (TSH-R) in cooperation with thyroid transcription factor-1 (TTF-1)³³). Expression of Pax-8 and TTF-1 has also been observed in differentiated thyroid carcinoma³⁴). With the dedifferentiation of thyroid cancer, these differentiation markers and transcriptional factors would decrease and cancer cells would be acquired to change into an unfavorable phenotype³⁴).

In this study, we detected the expression level of WT1 in papillary thyroid carcinomas including poorly differentiated ones and in anaplastic thyroid carcinomas using an immunohistochemical method. We investigated the relationship between clinical behavior and WT1 expression to determine whether WT1 expression in thyroid carcinomas is related to differentiation. We also elucidated the expression levels of Pax-8 and TTF-1 in such carcinomas and analyze the relationships among them and WT1.

MATERIALS AND METHODS

Materials

The subjects of this study were 45 papillary thyroid carcinoma patients (15 of well differentiated type without distant metastasis, 15 of well differentiated type with distant metastases, and 15 of poorly differentiated type), and 15 anaplastic thyroid carcinoma patients who were treated in our hospital. The thyroid function of all the patients was euthyroid before primary operations. All of the patients with papillary carcinoma had received thyroxin to suppress serum TSH during follow-up after the initial surgical treatment. Determination of the dose of thyroxin to maintain the serum TSH level under the lower limit for each patient was done by measuring the level every six months during follow-up. Distant recurrence of disease was defined by echography, chest X-ray films, and computed tomography and/or thallium or radioiodine scintigraphy during follow-up.

For this study, we used embedded paraffin sections of the primary tumor of each patient, which were resected at each initial surgical procedure, or percutaneous fine needle biopsy. Informed consent was obtained

from all enrolled patients.

Immunohistochemical procedure

The sections embedded in paraffin were cut into sections 4 μ m in thickness, and fixed on aminopropyltriethoxysilane-coated glass slides (Matsunami, Osaka, Japan). After typical deparaffinization, all the slides were autoclaved for 20 min at 121 °C in a 10mM citrate buffer (pH 6.0) for antigen retrieval. Then, the slides were incubated for two hours at room temperature with each of the following primary antibodies ; WT1 antibody, a mouse monoclonal antibody (sc-7385, Santa Cruz, CA, USA), at a concentration of 10 ug/ml, Pax-8 antibody, a goat polyclonal antibody (sc-16279, Santa Cruz, CA, USA), at a concentration of 10 ug/ml, and TTF-1 antibody, a goat polyclonal antibody (sc-8761, Santa Cruz, CA, USA), at a concentration of 10 ug/ml. After incubation, the slides were washed in PBS for 15 min, and a secondary antibody was applied using the LSAB plus kit (DAKO Corp., Carpinteria, CA, USA) according to the manufacturer's protocol. After washing, the colors were developed using new fuchsin and 5-bromo-4-chloro-3-indoxyl phosphate, nitro blue tetrazolium chloride, and indonitrotetrazolium (BCIP/NBT/INT) (DAKO Corp., Carpinteria, CA, USA). Then the slides were counterstained with hematoxylin (WAKO, Osaka, Japan), washed in distilled water for 5 min and mounted.

We used normal mouse serum instead of primary antibody in negative control studies, which were performed for all slides (data not shown).

Analysis of immunohistochemical detection

Immunostaining was evaluated by two repeated stainings of the same specimen. We blinded ourselves to the characteristics of the patients, the extent of the tumor and prognostic scores. The cells of a tissue section were evaluated as positive when they showed a distinctly specific stain when compared with the cells of negative control sections. Then, two investigators independently decided the level of intensity of positive cells of each slide. Only in the case of discordance among investigators, did we discuss and decide on the intensity together.

For these immunohistochemical analyses, except for that of WT1, we evaluated both the concentration and positive rate of stained nucleus in each section using the Allred scoring system³⁵. In brief, for evaluation of the concentration of staining, the stained cells in the largest population of the section were divided into four degrees (from zero to three). For evaluation of the positive rate of the stained nucleus, the major intensity of the nucleus was divided into six degrees, zero, one (population of the stained nucleus in less than 1/100 of the entire nucleus of a cell), two (1/100-1/10), three (1/10-1/3), four (1/3-2/3), and five (>2/3). Multiplication of both scores resulted in a final quotation ranging from 0-8, which was presented as the mean \pm S.D.

The expression of WT1 was evaluated with regard to both the concentration and distribution of stained cells, as previously described³⁶. For evaluation of the concentration of staining, the stained cells in the largest population of the section were divided into four degrees (from zero to three). For evaluation of the distribution of stained cells, the major intensity of cells was divided into four degrees, zero, one (observation of stained cells in less than 30% of the entire population of cells.), two (<60%), and three (\geq 60%). Multiplication of both scores resulted in a final quotation ranging from 0-9, which was presented as the mean \pm S.D.

Table 1. Background of patients

	PTC (without rec) n=15	PTC (with rec) n=15	poorly PTC n=15	ATC n=15
Age	54 (31-72)	57 (11-73)	57 (20-82)	71 (43-83)
Gender (male: female)	3:12	5:10	5:10	6:09
Tumor size (cm)	3 (2.3-8.5)	3 (0.8-7)	2.75 (1.8-6.5)	5.5 (2.5-11)
EX				
0	6	7	6	-
1	1	0	2	-
2	8	8	7	6
Recurrent site				
Lung	-	13	1	4
Lymph node	-	7	3	6
Bone	-	6	0	2
Pleura	-	1	0	2
Brain	-	1	0	0
DFI (m)	-	42 (2-135)	23.5 (5-94)	-
OS (m)	109 (51-202)	86 (38-214)	28 (5-94)	8 (2-35)

PTC; papillary thyroid carcinoma, ATC; anaplastic thyroid carcinoma

EX: extrathyroidal infiltration

DFI; disease free interval, OS; overall survival

Statistical analysis

For statistical analysis, the F test and Spearman's rank correlation test were used, and $p < 0.05$ was considered significant.

RESULTS

Table 1 shows the characteristics of patients enrolled in this study. The median ages of the well differentiated papillary carcinoma patients without recurrence or with recurrence, the poorly differentiated papillary carcinoma patients, and the anaplastic carcinoma patients were 54, 57, 57, and 71 years old, respectively. Forty-seven of the patients were women (68.3%) and 13 were men (31.7%). Among the papillary carcinoma patients, there were 23 patients with EX2 (infiltration to adjacent organs except muscles). Among the well-differentiated papillary carcinoma patients with recurrent tumors, metastatic sites appeared in the lung (86.7%), lymph nodes (46.7%), bone (40%), pleura (6.7%), and brain (6.7%). The disease-free interval and overall survival of patients with recurrent well differentiated papillary carcinomas and poorly differentiated papillary carcinomas were 42 and 86 months, and 23.5 and 28 months, respectively. The overall survival of patients with anaplastic carcinomas was eight months.

Immunohistochemical study

Positive staining of WT1 was found in the cytoplasm and nucleus of the cancer cells (Fig. 1A and 1C).

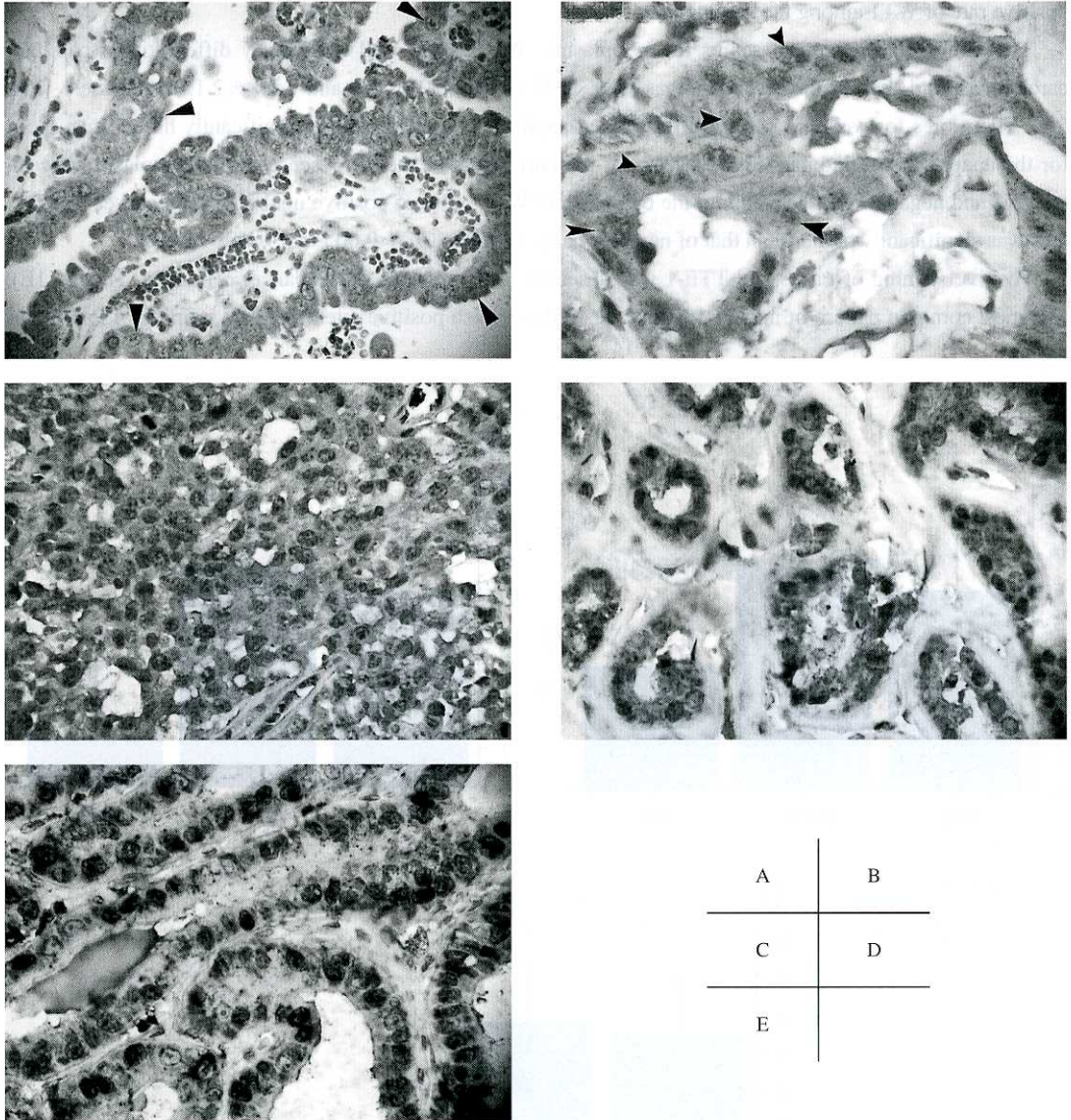


Fig. 1. Immunohistochemical results

Magnification of each figure is $\times 200$ (A and C) and $\times 400$ (B, D and E).

The positive staining for WT1 was developed using fuchsin, and for Pax-8 and TTF-1 by NBT/BCIP/INT. The slide for WT1 was counterstained with hematoxylin.

In all papillary thyroid carcinoma, positive staining of WT1 appears in the cytoplasm (A). Speckled nuclear staining can be seen in the same specimen. Representative nucleic staining is showed in B (Arrow head). WT1 staining in anaplastic carcinoma is shown in C. The brown staining of Pax-8 is mostly in the nucleus and partly in the cytoplasm in papillary carcinoma (D). Positive brown staining of TTF-1 appears in the nucleus (E).

Each positive rate of cytoplasmic staining of WT1 was 100% in the 45 papillary carcinomas, and 93.3% in the 15 anaplastic carcinomas, respectively. And the positive rate of nucleic staining was 37.8% in the 45 papillary carcinomas and 20% in the 15 anaplastic carcinomas. The most common pattern of nucleic staining was speckled (Fig. 1B). Diffuse staining of the nucleus was rare. There was no significant difference in the

positive rate of WT1 among the carcinomas.

The immunohistochemical scores for cytoplasmic WT1 in the well and poorly differentiated papillary carcinomas, and the anaplastic carcinomas were 5.90 ± 1.7 , 6.53 ± 1.7 , and 3.60 ± 2.1 , respectively (Fig. 2A). The scores for well and poorly differentiated papillary carcinomas were significantly higher than those for the anaplastic carcinomas (Fig. 2A). As for the correlation of the WT1 score of the cytoplasm between positive and negative nucleic staining, the cytoplasmic WT1 score of positive nucleic staining cases (6.55 ± 2.0) was significantly higher than that of negative cases (4.95 ± 1.9 , $p < 0.005$ by Fischer's test).

Positive staining of Pax-8 and TTF-1 was observed in the nucleus of the cancer cells (Fig. 1D and 1E). The most common pattern of nucleic staining was diffuse. Each positive rate of nucleic staining of Pax-8 was

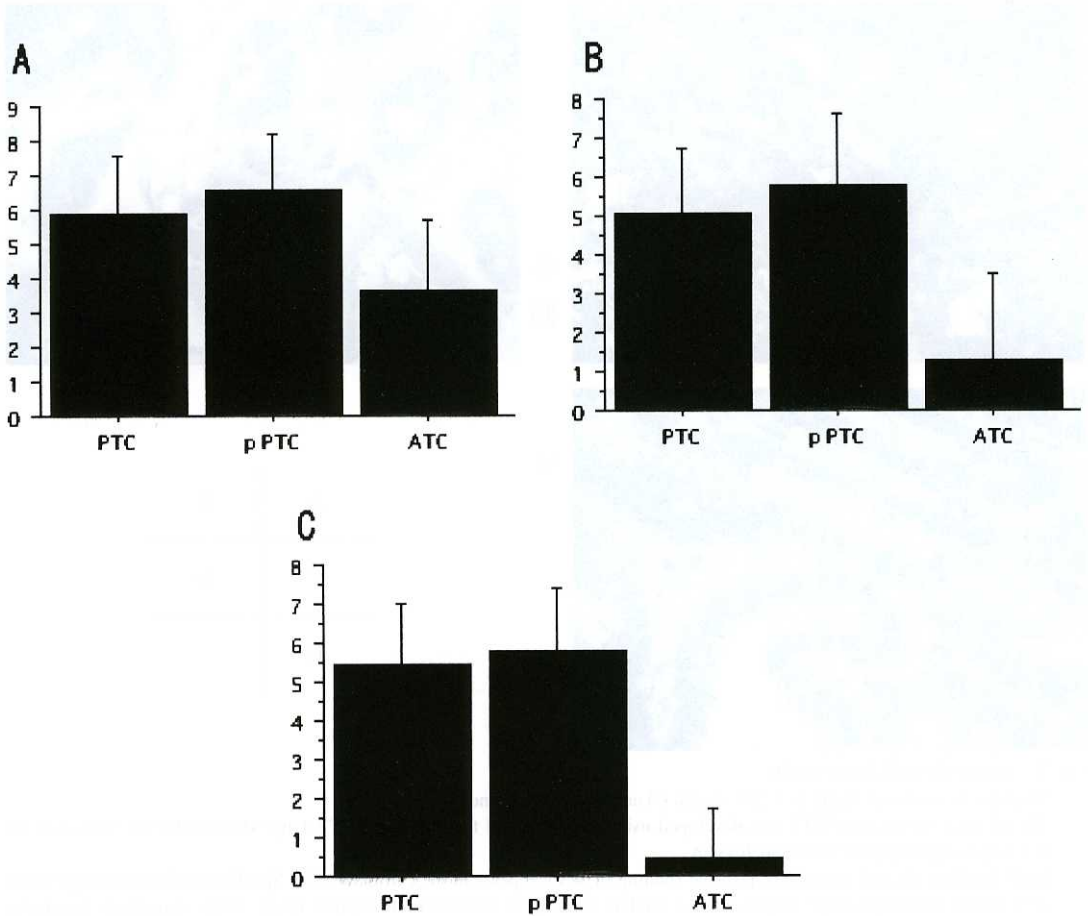


Fig. 2. The immunohistochemical scores of WT1 (A), Pax-8 (B), and TTF1 (C) among each histological type

The immunohistochemical scores for WT1 in well differentiated, and poorly differentiated papillary carcinomas, and anaplastic carcinomas were 5.90 ± 1.7 , 6.53 ± 1.7 , and 3.60 ± 2.1 , respectively. The scores for well and poorly differentiated papillary carcinomas were significantly higher than those for anaplastic carcinomas ($p < 0.001$, analyzed by Fischer's test, Fig. 2A). The immunohistochemical scores for Pax-8 in well and poorly differentiated papillary carcinomas, and anaplastic carcinomas were 5.07 ± 1.6 , 5.8 ± 1.8 , and 1.27 ± 2.3 , respectively (Fig. 2B). The scores of Pax-8 in anaplastic carcinomas were significantly lower than those in papillary carcinomas (Fig. 2B). The immunohistochemical scores for TTF-1 in well and poorly differentiated papillary carcinomas, and anaplastic carcinomas were 5.47 ± 1.5 , 5.8 ± 1.6 , and 0.47 ± 1.2 , respectively (Fig. 2C). The scores of TTF-1 in each histological type showed similar significance in Pax-8.

100% in the 45 papillary carcinomas, and 13.3% in the 15 anaplastic carcinomas. Each positive rate of nucleic staining of TTF-1 was 100% in the 45 papillary carcinomas, and 6.7% in the 15 anaplastic carcinomas. The positive rates for Pax-8 and TTF-1 in the anaplastic carcinomas were significant lower than those in the papillary carcinomas. The immunohistochemical scores for Pax-8 in the well and the poorly differentiated papillary carcinomas, and the anaplastic carcinomas were 5.07 ± 1.6 , 5.8 ± 1.8 , and 1.27 ± 2.3 , respectively (Fig. 2B). The Pax8 score in the anaplastic carcinomas was significantly lower than that in papillary carcinomas (Fig. 2B). The immunohistochemical scores for TTF-1 in the well and the poorly differentiated papillary carcinomas, and the anaplastic carcinomas were 5.47 ± 1.5 , 5.8 ± 1.6 , and 0.47 ± 1.2 , respectively (Fig. 2C). The TTF-1 score in the anaplastic carcinomas was significantly lower than that in papillary carcinomas (Fig. 2C).

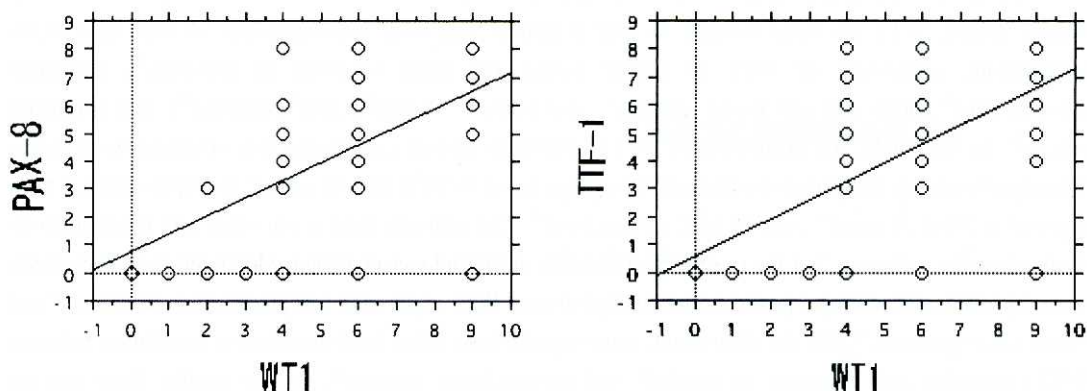


Fig. 3. The correlation of immunohistochemical scores among WT1 and Pax-8 and TTF-1 in thyroid cancers

The expression level of WT1 showed a significant correlation with both Pax-8 (Fig. 3A, $p < 0.0001$) and TTF-1 (Fig. 3B, $p < 0.0001$). The correlation coefficients were 0.529 and 0.531.

Table 2. Correlation between WT1 expression and clinical data in papillary carcinoma patients

	WT1 score	p value
Age	-	$p = 0.645$
Gender		
male (n=13)	6 ± 1	$p = 0.507$
female (n=32)	6 ± 2	
Tumor Size	-	$p = 0.398$
EX		
0 or 1 (n=23)	6 ± 2	$p = 0.938$
2 (n=22)	6 ± 2	
Presence of Recurrence		$p = 0.209$
yes (n=18)	6 ± 1	
no (n=27)	6 ± 2	

WT1 score: mean \pm S.D.

Correlation among the expression levels of WT1, Pax-8, and TTF-1 (Fig. 3)

The expression level of WT1 in thyroid cancer showed a significant correlation with both Pax-8 and TTF-1 (Fig. 3A and 3B). The correlation coefficients were 0.529 and 0.531, respectively.

Relationship between clinical data and the expression level of WT1 (Table 2)

Table 2 shows the relationship between clinical data and the expression level of WT1 in only patients with papillary thyroid carcinomas. With regards to age, gender, tumor size, EX, presence of recurrence, and overall survival, there was no significant relationship with the expression of WT1.

DISCUSSION

The role of WT1 in the initiation and/or progression of cancer is still unknown except in hereditary Wilms' tumor. WT1 has been thought to play a critical role with overexpression of wild-type WT1. Furthermore, expression of WT1 in cancer tissues has been observed in leukemia⁴⁾, malignant mesothelioma⁵⁾, bone and soft tissue sarcoma⁶⁾, and breast⁷⁾, ovarian⁸⁾, lung⁹⁾, thyroid¹⁰⁾, and colorectal cancer¹¹⁾. In this study, the expression of WT1 in papillary thyroid carcinomas was observed in all cases. There has been only two reports concerning the expression of WT1 in thyroid cancer. Its expression has been reported in 20 of 21 cases¹⁰⁾, and in none of ten cases³⁷⁾. The antibody used in our study and these previous studies was the same one, but the positivity of staining in thyroid cancer varied. Moreover, there have been some reports demonstrating a positive relationship between WT1 expression and prognosis in leukemia³⁸⁾ and breast cancer patients³⁹⁾. On the other hand, some reports have failed to demonstrate a correlation between WT1 expression and prognosis in ovarian⁸⁾ and mesothelioma patients⁴⁰⁾. In our results, there was no significant relationship between the expression of WT1, the clinical parameters and prognosis. No difference was observed in the expression levels of WT1, Pax-8, and TTF-1 between well and poorly differentiated thyroid carcinomas. Therefore, there may be no relationship between WT1 and clinical behavior.

In our findings, although the localization of WT1 was mainly cytoplasmic, in about one third, it was also in the nucleus. Reports have shown the main localization of WT1 to be the nucleus in breast cancers⁷⁾, ovarian tumors⁸⁾, malignant mesotheliomas^{37), 40)}, and desmoplastic small round cell tumors⁴¹⁾. On the other hand, Oji *et al.* reported its expression only in the cytoplasm of thyroid¹⁰⁾, and colorectal cancers¹¹⁾. As with our findings, both staining of the cytoplasm and the nucleus has been reported in a small population of breast cancer⁷⁾ and mesothelioma patients⁹⁾. The difference in the localization of WT1 in each organ has been demonstrated as a model of shuttling of multifunctional pre-mRNA binding proteins between the nucleus and the cytoplasm^{42)~44)}.

The presence or absence of three amino acids (lysine, threonine, and serine ; KTS) between the third and fourth zinc fingers has been thought to be important in isoforms of WT1, because, the insertion or omission of KTS affects DNA-binding affinity and the specificity of WT1^{13)~15)}. These differences of alternative splicing are more important in the biological behavior of WT1. Englert *et al.* reported that WT1 (+KTS) or WT1 mutants exhibited a speckle pattern within the nucleus⁴⁵⁾. In our findings, nuclear staining of WT1 showed a speckled pattern. Therefore, overexpression of WT1 in thyroid cancer might be depended on WT1 (+KTS) or WT1 mutants.

Although the regulatory factors for expression of WT1 are not as yet well known, paired-box (Pax)-2 and

-8^{27), 28)} was demonstrated as the regulatory factor. Pax-8 is also essential for the maintenance of differentiation of thyroid follicular cells. Even in adults, Pax-8 is still expressed in normal thyroid and kidney. In the thyroid, in particular, Pax-8 plays an important role as a promoter activator of differentiation markers such as Tg, TPO, and TSH-R. Like Pax-8, TTF-1 is also essential for the differentiation of thyroid cells³³⁾. In adults, TTF-1 expression is still expressed in thyroid, lung³³⁾. Thus, the thyroid retains genes that act mainly in development. In this study, we evaluated the relationship between the expression levels of Pax-8 and WT1, and found a significant correlation between them. There was also a significant correlation between expression levels of TTF-1 and WT1. Eccles *et al* investigated the expression levels of WT1, Pax-2, and Pax-8 in Wilms' tumor and fetal kidney by in situ hybridization⁴⁶⁾, but they failed to demonstrate a significant correlation among them. To the best of our knowledge, this is the first report to demonstrate a positive correlation of the expressions of WT1, Pax-8, and TTF-1 in clinical cases.

In our findings, the highest expression of WT1 was seen in papillary carcinomas. Anaplastic carcinomas showed lower expression. In thyroid cancer, the expression of WT1 seemed to decrease with dedifferentiation in the same manner as the expression levels of Pax-8 and TTF-1. In the thyroid, which expresses Pax-8 even in adults, Pax-8 is assumed to play an important role in the regulation of WT1.

ACKNOWLEDGMENTS

This work was supported in part by a grant from the Kawasaki Medical School Study Project 2003 (No. 15-504)

REFERENCES

- 1) Gessler M, Poustka A, Cavenee W, Neve RL, Orkin SH, Bruns, GAP : Homozygous deletion in Wilms tumours of a zinc-finger gene identified by chromosome jumping. *Nature* 343 : 774-778, 1990
- 2) Call KM, Glaser T, Ito CY, Buckler AJ, Pelletier J, Haber DA, Rose EA, Kral A, Yeger H, Lewis WH, Jones C, Housman DE : Isolation and characterization of a zinc finger polypeptide gene at the human chromosome 11 Wilms' tumor locus. *Cell* 60 : 509-520, 1990
- 3) Madden SL, Cook DM, Morris JF, Gashler A, Sukhatme VP, Rauscher III FJ : Transcriptional repression mediated by the WT1 Wilms tumor gene product. *SCIENCE* 253 : 1550-1553, 1991
- 4) Miwa H, Beran M, Saunders GF : Expression of the Wilms' tumor gene (WT1) in human leukemia. *Leukemia* 6 : 405-409, 1992
- 5) Amin KM, Litzky LA, Smythe WR, Mooney AM, Morris JM, Mews DJY, Pass HI, Kari C, Rodeck U, Rauscher III FJ, Kaiser LR, Albelda SM : Wilms' tumor 1 susceptibility (WT1) gene products are selectively expressed in malignant mesothelioma. *Am J Pathol* 146 : 344-356, 1995
- 6) Ueda T, Oji Y, Naka N, Nakano Y, Takahashi E, Koga S, Asada M, Ikeba A, Nakatsuka S, Abeno S, Hosen N, Tomita Y, Aozasa K, Tamai N, Myoui A, Yoshikawa H, Sugiyama H : Overexpression of the Wilms' tumor gene WT1 in human bone and soft-tissue sarcomas. *Cancer Sci* 94 : 271-276, 2003
- 7) Silberstein GB, Van Horn K, Strickland P, Roberts Jr CT, Daniel CW : Altered expression of the WT1 Wilms tumor suppressor gene in human breast cancer. *Proc Natl Acad Sci USA* 94 : 8132-8137, 1997
- 8) Shimizu M, Toki T, Takagi Y, Konishi I, Fujii S : Immunohistochemical detection of the Wilms' tumor gene (WT1) in epithelial ovarian tumors. *Int J Gynecol Pathol* 19 : 158-163, 2000

- 9) Focter MR, Johnson JE, Allred DC : Immunohistochemical analysis of nuclear versus cytoplasmic staining of WT1 in malignant mesotheliomas and primary pulmonary adenocarcinomas. *Arch Pathol Lab Med* 125 : 1316-1320, 2001
- 10) Oji Y, Miyoshi Y, Koga S, Nakano Y, Ando A, Nakatsuka S, Ikeba A, Takahashi E, Sakaguchi N, Yokota A, Hosen N, Ikegame K, Kawakami M, Tsuboi A, Oka Y, Ogawa H, Aozasa K, Noguchi S, Sugiyama H : Overexpression of the Wilms' tumor gene WT1 in primary thyroid cancer. *Cancer Sci* 94 : 606-611, 2003
- 11) Oji Y, Yamamoto H, Nomura M, Nakano Y, Ikeba A, Nakatsuka S, Abeno S, Kiyotoh E, Jomgeow T, Sekimoto M, Nezu R, Yoshikawa Y, Inoue Y, Hosen N, Kawakami M, Tsuboi A, Oka Y, Ogawa H, Souda S, Aozasa K, Monden M, Sugiyama H : Overexpression of the Wilms' tumor gene WT1 in colorectal adenocarcinoma. *Cancer Sci* 94 : 712-717, 2003
- 12) Haber DA, Sohn RL, Buckler AJ, Pelletier J, Call KM, Housman DE : Alternative splicing and genomic structure of the Wilms tumor gene WT1. *Proc Natl Acad Sci USA* 88 : 9618-9622, 1991
- 13) Rauscher III FJ, Morris JF, Tournay OE, Cook DM, Curran T : Binding of the Wilms' tumor locus zinc finger protein to the EGF-1 consensus sequence. *SCIENCE* 250 : 1259-1262, 1990
- 14) Bickmore WA, Oghene K, Little MH, Seawright A, van Heyningen V, Hastie ND : Modulation of DNA binding specificity by alternative splicing of the Wilms tumor wt1 gene transcript. *SCIENCE* 257 : 235-237, 1992
- 15) Larsson SH, Charlier J-P, Miyagawa K, Engelkamp D, Rassoulzadegan M, Ross A, Cuzin F, van Heyningen F, Hastie ND : Subnuclear localization of WT1 in splicing or transcription factor domains is regulated by alternative splicing. *Cell* 81 : 391-401, 1995
- 16) Murata Y, Kudoh T, Sugiyama H, Toyoshima K, Akiyama T : The Wilms tumor suppressor gene WT1 induces G1 arrest and apoptosis in myeloblastic leukemia M1 cells. *FEBS Lett* 409 : 41-45, 1997
- 17) Laity JH, Dyson HJ, Wright PE : Molecular basis for modulation of biological function by alternate splicing of the Wilms' tumor suppressor protein. *Proc Natl Acad Sci USA* 97 : 11932-11935, 2000
- 18) Drummond IA, Madden SL, Rohwer-Nutter P, Bell GI, Sukhatme VP, Rauscher III FJ : Repression of the insulin-like growth factor II gene by the Wilms tumor suppressor WT1. *SCIENCE* 257 : 674-678, 1992
- 19) Gashler AL, Bonthron DT, Madden SL, Rauscher III FJ, Collins T, Sukhatme VP : Human platelet-derived growth factor A chain is transcriptionally repressed by the Wilms tumor suppressor WT1. *Proc Natl Acad Sci USA* 89 : 10984-10988, 1992
- 20) Werner H, Re GG, Drummond IA, Sukhatme VP, Raucher III FJ, Sens DA, Garvin AJ, LeRoith D, Roberts Jr. CT : Increased expression of the insulin-like growth factor I receptor gene, IGF1R, in Wilms tumor is correlated with modulation of IGF1R promoter activity by the WT1 Wilms tumor gene product. *Proc Natl Acad Sci USA* 90 : 5828-5832, 1993
- 21) Englert C, Hou X, Maheswaran S, Bennett P, Ngwu C, Re GG, Garvin AJ, Rosner MR, Haber DA : WT1 suppresses synthesis of the epidermal growth factor receptor and induces apoptosis. *EMBO J* 14 : 4662-4675, 1995
- 22) Harrington MA, Konicek B, Song A, Xia X, Fredericks WJ, Rauscher III FJ : Inhibition of colony-stimulating factor-1 promoter activity by the product of the Wilms' tumor locus. *J Biol Chem* 268 : 21271-21275, 1993
- 23) Hewitt SM, Hamada S, McDonnell TJ, Rauscher III FJ, Saunders GF : Regulation of the proto-oncogenes bcl-2 and c-myc by the Wilms' tumor suppressor gene WT1. *Cancer Res* 55 : 5386-5389, 1995
- 24) Hosono S, Gross I, English MA, Hajra KM, Fearon ER, Licht JD : E-cadherin is a WT1 target gene. *J Biol Chem* 275 : 10943-10953, 2000
- 25) Loeb DM, Korz D, Katsnelson M, Burwell EA, Friedman AD, Sukumar S : Cyclin E is a target of WT1 transcriptional repression. *J Biol Chem* 277 : 19627-19632, 2002
- 26) Rae FK, Martinez G, Gillinder KR, Smith A, Shooter G, Forrest AR, Grimmond SM, Little MH : Analysis of complementary expression profiles following WT1 induction versus repression reveals the cholesterol/fatty acid synthetic pathways as a possible major target of WT1. *Oncogene* 23 : 3067-3079, 2004
- 27) Tagge EP, Hanson P, Re GG, Othersen Jr. HB, Smith CD, Garvin AJ : Paired box gene expression in Wilms' tumor. *J Pediat Surg* 29 : 134-141, 1994
- 28) Dehbi M, Pelletier J : PAX8-mediated activation of the wt1 tumor suppressor gene. *EMBO J* 15 : 4297-4306, 1996
- 29) Wagner K-D, Wagner N, Wellmann S, Schley G, Bondke A, Theres H, Scholz H : Oxygen-regulated expression of the

- Wilms' tumor suppressor Wt1 involves hypoxia-inducible factor-1 (HIF-1). *FASEB J* 17 : 1364-1366, 2003
- 30) Bentov I, Leroith D, Werner H : The WT1 Wilms' tumor suppressor gene : a novel target for insulin-like growth factor-I action. *Endocrinology* 144 : 4276-4279, 2003
 - 31) Cohen HT, Bossone SA, Zhu G, McDonald GA, Sukhatme VP : Spl is a critical regulation of the Wilms' tumor-I gene. *J Biol Chem* 272 : 2901-2913, 1997
 - 32) Plachov D, Chowdhury K, Walther C, Simon D, Guenet J-L, Gruss P : Pax8, a murine paired box gene expressed in the developing excretory system and thyroid gland. *Development* 110 : 643-651, 1990
 - 33) Kambe F, Seo H : Thyroid-specific transcription factors. *Endocrine J* 44 : 775-784, 1997
 - 34) Fabbro D, Di Loreto C, Beltrami CA, Belfiore A, Di Lauro R, Damante G : Expression of thyroid-specific transcription factors TTF-1 and PAX-8 in human thyroid neoplasms. *Cancer Res* 54 : 4744-4749, 1994
 - 35) Allred DC, Harvey JM, Berardo MD, Clark GM : Prognostic and predictive factors in breast cancer by immunohistochemical analysis. *Mod Pathol* 11 : 155-168, 1998
 - 36) Tanaka K, Sonoo H, Kurebayashi J, Nomura T, Ohkubo S, Yamamoto Y, Yamamoto S : Inhibition of infiltration and angiogenesis by thrombospondin-1 in papillary thyroid carcinoma. *Clin Cancer Res* 8 : 1125-1131, 2002
 - 37) Ordonez NG : Value of thyroid transcription factor-1, E-cadherin, BG8, WT1, and CD44S immunostaining in distinguishing epithelial pleural mesothelioma from pulmonary and nonpulmonary adenocarcinoma. *Am J Surg Pathol* 24 : 598-606, 2000
 - 38) Garg M, Moore H, Tobal K, Yin JAL : Prognostic significance of quantitative analysis of WT1 gene transcripts by competitive reverse transcription polymerase chain reaction in acute leukemia. *Br J Haematol* 123 : 49-59, 2003
 - 39) Miyoshi Y, Ando A, Egawa C, Taguchi T, Tamaki Y, Tamaki H, Sugiyama H, Noguchi, S : High expression of Wilms' tumor suppressor gene predicts poor prognosis in breast cancer patients. *Clin Cancer Res* 8 : 1167-1171, 2002
 - 40) Kumar-Singh S, Segers K, Rodeck U, Backhovens H, Bogers J, Weyler J, van Broeckhoven C, van Marck E : WT1 mutation in malignant mesothelioma and WT1 immunoreactivity in relation to p53 and growth factor receptor expression, cell-type transition, and prognosis. *J Pathol* 181 : 67-74, 1997
 - 41) Barnoud R, Sabourin J-C, Pasquier D, Ranchere D, Bailly C, Terrier-Lacombe M-J, Pasquier B : Immunohistochemical expression of WT1 by desmoplastic small round cell tumor. *Am J Surg Pathol* 24 : 830-836, 2000
 - 42) Pinol-Roma S, Dreyfuss G : Shuttling of pre-mRNA binding proteins between nucleus and cytoplasm. *Nature* 355 : 730-732, 1992
 - 43) Wilkinson MF, Shyu A-B : Multifunctional regulatory proteins that control gene expression in both the nucleus and the cytoplasm. *BioEssays* 23 : 775-787, 2001
 - 44) Niksic M, Slight J, Sanford JR, Caceres JF, Hastie ND : The Wilms' tumour protein (WT1) shuttles between nucleus and cytoplasm and is present in functional polysomes. *Hum Mol Genet* 13 : 463-471, 2004
 - 45) Englert C, Vidal M, Maheswaran S, Ge Y, Ezzell RM, Isselbacher KJ, Haber DA : Truncated WT1 mutants alter the subnuclear localization of the wild-type protein. *Proc Natl Acad Sci USA* 92 : 11960-11964, 1995
 - 46) Eccles MR, Yun K, Reeve AE, Fidler AE : Comparative in situ hybridization analysis of PAX2, PAX8, and WT1 gene transcription in human fetal kidney and Wilms' tumors. *Am J Pathol* 146 : 40-45, 1995