

Effect of shellfish culture on phytodetritus vertical fluxes in tropical waters - southern Brazil

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- **Abstract:** Mussel culture is an expanding activity in shallow and sheltered bays along the coast of Santa Catarina, southern Brazil. Although mussel culture generates benefits, several environmental impacts are associated with this kind of activity. Its success depends on factors which include the environmental impact and the carrying capacity of the system. One conspicuous effect of mussel culture is the enhancement of vertical particle flux to the bottom sediment and depletion of water column phytoplankton biomass. Phytodetritus vertical fluxes was evaluated in a mussel culture area by collecting particles with sediment traps and analysing plant pigment by high performance liquid chromatography (RF-HPLC). Results showed that mussel culture, in average, enhances almost 5 times the vertical phytodetritus flux as compared to the reference site. Pheophorbide like pigments were the main chlorophyll-*a* degradation products collected by the traps. Given the high phytodetritus production, compared to the low water column phytoplankton biomass observed, it is suggested that allocthonous phytoplankton advected to the cultured area is an important process to sustain the mussel growth in the area.
 - **Resumo:** O cultivo de moluscos ao longo da costa de Santa Catarina tem crescido de forma acelerada nos últimos anos. Embora os benefícios sejam muitos, vários tipos de impactos ambientais podem ser decorrentes do cultivo de moluscos. Desta forma, o sucesso da atividade depende de fatores que envolvem a capacidade suporte do meio e do grau que o ambiente é impactado. Um dos mais evidentes impactos associados ao cultivo de moluscos marinhos é o aumento da taxa de fluxo vertical de partículas associado a redução da biomassa fitoplanctônica na coluna de água. O fluxo vertical de fitodetrítos produzidos em uma área de cultivo foi avaliado por meio de análise por cromatografia líquida de alta eficiência (CLAE) de pigmentos fotossintéticos e produtos de degradação do material coletado por armadilhas de sedimento. Os resultados indicam que o cultivo de moluscos aumenta em média 5 vezes o fluxo vertical de fitodetrítos, sendo que a maior parte da degradação da clorofila-*a* ocorre na forma de feoforbídeos. Dado o expressivo fluxo vertical de fitodetrítos, comparado a pequena biomassa na coluna de água, sugere-se que o fitoplâncton alóctono, trazido por processos advectivos, é importante para a manutenção do crescimento dos moluscos na área estudada.
 - **Descriptors:** Pheopigment, HPLC, Detritus, Mussel culture, Sedimentation.
 - **Descritores:** Detrito, Cultivo de moluscos, Sedimentação, Feopigmentos, CLAE, Águas costeiras, Santa Catarina.
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Introduction

Aquaculture has become an important activity for developing economic growth in coastal areas of Santa Catarina State, southern Brazil. During the last ten years, total mussel production increased from 190 to 6000 metric tons, making Santa Catarina state the largest cultured mussel producer in the country (EPAGRI, 1997). The production is rapidly increasing due to cultured area expansion, introduction of new technologies and species, and more investment from state government and seafarmers. The economic aspect of mussel culture in Santa Catarina is socially important as it is carried out mostly by former artisanal fishermen which have been facing a decrease in fish catch (Medeiros *et al.*, 1997). The blue mussel *Perna perna* is the main cultured species, followed in a much less extension by the exotic oyster *Crassostrea gigas*. Culture sites along the northern coast of Santa Catarina are located in sheltered and shallow bays. Besides its benefits, environmental impacts are expected as mussel culture changes several natural features (Dhalbäck & Gunnardson, 1981; Ottman & Sornin, 1985; Hatcher *et al.*, 1994; Grant *et al.*, 1995; Dankers & Zuidema, 1995).

The environmental impacts generated by mussel culture include: 1) the aesthetics, as culture structure changes the landscape; 2) hydrological, as culture structures modify current patterns; 3) biological, with introduction of new substrate and, in some cases, new species; 4) chemical and geological, as mussel filtration activity clears the water and enhances the rate of particle sedimentation throughout faecal material production. The last item which has been studied in different environments, most from temperate regions, was recognised to affect nutrient cycling, the water column oxygen budget and particle fluxes (e.g.: Ottman & Sornin, *op. cit.*; Grant *et al.*, *op. cit.*).

Mussel feeding relies on the low selective filtering process, which leads to small particle aggregation into larger faecal pellets, enhancing local vertical particle transport (Dankers & Zuidema, *op. cit.*). The organic matter in the aggregated material can be used as substrate for heterotrophic growth, which occurs either in the water column during settling and resuspension, or at the sediment-water interface (Dhalbäck & Gunnardson, *op. cit.*). In some case, where currents are not strong enough to transport this material, bottom oxygen can be depleted, leading to anoxia of the sediment and the overlying water. A large fraction of this organic matter is derived from filtered phytoplankters which is added to the detritus pool. As mussel filtration is virtually continuous, the process represents an increase in the overall phytoplankton sedimentation

and a loss of autotrophic water column biomass. Mussel filtering rates, food availability and its vertical flux are therefore key processes to evaluate the carrying capacity of the system and associated environmental impacts.

In this study, the effect of mussel culture on vertical fluxes of phytoplankton derived detritus was evaluated on a seasonal basis in a tropical coastal area. Vertical fluxes were estimated by sediment trap experiments and phytodetritus was evaluated from the quantitative analysis of chlorophyll-*a*, some of its degradation products and carotenoids by reversed phase high performance liquid chromatography (RF-HPLC) from the collected material.

Study area

The sediment trap experiments were carried out in the "Armação do Itapocoroy" Bight (AIB), located at the northern shore of Santa Catarina State, southern Brazil (Fig. 1). The south-eastern portion of the bight is sheltered from the direct influences of southerly winds and waves, related to the passage of cold fronts, making the region suitable for culture. The area is shallow, reaching up to 10 m depth at its outermost portion.

The regional tide pattern is semi-diurnal with slight inequalities. The mean range is about 0.8 m, varying from 0.6 to 1.2 m during the neap and spring tides. Tide currents may reach up to 0.3 m s⁻¹ during spring tides, but are generally slower than 0.2 m s⁻¹. The circulation in the bight is strongly driven by winds, which usually comes from the NE sector. The salinity is generally high, but during periods of high riverine runoff, mainly from the Itajaí-Açu Estuary located about 20 km to the South, it can decrease to less than 25. Water temperature shows a seasonal pattern and ranges from 18 to 28°C during the year.

Total annual mussel production in 1997 was estimated at 1,300 tons, distributed in an area of approximately 4,000 m², where about 95% is used to culture the *Perna perna* (Mytilidae) and the remaining 5% reserved to culture the oyster *Crassostrea gigas*. Mussels are cultured using the long-line method, where individuals are fixed on a vertical rope, sustained by buoys attached to a long horizontal line. These lines are perpendicular to the coast to minimise the stress generated by swells. Phytoplanktonic biomass and production in the culture area is relatively low and dominated by diatoms. Nutrient inputs comes directly to the area from small creeks and from the Itajaí-Açu Estuary. These overall conditions enable a high mussel growth rate in the area. Commercial size (7 cm) of *P. perna* is attained in about 7 months.

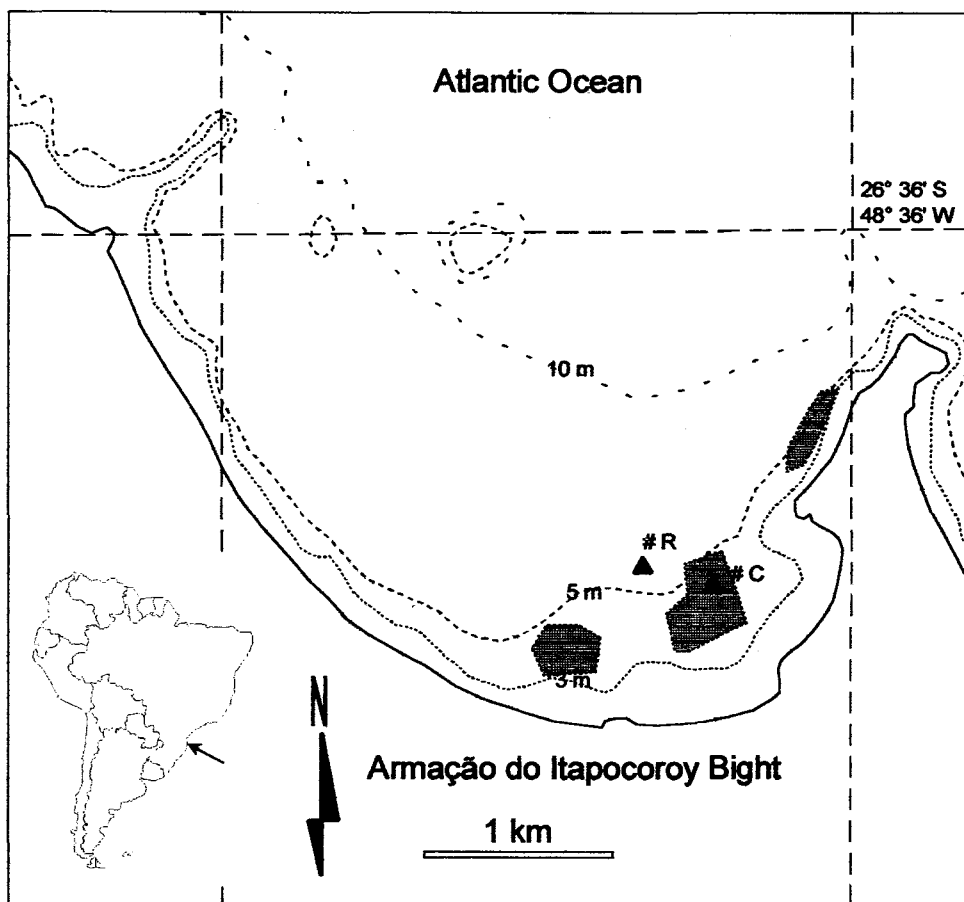


Fig. 1. Armação do Itapocoroy Bight showing culture areas (dark hatched) and trap deployment sites (R - reference and C - culture).

Materials and methods

Sediment trap experiments were carried out every two months, during neap tide periods at two sites: under the culture (#C) and outside (reference site, #R) from February to December 1996 (Fig. 1). The traps, installed and recovered by divers, consisted of two PVC tubes, attached to a submersed buoy at 1 m above the bottom (Fig. 2). Local depth ranged from 4 to 6 m at station #C and station #R, respectively. The trap system was adapted from Larson *et al.* (1986) and measured 20 cm in height and 4 cm in width, with a ratio of 5:1 (Schettini *et al.*, 1997). After 25 hours of deployment (two tidal cycles), the traps were sealed under water and brought to the laboratory within approximately one hour for sample fractionation.

The trapped material was transferred to a beaker and the volume adjusted to 200 ml. Under magnetic stirring, four to five sub-samples were taken using a syringe like device. The quartered material was retained on Whatman glass fiber GF/F by filtration under gentle pressure. Total trapped

material was estimated by weighing the filters prior and after filtration. In situ trapping reproducibility was previously tested using four cylinders and showed satisfactory results (Schettini *et al.*, 1997). The efficiency of the subsampling protocol described above was also checked and generally showed little variation among aliquots (i.e. variation coefficient usually around 3 and always less than 10%).

From a subsample, plant pigment concentration, chlorophyll-*a* (chl-*a*), its degradation products and some carotenoids were determined by reversed-phase liquid chromatography (RF-HPLC). Before grinding the filters to extract the pigments, they were soaked in 2 or 3 ml of 90 % aqueous acetone solution and left overnight in the freezer. Although this procedure may cause incomplete extraction, and better methods, such as sonication could be used, it reduces degradation of chl-*a* (Wright *et al.*, 1997). The chromatography was carried out according to Wright *et al.* (1991). The three solvent gradient elution occurred in a FLD-ODS 4 μ m column (4.5 cm x 4.5 mm, Shimadzu) as shown in Table 1.

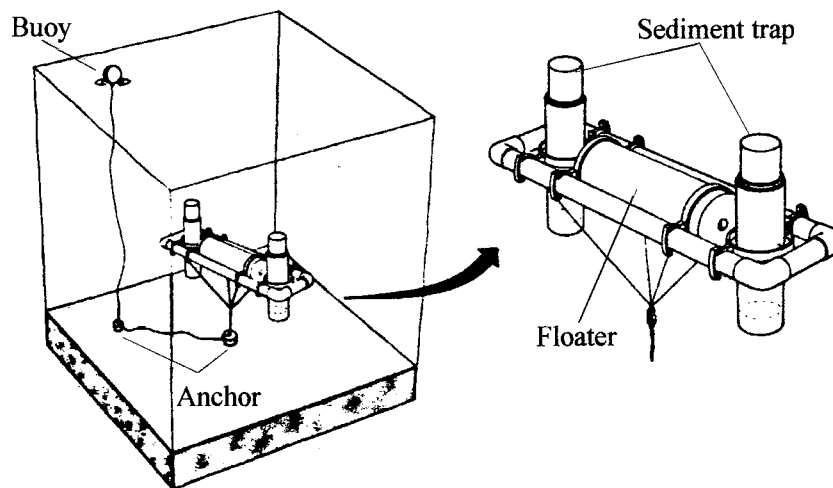


Fig. 2. The sediment trap system, after Larsson *et al.*, 1986.

Table 1. Gradient program for RF-HPLC analysis

Time (min)	Flux (ml m ⁻¹)	A (%)	B (%)	C (%)	D (%)
0	1	100	0	0	0
3	1	0	90	0	10
18	1	0	18	80	2
20	1	0	90	0	10
30	1	100	0	0	0

A: methanol : 1 M ammonium acetate (8:2)

B: acetonitrile

C: ethyl acetate

D: water

Before injecting a subsequent sample, the chromatographic initial condition was obtained by flushing the column with solvent A (methanol: 1 M ammonium acetate, 8:2) for 10 minutes. The HPLC equipment consisted in a Shimadzu[®] LC10 system composed by a quaternary solvent delivery module, a Rheodine[®] manual injector with a 100 µl sample loop, a diode array detector (200-600 nm) and a spectrofluorometer, set at 407 and 607 nm of excitation and emission, respectively. The system was controlled by a Shimadzu[®] CBM 10A controller and data acquired by a personal computer loaded with the Shimadzu[®] LC10 software. Chl-*a* peak identification and its response factor was obtained injecting pure chl-*a* from Sigma Co. standardised by spectrophotometry using the trichromatic equation from Jeffrey & Humphrey (1975).

Other chlorophyll and some carotenoid retention times were obtained analysing samples from cultured algae *Skeletonema costatum* (diatom), *Amphidinium carterae* (dinoflagellate) and *Tetraselmis suecica* (green algae, Proença, 1997). Peak identification was confirmed by their elution order and spectral characteristics. Carotenoid concentration was estimated using their specific absorption coefficient (Mantoura & Llewelyn, 1983; Descy & Mérens, 1996). Fluorescent degradation

products eluting prior and after chl-*a* were separated into two main groups: pheophorbides and pheophytins, respectively (Barlow *et al.*, 1993). Chl-*a* degradation product concentrations were estimated from the fluorescence response of acidified chl-*a* (pheophytin-*a*). The fluorometric response was assumed to be the same for all degradation products considered. To calculate the concentration of the degradation products, a factor was applied to convert chl-*a* mass loss (MW= 893,5) to pheophorbide like (MW= 592,7) and pheophytin like (MW= 871,2). The fluxes Q_t (in mg day⁻¹) were calculated as follows:

$$Q_t: (f A_p (MW_p / MW_{chl-a}) v_e / (v_f v_i)) 0.96,$$

where f is the pheophytin-*a* fluorescent response factor, A_p is the chromatographic area of pigment p in $\mu V s^{-1}$, MW is the molecular weight of p and of pheophytin-*a*, v_e is the extract volume in μl , v_f is the total trapped volume in l and v_i is the injected volume in μl . The flux equivalent of the area of the trap ($1.52 \times 10^{-3} m^2$) was then converted to m^2 .

Results

Figure 3 shows a typical chromatogram from trap samples. Several unidentified peaks were observed in both absorbance and fluorometric chromatograms, turning the identification difficult for some pigments. The absorbance chromatograms were particularly complex in the region closed to fucoxanthin retention time, between 7 to 13 minutes. The complexity was related to the presence of several carotenoid degradation products with similar spectral characteristics to their parent pigments. Another source of bias included the presence of pheophorbides eluting in this region. Although these pigments do not have a strong absorbance at 440 nm (Fig. 4), the high concentration in the samples

interferes in the absorbance of carotenoids eluting at a similar chromatographic retention time. On the other hand, fluorometric chromatograms were easier to interpret once they only registered chlorophylls and degradation products. Carotenoids do not fluoresce, therefore the fluorescent chromatogram appears much clear (Rowan, 1989).

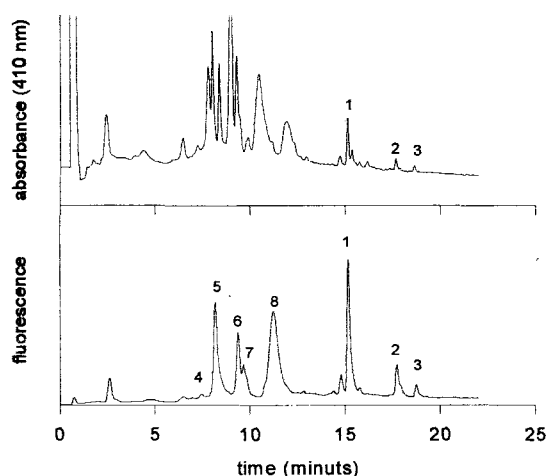


Fig. 3. Pigment separation by RF-HPLC from a 24 hours trap sample under culture (23/08/96), showing chlorophyll-*a* (1), pheophytins (2 and 3) and pheophorbides (4,5,6,7,8).

Degradation products were separated into two main groups. Five major pigments eluting between chlorophyll *c2* and *chl-a*, (within 5 and 15 minutes), with maximum absorption in the eluant around 407 nm, were named pheophorbides like pigments according to their elution order (Barlow *et al.*, 1993). They were present in all samples but varied in concentration. The other group included two pheophytin (magnesium free) pigments which eluted after *chl-a*. Diode array spectra (350 - 600 nm) in the eluant from some of these pheopigments are shown in Figure 4. Among pheophorbides and pheophytins, the former were the dominant *chl-a* degradation products in all trap samples, either from the culture or reference site.

Vertical fluxes of pigments are depicted in Figure 5. Results indicate that mussel culture increased the overall pigment vertical flux, but the proportion in which it occurs depends on the pigment or period of the year analysed. *Chl-a* fluxes ranged from 1.2 to 27.0 mg *chl-a* m⁻² d⁻¹ under the culture and from 0.4 to 5.6 mg *chl-a* m⁻² d⁻¹ at the reference site. Just in one occasion, in April, the flux of *chl-a* at the reference site was higher than under the culture array. Mussel culture enhanced the vertical *chl-a* flux up to 11 times as measured in

February 1996. Apparently, a seasonal signal was detected with lower fluxes in June and August, austral winter months. Although this behaviour may be related to other seasonal variables, such as phytoplankton biomass or mussel growth rate, data are not sufficient in number to obtain a significant correlation.

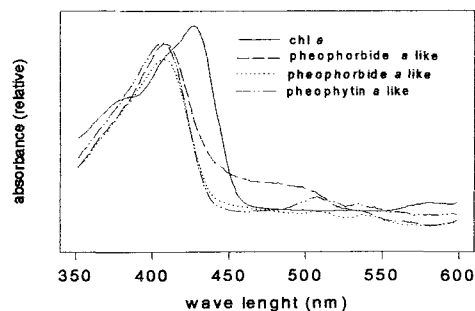


Fig. 4. Absorption spectral in eluant of chlorophyll-*a*, pheophorbides and pheophytins separate by RF-HPLC from trap samples.

Fluxes of magnesium free *chl-a* degradation products were always higher under the mussel culture. Fluxes of pheophorbides ranged from 4.6 to 55.4 mg m⁻² d⁻¹, and from 0.5 to 6.6 mg m⁻² d⁻¹ at the culture and reference sites, respectively. As shown in Figure 5, these products, but pheophytins, were the main fluorescent pigment found in trap samples under the culture with values and even higher than *chl-a*. This behaviour was slightly different at the reference site, where *chl-a* fluxes were higher in April, June and December. Pheophytins accounted for a smaller portion of total pheopigment fluxes both under the culture and the reference site.

Table 2 summarises the results obtained during the experiments. It is evident that the fluxes of all pigments analysed were increased due to the culture, but not at the same proportion. While pheophorbide flux was in average 8 times higher under the culture, pheophytins had an increment of only 50%. *Chl-a* flux was in average 3 times higher under the culture site. The effect of the culture is evident in the total particle vertical flux as well, which averaged 44.8 and 110.3 g m⁻² d⁻¹ at the reference and culture sites, respectively. From these values, we estimate the average *chl-a* and pheopigment content in the settling particles from the two sites. For *chl-a* a quite similar value was observed between the reference and culture sites, 5.4 x 10⁻³ and for 6.3 x 10⁻³ %, respectively. On the other hand, pheopigment (pheophorbides plus pheophytins) content as percentage of the total settling particles fluxes, was two times higher under the culture.

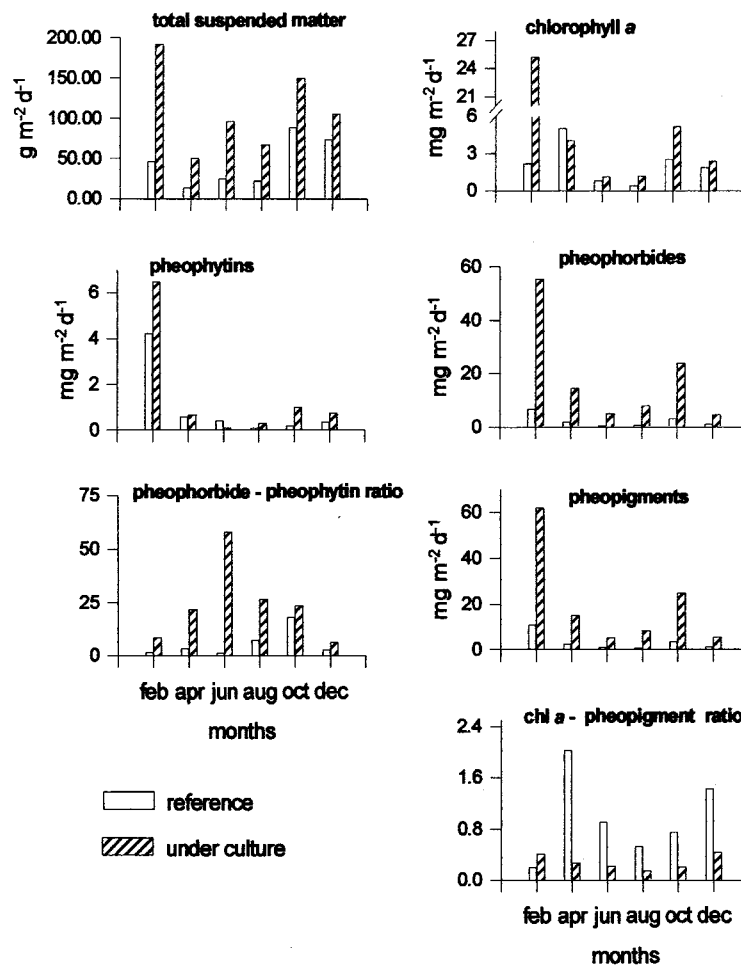


Fig. 5. Monthly fluxes of total particulate matter, chlorophyll-*a*, pheophytins, pheophorbides, total pheopigments and the ratios of pheophytins to pheophorbides and chlorophyll-*a* to pheopigments obtained by trap samples at culture area and at the reference sites at Armação do Itapocoroy Bight during 1996.

Table 2. Averaged pigment fluxes and pigment ratios obtained from trap samples in side of culture area and at the reference site.

Site	Total Suspended matter $\text{g m}^{-2} \text{d}^{-1}$	Chl- <i>a</i> $\text{mg m}^{-2} \text{d}^{-1}$	pheophorbides $\text{mg m}^{-2} \text{d}^{-1}$	pheophytin $\text{mg m}^{-2} \text{d}^{-1}$	pheopigment $\text{mg m}^{-2} \text{d}^{-1}$	Chl- <i>a</i> / pheopigment
Culture (c)	110.3	6.9	18.5	1.5	20.1	0.3
Reference (r)	44.8	2.4	2.3	1.0	3.3	0.7
c / r	2.5	2.9	8.0	1.5	6.1	

Discussion

Mussels cause the deposition of a great amount of suspended matter in which phytodetritus is included. Generally, methods applied to evaluate chl-*a* concentration in the water column, such as the

spectrophotometric and fluorometric, are used to quantify the phytodetritus portion in the settling material. This procedure has some drawbacks due to interference of degradation products, which are avoided with the use of HPLC method. In this study, apart from chl-*a*, we have found several degradation

products in the settling material produced during mussel filtration. The most common products of the degradation of the chlorophylls are the Mg-free derivatives, pheophytins or pheoporphyrins, formed rapidly when the pH is lowered. Loss of the phytyl chain by hydrolysis is another common degradation route, forming chlorophyllide. When both the phytyl chain and Mg are lost, the product is a pheophorbide (Rowan, 1989). While pheophorbides are major degradation products generated during animal grazing (Vernet & Lorenzen, 1987), pheophytin-*a* can play a role in the photosynthesis and is always present as a trace pigment in natural populations (Porra *et al.*, 1997). Although pheophorbides can dominate the pheopigment content in seawater, significant amount of pheophytin can be produced by some specific grazers (Vernet & Lorenzen, *op. cit.*). Our results show that pheophorbides are the major degradation product under the mussel culture and pheophytins the minor, being this degradation product found in up to a 58 times smaller concentration, as observed in June.

In contrast to the data obtained by fluorometric detection, not much can be inferred from the analysis of the absorbance chromatogram of the trapped material due to the large concentration of degradation products. In samples from the water column, usually with a low level of degradation products, useful information such as the dominant algal group, can be easily obtained by the analysis of the accessory carotenoids given by the absorbance chromatogram. But carotenoids, as chlorophylls, can be degraded during mussel filtration and several products formed, increasing the complexity of the chromatographic signal, as also observed in this study. Although we used a diode array detector, which could furnish an additional information, the absorption spectra within carotenoids or their degradation products are quite similar. Therefore, they must be chromatographically well resolved to be properly analysed. Apparently, fucoxanthin was a dominant carotenoid in the samples. This pigment is a marker used for diatoms (i.e. Mantoura & Llewellyn, 1983) and its presence indicates that this group is among the main algae filtered by the cultured mussel. The dominance of diatoms within the water column has been previously observed by the distribution of fucoxanthin, confirmed by microscopy (unpublished data).

Several factors may have affected the pigment vertical flux distribution observed during this study (Fig. 5). They include: particle availability, amount and quality, mussel biomass and filtration rate, advection and resuspension, among others. The Itajaí-Açu River, located 20 km southwards, has been identified as an potentially important source of

particles and nutrients to the bight. The dispersion of the Itajaí-Açu River plume is largely driven northwards (Schettini *et al.*, 1998). Thus, the water quality of AIB is expected to vary according to Itajaí-Açu River runoff pulses and cross shelf exchange, affecting the observed temporal vertical particle fluxes. The extent of these meso-scale process is still difficult to assess. However, salinity revealed low impact, and showed little variation during the sampling days, ranging between 31 and 35. Most probably, the effect of the Itajaí-Açu River and other estuaries occurs in a more indirect way.

Another factor which may have influenced the temporal variation of the vertical fluxes was bottom resuspension. Sediment granulometric measurements revealed a low percentage of fine sediments, with a predominance of sand and a high percentage of carbonates (Schettini, unpublished data). This indicates that although vertical fluxes are enhanced below the mussel culture array, particles may be resuspended to the water column and flushed out by advection. Resuspension might have been the cause for the extreme high vertical flux observed in February, as the experiment was carried out under high external physical energy, with strong winds and long period waves.

The effect of resuspension on the mussel feeding must be addressed in the future to make possible a better estimation of the fate and origin of the particles. On the other hand, the present data may be used to formulate some hypotheses. The growth rate of *P. perna* at AIB is high, attaining commercial size (e.g. 7 cm length) in approximately seven months. Food availability has been identified as one of the most important factor for this rapid growth. The high filtration rates and food requirements certainly contributes to the relatively low chl-*a* values found in the water column at Armação do Itapocoroy during this study, which averaged 1.0 mg m^{-3} ($n=40$). Averaged chl-*a* plus pheopigment flux under the culture attained $27.0 \text{ mg m}^{-2} \text{ d}^{-1}$. This value is about 5 times greater than the averaged instantaneous chl-*a* concentration in the water column (1.0 mg m^{-2} and 6 m depth). Even considering a fast phytoplankton doubling rate, e.g. $\mu_{\text{max}}: 1.0 \text{ d}^{-1}$ for eutrophic coastal waters (Parsons *et al.*, 1984), in situ production would not be enough to sustain the observed phytodetritus vertical flux. If in situ production is not sufficient, the phytoplankton settled biomass must come from elsewhere. Hydrological measurements showed that water advection is a major process at AIB (Schettini *et al.*, 1997). It can be inferred that this process is responsible for influx of allochthonous phytoplankton biomass to AIB, which will sustain the high mussel growth rates and phytodetritus fluxes observed. On

the other hand, resuspension caused by wave action may also play a role in the process. Particles already settled could be resuspended and filtered, thus diminishing the importance of the allochthonous input of particles.

It is important to point out that the pigments analysed are not the only degradation products of chl-*a*. Several other pathways are possible, including the production of colourless products (Svec, 1978). Therefore, the phytodetritus vertical flux may have been even greater than the sum of those pigments analysed. Comparable results were obtained by Dahlbäck & Gunnarsson (1981) investigating vertical fluxes under a similar mussel culture at the Swedish west coast in a shallow, 8-13 m deep, bay. Their calculated ratios referred to the fall season in a temperate ecosystem. Chl-*a* averaged vertical fluxes at AIB under the culture and reference sites were approximately 3 times higher than that measured by Dahlbäck & Gunnarsson (*op. cit.*), even though the ratio between culture and reference was quite similar as well the ratio of chl-*a* / pheopigment.

Our results showed that mussel culture affects the chl-*a* to pheopigment (mainly pheophorbides) ratio in the particulate matter changing it from 1:1.4 to 1:3. This change is even greater when compared to the averaged value of 13:1 found in the water column. This finding indicates that the mussel culture at AIB has an impact not only on the bulk of particle flux, but in its quality as well. This fact has an implication on chl-*a* measurements within culture areas based on fluorometric or spectrophotometric methods. Both standard methods are biased by the presence of degradation products (Mantoura *et al.*, 1997). Therefore, an overestimation of water column chl-*a* at mussel culture areas is to be expected. If proper measurements of the chl-*a* to degradation products are taken within the area, they could be used as indicative for mussel derived phytodetritus.

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