

# PLANKTON STUDIES IN A MANGROVE ENVIRONMENT. VII. SIZE FRACTIONATION OF THE PHYTOPLANKTON: SOME STUDIES ON METHODS

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## SYNOPSIS

A series of experiments on the different methods of measuring the relative uptake of  $C^{14}$ , by the various size fractions of the phytoplankton, were carried out in a mangrove swamp of the estuarine type at 25° south latitude. As a complement, some factors which might influence the relative uptake of the different fractions in inshore waters, were studied. These factors are related, mainly, with the increase on the amount of labelled particulate matter during incubation period, the effect of different pressures in the filtration of the size fractions of the phytoplankton, the use of a range of volumes for the incubation of samples, and the effect of formalin on the fixation of samples, after the incubation period. Based on these results, some suggestions are made for the use of the  $C^{14}$  method in inshore waters.

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## INTRODUCTION

The separation of the phytoplankton in constituents of various size range, has been object of intensive studies, mainly, in the last few years. The investigations carried out by ATKINS (1945), WOOD & DAVIS (1956), STEEMANN-NIELSEN & JENSEN (1957), HOLMES (1959), YENTSCH & RYTHER (1959), TEIXEIRA (1963), HOLMES & ANDERSON (1963), ANDERSON (1964), MULLIN (1965), ANDERSON (1965), TEIXEIRA, TUNDISI & SANTORO (1967) demonstrated that a great part of the phytoplankton will pass through the pores of a fine net.

The significant role of the nannophytoplankton in the primary production, in the oceans and lakes, has been emphasized by SAIJO (1964), SAIJO & TAKESUE (1965) and GESSNER (1959).

By the other hand, these studies have considerable interest, in the possible role played by the different fractions, in the feeding of herbivores. In fact, the knowledge of the various size fractions which are offered to the herbivores is of prime importance in the studies of some feeding relationships as selective grazing.

BALLANTINE (1953) made a intensive study of the procedures for the estimation of nannophytoplankton, by comparing six different methods; it was de-

monstrated that centrifugation of a living samples followed by determination of the number of organisms, in the concentrate, was the most satisfactory method.

Research on the comparison of the different methods for the assessment of the relative importance of the fractions of the phytoplankton is scarce. In the investigations carried out in Vineyard Sound, YENTSCH & RYTHER (1959) estimated the relative importance of net-nannophytoplankton, by chlorophyll content, photosynthesis and cell numbers. The net portion constituted a small percentage of the population, and exhibited marked seasonal trends.

These results agree well, with a seasonal study at 25° south latitude, (TEIXEIRA *et al.*, 1967) during 1965; the bulk of the population was composed by nannophytoplankton smaller than 50  $\mu$ , and the net-phytoplankton showed very defined seasonal variation, as demonstrated by the relative uptake of  $C^{14}$ .

In the present investigation, attention has been paid to the methods of measuring relative uptake of  $C^{14}$  by the different fractions of the phytoplankton given by STEEMANN-NIELSEN & JENSEN (1957) and YENTSCH & RYTHER (1959). As a complement, some factors which might influence the relative uptake of  $C^{14}$ , by the different fractions, in inshore waters, were studied.

This research was carried out at three stations in a mangrove region of the estuarine type, at 25° south latitude, during October and November, 1966.

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## MATERIAL AND METHODS

### 1 — Area studied

Previous investigations, (TEIXEIRA & KUTNER, 1963; TEIXEIRA, TUNDISI & KUTNER, 1965) demonstrated that the region studied, has different environmental characteristics, at different geographical positions. Thus, three stations were selected: one, in the inner parts of the lagoon (Maria Rodrigues river), one in the middle region, and one in the entrance of the estuary, subjected to a more marked influence of the coastal water. Sketches of the lagunar region, have been published elsewhere in earlier papers (TEIXEIRA *et al.*, 1968; TUNDISI & TUNDISI, 1968).

Besides the comparison of methods in these stations, an attempt was made to obtain a complementary information, on the relative importance of the distinct fractions in the area studied.

### 2 — Methods

Surface water, was collected at each station in volumes of 8-10 litres, in plastic bottles, and then, brought back immediately to the laboratory to be submitted to the different procedures.

I — a) The sea water was placed into clean, 125 ml pyrex bottles, and inoculated with radioactive sodium bicarbonate. After this procedure, the samples were incubated, during four hours at  $25 \pm 1^\circ\text{C}$  with approximately 10,000 Lux light intensity, and later filtered through a silk net with  $25 \mu$  pore size, and a silk net with  $50 \mu$  pore size (STEEMANN-NIELSEN & JENSEN, 1957).

After filtration, through the nets, the samples were filtered through H. A. Millipore filters, 25 mm diameter,  $0.45 \mu$  pore size.

A similar treatment was given to another samples, of the same sea water, only that the filtration through the nets, was done before inoculation of radioactive  $\text{C}^{14}$ . Incubation was done at the same temperature and light intensity.

The phytoplankton was thus divided into two fractions: smaller than  $50 \mu$  (from  $0.45 - 50 \mu$ ) and smaller than  $25 \mu$  (from  $0.45 - 25 \mu$ ).

b) One litre of the sea water was filtered through a net with  $50 \mu$  pore size, and washed in one litre of the same sea water, Millipore filtered ( $0.8 \mu$  pore size, and 45 mm diameter) (YENTSCH & RYTHER, 1959). 125 ml pyrex bottles, were then filled with this sea water. The remaining net-filtered sea water (i.e. the fraction smaller than  $50 \mu$ ) was used to fill another series of bottles.

The samples were inoculated with radioactive sodium bicarbonate, and incubated at approximately 10,000 Lux light intensity during 4 hours.

The same treatment was given to sea water which have been passed through nets with  $25 \mu$  pore size.

For each experiment of the *a* and *b* series, replicates were prepared, as well as a dark bottle for correction.

II — A study on the effect of the dilution of the phytoplankton population, when filtered through a net and washed in Millipore filtered sea water, was included: for each station, 50 litres of surface sea water, were filtered through a  $50 \mu$  pore size net silk, and diluted in 100 ml of Millipore ( $0.45 \mu$  pore size-HA) filtered sea water. 10 ml of this water, were diluted to 100 ml, 250 ml and 500 ml of the same sea water.

Each sample was inoculated with the corresponding amount of radioactive sodium bicarbonate, necessary to give approximately the same activity for the different volumes.

III — ANDERSON (1965) found that 1.2 to 6.6% of the total  $\text{C}^{14}$  uptake in raw sea water, occurred in samples previously filtered, through filters with  $0.8 \mu$  pore size. To investigate this possibility, some experiments were conducted in the following way:

The sea water was first filtered, through filter paper, Whatmann n.º 1, then through glass filters, with  $9 \mu$ , pore size, and through a Millipore filter  $0.8 \mu$  pore size. 125 ml pyrex bottles, were filled with this sea water and inoculated with radioactive sodium carbonate. After the incubation period during 4 hours, and 10,000 Lux approximately, 100 ml of this samples was filtered through Millipore filters of  $0.45 \mu$  pore size.

Another series was prepared using different volumes of sea water, under the same conditions of illumination, and the same time for incubation. Volumes used were 50 ml, 100 ml, 250 ml and 500 ml.

IV — The effect of positive and negative pressure in the different fractions.

Some studies which have been made (LASKER & HOLMES, 1957) have demonstrated that strong pressures can present a problem in the filtration of samples inoculated with radioactive carbon.

GUILLARD & WANGERSKY (1958) and TUNDISI (1965) suggested that the pressure applied, can be responsible for the liberation of cellular material in old cultures of flagellates. Hence, to investigate a possible effect of positive and negative pressure in the different fractions of phytoplankton, during filtration, two series of experiments were prepared:

The sea water was filtered through a silk net of  $50 \mu$ , inoculated with radioactive sodium bicarbonate, and incubated for 4 hours at approximately 10,000 Lux light intensity. After this time, the samples were filtered through a Millipore filter,  $0.45 \mu$  pore size, under positive pressure, of 1.0, 1.5, 2.0  $\text{Kg}/\text{cm}^2$ . The same treatment was given to sea water which was passed through a silk net of  $25 \mu$ , and to total samples of phytoplankton (i.e. not filtered sea water).

V — The effect of formaldehyde in the different fractions.

Some authors (STRICKLAND & PARSONS, 1965) recommended the use of formalin, immediately at the end of the incubation period to preserve the samples. It is indicated that the formaldehyde may affect the more delicate algae. Thus, to investigate this possibility, and the effect of formaldehyde, on the different fractions, the following series of experiments was prepared:

Twelve samples of sea water, were inoculated with radioactive sodium bicarbonate in 125 ml pyrex and incubated for 4 hours. After this period six samples were filtered through H.A. Millipore filter 0.45  $\mu$  pore size and to the remaining six of them, formaldehyde was added. Both the batches of samples, were passed through nets of 50  $\mu$  and 25  $\mu$  pore size. Thus, any effect caused by the use of formaldehyde to stop photosynthesis could be detected in comparison with the other samples.

An end-window Geiger Müller tube (Phillips) was used to measure the radioactivity in the filters, for all the samples. All the results are given in counts per minute.

## RESULTS

### I — Methods of fractionation

Table I compares the results given by the first method used (STEEMANN-NIELSEN & JENSEN, 1957) in the three stations. This method seems to give always greater results, for the fractions smaller than 50  $\mu$ , unless at station II, where the average results were almost similar.

TABLE I — Results for the fractionation of total samples of phytoplankton, with the first method used (description in the text)

STATION I		cpm
Size fraction		
	Total .....	2.289
a)	< 50 $\mu$ .....	2.099
	< 25 $\mu$ .....	1.567
		cpm
b)	< 50 $\mu$ .....	1.745
	< 25 $\mu$ .....	2.042
STATION II		cpm
Size fraction		
	Total .....	1.059
a)	< 50 $\mu$ .....	255
	< 25 $\mu$ .....	295
		cpm
b)	< 50 $\mu$ .....	358
	< 25 $\mu$ .....	377
STATION III		cpm
Size fraction		
	Total .....	1.179
a)	< 50 $\mu$ .....	761
	< 25 $\mu$ .....	404
		cpm
b)	< 50 $\mu$ .....	762
	< 25 $\mu$ .....	561

a = Filtered before inoculation and incubation.  
b = Filtered after inoculation and incubation.

In all the three stations the average of the cpm was greater in the samples filtered through the nets after incubation. Table II, shows the results obtained with the second method (YENTSCH & RYTHER, 1959).

TABLE II — Results for the fractionation of total samples of phytoplankton, with the second method used (description in the text)

STATION I		cpm
Size fraction		
	Total .....	1.365
	< 50 $\mu$ .....	1.271
	< 25 $\mu$ .....	374
STATION II		cpm
Size fraction		
	Total .....	570
	< 50 $\mu$ .....	204
	< 25 $\mu$ .....	456
STATION III		cpm
Size fraction		
	Total .....	585
	< 50 $\mu$ .....	359
	< 25 $\mu$ .....	436

There were fluctuations in the number of cpm in all the stations studied. The fraction smaller than 50  $\mu$ , was the most important at station I. The fraction greater than 25  $\mu$ , was the most important at station II and III.

It was demonstrated that the fraction greater than 50  $\mu$ , was the less important regarding the C<sup>14</sup> uptake with the two methods studied.

### II — Experiments with dilution of the phytoplankton populations

In the three stations studied, concentration of the phytoplankton, and subsequent dilution in different volumes, demonstrated that differential C<sup>14</sup> uptake occurs.

In some experiments the level of C<sup>14</sup> uptake was depressed at volumes of 0.5 l, when compared with the level of C<sup>14</sup> uptake at volumes smaller than 0.5 l.

Table III, summarizes, the value obtained, for the three stations studied, and with increasing volumes of Millipore filtered sea water.

TABLE III — Effect of dilution of phytoplankton, in samples of different volume

STATION I		cpm
DILUTION		
Volume used		
	50 ml .....	368
	100 ml .....	500
	100 ml .....	380
	250 ml .....	569
	500 ml .....	567

STATION II

Volume used	cpm
100 ml .....	238
100 ml .....	294
250 ml .....	471
250 ml .....	365
500 ml .....	141
500 ml .....	251

STATION III

Volume used	cpm
100 ml .....	1.107
250 ml .....	1.216
500 ml .....	922

III — Particulate matter

The results for the series of experiments in which the labelled particulate matter is formed during experimental hours, are summarized in Table IV. It seems clear, that a certain amount of labelled particulate matter is present at the end of the incubation period.

TABLE IV — Particulated matter formed during experimental hours

Volume used	cpm
50 ml .....	21.2
100 ml .....	44.0
a) 250 ml .....	95.0
500 ml .....	134.8
Volume used	cpm
b) 100 ml .....	3.4
250 ml .....	4.6
500 ml .....	19.4

a = Samples submitted to illumination  
 b = Dark bottle.

When the samples were filtered twice, in the same series of filters used, there was a reduction of the order of 27% in the "blank" (Table V).

The percentage of cpm in the raw sea water of different volumes used ranged from 0.3 to 1.6%.

TABLE V — Particulate matter formed during experimental hours

Volume used	cpm
a) 100 ml .....	52
100 ml .....	45
100 ml .....	28 (black bottle)
b) 100 ml .....	12
100 ml .....	15
100 ml .....	10 (black bottle)

a = Filtered once Whatmann n.º 1, filter paper; glass filter M.F., Millipore filter.

b = Filtered twice, Whatmann n.º1, filter paper; glass filter M.F., Millipore filter.

IV — Effect of pressure

I — Different positive pressure applied on total samples, at the three stations studied, showed differences in the number of counts per minute. This was particularly marked at station II, where, with a pressure of 2.0 kg/cm<sup>2</sup> there was a considerable reduction in the number of counts. At stations I and III a less marked effect of pressure was observed (Table VI).

TABLE VI — Effect of different positive pressures on total samples

STATION I	Kg/cm <sup>2</sup>	cpm (average of three replicates)
	0.5 .....	2.032
	1.0 .....	2.364
	1.5 .....	2.227
	2.0 .....	2.109

STATION II	Kg/cm <sup>2</sup>	cpm (average of three replicates)
	0.5 .....	1.224
	1.0 .....	726
	1.5 .....	950
	2.0 .....	441

STATION III	Kg/cm <sup>2</sup>	cpm (average of three replicates)
	0.5 .....	1.135
	1.0 .....	882
	1.5 .....	956
	2.0 .....	1.118

2 — A more marked effect was observed when different positive pressures, were applied in the different fractions. The fraction smaller than 25 μ showed a reduction of approximately 50% in cpm at 2.0 kg/cm<sup>2</sup> in relation to the other pressures applied (Table VII).

TABLE VII — The effect of different positive pressure in the different fractions

Pressure Kg/cm <sup>2</sup>	cpm *	Size fraction
1.0	389 .....	Total
	307 .....	50 μ
	244 .....	25 μ
1.5	319 .....	Total
	273 .....	50 μ
	322 .....	25 μ
2.0	428 .....	Total
	282 .....	50 μ
	164 .....	25 μ

\* Average of two replicates.

The composition of the fractions was not studied, but it seems thus that the fraction smaller than 25  $\mu$  was constituted by more fragile cells.

#### V — The effect of formalin

Table VIII shows the differences obtained when formalin is used to stop photosynthesis after the samples were incubated during four hours.

TABLE VIII — Fixation with formalin\* 4% after incubation

Size fraction	cpm with formalin added	cpm with no formalin added
25 $\mu$ .....	2.612 .....	4.116
25 $\mu$ .....	2.368 .....	3.994
50 $\mu$ .....	3.544 .....	4.574
50 $\mu$ .....	3.880 .....	5.126
Total sample	3.880 .....	5.126
	3.400 .....	4.850

\* Formalin prepared according to Strickland & Parson, 1965.

#### DISCUSSION

It can be postulated, that, to some extent, the first method used in the study of size distribution of phytoplankton is more reliable. The fact that the different fractions are separated before the incubation period, might be the cause of differential results. However, it is not clear such fact from the above results on the comparison of the two methods, but, filtration before the incubation period, may affect the phytoplankton causing a damage in the different fractions, lowering the  $C^{14}$  uptake.

SALJO (1964), pointed out that some differences which were found in his results, and KAWAMURA'S (1960) might be due in part to this difference in technique (i.e. filtration before and after incubation) though these differences were due also to the geographical variation of phytoplankton species.

When the filtered phytoplankton, is confined in different volumes there are differences in the  $C^{14}$  uptake too. These differences might arise from a "group effect" or the availability of nutrients to the different fractions. Also, it was clearly demonstrated that the greater the volume used, the greater the amount of particulate matter formed during the incubation period. It was showed that filtration through a series of filters is efficient in removing most of the particulate matter, but even so, this could not be prevented.

The causes which are responsible for the presence of higher amounts of particulate matter at the end of the incubation period are unknown. Perhaps the greater surface in the glass bottles would contribute to the formation of labelled organic aggregates. The results obtained, are lower than those quoted by ANDERSON (1965).

It seems, thus, from these experiments, that the use of a "blank" in certain types of experiments in estuarine waters, is indicated. This blank could be made by filtering the raw sea water, in a series of filters of different pore size, and then on Millipore or Membrane filters. The possibility of increasing the amount of labelled particulate matter during incubation is prevented but not at all excluded. This depends on the region studied. Regions with higher content of "yellow substances", might give certain amounts of labelled particulate matter after incubation. The use of formaldehyde, may be responsible for the loss of organic matter from the more delicate algae; it is clear from these experiments that the fraction smaller than 25  $\mu$ , is the most affected by this treatment. Further, it is generally known from earlier work, that, this fraction is composed mainly by small flagellates during the time when the study was done. It would be useful to compare this effect on the small diatoms.

Finally, in comparing the size distribution of phytoplankton in the area studied, it seems that the fraction smaller than 50  $\mu$ , represents the greater percentage regarding the uptake of  $C^{14}$ . This is in agreement with earlier results (TEIXEIRA, TUNDISI & SANTORO, 1967), as well as some results obtained this year (1967, unpublished results).

These results are also in agreement with the data obtained by SALJO (1964), SALJO & TAKESUE (1965) for the Indian Ocean. COLLIER & MURPHY (1962), called attention to the fact that very small diatoms and flagellates are characteristic of waters with low concentration of nutrients. Rapid multiplication of these forms would occur even under poor nutritional conditions.

Studies on the seasonal variation of the different fractions were already done (TEIXEIRA *et al.*, 1967), and data from a more detailed investigation, are being worked (TUNDISI, in preparation). The causes which could influence this composition might be related to seasonal differences in the influx of nutrients. Also, the amount of humic substances added from land drainage, could be of great importance, to this seasonal variation (PRAKASH & HODGSON, 1966; TEIXEIRA, TUNDISI & SANTORO, 1968).

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#### RESUMO

Como parte do programa de estudos do plâncton, da região lagunar de Cananéia, foi levado a efeito uma série de experimentos com o método do  $C^{14}$ , utilizado para a determinação da fixação relativa do isótopo, em cada fração do fitoplâncton. Foram testados principalmente os métodos utilizados por STEEMANN-NIELSEN & JENSEN (1957) e YENTSCH & RYTHER (1959). Como complemento, foram realizados experimentos para verificação do efeito da pressão negativa sobre as diversas frações do fitoplâncton, o

uso de diferentes volumes para incubação das amostras, e o efeito da fixação das amostras, com formol 4% depois do período de incubação, e antes da filtração. Atenção especial foi dada ao problema de formação da matéria particulada durante o período de incubação. Com base nos resultados, são feitas sugestões para o uso do método do  $C^{14}$  em águas estuarinas.

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