

PHYTOEXTRACTION AND FRACTIONATION OF HEAVY METALS IN SOIL AFTER MULTIPLE APPLICATIONS OF NATURAL CHELANTS

Évio Eduardo Chaves de Melo¹; Clístenes Williams Araújo do Nascimento^{1*}; Adriana Maria de Aguiar Accioly²; Ana Cristiane Queiroz Santos¹

¹UFRPE - Depto. de Agronomia - R. Dom Manuel de Medeiros, s/n - Dois Irmãos - 52171-900 - Recife, PE - Brasil.

²Embrapa Mandioca e Fruticultura Tropical - Embrapa, s/n - 44380-000 - Cruz das Almas, BA - Brasil.

*Corresponding author <clistenes@pq.cnpq.br>

ABSTRACT: Chelate-assisted phytoextraction of heavy metals is a promising approach to clean up polluted soils. However, the most successful chelants tested so far are synthetic agents that barely degrade in soil, increasing the metal leaching risks. Natural organic acids have been proposed to enhance phytoextraction due to their higher biodegradability, but they can also be a drawback for efficient phytoextraction. This work was carried out to compare the effectiveness of multiple applications of citric and gallic acids on the availability in soil and accumulation of Cd, Pb, Cu, and Zn by velvet bean (*Stizolobium aterrimum*) plants. The organic acids were added as follows: 5 mmol kg⁻¹ in the 28th cultivation day; two doses of 5 mmol kg⁻¹ at the 28th and 31st day; and three applications at the same rate at the 28th, 31st, and 34th day of velvet bean cultivation. Soil samples were sequentially extracted and soil solution metal concentrations assessed. Neither citric acid nor gallic acid was efficient for metals phytoextraction. In general, low molecular weight organic acids (LMWOA) application increased the metal concentrations in the water soluble and exchangeable fractions. Zinc and Cu were retained mostly in the organic matter fraction. Zn was remobilized from the organic matter fraction through LMWOA application into the water soluble and exchangeable fractions. LMWOA mobilized Pb and Cu from iron oxides, but such an increase in solubility was not high enough to affect phytoextraction.

Key words: phytoremediation, soil pollution, soil remediation, organic acids

FITOEXTRAÇÃO E FRACIONAMENTO DE METAIS PESADOS EM SOLO APÓS MÚLTIPLAS APLICAÇÕES DE ÁCIDOS ORGÂNICOS

RESUMO: Fitoextração de metais induzida por quelantes é uma técnica promissora para despoluição de solos. Entretanto, os quelantes mais eficientes são sintéticos e apresentam baixa taxa de biodegradação, o que aumenta o risco de lixiviação de metais. Ácidos orgânicos têm sido propostos para auxiliar na fitoextração, mas a sua rápida degradação pode ser limitante para a eficiência desta técnica. O objetivo do trabalho foi comparar a eficiência de aplicações sucessivas dos ácidos cítrico e gálico, na disponibilidade, absorção e acumulação de Cd, Pb, Cu e Zn pela mucuna preta (*Stizolobium aterrimum*). O solo foi contaminado com Cd, Pb, Cu e Zn nas doses 20, 150, 100 e 150 mg kg⁻¹, respectivamente, e cultivados por 35 dias. Os ácidos foram aplicados de três formas: uma aplicação de 5 mmol kg⁻¹ no 28^o dia de cultivo da mucuna; duas aplicações de 5 mmol kg⁻¹ no 28^o e 31^o dia; e três aplicações de 5 mmol kg⁻¹ no 28^o, 31^o e 34^o dia de cultivo. As amostras de solo foram submetidas à extração e fracionamento químicos. Os ácidos orgânicos testados não foram eficientes na indução da fitoextração. Em geral, a aplicação dos ácidos aumentou as concentrações de metais tanto nas frações solúveis como em água e também na trocável. Zinco e cobre ficaram retidos principalmente na fração matéria orgânica. O Zn, após aplicação dos ácidos, foi remobilizado da fração matéria orgânica para as frações solúvel em água e trocável. Os ácidos orgânicos mobilizaram Pb e Cu dos óxidos de ferro, mas este aumento na solubilidade não foi suficiente para favorecer a fitoextração.

Palavras-chave: fitorremediação, poluição do solo, remediação do solo, ácidos orgânicos

INTRODUCTION

Environmental restoration of polluted soils by conventional technologies demands large economic re-

sources (Cunningham & Berti, 2000; Ensley, 2000; Nascimento & Xing, 2006). Thus a more affordable approach to decontaminate soils are increasingly gaining interest. Phytoextraction - the use of plants to remove

metals from soils by storing them in above-ground biomass - has been developed as an economically and environmentally attractive technology to decontaminate heavy metal polluted soils (Raskin et al., 1997; Terry & Bañuelos, 2000). So far, two phytoextraction strategies have been developed. The first one is the use of hyperaccumulating plants that naturally concentrate metals in their aboveground tissues. The other strategy makes the use of high biomass plants that are induced to accumulate metals through chelants applications in soils (Blaylock et al., 1997; Huang et al., 1997; Cooper et al., 1999; Wu et al., 1999; Shen et al., 2002; Melo et al., 2006). The main drawback for the public acceptance of this technique is the risk that synthetic chelants pose to environment due to a possible excessive metal solubilization and consequent leaching (Shen et al., 2002; Chen et al., 2003).

The use of low molecular weight organic acids (LMWOA) which are easily biodegradable has been proposed as an alternative to the synthetic chelants. However, many authors have found low effectiveness of LMWOA on inducing metals accumulation in plants (Lombi et al., 2001; Kos & Lestan, 2004; Wu et al., 2004; Nascimento et al., 2006). This low LMWOA effectiveness has been attributed to their rapid mineralization by soil microorganisms (Krishnamurti et al., 1997; Römkens et al., 2002; Meers et al., 2005). For instance, Ström et al. (2001) reported that one third of citric acid added to soil was degraded after a 24-hour period.

Taking into account the low efficiency of a single application of LMWOA on phytoextraction enhancement, we studied the effect of multiple applications of citric and gallic acids on the phytoextraction of Cd, Pb, Cu, and Zn by velvet bean plants (*Stizolobium aterrimum*). Additionally, a soil fractionation was performed to determine the metal soil pools most affected by organic acids and how this effect can be related to the phytoextraction.

MATERIAL AND METHODS

The soil material used in this study was classified as Ustic Kandihumult. Selected chemical and physical properties of the soil are: pH = 4.3 (water/soil 1:2.5); exchangeable acidity = 8.2 (Al, in $\text{mmol}_c \text{dm}^{-3}$); exchangeable basis = 5.5 (Ca and Mg, respectively in $\text{mmol}_c \text{dm}^{-3}$) and 5.0; extractable phosphorus = 12.0 mg dm^{-3} ; exchangeable potassium, sodium and total activity = 0.5, 1.4 and 49.5, respectively, (in $\text{mmol}_c \text{dm}^{-3}$); organic matter = 566 g kg^{-2} ; and sand, silt and clay (in g kg^{-1}) = 566, 48 and 386, respectively.

The soil sample was air dried, passed through a 2 mm sieve and limed to pH 6.5 - 7.0 with carbon-

ates of Ca and Mg (3:1 molar ratio solution) before further use. Samples of 1 kg were amended with CdCl_2 , PbCl_2 , $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$, and ZnCl_2 to achieve the concentrations of 20, 150, 100, and 150 mg kg^{-1} of Cd, Pb, Cu, and Zn, respectively. These metal concentrations were chosen to simulate a soil moderately polluted, assuming that phytoextraction would be an appropriate technique. The metal-amended samples were kept incubated for a 30-day period at 80% of the water holding capacity. The samples of the soil material were fertilized as follows: 250, 240, 150, and 100 mg kg^{-1} of N, P, K, and S, respectively, added as NH_4SO_2 , $\text{NH}_4\text{H}_2\text{PO}_4$, and KNO_3 ; micronutrients Fe ($\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$), Mn ($\text{MnCl}_2 \cdot 4\text{H}_2\text{O}$), B (H_3BO_3), and Mo ($\text{Na}_2\text{MoO}_4 \cdot 2\text{H}_2\text{O}$) were applied at the concentrations of 2, 4, 1, and 0.2 mg kg^{-1} , respectively. Then, the samples were transferred into plastic pots set as a greenhouse experiment.

The soil samples in the pots were cultivated for 35 days with velvet bean (*Stizolobium aterrimum*), a high biomass green manure species. Velvet bean was chosen because its potential for metal accumulation when grown on a soil contaminated with heavy metals as compared to seven other high biomass producing species (Santos, 2006). This species has been also reported as tolerant to herbicides (Procópio et al., 2005). Plants were watered to 80% of their water holding capacity on a daily basis by weighing the pots and adding water to replace any weight loss. Two LMWOA, citric acid (aliphatic) and gallic acid (aromatic), were applied to the samples at a 5 mmol kg^{-1} rate in three ways: a single application at the 28th day of growing; two applications at the 28th and 31st day; and three successive applications at the 28th, 31st, and 34th growing day. A control pot (contaminated soil with no LMWOA application) and a blank (soil with no metals neither LMWOA) were used to test the effects of LMWOA and metals on the plant dry matter production.

To study the effects of chelants on the solubilization of the soil metals, one soil moisture sampler (Rhizon[®] SMS: Rhizosphere Research Products, Eijkelkamp, The Netherlands) was placed in the center of each pot. Soil solution was sampled by pressure suction in a syringe at the following day each chelant addition. The concentration of metals in the soil solutions were summed up.

Plants were harvested at the 35th day of growing by cutting the shoots on the soil surface and the total shoot dry biomass was determined for each pot. The soil was then broken up and roots were harvested by hand. The roots were abundantly washed in tap water until free of soil and then washed with distilled water and dried in an oven at 70°C. Both shoot and

root dried materials were further ground and digested in a mixture of $\text{HNO}_3:\text{HClO}_4$ (3:1 v/v). The concentrations of Cd, Pb, Cu, and Zn in the extracts were determined by atomic absorption spectrometry. The net removal of metals was calculated by multiplying the concentration of metals in the shoots by the biomass.

Water soluble metals were extracted by CaCl_2 0.01 mol L^{-1} according to Novozamsky et al. (1993). The metal contents in the exchangeable fractions, organic matter, and crystalline iron oxides were determined based on Shuman (1985), whereas the amorphous iron oxides fraction was extracted according to Chao & Zhou (1983). The fractionation method consisted of the following steps: (1) Water soluble (WS) - 5.0 grams of soil were placed in a 50-mL centrifuge tube in which were added 20 mL of CaCl_2 0.01 mol L^{-1} . The mixture was agitated for 2 h and centrifuged for 10 min at $2000 \times g$. The supernatant was filtered and separated. The fractionation continued for the remaining sample in the tube; (2) Exchangeable Fraction (Ex) - 20.0 mL of $1 \text{ mol L}^{-1} \text{ Mg}(\text{NO}_3)_2$ were added to the tube and the set was agitated for two hours, the mixture was then centrifuged for 10 min at $2,000 \times g$ and the supernatant filtered and separated; 20.0 mL of distilled water was added; then another agitation was performed for 3 min followed by a new centrifugation, and the supernatant filtered and separated again. The two supernatant liquids were combined and stored for analysis. The sequential extraction continued for the remaining soil sample; (3) Organic Matter Fraction (OM) - 10.0 mL of NaClO , 5 - 6% at pH 8.5, were added to the Ex-free sample; the system was heated to 100°C for 30 min, and agitated periodically; thereafter, the sample was centrifuged, and the supernatant was filtered. This procedure was repeated twice, and the three separated supernatants were combined. Ten mL of distilled water were added in the centrifuge tube, and after shaking for 3 min, it was centrifuged, filtered, and the separated supernatant was added to the three combined extracts from the NaClO extraction and stored for analysis. The sequential extraction continued for the remaining soil sample; (4) Amorphous Iron Oxide Fraction (AFeOx) - 30.0 mL of $\text{NH}_2\text{OH.HCl}$ (hydroxylamine hydrochloride), 0.1 mol L^{-1} plus HCl 0.25 mol L^{-1} , at pH 3.0, were added to the OM-free soil sample, and the mixture agitated for 30 min, centrifuged and filtered; the separated supernatant was stored for analysis. To continue the sequential extraction, 15 mL of distilled water were added to the centrifuge tube and shaken for 3 min. The supernatant was then discarded; (5) Crystalline Iron Oxide Fraction (CFeOx) - 30.0 mL of $(\text{NH}_4)_2\text{C}_2\text{O}_4$ (ammonium oxalate), $0.2 \text{ mol L}^{-1} + \text{H}_2\text{C}_2\text{O}_4$ (oxalic acid) 0.2

$\text{mol L}^{-1} + \text{ascorbic acid}$ 0.01 mol L^{-1} , at pH 3.0, were added to the AFeOx-free soil sample, the mixture was heated to 100°C for 30 min, agitated periodically, and thereafter was centrifuged and filtered; the separated supernatant was stored for analysis.

The experiment was arranged in a completely randomized design. All the data comprise averages of three replicates. Data were statistically evaluated by the ANOVA and significantly different means assessed by the Scott-Knot test.

RESULTS AND DISCUSSION

Plants produced less biomass on the heavy metal contaminated soil when no acids were added (control pot) as compared to the ones grown on the non-contaminated soil (Table 1). The metal phytotoxicity induced a 79% reduction in the root growth and 86% in the shoots. Of course, such a reduction in biomass production can be a drawback for efficient phytoextraction. Toxicity symptoms such as chlorosis and necrosis were observed during the plants growth in all the pots with contaminated soil, even before chelants application.

Except for gallic acid applied three times, LMWOA did not reduce the plants dry matter production (Table 1). Luo et al. (2005); Tamura et al. (2005) and Evangelou et al. (2006) also found that citric acid application did not affect the shoots dry matter production, which indicates the lower phytotoxicity of this organic acid. Citric acid could be successively applied on field conditions aiming to hasten metal phytoextraction without affecting shoot biomass. As it will be discussed later, three applications of citric acid can be effective in removing metals from soil, especially owing to its low toxicity to shoots. Gallic acid, on the other hand, can be applied up to two times at a 5 mmol kg^{-1} dose without decreasing the shoots dry matter of the plants, but with little effect on metal removal from soils.

The LMWOA increased the heavy metal concentrations in the soil solution (Figure 1), but gallic acid solubilized a much smaller amount of metals than citric acid. Nascimento et al. (2006) also reported that citric acid was more efficient to solubilize Zn and Cu in soil than gallic acid. This was probably due to the lower solubility of gallic acid in soil. However, it should be noted, that the concentrations of Pb and Cu brought into solution by citric acid were lower than Cd and Zn, which probably results from the differences on solubility of such metal compounds in soils. The concentrations of Cd, Zn, and notably Pb and Cu in soil solution increased linearly following the successive applications of citric acid.

Table 1 - Dry matter production of velvet bean roots and shoots after chelants successive applications to a metal multi-contaminated soil material.

Blank	Control	Citric acid			Gallic acid		
		1 Appl.	2 Appl.	3 Appl.	1 Appl.	2 Appl.	3 Appl.
----- g per pot -----							
Roots							
3.41 a	0.69 c	0.66 c	0.61 c	0.67 c	0.81 b	0.81 b	0.57 c
Shoots							
10.47 a	1.42 b	1.37 b	1.50 b	1.48 b	1.56 b	1.50 b	0.99 c

Values followed by the same letter in lines are not different (Scott-Knott com $P < 0.05$).

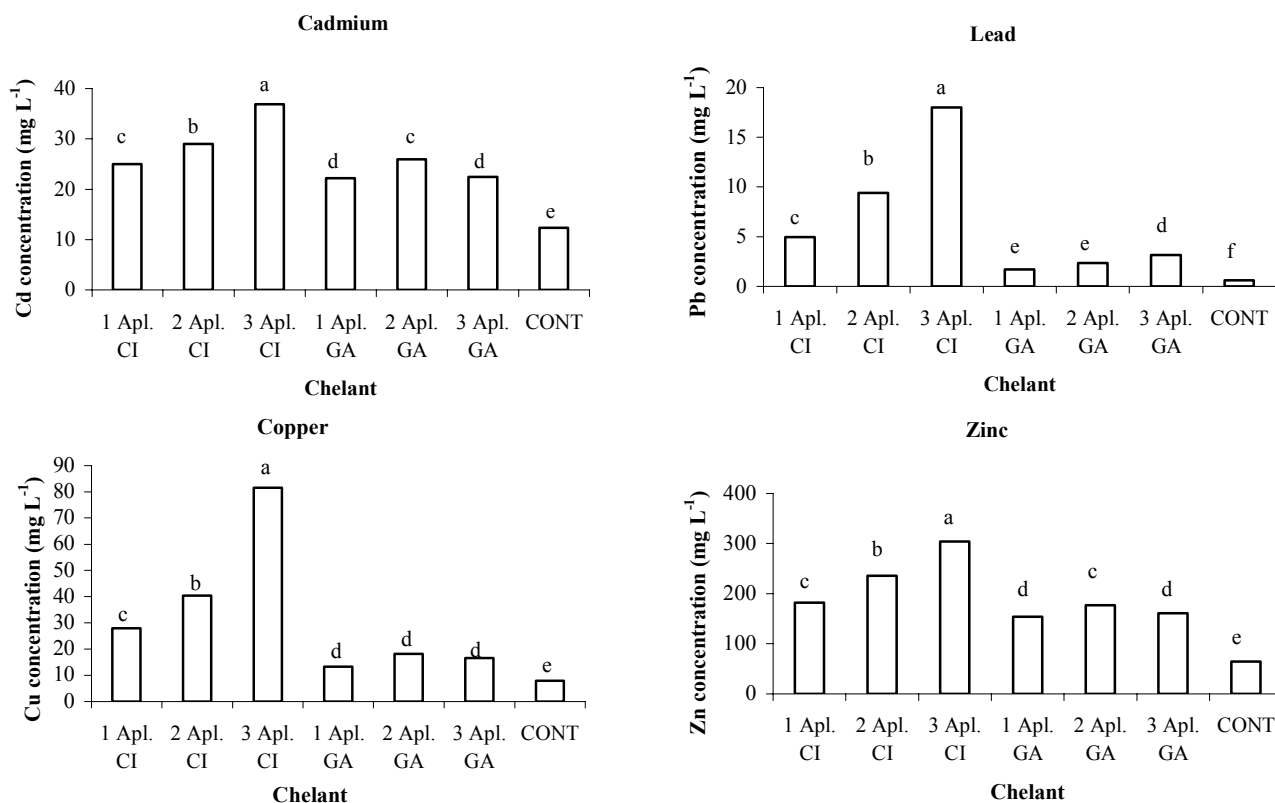


Figure 1 - Concentrations of Cd, Pb, Cu, and Zn in soil solution as a result of three successive applications of citric acid (CI) and gallic acid (GA) to the soil material. Values followed by the same letter are not different ($P < 0.05$).

Treatments with a single application of organic acids resulted in low metal solubility (Figure 1). This is clearly due to the high biodegradability of natural organic acids in soil (Krishnamurti et al., 1997; Ström et al., 2001). The citric acid three-application treatment presented the highest Cd concentration in solution (36.90 mg L^{-1}) whereas one single application of such acid resulted in a soluble Cd concentration 32% lower (25 mg L^{-1}). Successive applications of citric acid can suppress the negative effect of its high degradation rate, increasing the efficiency of chemically-assisted phytoextraction. On the other hand, the highest concentration of Cd in soil solution after gallic acid addition resembled the one obtained by the single citric acid application. This dem-

onstrates the low ability of gallic acid to solubilize Cd in soils.

The Pb concentration in soil solution increased 97 and 81% after three applications of the citric and gallic acid, respectively (Figure 1). For the citric acid treatments, the largest Pb concentration was 18 mg L^{-1} , and the smallest 5 mg L^{-1} , representing a 72% reduction in six days after the chelant application. Such decrease in Pb concentration was lower for the gallic acid (46%), i.e., from 3.15 mg L^{-1} to 1.7 mg L^{-1} in the same stage. This demonstrates both the lower ability to solubilize Pb and the slower biodegradability of gallic acid as compared to citric acid. In spite of the high percentage increase, neither citric nor gallic acid seem to be able to solubilize Pb at suitable levels for effi-

cient phytoextraction, even when applied three successive times. Tamura et al. (2005), Nascimento (2006) and Evangelou et al. (2006) demonstrated the low ability of LMWOA to bring less soluble metals (such as Pb) into soil solution. This is a serious drawback for phytoextraction with these chelants in Pb contaminated soils. So far only synthetic chelants such as EDTA (Blaylock et al., 1997), EDDS (Kos & Lestan, 2004) and MGDA (Tamura et al., 2005) are effective for lead phytoextraction.

The single, double and triple application of gallic acid did not differ regarding Cu concentration in soil solution (Figure 1), although they get a 50% increase in the metal solubility as compared to control. On the other hand, the citric acid three-time application was significantly more efficient in solubilizing Cu (81.53 mg L⁻¹). Accordingly, high Cu mobility in soil treated with organic acids, especially citric acid, was observed (Evangelou et al., 2006). In such a case, the Cu concentration in the soil solution increased 90% as compared to control. The Cu concentration between the first and third application decreased 66%, indicating the high citric acid degradability. The lowest value of Zn concentration in soil solution after citric acid application was 182 mg L⁻¹; after three applications, this figure reached 304 mg L⁻¹. Similarly to Cd, the lowest concentration of Zn in citric acid treatments was equivalent to the highest one obtained by gallic acid application. Once more this demonstrates the low potential of gallic acid when used in phytoextraction, even for fairly soluble metals.

LMWOA application to soil did not promote an increase in the velvet shoots heavy metal concentrations (Table 2). Similar results were reported to other species by Chen et al. (2003), Turgut et al. (2004),

Luo et al. (2005) and Meers et al. (2005). It can be explained as an effect of the fast mineralization of soil organic acids (Römken et al., 2002). Synthetic chelators applied to soil at 5 mmol kg⁻¹ can solubilize much higher amounts of metals than LMWOA applied at 20 mmol kg⁻¹ rates (Nascimento, 2006). The low root biomass production resulting from metal toxicity (Table 1) limited the efficiency of metal extraction by velvet bean, since there was a substantial increase in the metals availability via LMWOA addition (Figure 1). It also suggests that using tolerant or hyperaccumulator species, associated with successive applications of LMWOA, could be a viable approach. Plant's tolerance to metal stress is certainly a key factor to successful phytoextraction and hampers the feasibility of this technique.

Root metal concentration increase through organic acids application was observed only to Pb (Table 2). This is probably a consequence of the naturally low mobility of such metal in soil. However, the low affinity ($\log K_s = 6.5$) of Pb for citric acid (Römken et al., 2002) suggests their low ability to substantially increase the Pb translocation to shoots. Actually, only synthetic chelants seem to be capable of hastening the Pb transport to shoots up to levels proper to phytoextraction (Blaylock et al., 1997; Tamura et al., 2005).

The phytoextraction efficiency can be evaluated by the net removal of metals from soils rather than by the concentration of metals in shoots. As it can be seen in Figure 2 LMWOA increased the removal of Pb, Cu and Zn from soil, except for a single application of citric acid for Zn. The amount of Cd removed by plants upon LMWOA addition was not higher than in control. This can be related to the soil high avail-

Table 2 - Metals concentration in velvet bean shoots and roots, according to three different application times (one, two, and three applications).

Metal	Control	Citric acid			Gallic acid		
		1 Appl.	2 Appl.	3 Appl.	1 Appl.	2 Appl.	3 Appl.
----- $\mu\text{g kg}^{-1}$ -----							
Shoots							
Cd	82.0 a	80.7 a	72.7 a	80.6 a	72.3 a	86.3 a	83.0 a
Pb	11.5 a	14.9 a	12.1 a	13.9 a	10.2 a	11.3 a	7.8 a
Cu	52.2 a	52.6 a	49.4 a	51.0 a	47.5 a	51.1 a	50.1 a
Zn	670.4 a	695.6 a	701.5 a	715.2 a	704.7 a	772.3 a	823.7 a
Roots							
Cd	717.9 a	709.5 a	674.9 a	666.4 a	632.8 a	660.8 a	677.9 a
Pb	975.3 c	1,650.6 a	1,294.4 b	1,698.1 a	1,393.5 b	1,478.4 a	1,735.9 a
Cu	4,287.8 a	5,049.5 a	4,287.1 a	4,616.1 a	4,451.7 a	4,225.4 a	4,648.7 a
Zn	3,686.5 a	3,742.9 a	3,514.2 a	3,669.1 a	3,565.9 a	3,818.9 a	3,974.0 a

Values followed by the same letter in rows are not different (Scott-Knott com $P < 0.05$).

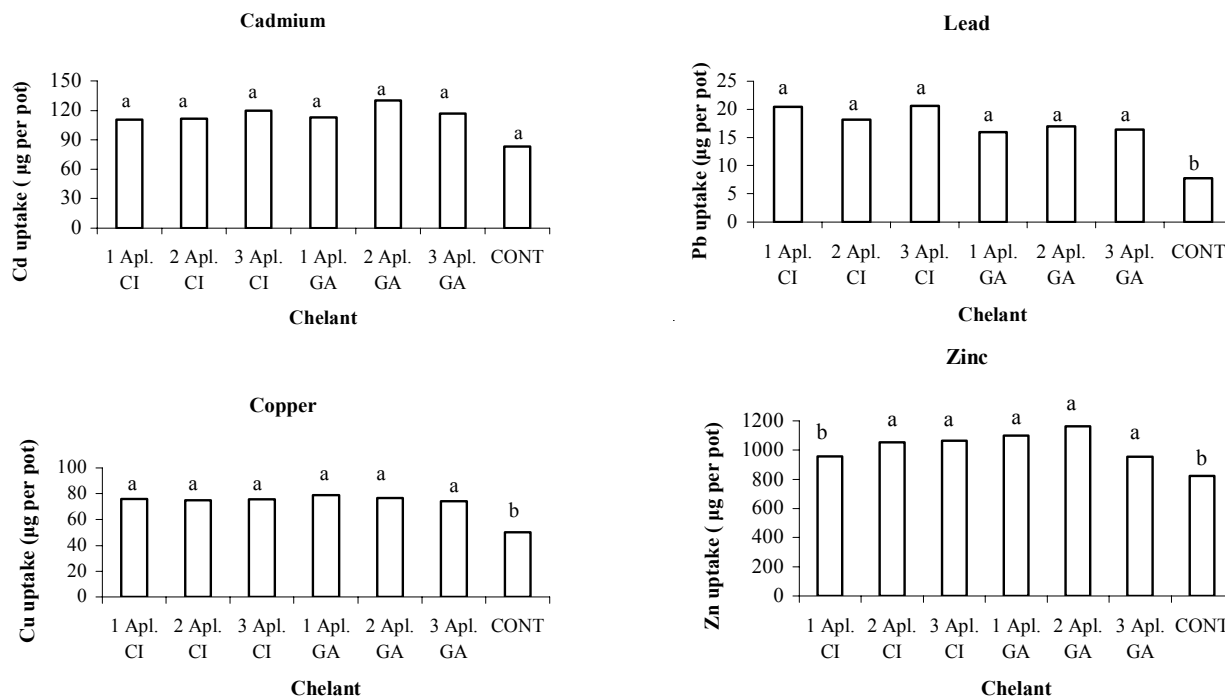


Figure 2 - Removal of Cd, Pb, Cu, and Zn in velvet bean shoots ($\mu\text{g per pot}$) grown in soil material treated with gallic (GA) and citric (CI) acids applied in three forms (1, 2, and 3 applications) at a 5 mmol kg^{-1} rate. Values followed by the same letter are not different ($P < 0.05$).

ability of Cd. Gallic and citric acid are not able to increase the phytoextraction of Pb from soil (Nascimento et al., 2006). For instance, citric acid had no effect on Pb uptake by common buckwheat even when the soil was treated with a high concentration (20 mmol kg^{-1}) of this chelant (Tamura et al., 2005). These results corroborate the low potential of LMWOA for Pb phytoextraction.

Cadmium, Pb, Cu, and Zn water soluble contents increased accordingly to the successive organic acids applications to soil (Table 3). Contents of Cd, Pb, and Zn were highest for the three-application treatment of citric acid. The low content of Cd in iron oxides indicates this metal low affinity for sites of specific adsorption and demonstrates the relatively high solubility of Cd in soils. The exchangeable concentration of metals was also increased owing to LMWOA addition.

Organic matter fraction retained most of the Cu and Zn in soil (Table 3). The concentration of organic matter bounded Cu and Zn was reduced as a result of organic acids application. In the more bioavailable fractions (WS and Ex) Zn seemed to be incremented by remobilization from organic matter bounded Zn, which substantially diminished by LMWOA. On the other hand, lead and copper contents in organic matter were almost unaffected by acids addition. This is probably due to the organic matter affinity of Pb and Cu which is higher than

Zn making the Pb- and Cu-complexes to be more stable.

The LMWOA caused a mobilization of Pb and Cu from oxide fractions in soil (Table 3). LMWOA induced metal dissolution in crystalline and/or poorly crystallized soil minerals. Carboxylic groups of LMWOA can dissociate over a wide range of pH, weathering primary minerals and dissolving then by complexation (Sposito, 1989). This is particularly important for poorly available soil metals since they depend on solubilization from stable pools in order to be taken up by plants. However, LMWOA metal mobilization from iron oxides, as observed here, is less impressive when compared to the ability of synthetic chelating agents, such as EDTA (Barona et al., 2001). Thus, if the targets for phytoextraction are metals tightly bound to oxides and to soil residual fraction, the LMWOA tested here do not seem to be a good bet to improve phytoextraction.

CONCLUSIONS

Multiple applications of citric acid to soil had a positive effect on the mobilization of Cd, Pb, Cu, and Zn. However, neither citric acid nor gallic acid was efficient to induce metals phytoextraction. In general, LMWOA application increased the metal concentrations in the water soluble and exchangeable fractions. Zinc and Cu were retained mostly in the

Table 3 - Metal concentration in five different soil fractions: water soluble (WS), exchangeable (Ex), organic matter (OM), amorphous iron oxides (AFeOx) and crystalline iron oxides (CFeOx) after addition of chelants to soil (one, two and three applications).

	Control	Citric acid			Gallic acid		
		1 Appl.	2 Appl.	3 Appl.	1 Appl.	2 Appl.	3 Appl.
Cadmium (mg dm ⁻³)							
WS	6.4 d	6.8 c	6.9 c	7.6 a	7.2 b	7.3 b	6.9 c
Ex	1.4 b	1.6 a	1.6 a	1.6 a	1.6 a	1.6 a	1.4 b
OM	0.6 a	0.5 b	0.5 b	0.4 c	0.5 b	0.5 b	0.5 b
AFeOx	< DL	< DL	< DL	< DL	< DL	< DL	< DL
CFeOx	< DL	< DL	< DL	< DL	< DL	< DL	< DL
Lead (mg dm ⁻³)							
WS	2.4 d	5.1 b	4.5 b	5.6 a	4.0 c	4.0 c	4.5 b
Ex	0.5 b	1.1 a	1.0 a	1.1 a	1.0 a	1.0 a	1.1 a
OM	1.3 d	2.0 a	1.4 c	1.6 b	1.4 c	1.4 c	1.3 d
AFeOx	1.9 a	1.6 b	1.4 b	1.5 b	1.8 a	1.5 b	1.4 b
CFeOx	2.5 a	1.9 c	1.8 c	1.9 c	2.0 c	2.2 b	2.2 b
Copper (mg dm ⁻³)							
WS	2.1 d	3.0 b	2.7 c	3.2 a	2.6 b	3.3 a	3.3 a
Ex	1.4 c	1.7 a	1.4 c	1.5 b	1.3 c	1.5 b	1.7 a
OM	4.3 a	4.1 a	4.4 a	3.8 a	3.9 a	4.3 a	4.4 a
AFeOx	1.7 a	1.4 c	1.4 c	1.2 d	1.5 b	1.3 d	1.4 c
CFeOx	1.4 a	1.2 c	1.4 a	1.3 b	1.3 b	1.2 c	1.1 d
Zinc (mg dm ⁻³)							
WS	13.5 d	14.0 c	13.0 d	17.6 a	15.9 b	16.2 b	14.7 c
Ex	7.7 d	8.4 c	8.9 c	9.7 b	7.7 d	10.7 a	8.4 c
OM	65.2 a	52.3 c	54.37 b	40.99 e	54.79 b	52.29 c	49.77 d
AFeOx	0.1 a	0.1 a	0.1 a	0.1 a	0.1 a	0.1 a	0.1 a
CFeOx	0.3 a	0.3 a	0.2 b	0.2 b	0.3 a	0.3 a	0.3 a

Values followed by the same letter in each row are not different (Scott-Knott $P < 0.05$). DL - Below detection limit.

organic matter fraction, from where Zn was remobilized through LMWOA application into the more available water soluble and exchangeable fractions. LMWOA mobilized Pb and Cu from iron oxides, but such an increase in solubility was not high enough to affect the phytoextraction. So far the LMWOA tested are not suitable to enhance phytoextraction, even when successively applied, suggesting that more detailed information about a natural LMWOA able to replace synthetic chelants that induce metal phytoextraction is required. Studies on amounts and time of application of LMWOA are also important. Therefore, research must continue searching for natural LMWOA to replace synthetic chelants for metal phytoextraction. More studies on doses and application timing of LMWOA are also needed to make these acids a viable alternative to synthetic chelants.

REFERENCES

- BARONA, A.; ARANGUIZ, I.; ELÍAS, A. Metal associations in soils before and after extractive decontamination: implications for the effectiveness of further clean-up procedures. *Environmental Pollution*, v.113, p.79-85, 2001.
- BLAYLOCK, M.J.; SALT, D.E.; DUSHENKOV, S.; ZAKHAROVA, O.; GUSSMAN, C.; KAPULNIK, Y.; ENSLEY, B.D.; RASKIN, I. Enhanced accumulation of Pb in Indian mustard by soil applied chelating agents. *Environmental Science and Technology*, v.31, p.860-865, 1997.
- CHAO, T.T.; ZHOU, L. Extraction techniques for selective dissolution of amorphous iron oxides from soils and sediments. *Soil Science Society of America Journal*, v.47, p.224-232, 1983.
- CHEN, Y.X.; LIN, Q.; LUO, Y.M.; HE, Y.F.; ZHEN, S.J.; YU, Y.L.; TIAN, G.M.; WONG, M.H. The role of citric acid on the phytoremediation of heavy metal contaminated soil. *Chemosphere*, v.50, p.807-811, 2003.
- COOPER, E.M.; SIMS, J.T.; CUNNINGHAM, S.D.; HUANG, J.W.; BERTI, W.R. Chelate-assisted phytoextraction of lead from contaminated soils. *Journal of Environmental Quality*, v.28, p.1709-1719, 1999.

- CUNNINGHAM, S.D.; BERTI, W.R. **Phytoextraction and phytoestabilization: technical, economic, and regulatory considerations of soil-lead issue**. In: TERRY, N.; BAÑUELOS, G. (Ed.) Phytoremediation of contaminated soil and water. Boca Raton: Lewis, 2000. p.359-376
- EVANGELOU, M.W.H.; EBEL, M.; SHAEFFER, A. Evaluation of the effect of small organic acids on phytoextraction of Cu and Pb from soil with tobacco *Nicotiana tabacum*. **Chemosphere**, v.63, p.996-1004, 2006.
- HUANG, J.W.; CHEN, J.; BERTI, W.R.; CUNNINGHAM, S.D. Phytoremediation of lead-contaminated soils: role of synthetic chelates in lead phytoextraction. **Environmental Science and Technology**, v.31, p.800-805, 1997.
- KOS, B.; LESTAN, D. Chelator induced phytoextraction and in situ soil washing of Cu. **Environmental Pollution**, v.132, p.333-339, 2004.
- KRISHNAMURTI, G.S.R.; CIESLINSKI, G.; HUANG, P.M.; VAN PEES, K.C.J. Kinetics of cadmium release from soils as influenced by organic acids: implication in cadmium availability. **Journal of Environmental Quality**, v.26, p.271-277, 1997.
- LOMBI, E.; ZHAO, F.J.; DUNHAM, S.J.; McGRATH, S.P. Phytoremediation of heavy-metal contaminated soils: natural hyperaccumulation versus chemically enhanced phytoextraction. **Journal of Environmental Quality**, v.30, p.1919-1926, 2001.
- LUO, C.; SHEN, Z.; LI, X. Enhanced phytoextraction of Cu, Pb, Zn and Cd with EDTA and EDDS. **Chemosphere**, v.59, p.1-11, 2005.
- MEERS, E.; RUTTENS, A.; HOPGOOD, M.J.; SAMSON, D.; TACK, F.M.G. Comparison of EDTA and EDDS as potential soil amendments for enhanced phytoextraction of heavy metals. **Chemosphere**, v.58, p.1011-1022, 2005.
- MELO, E.E.C.; NASCIMENTO, C.W.A.; SANTOS, A.C.Q. Solubilidade, fracionamento e fitoextração de metais pesados após aplicação de agentes quelantes. **Revista Brasileira de Ciência do Solo**, v.30, p.1051-1060, 2006.
- NASCIMENTO, C.W.A. Organic acids effects on desorption of heavy metals from a contaminated soil. **Scientia Agricola**, v.63, p.276-280, 2006.
- NASCIMENTO, C.W.A.; AMARASIRIWARDENA, D.; XING, B. Comparison of natural organic acids and synthetic chelates at enhancing phytoextraction of metals from a multi-metal contaminated soil. **Environmental Pollution**, v.140, p.114-123, 2006.
- NASCIMENTO, C.W.A.; XING, B. Phytoextraction: a review on enhanced metal availability and plant accumulation. **Scientia Agricola**, v.63, p.299-311, 2006.
- NOVOZAMSKY, I.; LEXMOND, T.M.; HOUBA, V.J.G. A single extraction procedure of soil for evaluation of uptake of some heavy metals by plants. **International Journal of Environmental and Analytical Chemistry**, v.51, p.47-58, 1993.
- PROCÓPIO, S.O.; SANTOS, J.B.; PIRES, F.R.; SILVA, A.A.; SANTOS, E.A.; FERREIRA, L.R. Fitorremediação de solos contaminados com tebutiuron utilizando-se espécies cultivadas para adubação verde. **Planta Daninha**, v.23, p.719-724, 2005.
- RASKIN, I.; SMITH, R.D.; SALT, D.E. Phytoremediation of metals: using plants to remove pollutants from the environment. **Current Opinion in Biotechnology**, v.8, p.221-226, 1997.
- RÖMKENS, P.; BOUWMAN, L.; JAPENGA, J.; DRAAIMA, C. Potentials and drawbacks of chelate-enhanced phytoremediation of soils. **Environmental Pollution**, v.116, p.109-121, 2002.
- SANTOS, A.C.Q. **Metais pesados em solos: seleção de espécies acumuladoras, competição entre elementos e uso de silício na fitoextração induzida**. Recife: UFRPE, 2006. 62p. Dissertação (Mestrado).
- SHEN, Z.G.; LI, X.D.; WANG, C.C.; CHEN, H.M.; CHUA, H. Lead phytoextraction from contaminated soil with high-biomass plant species. **Journal of Environmental Quality**, v.31, p.1893-1900, 2002.
- SHUMAN, L.M. Fractionation method for soil microelements. **Soil Science**, v.140, p.11-22, 1985.
- SPOSITO, G. The chemistry of soils. New York: Oxford University Press, 1989. 277 p.
- STRÖM, L.; OWEN, A.G.; GODBOLD, D.L.; JONES, D.L. Organic acid behaviour in a calcareous soil: sorption reactions and biodegradation rates. **Soil Biology and Biochemistry**, v.33, p.2125-2133, 2001.
- TAMURA, H.; HONDA, M.; SATO, T.; KAMACHI, H. Pb hyperaccumulation and tolerance in common buckwheat (*Fagopyrum esculentum* Moench). **Journal of Plant Research**, v.118, p.355-359, 2005.
- TERRY, N.; BAÑUELOS, G. **Phytoremediation of contaminated soil and water**. Boca Raton: Lewis, 2000. p.189-200.
- TURGUT, C.; PEPE, M.K.; CUTRIGHT, T.J. The effect of EDTA and citric acid on phytoremediation of Cd, Cr, and Ni from soil using *Helianthus annuus*. **Environmental Pollution**, v.131, p.147-154, 2004.
- WU, J.; HSU, F.C.; CUNNINGHAM, S.D. Chelate-assisted Pb phytoremediation: Pb availability, uptake, and translocation constraints. **Environmental Science and Technology**, v.33, p.1898-1904, 1999.

Received September 13, 2006

Accepted September 26, 2007