

Rev. Inst. Med. Trop. Sao Paulo  
55(6):377-383, November-December, 2013  
doi: 10.1590/S0036-46652013000600002

## DISTRIBUTION OF DERMATOPHYTES FROM SOILS OF URBAN AND RURAL AREAS OF CITIES OF PARAIBA STATE, BRAZIL

Zélia Braz Vieira da Silva PONTES(1), Aurylene Carlos de OLIVEIRA(1), Felipe Queiroga Sarmiento GUERRA(1), Luiz Renato de Araújo PONTES(2) & Jozemar Pereira dos SANTOS(3)

### SUMMARY

The dermatophytes, keratinophilic fungi, represent important microorganisms of the soil microbiota, where there are cosmopolitan species and others with restricted geographic distribution. The aim of this study was to broaden the knowledge about the presence of dermatophytes in soils of urban (empty lots, schools, slums, squares, beaches and homes) and rural areas and about the evolution of their prevalence in soils of varying pH in cities of the four mesoregions of Paraíba State, Brazil. Soil samples were collected from 31 cities of Paraíba State. Of 212 samples, 62% showed fungal growth, particularly those from the Mata Paraibana mesoregion (43.5%), which has a tropical climate, hot and humid. Soil pH varied from 4.65 to 9.06, with 71% of the growth of dermatophytes occurring at alkaline pH (7.02 - 9.06) ( $p = 0.000$ ). Of 131 strains isolated, 57.3% were geophilic species, particularly *Trichophyton terrestre* (31.3%) and *Mycrosporium gypseum* (21.4%). *M. nanum* and *T. ajelloi* were isolated for the first time in Paraíba State. The zoophilic species identified were *T. mentagrophytes* var. *mentagrophytes* (31.3 %) and *T. verrucosum* (7.6 %), and *T. tonsurans* was isolated as an anthropophilic species. The soils of urban areas including empty lots, schools, slums and squares of cities in the mesoregions of Paraíba State were found to be the most suitable reservoirs for almost all dermatophytes; their growth may have been influenced by environmental factors, soils with residues of human and/or animal keratin and alkaline pH.

**KEYWORDS:** Dermatophytes; Keratinophilic fungi; Soil; pH conditions; Brazil.

### INTRODUCTION

The dermatophytes (*Trichophyton*, *Microsporium* and *Epidermophyton*), keratinophilic fungi, represent important microorganisms of the soil microbiota, where there are cosmopolitan species and others with restricted geographic distribution<sup>1,2,6,10,17,21</sup>. There have been reports of the isolation of *T. ajelloi*, *T. rubrum*, *T. mentagrophytes*, *T. verrucosum*, *T. terrestre*, *T. tonsurans*, *T. simii*, *T. schoenleinii*, *M. gypseum*, *M. canis*, *M. audouinii*, *M. nanum*, *M. cookei* and/or *E. floccosum*, from the soils of various Brazilian states and locals around the world<sup>8,20,24,25,30,32,34</sup>.

The occurrence of fungi in the soil can also be influenced by non-biological factors such as soil temperature, humidity, rainfall, environmental light, climate, chemical composition, quantity of organic matter in the soil and pH. Some have a wide range of tolerance for acidic to alkaline soils<sup>2,7,14,16</sup>. However, studies of soil pH in relation to occurrence of dermatophytes are uncommon in Brazil.

The study of the diversity of dermatophytes in the soil is important because changes in the distribution of species of dermatophytes due to ecological factors, socio-economic, therapeutic, and migration processes

of livestock populations, reflect the epidemiology of dermatophytosis, which are one of the source infections of the soil<sup>2,3,16,18,31</sup>. Thus, the aim of this study was to broaden the study into the presence of dermatophytes from soils of urban and rural areas of cities of four mesoregions of Paraíba State and the influence of pH on fungi growth.

### MATERIALS AND METHODS

The state of Paraíba is situated in the eastern portion of Northeast Brazil, with coordinates between 6° and 8° S and between 34° and 38° W; therefore, it is included in the tropical zone. It comprises an area of 56,372 km<sup>2</sup> and is divided into four mesoregions (Mata Paraibana, Borborema, Agreste Paraibano and Sertão Paraibano) and into 23 geographic microregions, including a total of 223 cities. In the Mata Paraibana, the predominant climate is warm, humid tropical (As') with an average annual rainfall of 1,800 mm, temperature of 26 °C and relative humidity of 80%. The soils are sandy and muddy, which are influenced by sea water and have especially coastal vegetation of mangrove swamp, rainforest and cerrado. In Borborema, the predominant climate is semi-arid (Bsh), warm and dry with average annual rainfall of 500 mm, temperature of 26 °C and relative humidity of 75%. The soils are shallow stony soil with caatinga

(1) Laboratory of Mycology, Department of Pharmaceutical Sciences, Federal University of Paraíba, João Pessoa, PB, Brazil.

(2) Laboratory of Ceramic, Department of Mechanical Engineering, Federal University of Paraíba, João Pessoa, PB, Brazil.

(3) Department of Statistic, Federal University of Paraíba, João Pessoa, PB, Brazil.

**Correspondence to:** Felipe Queiroga Sarmiento Guerra, Tel.: 55.83.9602-1666. E-mail: felipeqsguerra@gmail.com

vegetation. The climate Bsh, together with As' are observed in Agreste Paraibano. However, in Sertão Paraibano, the predominant climate is semi-humid (Aw') with an average annual rainfall of 800 mm, temperature of 27 °C and relative humidity of 70%. In the two last mesoregions, a slow development of soils with caatinga vegetation (Fig. 1)<sup>28</sup>.

An ecological study was performed with a total of 212 soil samples. The sampling was non-probabilistic, as it was done by convenience and accessibility to the members of the team, taking into consideration conglomerates of cities in Paraíba mesoregions. Each mesoregion was represented by a city of great geographical and population density: João Pessoa for Mata Paraibana, Monteiro for Borborema, Campina Grande for Agreste Paraibano and Patos for Sertão Paraibano. The other cities were randomly included.

Soil samples were selected from urban (empty lots, schools, slums, squares, homes and beaches) and rural areas of cities. The sampling sites were selected on the basis of the likely presence of soil with keratin residues from humans and animals.

The collection, processing and pH of soil solutions were according to the techniques described by VANBREUSEGHEM<sup>33</sup>. Approximately 100g of soil at a depth of three to five centimeters was collected, placed in polyethylene bags and brought to be processed at the Laboratory of Mycology in the Department of Pharmaceutic Sciences and Laboratory of Ceramic, Department of Mechanical Engineering at the Federal University of Paraíba.

Using a pHmeter, the pH of each soil sample (20 g) was measured

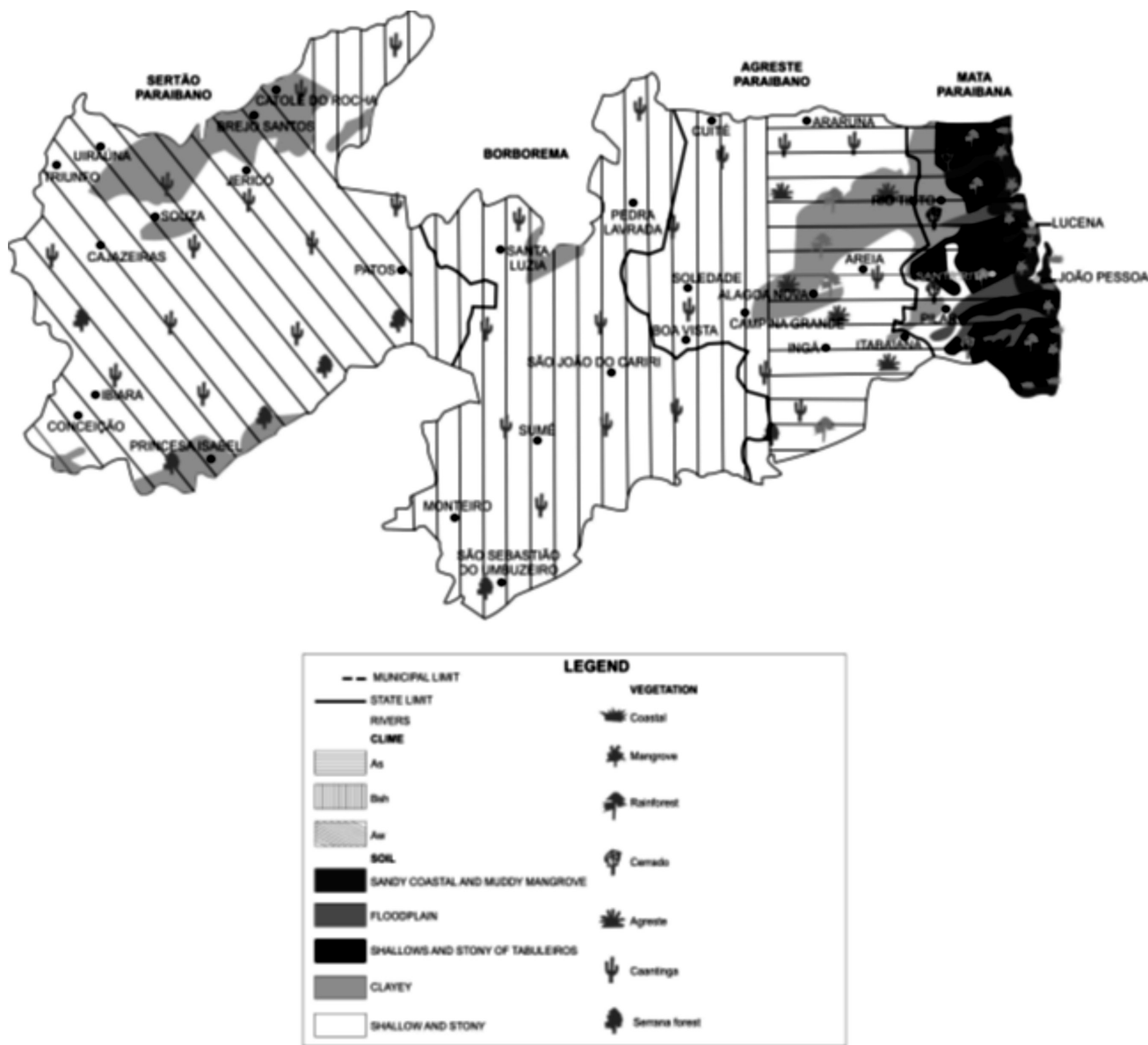


Fig. 1 - Location of 31 cities, according to four mesoregions, soils type, vegetation and climate of the state of Paraíba, Brazil. Adapted from RODRIGUEZ<sup>28</sup>.

after dilution in distilled sterile water (20 mL) with 20 minutes of agitation and decantation. Each sample was distributed in sterile Petri plates, moistened with sterile water (20 mL) and some sterile human hair strips were placed over each surface. The plates were identified and incubated (27-30 °C) and from the 5<sup>th</sup> to the 70<sup>th</sup> day the hair strips were regularly observed with magnifying glasses for signs of fungal growth. Hair strips with a development of prominent fungal growth around them, were placed between slide and cover slid, colored in lactophenol blue cotton and examined in a microscope (10X and 40X). They were cultivated in Sabouraud dextrose agar<sup>®</sup> medium with chloramphenicol (0.05 mg mL<sup>-1</sup>) and in Mycobiotic agar<sup>®</sup> and incubated at room temperature for another minimum period of two weeks.

The identification of the species was based on macromorphology and micromorphology features (slide-culturing) and physiological tests (urea hydrolysis, *in vitro* hair perforation, vitamin requirement and sensitive media). The classification was based on BARNETT & HUNTER<sup>5</sup>, REBELL & TAPLIN<sup>27</sup> and HOOG *et al.*<sup>12</sup>.

The data were subjected to statistical analysis, which consisted of the Binomial test. The process was carried out by computing SPSS 13<sup>22</sup>, allowing to verify if the dermatophytes growth soil acidic pH is equal to alkaline pH.

## RESULTS

In 31 cities of four mesoregions of the state of Paraíba (Fig. 1), 62% of the growth of dermatophytes occurred in soil with different pH. In cities from Mata Paraibana, isolations were observed in 43.5% of samples, where this rate was 84% in the capital, João Pessoa. In cities from Sertão Paraibano, the isolation rate was 20.6%, whereas 23.7% in cities from Agreste Paraibano and 12.2% in cities from Borborema (Table 1).

A total of 131 strains of dermatophytes were isolated, where 57.3% of the geophilic species were identified. *T. terrestre* (31.3%) was the most common species, followed by *M. gypseum* (21.4%), *M. nanum* (3%), *T. ajelloi* (0.8%) and *Anthroderma gypsea* (0.8%), a teleomorph form of *M. gypseum*, observed in sample soil. *M. nanum* and *T. ajelloi* were isolated for the first time in Paraíba State. The zoophilic species identified included *T. mentagrophytes* var. *mentagrophytes* (31.3%) and *T. verrucosum* (7.6%). *T. tonsurans* (3.8%) was the only anthropophilic species isolated. The growth of more than one fungal species was observed in 13 samples (Table 1).

The soils that showed the highest rates of dermatophytes were those of urban areas (95%), especially in soils of empty lots (25.2% of isolations), around schools (22.9%), in slums (21.4%) and squares (19.8%), compared to around homes (3.8%) and on beaches (2.3%) (Table 2).

Dermatophytes developed in a wide pH range: acid to alkaline (4.65 - 9.06), with 71% in alkaline pH (7.02 - 9.06). *T. terrestre* develops within the pH range of 5.76 - 8.90. *T. mentagrophytes* var. *mentagrophytes* and *M. gypseum* develop within the pH range 4.65 - 9.06 and 5.77 - 8.31, respectively and *T. verrucosum* was reported from urban areas at pH 6.65 - 8.05. In acid pH soil, an inhibition of growth *M. nanum*, *A. gypsea* and *T. ajelloi* was observed. The dermatophytes growth in soil of alkaline pH was significantly different from the acidic pH ( $p = 0.000$ ) (Table 3).

## DISCUSSION

Studies worldwide have examined various variables, such as soil type, pH, climate, temperature, moisture and organic matter content, and have revealed the presence of dermatophytes and other keratinophilic fungi in soil<sup>1,3,6,9,14,21,31</sup>. In Brazil, there are few reports on the isolation of dermatophytes in soil, specifically in the Northeast region<sup>16,26,32</sup>. In the mesoregion of Mata Paraibana, with an As' climate and sandy and muddy soils<sup>28</sup>, dermatophytes were isolated in 43.5% of samples. A previous study reported that 55.7% of 68 soil samples from the city of João Pessoa-Paraíba State (PB), showed the growth of dermatophytes<sup>26</sup>. In Borborema, the isolation rate was 12.2%. This area has a Bsh climate and shallow rocky soil. In other mesoregions, the lack of water for prolonged periods accounts for the slow development of soil. The distribution of climates is related to the geographic localization, that is, the closer to the coast the more humid and the farther from the coast the drier. The four mesoregions of Paraíba have predominantly caatinga vegetation, except Mata Paraibana<sup>28</sup>. Although the roles of fungi in ecosystems have been well documented, knowledge about their population dynamics and community structure and of the diversity of soil fungi is still poor. Further studies of Paraíba soils are necessary to analyze the changes and influence of variables such as types of climate, soil and vegetation on the development of dermatophytes.

The pH range of 7.2 - 8.0 is favorable for the production of proteolytic enzymes (keratinases) by keratinophilic fungi, which are necessary for their growth, along with other soil conditions<sup>15</sup>. However, the results of this study indicate the growth of dermatophytes in acid and alkaline pH, where 71% of isolations were observed in the alkaline pH range between 7.02 and 9.06 ( $p = 0.000$ ). These results, obtained with different soil samples, confirm the importance of pH in the habitat to the occurrence and distribution of dermatophytes. In acidic soils, there is growth inhibition of dermatophytes and other keratinophilic fungi, but soils that are weakly acidic to neutral or alkaline are optimal for their growth<sup>14,16,21,23</sup>. In this investigation, in acid pH soils, the growth of *A. gypseum*, *M. nanum* and *T. ajelloi* was inhibited. Some authors<sup>6</sup> observed that the frequency of *T. ajelloi* (33%) increased with a decrease in pH, reaching a maximum in strongly acidic soil.

Eight species of dermatophytes were identified in the soils of cities in Paraíba. Of the geophilic species (57.3%), *T. terrestre* (31.3%) was especially found in soils from squares, empty lots, schools, slums and beaches. This variable distribution rate can be related to the sampling sites, where the presence of people and animals are frequent, providing residues of organic matter, which are essential for the growth of these fungi. The results obtained are close to those for other cities in Brazil such as: Belo Horizonte and São Paulo<sup>29</sup> and in soils of countries such as Germany and Argentina<sup>7,21</sup>. However, the frequency of this species was low in Italy<sup>25</sup> and India<sup>31</sup>. *T. terrestre* has been found to be a pathogen particularly in pets and humans including the elderly who exhibit complications related to immunological factors<sup>25</sup>.

Other geophilic species that were isolated included *M. gypseum* (21.4%), *M. nanum* (3%), *T. ajelloi* (0.8%) and *A. gypsea* (0.8%) at alkaline pH, except *M. gypseum*, which also showed growth at acid pH. Similar results were obtained in soils from the Brazilian states of Rio de Janeiro (31%)<sup>10</sup>, São Paulo (30%)<sup>29</sup> and Bahia (28.8%)<sup>32</sup>. However, in Recife, Pernambuco State, 5.6% isolation was observed for this species

**Table 1**  
Dermatophytes isolated from urban and rural soil samples from 31 cities in four mesoregions of Paraíba State

Mesoregions	Cities	Soil* n	Dermatophytes**								Total n
			<i>A.g</i> n	<i>M.g</i> n	<i>M.n</i> n	<i>T.a</i> n	<i>T.m</i> n	<i>T.te</i> n	<i>T.t</i> n	<i>T.v</i> n	
Mata Paraibana	João Pessoa	68	1	10	-	-	18	12	2	5	48
	Lucena	4	-	-	-	-	1	1	-	-	2
	Pilar	5	-	1	-	-	1	1	-	-	3
	Rio Tinto	5	-	-	-	-	1	-	-	-	1
	Santa Rita	3	-	1	-	-	-	1	-	1	3
<b>Subtotal</b>		85	1	12	-	-	20	15	3	6	57
Agreste Paraibano	Alagoa Nova	7	-	1	-	-	-	-	-	1	2
	Araruna	4	-	1	1	-	-	3	-	-	5
	Areia	3	-	-	-	-	-	1	-	-	1
	Boa vista	4	-	1	-	-	3	-	-	-	4
	C. Grande	6	-	1	-	-	-	-	-	-	1
	Cuité	4	-	2	-	-	2	-	-	1	5
	Ingá	4	-	-	-	-	-	2	-	-	2
	Itabaiana	8	-	-	1	-	-	2	-	1	4
Soledade	3	-	1	-	-	-	2	-	-	3	
<b>Subtotal</b>		43	-	7	2	-	5	10	-	3	27
Borborema	Monteiro	4	-	-	-	-	1	-	-	-	1
	Pedra Lavrada	2	-	-	-	-	-	2	-	-	2
	São João Cariri	6	-	-	-	-	3	-	-	1	4
	S. S.Umbuzeiro	3	-	-	-	-	-	2	-	-	2
	Santa Luzia	4	-	-	1	-	1	1	-	-	3
	Sumé	12	-	-	-	1	2	1	-	-	4
<b>Subtotal</b>		31	-	-	1	1	7	6	-	1	16
Sertão Paraibano	Brejo Santos	7	-	1	-	-	-	1	1	-	3
	Cajazeiras	4	-	1	-	-	4	-	-	-	5
	Catolé Rocha	5	-	3	-	-	-	1	-	-	4
	Conceição	6	-	-	-	-	-	2	-	-	2
	Ibiara	4	-	-	1	-	-	-	-	-	1
	Jericó	3	-	1	-	-	3	-	-	-	4
	Patos	4	-	-	-	-	-	1	-	-	1
	Princesa Isabel	6	-	1	-	-	1	3	-	-	5
	Souza	3	-	2	-	-	-	-	-	-	2
	Triunfo	6	-	-	-	-	1	2	-	-	3
Uiraúna	5	-	-	-	-	-	-	1	-	1	
<b>Subtotal</b>		53	-	9	-	-	9	10	2	-	31
<b>Total</b>		212	1	28	4	1	41	41	5	10	131

\* Some soil samples showed growth of more than one species of dermatophyte. \*\* Dermatophytes: *A.g* - *Arthroderma gypseum*; *M.g* - *Microsporium gypseum*; *M.n* - *M. nanum*; *T.a* - *Trichophyton ajelloi*; *T.m* - *T. mentagrophytes* var. *mentagrophytes*; *T.te* - *T. terrestre*; *T.to* - *T. tonsurans*; *T.v* - *T. verrucosum*.

at alkaline pH<sup>16</sup>. High rates of *M. gypseum* were observed in soils from Rio Grande do Sul, Brazil (79%)<sup>8</sup>, Argentina (89%)<sup>13</sup>, India (64%)<sup>4</sup>, Kuwait (50%) in parks and gardens<sup>1</sup>, and Italy (39%)<sup>25</sup>.

*M. gypseum* has a universal distribution, and it is the etiological agent of *tinea capitis* and *tinea corporis* in humans and animals,

where dogs, horses and rodents are common reservoirs of keratin<sup>1</sup>. In this investigation, it was found in soils of empty lots, slums, schools, squares, homes and rural areas. HAYASHI & TOSHITANI<sup>11</sup> reported, in Japan, 271 cases of human infection by this fungal species. A case of *tinea capitis* due to infection by this species, has been diagnosed in João Pessoa-PB<sup>18</sup>.

**Table 2**  
Distribution of dermatophytes from soil samples of urban and rural areas of cities of Paraiba State

Soils Samples	Urban Area						Rural Area n	Total n (%)
	School* n	Square* n	Empty lot* n	Slum* n	Residence n	Beach n		
Negative	21	23	15	12	07	06	14	94 (44.3)
Positive	28	24	29	23	05	03	06	118 (55.7)
<b>Dermatophytes</b>								
<i>Trichophyton terrestre</i>	10	12	11	07	-	01	-	41 (31.3)
<i>T. mentagrophytes var. mentagrophytes</i>	11	08	08	07	02	01	04	41 (31.3)
<i>T. verrucosum</i>	01	02	03	04	-	-	-	10 (7.6)
<i>T. tonsurans</i>	02	-	01	01	-	01	-	05 (3.8)
<i>T. ajelloi</i>	01	-	-	-	-	-	-	01 (0.8)
<i>Microsporum gypseum</i>	03	03	09	08	03	-	02	28 (21.4)
<i>M. nanum</i>	02	01	01	-	-	-	-	04 (3.0)
<i>Anthroderma gypsea</i>	-	-	-	01	-	-	-	01 (0.8)
<b>Total</b>	30	26	33	28	05	03	06	13
(%)	(22.9)	(19.8)	(25.2)	(21.4)	(3.8)	(2.3)	(4.6)	(100.0)

\* Some soil samples showed growth of more than one species of dermatophytes.

**Table 3**  
Distribution of dermatophytes, with reference to soil pH

Dermatophytes	Soil pH		
	Acid 4.65-6.65	Alkaline 7.02-9.06	Total n (%)
<i>Trichophyton terrestre</i>	09	32	41 (31.3)
<i>T. mentagrophytes var. mentagrophytes</i>	16	25	41 (31.3)
<i>T. verrucosum</i>	02	08	10 (7.6)
<i>T. tonsurans</i>	01	04	05 (3.8)
<i>T. ajelloi</i>	-	01	01 (0.8)
<i>Microsporum gypseum</i>	10	18	28 (21.3)
<i>M. nanum</i>	-	04	04 (3.1)
<i>Anthroderma gypsea</i>	-	01	01 (0.8)
<b>Total n (%)</b>	38 (29%)	93 (71%)	131 (100.0)

Binomial test.  $H_0$ : acid pH = alkaline pH and  $H_1$ : acid pH  $\neq$  alkaline pH;  $\rho = 0.000 \leq 0.05$ , reject  $H_0$ .

*M. nanum* (3%) was isolated for the first time from soil of schools, beaches and empty lots in Paraiba State. In a study carried out on soil of a swimming resort, in Mexico, its isolation rate was 5%<sup>19</sup>.

*T. ajelloi* was isolated from soils of the South and Southeast regions of Brazil<sup>8,10,29</sup>. ALVAREZ *et al.*<sup>2</sup> reported an isolation rate of 66% for this

fungus in soil of Argentina. In this study, the first and only isolation of this species (0.8%) was observed in soil around a school.

Among the zoophilic species, *T. mentagrophytes var. mentagrophytes* was the species of highest incidence in soils of various places (schools, gardens, parks, beaches, caverns, chicken coops, pens and homes) in some Brazilian states such as Amazonas, São Paulo and Goiás<sup>29,34,35</sup>, as well as soils of Mexico, Iran, Nigeria and India<sup>4,19,24,30,31</sup>. In this study, this species (31.3%) was isolated from all soils of urban and rural areas, and one strain of this species was reported in highly acidic soil at pH 4.65. In Berlin, the average pH of positive keratinophilic fungal samples was 5.8<sup>7</sup>, and in India, it was the most common isolated species from pH 6.5 to 9.5 soils<sup>14</sup>.

*T. verrucosum* is a zoophilic species cited as the agent encountered in the case of cattle, which can be transmitted to humans. It is usually highly inflammatory involving the scalp, beard or exposed area of body<sup>3,18</sup>. In this investigation, *T. verrucosum* was reported from urban areas at pH 6.65 - 8.05.

The isolation rate of *T. tonsurans* as an anthropophilic species was 3.8% in soils of schools, slums, beaches and empty lots and 80% at alkaline pH. GOULART *et al.*<sup>10</sup> also reported the isolation of this species in the soil of Rio of Janeiro. In Recife, an epidemiological correlation has been observed between *T. tonsurans* isolated from soils of parks I (28%) and II (20%) and dermatophytosis agents<sup>16,18</sup>.

## CONCLUSION

The soils of urban areas within empty lots, schools, slums and

squares of cities of mesoregions of Paraíba State were found to be the most suitable reservoirs for almost all dermatophytes. Its growth may have been influenced by environmental factors such as residues of human and/or animal keratin and alkaline pH.

## RESUMO

### Distribuição de dermatófitos isolados de solos de cidades do Estado da Paraíba, Brasil

Os dermatófitos, fungos queratinofílicos, representam importantes microrganismos da microbiota do solo, onde existem espécies cosmopolitas e outras de distribuição geográfica restrita. Este estudo teve como objetivo ampliar o conhecimento da distribuição de dermatófitos do solo proveniente de áreas urbanas (terrenos baldios, escolas, favelas, praças, praias e residências) e rurais de quatro mesorregiões paraibanas e da influência do pH na adaptação desse grupo de fungos. Amostras de solos urbanos e rurais foram coletadas de 31 cidades do estado da Paraíba, Brasil. De 212 amostras 62% apresentaram crescimento fúngico, destacando-se a Mesorregião da Mata Paraibana (43.5%), a qual apresenta clima tropical, quente e úmido. O pH das amostras de solo variou de 4.65 a 9.06, com crescimento de 71% dos dermatófitos em pH alcalino (7.02 - 9.06) ( $p = 0.000$ ). Das 131 cepas isoladas 57.3% eram espécies geofílicas, destacando-se *Trichophyton terrestre* (31.3%) e *Microsporium gypseum* (21.4%). *M. nanum* e *T. ajelloi* foram isolados pela primeira vez no estado da Paraíba. Entre as espécies zoofílicas foram identificadas *T. mentagrophytes* var. *mentagrophytes* (31.3%) e *T. verrucosum* (7.6%) e como espécie antropofílica foi isolada *T. tonsurans*. Os solos de terrenos baldios, escolas, favelas e praças de cidades paraibanas são os reservatórios mais adequados dos dermatófitos, cujo crescimento pode ter sido influenciado por fatores ambientais, solos com resíduos de queratina humana e ou animal e pH alcalino.

## ACKNOWLEDGEMENTS

The authors would like to thank to the Laboratory of Ceramics for collecting and measuring the pH of soils samples.

## REFERENCES

1. Al-Musallam AA, Al-Zarban SS, Al-Sanè NA, Ahmed TM. A report on the predominant occurrence of a dermatophytes species in cultivated soil from Kuwait. *Mycopathologia*. 1995;130:159-61.
2. Alvarez DP, Luque AG, Marini P. Influencia del sustrato queratinoso de suelos de Pradera sobre la colonización por dermatofitos geofílicos. *Bol Micol*. 1986;3:25-9.
3. Amaral CDP, Pereira DIB, Meireles MCA. Caracterização da microbiota por fungos filamentosos no tratamento hípico de bovinos de corte. *Ci Rural*. 2011;41:2137-42.
4. Anbu P, Hilda A, Gopinath SC. Keratinophilic fungi of poultry farm and feather dumping soil in Tamil Nadu, India. *Mycopathologia*. 2004;158:303-9.
5. Barnett HL, Hunter BB. Illustrated genera of imperfect fungi. 4<sup>th</sup> ed. New York: Burgess; 1986.
6. Bohacz J, Kowalska TK. Species diversity of keratinophilic fungi in various soil types. *Cent Eur J Biol*. 2012;7:259-66.
7. Böhme H, Ziegler W. The distribution of geophilic dermatophytes and other keratinophilic fungi in relation to the pH of the soil. *Mycopathol Mycol Appl*. 1969;38:247-55.
8. Fischman O, Ramos CD. Geophilic dermatophytes recovered from Rio Grande do Sul soil. *Mycopathol Mycol Appl*. 1967;33:157-60.
9. Ganaie MA, Sood S, Rizvi G, Khan TA. Isolation and identification of keratinophilic fungi from different soil samples in Jhansi city (India). *Plant Pathol J*. 2010;9:194-7.
10. Goulart EG, Lima SMF, Carvalho MA, Oliveira JA, Jesus MM, Campos RE, et al. Isolamento de fungos patogênicos do solo no município do Rio de Janeiro, RJ, Brasil. *Folha Méd*. 1986;93:15-20.
11. Hayashi N, Toshitani S. Human infections with *Microsporium gypseum* in Japan. *Mykosen*. 1983;26:337-45.
12. Hoog GS, Guarro J, Figueras MJ. Atlas of clinical fungi. 2<sup>nd</sup> ed. Utrecht: Centraalbureau voor Schimmelcultures; 2000. 1126p.
13. Iovannitti CA, Malliarchuk O, Casanova A, Dawson M. Estudio micológico en muestras de tierra de la ciudad de la Plata. *Rev Argent Micol*. 1985;8:9-11.
14. Jain N, Sharma M. Distribution of dermatophytes and other related fungi in Jaipur city, with particular reference to soil pH. *Mycoses*. 2011;54:52-8.
15. Kaul S, Sumbali G. Impact of some ecological factors on the occurrence of poultry soil-inhabiting keratinophiles. *Mycopathologia*. 1998;143:155-9.
16. Leal AFG, Macêdo DPC, Laranjeira D, Souza-Motta CM, Fernandes MJS, Magalhães OMC, et al. Correlação epidemiológica entre fungos queratinofílicos isolados do solo e agentes de dermatomicoses. *Rev Soc Bras Med Trop*. 2009;42:471-3.
17. Lee MJ, Park JS, Chung H, Jun JB, Bang YJ. Distribution of soil keratinophilic fungi isolated in summer beaches of the east sea in Korea. *Korea J Med Mycol*. 2011;16:44-50.
18. Lima OE, Pontes ZBVS, Oliveira NMC, Carvalho MFFP, Guerra MFL, Santos JP. Freqüência de dermatofitoses em João Pessoa, Paraíba, Brasil. *An Bras Dermat*. 1999;74:127-32.
19. López Martínez R. Investigación de algunas fuentes de infección en las dermatofitosis: estudio de suelos, animales y hombre. *Gac Méd Méx*. 1986;122:167-72.
20. Mahmoudabadi AZ, Zarrin M. Isolation of dermatophytes and related keratinophilic fungi from the two public parks in Ahvaz. *Jundishapur J Microbiol*. 2008;1:20-3.
21. Mangiaterra ML, Alonso JM. Keratinophilic fungi in soils of Corrientes city (Argentina). *Bol Micol*. 1989;4:129-33.
22. Norusis MJ. SPSS for Windows-Base System User's Guide, Release 13.0. Chicago: SPSS.
23. Ogbonna CI, Pugh GJ. Keratinophilic fungi from Nigerian soil. *Mycopathologia*. 1987;99:115-8.
24. Oyeka CA, Okoli I. Isolation of dermatophytes and non-dermatophytic fungi from soil in Nigeria. *Mycoses*. 2003;46:336-8.
25. Papini R, Mancianti F, Grassotti G, Cardini G. Survey of keratinophilic fungi isolated from city park soils of Pisa, Italy. *Mycopathologia*. 1998;143:17-23.
26. Pontes ZBVS, Oliveira AC. Dermatophytes from urban soils in João Pessoa, Paraíba, Brazil. *Rev Arg Microbiol*. 2008;40:161-3.
27. Rebell G, Taplin D. Dermatophytes: their recognition and identification. Coral Gables: University of Miami; 1974.
28. Rodríguez JL. Atlas escolar da Paraíba. 3. ed. João Pessoa: Grafiset; 2002.
29. Rogers AL, Beneke EJ. Human pathogenic fungi recovered from Brazilian soil. *Mycopathol Mycol Appl*. 1964;22:15-20.

30. Shadzi S, Chadeganipour M, Alimoradi M. Isolation of keratinophilic fungi from elementary schools and public parks in Isfahan, Iran. *Mycoses*. 2002;45:496-9.
31. Sharma M. Incidence of dermatophytes and other keratinophilic fungi in the schools and college playground soils of Jaipur, India. *Afr J Microbiol Res*. 2010;4:2647-54.
32. Silva ME. Ocorrência de *Cryptococcus neoformans* e *Microsporium gypseum* em solos da Bahia, Brasil. *Bol Fund Gonçalo Moniz*. 1960;17:1-14.
33. Vanbreuseghem R. Technique biologique pour l'isolement des dermatophytes de sol. *Ann Soc Belge Méd Trop*. 1952;32:173-8.
34. Vilela EM, Moraes MAP. Isolamento de *Microsporium gypseum* e *Trichophyton mentagrophytes* no solo da cidade de Manaus, Amazonas (Brasil). *Rev Inst Med Trop Sao Paulo*. 1962;4:299-301.
35. Zampronha VCC, Oliveira IP, Monteiro MSR, Souza H, Santos KJG, Araújo AA. Isolamento e identificação de dermatófitos presentes no contínuo do solo de cerrado do campus II da Universidade Católica de Goiás. *Rev Eletrôn Fac Montes Belos*. 2005;1:37-46.

Received: 16 January 2012

Accepted: 28 March 2013