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Color stability, surface roughness and microhardness of composites submitted to mouthrinsing action

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ABSTRACT

bjective: The purpose of this study was to evaluate the effect of mouth rinse solutions on color stability, surface roughness and microhardness of two composite resins. Material and Methods: Fifty test specimens of each composite (Filtek Z250 and Z350; 3M ESPE) were made using a teflon matrix (12x2 mm). Color, surface roughness and Knoop microhardness baseline measurements of each specimen were made and specimens (n=10)were immersed in 5 mouth rinse solutions: G1: distilled water (control), G2: Plax Classic, G3: Plax alcohol-free; G4: Periogard, and G5: Listerine. Final measurements of color, roughness and microhardness were performed and the results submitted to statistical analysis (2-way ANOVA, Bonferroni's test; p<0.05). Results: The most significant color change was observed for Z250 when immersed in Listerine (p < 0.05). Z350 showed greater color change when immersed in Plax alcohol-free (p<0.05), but with no significant difference for Listerine (p>0.05). With regard to roughness, both composites showed significant changes when immersed in Listerine in comparison with Plax alcohol-free (p<0.05). Microhardness of Z350 was shown to be significantly changed when the composite was immersed in Plax alcohol-free (p < 0.05). Conclusion: Composite changes depended on the material itself rather than the mouth rinse solution used.

Key words: Composite resins. Oral hygiene. Dental materials. Physical properties.

INTRODUCTION

Color and surface roughness are very important properties in aesthetics, characterizing a smile²¹. Since the introduction of composites in 1960, efforts have been made to increase the longevity of composite restorations. Although some progress has been made, optical properties in this type of materials need to be improved⁵.

The color change of composites is multifactorial, being associated with intrinsic discoloration and extrinsic staining of the material. Intrinsic factors involve changes in the chemical stability of resin matrix and matrix/particles interface, and extrinsic factors are related to absorption of staining solutions from exogenous sources related to hygiene habits, food, and smoking⁸.

Composite structure and the characteristics of the inorganic fillers have a direct impact on composite resin surface smoothness²⁶ and susceptibility to extrinsic staining²⁴. Several studies have shown that composite resins are susceptible to color alteration when exposed to staining solutions, especially red wine, coffee, cola, tea and whisky^{2,9}. Furthermore, this property depends on both water absorption of the composite and its hydrophilic resin matrix to allow permeation of staining agents, thus resulting in greater color changes².

Today, the number of people using mouth rinse

solutions for anti-microbial control has increased not only because of professional recommendations, but also due to the capacity of such materials to provide cooling sensation and to reduce halitosis⁷. Mouth rinse solutions have various components such as detergents, emulsifiers, organic acids, dyes and alcohol. It is known that composite resins exposed to ethanol exhibit lower microhardness values compared to non-exposed materials²³. According to Sarret, et al.²² (2000), alcohol acts as a plasticizer of the polymeric matrix, making the material more ductile. In addition, ethanol can reduce bonding between resin matrix and inorganic fillers, which might decrease erosion resistance and cause staining of resin matrix²³.

Thus, the purpose of this study was to evaluate the effect of mouth rinse solutions on color stability, surface roughness and microhardness of composite resins. The null hypothesis stated that mouth rinse solutions would not promote changes in the properties of the studied composites.

MATERIAL AND METHODS

Samples preparation

Two direct composite resins (shade A3) currently indicated for esthetic anterior and posterior restorations were used in the present study. Information regarding composite type, composition and manufacturer is given in Figure 1.

Fifty cylindrical test specimens (12 mm diameter x 2 mm thick) of each composite were made using a Teflon matrix. Each material was inserted into the matrix in 2 increments and light-activated by a LED device (FLASHlite 1401, Discus Dental, Culver City, CA, USA, light intensity \geq 1100 mW/ cm², wavelength in the range between 460 and 480 nm), for 20 seconds, according to manufacturer's recommendations. The specimens were polished with 320, 600, 1200 and 2000-grit sandpapers. The thickness of every test specimen was checked with a digital caliper (Digimess, São Paulo, SP, Brazil). Next, the test specimens were randomly separated into 5 groups (n=10) according to the mouth rinse solution in which they had been immersed (Figure

2): Group 1 - distilled water (control); Group 2 -Plax classic; Group 3 - Plax alcohol-free; Group 4 - Periogard; and Group 5 - Listerine Cool Mint.

To simulate the use of mouth rinse solutions once a day during a period of 1 year, the test specimens were submitted to 12 cycles of 1-min immersion, then washed in running water and immersed in distilled water for 29 min at 37°C, during 30 days, totaling 360 cycles¹¹.

Color stability

Before immersion, color reading of the test specimens was performed according to the CIE (Commission Internationale de l'Eclairage) L*a*b* system, against a white background (Standard for 45/0°; Gardner Laboratory Inc., Bethesda, MD, USA) in a reflection spectrophotometer (PCB 6807, BYK Gardner, Geretsried, Germany). This equipment is specific for color reading and has 30 LED lamps with 10 different colors arranged in a circle, which directs a light bundle at 45° with the material surface. This light bundle is reflected 0° back to the equipment, which captures and records the L*, a* and b* values of each specimen. The L* axes refers to the lightness coordinate and its value ranges from zero (black) to 100 (white). The axis a* and b* are chromaticity coordinates in the redgreen axis and the yellow-blue axis, respectively. Positive a* values indicate a shift to red and negative values indicate a shift to green. Similarly, positive b* values indicate the yellow color range and negative values indicate the blue color range.

After immersion in mouth rinse solutions, specimens color was measured by the spectrophotometer, as previously described. Based on the L*, a*, b* values, color variation (ΔE) was determined using the following equation:

 $\Delta E = [(\Delta L^*)^2 + (\Delta a^*)^2 + (\Delta b^*)^2]^{1/2}$

Values of $\Delta E \ge 3.3$ were considered clinically unacceptable²⁰.

Surface roughness and Knoop microhardness Surface roughness (R₂) and Knoop microhardness

Commercial	Туре	Monomers	Load particles		Manufacturer
name			Size (µm)	Volume %	
Filtek Z250	Microhybrid	Bis-GMA, UDMA,	Zirconia/Silica 0.6 µm	60%	3M ESPE Dental Products,
		Bis-EMA			St Paul, MN, USA
Filtek Z350	Nanoparticulate	Bis-GMA, Bis-	Zirconia/Silica 5-20 nm	59.5%	3M ESPE Dental Products,
		EMA, UDMA and	with clusters of 0.6-1.4		St Paul, MN, USA
		TEGDMA	μm		

Bis-GMA: bisphenol A diglycidyl ether dimethacrylate; Bis-EMA: Ethoxylated bisphenol A dimethacrylate, TEGDMA:Triethylene glycol dimethacrylate; UDMA: urethane dimethacrylate

Figure 1- Tested composites

of the test specimens were performed before and after mouth rinse solution immersion. Surface roughness was measured with a Rugosimeter (Mitutoyo SJ-201P, Mitutoyo, Tokyo, Japan), cutoff - 0.25 mm, L_c parameter - 1.25, speed - 0.1 mm/s. The rugosimeter needle (10 µm diameter) was positioned over each test specimen, performing three readings in different locations of the sample surface, after which, the mean roughness of the test specimens were obtained. Surface roughness changes were calculated by the differences between mean values obtained before and after immersion.

The Knoop microhardness of the test specimens was measured (Shimadzu device, HMV Shimadzu, Kyoto, Japan) in three different points, with a 10 N/15 s load. After three readings, the microhardness mean values of the test specimens were obtained. For Knoop microhardness calculation, the baseline mean values were subtracted from those obtained after immersion.

Statistical analysis

For each property, data obtained were subjected to two-way ANOVA and the measurements were compared by the Bonferroni's test. All statistical testing was performed at a pre-set alpha of 0.05.

RESULTS

Color stability

The mean values and standard deviations for color stability (ΔE) changes are presented in Table 1. None of the studied composites exhibited color change with values above the clinically acceptable limit ($\Delta E \ge 3.3$)²⁰. With regard to Z250, comparison of the effects of different mouth rinse solutions revealed a small color change for specimens immersed in Listerine, with statistically significant difference compared with the other mouth rinse solutions (p<0.05). When immersed in Plax Classic solution, this composite exhibited an intermediate

Commercial name	Composition	рН	Manufacturer
Plax Classic	Triclosan, Gantrez, Sodium lauryl sulphate, Glycerin and Propylene, Sodium fluoride (225 ppm fluoride), Sodium methyl taurate, Disodium phosphate, Sodium saccharine, red dye, ethylic alcohol (6%) and water	5.8	Colgate Palmolive Ind. e Com. Ltda., São Bernardo do Campo, SP, Brazil
Plax Alcohol-free	Cetyl Pyridinium Chloride (CPC) 0.05 % 225 ppm fluoride, Gantrez, Sodium lauryl sulphate, Glycerin and Propylene, Sodium methyl taurate, phosphoric acid, Disodium phosphate, Sodium saccharine and water	4.96	Colgate Palmolive Ind. e Com. Ltda., São Bernardo do Campo, SP, Brazil
Periogard	0.12% gluconate (or digluconate formulation with chlorhexidine-free concentration (0.067%), water, glycerin, ethanol, polysorbate 20, aromatic composition with predominant flavor of mint, sodium saccharine, FD&C blue dye #1	5.0-7.0	Colgate Palmolive Ind. e Com. Ltda., São Bernardo do Campo, SP, Brazil
Listerine Cool Mint	Thymol, eucalyptol, methyl salicylate, menthol, excipients: purified water, sorbitol solution, alcohol (30%), poloxamer 407, benzoic acid, mint essence, sodium saccharine, sodium benzoate, green dye #3	3.7	Laboratório Pfizer Ltda., São Paulo, SP, Brazil

Figure 2- Tested mouth rinses

Table 1- Means (standard deviations) for ΔE values (2-way ANOVA, Bonferroni's test; p<0.05)

	Z250	Z350
Control	1.52 (0.37) ^{a,A}	0.96 (0.24) ^{b,AB}
Listerine	0.99 (0.29) ^{a,B}	1.25 (0.24) ^{a,AC}
Plax Classic	1.32 (0.46) ^{a,AB}	0.64 (0.23) ^{b,B}
Plax alchohol-free	1.42 (0.45) ^{a,A}	1.46 (0.30) ^{a,C}
Periogard	1.40 (0.71) ^{a,A}	0.93 (0.16) ^{b,AB}

Different lowercase letters in lines and uppercase letters in columns indicate statistically significant difference.

	Z250	Z350
Control	44.94 (0.34) ^{a,A}	27.60 (0.28) ^{a,A}
Listerine	78.46 (0.56) ^{a,AB}	61.99 (0.53) ^{a,AB}
Plax Classic	38.28 (0.18) ^{a,A}	13.36 (0.18) ^{a,A}
Plax alcohol-free	-4.22 (0.20) ^{a,AC}	-10.76 (0.27) ^{a,AC}
Periogard	64.43 (0.46) ^{a,AB}	58.22 (0.34) ^{a,AB}

Table 2- Means (standard deviations) for R₂ (%) values (2-way ANOVA, Bonferroni's test; p<0.05)

Different lowercase letters in lines and uppercase letters in columns indicate statistically significant difference.

Table 3- Means (standard deviations)) for Knoop microhardness	(%) values (2-way ANOVA	Bonferroni's test; p<0.05)

	Z250	Z350
Control	7.30 (0.17) ^{a,A}	22.28 (0.18) ^{a,A}
Listerine	-14.88 (0.10) ^{a,A}	1.20 (0.27) ^{a,A}
Plax Classic	-6.98 (0.08) ^{a,A}	15.83 (0.40) ^{a,A}
Plax alcohol-free	-5.03 (0.23) ^{a,A}	-25.73 (0.19) ^{a,B}
Periogard	-0.24 (0.18) ^{a,A}	14.80 (0.24) ^{a,A}

Different lowercase letters in lines and uppercase letters in columns indicate statistically significant difference.

 ΔE value without statistically significant difference compared to other mouth rinse solutions (p>0.05). With regard to Z350, the highest ΔE change was observed for Plax alcohol-free solution, with statistically significant difference compared with the other solutions (p>0.05), except for Listerine. Comparing the composites to each other, it was observed statistically significant difference (p<0.05) for the test specimens immersed in distilled water (control), with both Plax Classic and Periogard solutions allowing smaller color changes for Z350.

Surface roughness

The mean values and standard deviations for surface roughness (R_a) changes are presented in Table 2. For both composites, the greatest change in R_a occurred when the samples were immersed in Listerine solution, with statistically significant difference compared with Plax alcohol-free (p<0.05). When compared to each other, Z250 and Z350, showed no statistically significant difference (p>0.05) regarding any studied mouth rinse solution.

Knoop microhardness

The mean values and standard deviations for Knoop microhardness changes are presented in Table 3. When compared to each other, Z250 and Z350 showed no statistically significant difference (p>0.05) regarding any studied mouth rinse solution. Immersion of Z350 in Plax alcoholfree solution resulted in greater decrease of microhardness values, with statistically significant difference in relation to the other mouth rinse

solutions (p < 0.05).

DISCUSSION

The variability in our results was consistent with other studies, which showed that several factors, including composite resin type, mouth rinse solution and immersion cycle, had a significant influence on color stability, surface roughness and microhardness of composites²⁸. Therefore, the null hypothesis could not be accepted.

Color stability can be evaluated both visually and by specific instruments^{5,8,11}. The methodology used in the present study is according to previous studies that used spectrophotometry and the CIE L*a*b* coordinates system^{5,11}. The CIE L*a*b* system was chosen to evaluate color variation (Δ E) because it is appropriate for small color changes determination and have advantages such as repeatability, sensitivity and objectivity¹¹.

Several authors have reported that ΔE values ranging from 1 to 3 are perceptible to the naked eye¹⁸ and ΔE values greater than 3.3 are clinically unacceptable²⁰. Considering these concepts; the composite resins tested in the present study demonstrated acceptable color stability when stored in the different types of mouth rinse solutions (Table 1).

The affinity of the resin matrix for stains is modulated by its conversion degree and by some physical properties, such as water sorption^{2,6}. Water sorption of composite resins depends on the resinous matrix composition. It has been reported that water uptake in Bis-GMA based composite resins increased from 3 to 6%, as the proportion of TEGDMA increased from 0 to 1%¹³. UDMA seems to be more stain resistant than Bis-GMA¹⁵, and under normal curing conditions, UDMA based composite resins presented lower water sorption and higher color stability than other dimethacrylates in their resin matrix^{15,20}, which could be observed in the present study.

Moreover, it is important that the composite resin presents uniform filler particle distribution in the polymer network to minimize the formation of filler-rich and filler-depleted areas within the composites. This is especially important regarding the performance of composites in aqueous environments, such as mouth rinse solutions, since voids or nonbonding spaces at the filler/ matrix interface may increase the water sorption of composites²⁵.

Nanocomposites correspond to a class of new materials with nanoscale inorganic filler particles dispersed within the resinous matrix²⁷. In comparison with microhybrid composites, these materials have been reported to have improved properties, such as, elasticity modulus, mechanical strength and color stability. Furthermore, these improvements are achieved at low concentrations of the inorganic filler particles. This fact contrasts strongly with conventional filled composites, which generally require high loadings within the range of 60%^{11,19}. This is an important factor in aesthetic maintenance of non-particulate composites, since filler particles are responsible for spreading the light, which in turn, provides the opacity of the restorative resin materials. The larger the filler particle size the greater the light spread, and consequently, the greater the opacity¹⁶. The opacity of composites increases as the difference between the refraction indexes of resinous matrix and filler particles also increases¹⁶. In addition, the smaller the filler particle, the smaller the amount of water absorbed by the polymer network, which results in lower degradation of the interface matrix/particle, and consequently, lower color change¹⁰. The Z250 contains filler particles with average size of 0.6 µm, while Z350 contains filler particles ranging from 5 to 20 nm, which could explain the greater color stability for Z350²⁹. Also, according to Kawaguchi, et al.¹⁴ (1994), microhybrid composites present a lower coefficient of light transmission due to the various sizes of their particles, which contributed to the higher values of ΔE for Z250.

According to Villalta, et al.³⁰ (2006), the low pH and alcohol concentration of solutions affect the surface roughness of composite resins and cause staining. Nevertheless, the composite Z350 presented statistically significant difference regarding color stability, surface roughness and microhardness values when the composite was immersed in the alcohol-free mouth rinse solution (Plax alcohol-free), thus contradicting the results of a previous study³⁰. However, Miranda, et al.¹⁷ (2011) demonstrated that despite the absence of alcohol, Plax alcohol-free has phosphoric acid in its composition, which might alter the polymer matrix of composites by catalysis of the ester groups present in the dimethacrylate monomers. The hydrolysis of the ester groups forms alcohol and carboxylic acid molecules, which accelerate polymer network degradation by the decrease of pH inside the composite matrix^{10,17}. The degradation of the polymer network leads to a phenomenon called "plasticization", which decreases microhardness values in composites¹⁰.

According to the methodology used in the present study, the negative values regarding microhardness represent a smoothening of the resin matrix, and positive values, hardening. Despite the great decrease in the composite microhardness (-25.73), the surface roughness improved when compared to the pre-immersion values, except when the composite was immersed in the Plax alcohol-free solution. The same result was observed for Z250, since a significant change in surface roughness was found after immersion into the same mouth rinse solution. It is worth emphasizing, however, that there exists a critical value regarding surface roughness change ($R_{a} \ge 0.2$ μ m). According to Bollen, et al.³ (1997), a greater biofilm retention allows an increase in recurrent caries. Another critical value for R_a is 0.3 μ m, which can be detected when the patient's lips or tongue enters in contact with the restorative material, causing discomfort³. None of the studied composites showed values for surface roughness change above the critical limits, irrespective of the type of mouth rinse solution used.

Asmussen¹ (1984) reported that mouth rinse solutions with high alcohol content might soften the composite resin, especially Bis-GMA-based composites. However, Gürgan, et al.¹² (1997) showed that regardless of the alcohol concentration, both alcohol-containing and alcohol-free mouth rinse solutions could affect the properties of composite resins. Immersed of Z350 in Listerine, the mouth rinse solution with the highest alcohol concentration (30%), resulted in decrease in microhardness values, though with no significant difference from the other mouth rinse solutions. As for Z250, the results were the opposite; there was an increase in the microhardness values when immersed in Listerine and a decrease when immersed in Plax alcohol-free solution, with statistically significant difference. These results suggest that changes in the microhardness of composites do not depend on mouth rinse's ethanol concentration¹².

The same finding could be observed regarding

to color stability. Z350 showed significant changes when immersed in Listerine and Plax alcohol-free solutions, both having entirely different concentrations of ethanol¹². However, this fact could be explained, once again, by the presence of phosphoric acid in Plax alcohol-free composition. Studies have reported the release of by-products from polymer network degradation, such as methacrylic acid, formaldehyde and specific methacrylate molecules, which are capable of promoting color change in composites^{4,10,17}.

On the other hand, Z250 had greater color change when immersed in Listerine compared to the other solutions, unlike Celik, et al.⁴ (2008), who found no color change in the composites immersed in the same mouth rinse solution, despite the high ethanol concentration.

Clinically, the effect of mouth rinse solutions on restorative materials can be modified by several factors that were not replicated in this *in vitro* study. Among these factors, saliva could dilute or buffer pH of mouth rinse solutions, thus reducing the effect of resinous matrix plasticization¹⁰ and forming a pellicle that could have a protective effect on the composite surface, thus, decreasing material staining⁴. Based on such factors, further *in vivo* studies are needed to determine the effects of mouth rinse solutions on these properties of composites.

CONCLUSIONS

The results of the present study allow us to conclude that the changes observed in the composites depended on the material itself rather than the mouth rinse solution used.

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