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COMPARATIVE SEM EVALUATION OF THREE SOLVENTS USED IN ENDODONTIC RETREATMENT: AN *EX VIVO* STUDY

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ABSTRACT

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This study compared, by scanning electron microscopy (SEM), the efficacy of three solvents on the removal of filling materials from dentinal tubules during endodontic retreatment. Forty human maxillary canines with straight canals were prepared according to a crown-down technique and enlarged to a#30 apical file size, before obturation with gutta-percha and a zinc-oxide-eugenol based sealer. The samples were stored for 3 months before being randomly assigned to four groups: chloroform (n=10), orange oil (n=10), eucalyptol (n=10) and control (n=10). Solvents were applied to a reservoir created on the coronal root third using Gates Glidden drills. The total time for retreatment using the solvents was 5 minutes *per* tooth. Following retreatment the roots were split longitudinally for SEM evaluation. SEM images were digitized, analyzed using Image ProPlus 4.5 software, and the number of dentinal tubules free of filling material from the middle and apical thirds was recorded. No significant difference was found among the solvent groups regarding the number of dentinal tubules free of filling material (p<0.05). Under the tested conditions, it may be concluded that there was no significant difference among the solvents used to obtain dentinal tubules free of filling material remnants.

Uniterms: Chloroform. Orange oil. Eucalyptol. Endodontic retreatment.

INTRODUCTION

Endodontic retreatment requires removal of pre-existing endodontic filling material and disinfection of the root canal system. This is accomplished during chemomechanical reinstrumentation. In order to remove filling materials without damage to the tooth, chemical solvents are used to solubilise gutta-percha^{1,2,5}. Orange oil, eucalyptol, xylol, chloroform, halothane, and rectified turpentine have all been used as adjuncts to remove endodontic filling materials^{13,15,16,21}. Retreatment has also been done combining heat with a solvent, in order to remove filling material from thermafilobturated teeth³³. It has been shown that ultrasonic instrumentation required significantly less time to remove root fillings than hand instrumentation¹⁶ Others authors have shown that rotary instruments remove more filling materials during endodontic retreatment than manual instruments^{3,12,17}. Additionally, Nd:YAP laser was shown to be effective for root canal preparation during retreatment because it preserves the root canal dentin walls and does not cause temperature raise in the root, which could potentially harm the periodontal tissues⁷. Furthermore, orange oil has been shown not only to be effective¹¹, but also to be less cytotoxic than eucalyptol, xylol, chloroform and halothane²⁴, during endodontic retreatment procedures.

Studies have reported that it is essential to remove all root canal filling material from anatomic ramifications and dentinal tubules to ensure cleaner root canal walls²⁸. This facilitates chemomechanical preparation and antimicrobial dressing to access all ramifications of the entire root canal system during retreatment and decrease the residual microbial population^{4,6,28,30,32}. In spite of all different retreatment strategies, studies have shown that it is not possible to obtain root canal walls completely free of debris

and residual infection^{14, 31.}

The aim of the present study was to evaluate, by scanning electron microscopy (SEM), the efficacy of chloroform, orange oil and eucalyptol as adjuncts during endodontic retreatment on the removal of obturation material from dentinal tubules.

MATERIAL AND METHODS

Forty human maxillary canines were obtained from the tooth bank of the Federal Fluminense University. Teeth with straight roots were selected and then radiographed to ensure that they had a single canal, mature apex and less than 5 degree curvature.

Teeth were sectioned to provide remaining roots measuring 21 mm in length. Working length was set at 20 mm and the root canals were prepared using K-files according to a crown-down technique. Instrumentation began with Kfile size 80, and progressively smaller K-files were used to prepare to a final apical size #30 K-file (Dentsply Maillefer, Ballaigues, Switzerland)

Canals were irrigated with 2 mL 5.25% NaOCl (Formula & Ação Farmácia, São Paulo, SP, Brazil) at each change of instrument. When instrumentation was completed, canals were irrigated with 20 mL 10% citric acid (Formula & Ação Farmácia, São Paulo, SP, Brazil) to remove smear layer, followed by rinsing with 20 mL distilled water.

The canals were dried with paper points before obturation with gutta-percha and Endofill sealer (Dentsply, Petrópolis, RJ, Brazil). The root canals were obturated using the lateral compaction technique employing standardized gutta-percha master cones (Dentsply, Petrópolis, RJ, Brazil), finger spreaders and accessory cones. Excess gutta percha was removed at the canal entrance with heated instruments, before vertical compaction was used to condense gutta percha at the coronal root third. The coronal access cavities of the specimens were sealed with a temporary filling material (Coltosol; Coltene-Whaledent, Cuyahoga Falls, OH, USA).

Radiographs of each specimen were taken to ensure homogeneity of root canal obturation, lack of voids and proper length of the root canal filling material, before storage at 37°C in 100% humidity (artificial saliva) for 3 months. The teeth were assigned to four groups of ten roots each, according to the solvent employed: Group I: Control (no solvent), Group II: chloroform (Fórmula & Ação Farmácia de Manipulação, São Paulo, SP), Group III: orange oil (Fórmula & Ação Farmácia de Manipulação, São Paulo, SP), and Group IV: eucalyptol (Fórmula & Ação Farmácia de Manipulação, São Paulo, SP). In the control group, the filling material was not removed.

A reservoir for solvents was created within the coronal root third using #2 (0.70 mm) and #3 (0.09 mm) Gates Glidden drills to a depth of 3 mm beyond the canal entrance. 0.4 mL of each solvent was delivered with a syringe to sample reservoirs. After 2 minutes, to allow for solvent penetration, crown-down instrumentation was initiated to remove gutta percha using a size 60 K-file. Every 30 seconds, canal debris

was removed by rinsing with 2.0 mL 5.25 % NaOCl using a 30-gauge irrigation needle, before adding solvent and using sequentially smaller diameter files to remove gutta percha using a reaming motion. The endpoint of instrumentation was determined when a #30 K-file reached the working length. The total time required to remove the filling material was 5 minutes.

Apical patency was ensured with #10 K-file before final irrigation with 10 mL of citric acid and 10 mL with 5.25% NaOCl. All canals were then dried with paper points.

Longitudinal grooves were prepared on the external surface of all roots before splitting the samples into two halves. One half of each sample was mounted on an SEM specimen holder before placement in a vacuum chamber and coating with a 20-nm thick gold-palladium layer. Microscopic analyses were done using a scanning electron microscope (DSM940, ZEISS, Oberkochen, Germany). SEM images were obtained with an acceleration tension of 20kV, in modes of secondary (SE) and back-scattered (BSE) electrons. All sample preparation procedures were performed by the same operator.

For evaluation purposes, each specimen was divided into cervical, middle and apical thirds. A single evaluator, who was blinded to the specimen location, examined only the central region of the middle and apical thirds. SEM micrographs of the center of the middle and apical third regions were taken at X500 magnification that corresponded to an area of $367.2 \,\mu\text{m}^2$. The number of open dentinal tubules *per* area unit was quantified using image Pro Plus 4.5 software (Media Cybernetics, Silver Spring, MD, USA). The number of dentinal tubules free of filling materials per mm2 was statistically evaluated by Kruskall-Wallis test (p<0.05).

RESULTS

The median of the numbers of dentinal tubules free of filling materials *per* mm² is displayed on Table 1. Although there was a tendency for orange oil to yield a larger number of dentinal tubules free of filling material *per* mm² compared to chloroform and Eucalyptol, this difference was not statistically significant (p<0.05) for both the middle and the apical root canal thirds. However, all three tested solvents had significantly more visible dentinal tubules free of filling materials than the control group (p<0.05).

Figures 1 and 2 represent SEM micrographs of the evaluated areas, in the middle and apical root thirds respectively, showing dentinal tubules free of filling materials after the endodontic retreatment using the tested solvents. White arrows in these figures point to open dentinal tubules.

Third		Control Mean (Range)	Chloroform Mean (Range)	Orange oil Mean (Range)	Eucalyptol Mean (Range)	
Number	Middle	1332.25	6160.46*	6332.35*	6286.98*	
of tubules		(242)	(2027)	(1335)	(1345)	
	Apical	1053.12	3552.05**	3575.53**	3501.29**	
		(135)	(876)	(1000)	(946)	

TABLE 1- Means of the numb	er of dentinal tubules free	of filling materials p	<i>er</i> mm ² in each group
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*, **: No statistically significant difference (p>0.05) using Kruskall-Wallis test.

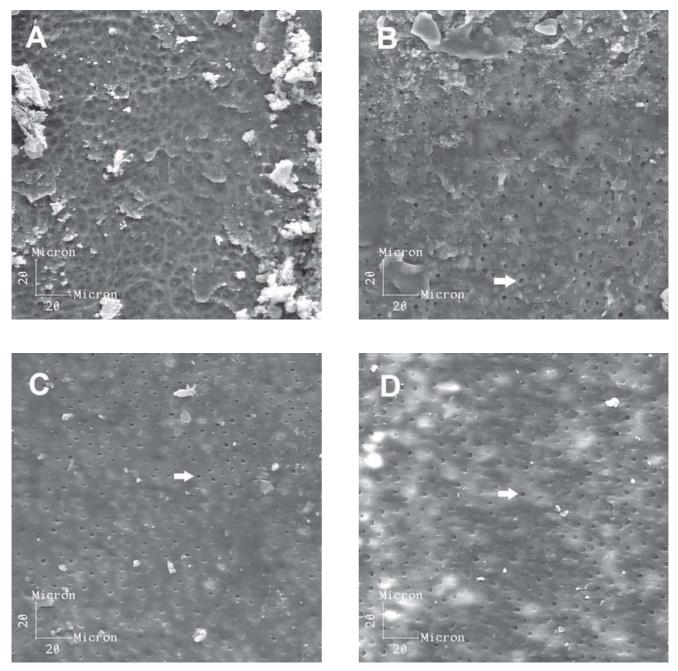


FIGURE 1- Representative SEM micrographs of samples from the middle root third: A) Control, B) Chloroform, C) Eucalyptol, D) Orange oil. Note the presence of only few open dentinal tubules *(arrows)*. Original magnification X500

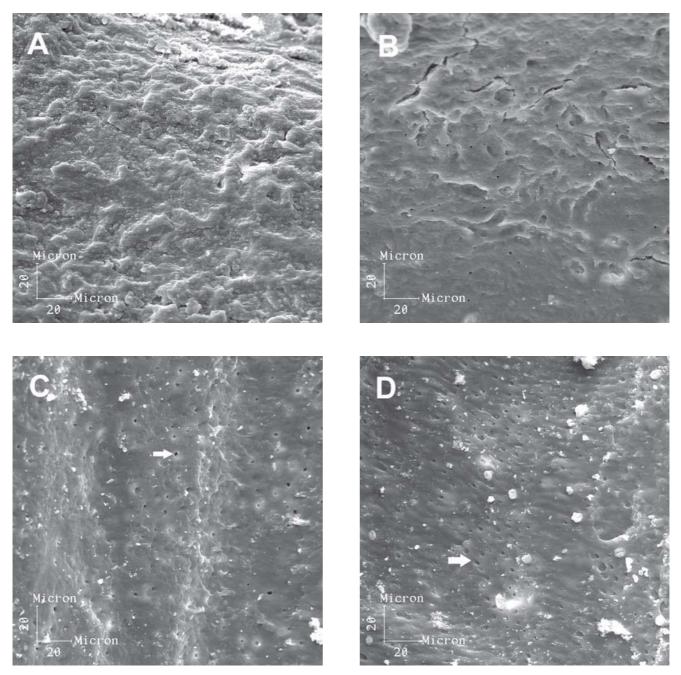


FIGURE 2- Representative SEM micrographs of samples from the apical root third: A) Control, B) Chloroform, C) Eucalyptol, D) Orange oil. Note the presence of only few open dentinal tubules *(arrows)*. Original magnification X500

DISCUSSION

The retreatment of a previously filled root canal is indicated when there is a persistent periradicular disease resulting from coronal microleakage, incomplete cleaning and shaping, and complex anatomy. Procedures for removal of root fillings include the use of stainless steel hand files, heat, ultrasonic, laser, rotary systems and solvents^{2,9,14,22,28,29,31}. In this study, 40 maxillary canines were chosen because of their round anatomic shape, which would facilitate the removal of the filling material.

Complications during retreatment procedures may be accounted for by the resistance of the filling material to

instrument penetration⁸. In this context, solvents serve as useful adjuncts to the action of the mechanical action instruments.

Although many endodontic retreatment studies have been reported, little clinical data are available regarding the time required to keep root canals soaked in solvents during retreatment. Removal of gutta-percha using hand files with or without solvent can be time-consuming, especially when the root canal filling material is well condensed ^{20.} A mean time of 1.5-10.8 minutes have been reported in laboratorial studies for retreatment of canals filled with laterally condensed gutta-percha and sealer ^{6, 18, 20, 30}. Our study allowed 5 minutes to remove the obturation material. In this study, samples were maintained at 100% humidity at 37°C for 3 months to simulate oral conditions²², prior to retreatment. However, studies have used different storage times, ranging from 2 to 3 weeks^{6,20}. It is conceivable that different storage methods and length of storage time may affect the cleaning of the root canal system.

The results indicated that there was no significant difference in the number of dentinal tubules free of filling materials among the tested solvents. Our study corroborates the results published elsewhere, which revealed that it is not possible to obtain root canal walls completely free of debris^{14, 31}.

SEM studies have been proved to allow examination of dentinal tubules with respect to the presence of filling materials^{6,26,27}. Complex root canal systems can make it difficult to clean the root canal system efficiently. In the present study, SEM micrographs showed dentinal tubules free of filling material remnants. The apical third presented the smallest number of clean dentinal tubules, compared to the middle root third.

The present work employed 10% citric acid to remove smear layer²³ prior to root canal system obturation. Other studies have examined sealer penetration into dentinal tubules after smear layer removal^{10,19}. These studies have found a greater penetration of filling materials into dentinal tubules and enhanced material adhesion to the canal walls. Such a mechanical plug may further become a problem during retreatment procedures.

Our results revealed that there was no difference in the number of clean dentinal tubules amongst the different tested solvents, in the middle or apical thirds. This is no consistent with the findings of a previous study, which showed that chloroform was more effective than eucalyptus oil in dissolving root canal sealers²⁵. Another study compared chloroform with 18 essential oils, including eucalyptol and orange oil¹⁵. These studies evaluated the capacity of dissolving sealers using standard ring moulds and gutta percha cone fragments, respectively. These authors found that only chloroform and rectified turpentine oil dissolved gutta-percha completely. The results of the present study were different because SEM was used only to evaluate dentinal tubules regarding the presence of root filling material.

Chloroform, halothane and turpentine oil have been described as cytotoxic solvents¹. In a recent study, orange oil was found to be more biocompatible than chloroform and eucalyptol²⁴. In the present study orange oil performed equally to chloroform and eucalyptol. Therefore, orange oil can be recommended as a suitable alternative to those products.

CONCLUSIONS

During endodontic retreatment of root canals filled with gutta-percha and sealer, orange oil, chloroform and eucalyptol performed equally for removal of the filling material from dentinal tubules, but superior to retreatment without the use of a solvent.

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