

IMPORTANCE OF BACTERIAL ENDOTOXIN (LPS) IN ENDODONTICS

A IMPORTÂNCIA DA ENDOTOXINA BACTERIANA (LPS) NA ENDODONTIA ATUAL

Mario Roberto LEONARDO¹, Raquel Assed Bezerra da SILVA², Sada ASSED³, Paulo NELSON-FILHO⁴

1- Chairman, Department of Endodontics, School of Dentistry of Araraquara, UNESP, Araraquara, São Paulo, Brazil.

2- Graduate Student in Pediatric Dentistry, Department of Pediatric, Preventive and Social Dentistry, School of Dentistry of Ribeirão Preto, University of São Paulo, Ribeirão Preto, São Paulo, Brazil.

3- Chairman, Discipline of Pediatric Dentistry, Department of Pediatric, Preventive and Social Dentistry, School of Dentistry of Ribeirão Preto, University of São Paulo, Ribeirão Preto, São Paulo, Brazil.

4- Associate Professor, Discipline of Pediatric Dentistry, Department of Pediatric, Preventive and Social Dentistry, School of Dentistry of Ribeirão Preto, University of São Paulo, Ribeirão Preto, São Paulo, Brazil.

Corresponding address: Mário Roberto Leonardo - Rua Humaitá, 1680 - Araraquara - SP - Brasil - CEP - 14801-903 - Fone:16-9782-6855
e-mail: nelson@forp.usp.br

Received: March 01, 2004 - Returned for modification: April 22, 2004 - Accepted: May 12, 2004

ABSTRACT

New knowledge of the structure and biological activity of endotoxins (LPS) has revolutionized concepts concerning their mechanisms of action and forms of inactivation. Since the 1980's, technological advances in microbiological culture and identification have shown that anaerobic microorganisms, especially Gram-negative, predominate in root canals of teeth with pulp necrosis and radiographically visible chronic periapical lesions. Gram-negative bacteria not only have different factors of virulence and generate sub-products that are toxic to apical and periapical tissues, as also contain endotoxin (LPS) on their cell wall. This is especially important because endotoxin is released during multiplication or bacterial death, causing a series of biological effects that lead to an inflammatory reaction and resorption of mineralized tissues. Thus, due to the role of endotoxin in the pathogenesis of periapical lesions, we reviewed the literature concerning the biological activity of endotoxin and the relevance of its inactivation during treatment of teeth with pulp necrosis and chronic periapical lesion.

UNITERMS: Bacterial endotoxin (LPS); Gram-negative bacteria, Calcium hydroxide.

RESUMO

O conhecimento mais aprofundado sobre a estrutura e atividade biológica das endotoxinas (LPS) revolucionou os conceitos sobre seu mecanismo de ação e formas de inativação. A partir da década de 80, os avanços tecnológicos na cultura e identificação microbiológica demonstraram que, em canais radiculares de dentes portadores de necrose pulpar e lesão periapical crônica, visível radiograficamente, predominam microrganismos anaeróbios, particularmente os gram-negativos. Como se sabe, os microrganismos gram-negativos, além de possuírem diferentes fatores de virulência e gerarem produtos e sub-produtos tóxicos aos tecidos apicais e periapicais, contêm endotoxina em sua parede celular. Esse conhecimento é particularmente importante, uma vez que a endotoxina é liberada durante a multiplicação ou morte bacteriana, exercendo uma série de efeitos biológicos relevantes, que conduzem a uma reação inflamatória e à reabsorção dos tecidos mineralizados. Tendo em vista o papel da endotoxina na patogênese das lesões periapicais, os autores realizaram uma revisão da literatura específica, abordando suas atividades biológicas e a importância de sua inativação durante o tratamento de dentes portadores de necrose pulpar e lesão periapical.

UNITERMOS: Endotoxina bacteriana; Gram-negativos; Hidróxido de cálcio.

INTRODUCTION

Bacterial endotoxin (LPS) has been amply studied. In fact, interest in knowledge concerning the structure of

bacterial endotoxin, its mechanism of action, and forms of inactivation in both the clinical and laboratory studies is obvious by the fact that in the past 10 years, a total of 28.100 articles have been reported on Medline (<http://>

www.ncbi.nlm.nih.gov/PubMed). In Dentistry, much research using different *in vivo* and *in vitro* methodologies has emphasized the importance of anaerobic bacteria and endotoxin in the etiology of chronic periapical lesions^{3,9,13,19,21,25,36,42,52,57,63}. However, only few articles have evaluated the effect of the presence of LPS in root canals on apical and periapical tissues^{8,10,32,37,43,55} and some articles have reported the inactivation of LPS toxic properties after endodontic procedures both *in vivo* and *in vitro*^{1,4,6,23,37,42,48,49,55,61,68}.

ROLE OF GRAM-NEGATIVE MICROORGANISMS AND ENDOTOXIN ON ETIOLOGY OF PERIAPICAL LESIONS

When dental pulp is exposed to the oral cavity due to caries or trauma, it is initially contaminated by predominantly aerobic and facultative microorganisms. Due mainly to the existing nutritional relationships between microorganisms together with the slow decrease of oxygen tension in root canals, a microbial shift takes place leading to a predominance of anaerobic microorganisms⁶⁰. Since the 1980's, technical advances in microbiological culture and identification have shown that anaerobic microorganisms predominate in root canals of teeth with pulp necrosis and radiographically visible chronic periapical lesion^{29,30,51,60}, especially Gram-negative⁴. This polymicrobial infection is located not only in the lumen of the root canal and dentinal tubules, but also in apical craters and the entire root canal system^{29,30,58}.

Gram-negative microorganisms have different virulence factors²⁶ and form products and sub-products that are toxic to apical and periapical tissues. They also contain endotoxin in their cell wall³³.

Endotoxin, present on all Gram-negative bacteria, is composed of polysaccharides (polymerized sugars), lipids (complexes containing fatty acids) and proteins. Endotoxin can be named lipopolysaccharide (LPS), emphasizing its chemical structure^{46,65}. Lipid A is the region of the endotoxin molecule responsible for its toxic effects^{11,27,33,34,65}. In 1993, Raetz⁴⁵ published a short review about the synthesis of lipid A and classified the endotoxins as extraordinary lipids.

Besides the chemical structure, much has been studied about the mechanism of action of endotoxins. When free to act, endotoxins do not cause cell or tissue lesions directly, but they stimulate competent cells to release chemical mediators. Researches showed that macrophages are the main target of endotoxins. Thus, endotoxins are not intrinsically venoms. Their effects depend on the host's response, as reported by Lewis Thomas, in *The Lives of a Cell*: "This oppressive uncontrolled and autodestructive behavior of the host is what makes endotoxin a venom." Furthermore, the same autor wrote: "Endotoxins are read by our tissues as the worst of news. When in contact with an endotoxin, our organism places all of its defenses at disposal with the idea to bombard, block and destroy all the tissue in the area. This appears to generate panic"⁴⁶.

During endodontic treatment this is particularly

significant because endotoxin (LPS) is released during multiplication or bacterial death causing a series of biological effects^{4,33}, which lead to an inflammatory reaction⁴⁶ and periapical bone resorption^{59,67}.

Even though the bacterial etiology of periapical lesions has already been proven since the classic study of Kakehashi, et al.²⁴, few investigations have evaluated the isolated effect of LPS in contact with apical and periapical tissues^{8,10,32,37,43,55}.

Among all animals, humans are the most sensitive to the effects of endotoxins⁶⁶, which makes the knowledge of their biological effects on tissues fundamentally important. Endotoxins from vital or nonvital, whole or fragmented bacteria act on macrophages⁴⁶, neutrophils³⁵ and fibroblasts⁹, leading to the release of a large number of bioactive or cytokine chemical inflammatory mediators³³, such as tumor necrosis factor (TNF)^{5,33,68}, interleukin-1 (IL-1)^{31,33,68}, IL-5³¹, IL-8³¹, alpha-interferon³³ and prostaglandins⁷. Furthermore, LPS is cytotoxic²⁰ and acts as a potent stimulator of nitric oxide production^{5,68}.

LPS also activates Hageman factor (factor XII of coagulation), has a lethal effect on animals³³, induces fever²¹, activates the complement system^{7,21,34}, thus acting in inflammatory response reactions by increasing vascular permeability, neutrophil and macrophage chemotaxis, lysozyme and lymphokine release³⁴, activation of the metabolic cycle of arachidonic acid^{7,33} being mitogenic for B lymphocytes³³ and causing mastocyte degranulation¹⁸. In infected root canals, endotoxin can contribute to an increased release of vasoactive neurotransmitter substances in the region of the nerve endings in periapical tissues, causing pain⁵¹.

According to Torabinejad, et al.⁶², the products of arachidonic acid metabolism and the activation of the complement system play an important role in bone resorption that is associated with periapical lesions in human teeth.

Besides causing an inflammatory reaction, LPS adheres irreversibly to mineralized tissues acting as a potent stimulator of bone resorption^{52,67}, acting on the synthesis and release of cytokines that activate osteoclasts^{22,23}, such as IL-1 and TNF, and stimulates the release of prostaglandin-E2 that also influences osteoclasts^{48,64}. In tissue culture, Nair, et al.³⁶ observed stimulation of bone resorption by endotoxin, confirming the role of LPS in the pathogenesis of periapical lesions seen by others^{4,10,32,52,59,67}.

Considering the discussed above, the major objective of the dental professional during treatment of root canals of permanent teeth with pulp necrosis and chronic periapical lesion should not be only bacterial death, but also the inactivation of lipid A, which is the toxic portion of endotoxin. This objective is not reached by using root canal antibacterial dressings, which only kill the bacteria remaining in the root canal system after biomechanical preparation.

Medical and dental literature have published studies that have attempted to obtain a medication or substance that inactivates bacterial endotoxin, eliminating its biologically toxic potential. Caustic soda^{10,39}, polymyxin B⁴⁴, neutrophilic enzymes³⁵, lysozyme⁴¹, formocresol⁵⁰, 1.25% chlorhexidine¹,

and sodium hypochlorite⁶ have been tested, with no significant results. Many of these products present inherent limitations due to their high toxicity causing damaging effects when in contact with vital tissues. Thus, their routine clinical use is limited. The action of laser on periapical bacterial biofilm has also been tested², however, its use is limited by the fact that there is no free access to the sites where the endotoxin is present, the root canal system of infected teeth, except when apical surgery is performed.

ROLE OF CALCIUM HYDROXIDE IN THE INACTIVATION OF BACTERIAL ENDOTOXIN

The first reference⁴⁰ to the introduction of calcium hydroxide in dentistry was in 1838. However, its clinical use progressed only after the studies by Hermann¹⁶ in 1920. Calcium hydroxide, which has a highly alkaline pH, has been used in numerous different clinical situations, i.e., direct pulp protection, pulpotomy in permanent or deciduous teeth, root canal dressing in the treatment of permanent teeth with incomplete rhizogenesis, filling sealer in root canals, root perforations, dental resorptions, and antiseptic intracanal dressing⁵⁶. This ample use has been attributed to its antibacterial activity^{12,29,30}, biocompatibility^{15,17,38,54}, hygroscopic property, ability to reduce periapical exudate¹⁶, and its capacity to induce mineralization^{28,54} and to dissolve necrotic tissue remnants after biomechanical preparation that can act as bacterial substrate¹⁴ leading to the stimulation of apical and periapical repair of teeth with chronic lesions.

Currently, one of the concerns of Endodontics is the treatment of teeth with pulp necrosis and periapical lesion because treatment failure is higher than in cases without periapical lesion. In teeth with chronic periapical lesion, there is a greater prevalence of Gram-negative anaerobic bacteria disseminated throughout the root canal system (dentinal tubules, apical craters and cementum lacunae), including apical bacterial biofilm. Because these areas are not reached by instrumentation, the use of an root canal dressing is recommended to aid in the elimination of these bacteria and increase the possibility of clinical success^{25,28,38,63}.

According to Leonardo, et al.³⁰, teeth with and without radiographically visible periapical lesion are different pathological entities requiring different treatment. In the first case, they recommend the use of an root canal dressing between treatment sessions, because the success of treatment in cases with a periapical lesion is directly related to the elimination of bacteria, products and subproducts from the root canal system. The procedures and medicaments used in root canal treatment should not only lead to bacterial death, but also to the inactivation of bacterial endotoxin.

Because of the lack of information concerning the effect of intracanal dressings on residual LPS that may adhere to mineralized tissues⁸, Safavi and Nichols⁴⁸ evaluated *in vitro* the effect of calcium hydroxide on bacterial LPS. They concluded that calcium hydroxide hydrolyzes the highly toxic lipid A molecule that is responsible for the damaging effects of endotoxin. In a later study, Safavi and Nichols⁴⁹

concluded that calcium hydroxide transforms lipid A into fatty acids and amino sugars which are atoxic components. These results were confirmed in recent studies by Barthel, et al.⁴ and Olsen, et al.⁴² who reported that calcium hydroxide detoxifies bacterial LPS *in vitro*.

In 2002, Nelson-Filho, et al.³⁷ carried out an *in vivo* study to evaluate radiographically the effect of endotoxin plus calcium hydroxide on apical and periapical tissues of dog's teeth. They observed that the endotoxin caused the formation of periapical lesions after 30 days and that calcium hydroxide inactivated bacterial LPS. Silva, et al.⁵⁵ analyzed histopathologically apical and periapical tissues of dog teeth in which the root canals were filled with bacterial LPS and calcium hydroxide. They reported that LPS caused the formation of periapical lesions and that calcium hydroxide detoxified this endotoxin *in vivo*.

More recently, Tanomaru, et al.⁶¹ evaluated the effect of biomechanical preparation using different irrigating solutions and a calcium hydroxide-based root canal dressing in dog teeth containing endotoxin. Biomechanical preparation with only irrigating solutions did not inactivate the endotoxin, however, the same treatment associated with the use of the calcium hydroxide root canal dressing (Calen[®], SS White Artigos Dentários Ltda – RJ - Brasil) was effective in the inactivation of the toxic effects of this endotoxin. With the objective of evaluating the production of TNF- α , IL-1 and nitrite in cultures of human monocytes incubated with different concentrations of LPS and associated with the calcium hydroxide-based paste (Calen[®]) or pure calcium hydroxide, Zuccolotto⁶⁸ showed that calcium hydroxide was capable of inactivating LPS.

Jiang, et al.²³ also evaluated the direct effects of LPS on osteoclastogenesis and the capacity of calcium hydroxide to inhibit the formation of osteoclasts stimulated by endotoxin. They reported that calcium hydroxide significantly reduced osteoclast differentiation.

This new knowledge has revolutionized concepts about root canal dressings, indicating calcium hydroxide as not only the medicament most indicated, but fundamentally the only one currently capable of inactivating the endotoxin present in the root canal system of teeth with pulp necrosis and chronic periapical lesion.

CONCLUSIONS

- Bacterial endotoxin (LPS), which is a component of Gram-negative cell wall, is present in all teeth with pulp necrosis and radiographically visible chronic periapical lesion. It plays fundamental role in the genesis and maintenance of periapical lesions due to the induction of inflammation and bone resorption;

- Calcium hydroxide inactivates the toxic effects of bacterial endotoxin, *in vitro* and *in vivo*, and is currently the only clinically effective medicament for inactivation of endotoxin.

REFERENCES

- 1- Aibel K, Stevens R. Effect of chlorhexidine on IL-6 induction by LPS. *J Endod* 1999; 25:282.
- 2- Araki AT, Lage-Marques JL, Ibaraki Y, Kawakami K. Ação do laser de Er:YAG no biofilme bacteriano periapical. *RPG* 1998; 5: 319.
- 3- Assed S, Ito IY, Leonardo MR, Silva LAB, Lopatin D. Anaerobic microorganisms in root canals of human teeth with chronic apical periodontitis detected by immunofluorescence. *Endod Dent Traumatol* 1996; 12:66-9.
- 4- Barthel CR, Levin LG, Reisner HM, Trope M. TNF-alpha in monocytes after exposure to calcium hydroxide treated *Escherichia coli* LPS. *Int Endod J* 1997; 30: 155-9.
- 5- Blix IJS, Helgeland K. LPS from *Actinobacillus actinomycetemcomitans* and production of nitric oxide in murine macrophages J774. *Eur J Oral Sci* 1998; 106: 576-81.
- 6- Buttler TK, Crawford JJ. The detoxifying effect of varying concentrations of sodium hypochlorite on endotoxins. *J Endod* 1982; 8: 59-66.
- 7- Cotran RS, Kumar V, Robbins SL. Robbins - patologia estrutural e funcional. Rio de Janeiro: Guanabara Koogan, 1991.
- 8- Dahlén G, Magnusson BC, Moller A. Histological and histochemical study of the influence of lipopolysaccharide extracted from *Fusobacterium nucleatum* on the periapical tissues in the monkey *Macaca fascicularis*. *Archs Oral Biol* 1981; 26:591-8.
- 9- Day AE, Langkamp HH, Bowen LL, Ascencio F, Agarwal S, Piesco NP. Signal transduction during LPS-mediated activation of pulp fibroblasts. *J Dent Res* 1998; 77:673.
- 10- Dwyer TG, Torabinejad M. Radiographic and histologic evaluation of the effect of endotoxin on the periapical tissues of the cat. *J Endod* 1981; 7:31-5.
- 11- Galanos C. Physical state and biological activity of lipopolysaccharides. Toxicity and immunogenicity of the lipid A component. *Z Immun - Forsch Bd* 1975; 149:214-29.
- 12- Georgopoulou M, Kontakiotis E, Nakou M. *In vitro* evaluation of the effectiveness of calcium hydroxide and paramonochlorophenol on aerobic bacteria from the root canal. *Endod Dent Traumatol* 1993; 9:249-53.
- 13- Hashioka K, Yamasaki M, Nakane A, Horiba N, Nakamura H. The relationship between clinical symptoms and anaerobic bacteria from infected root canals. *J Endod* 1992; 18:558-61.
- 14- Hasselgren G, Olsson B, Cvek M. Effects of calcium hydroxide and sodium hypochlorite on the dissolution of necrotic porcine muscle. *J Endod* 1998; 14:125-7.
- 15- Heithersay GS. Stimulation of root formation in incompletely developed pulpless teeth. *Oral Surg* 1970; 29:620-30.
- 16- Hermann BW. Calcium hydroxide as mitted zin behandeend und fullen von wurzel. Diss. Wurzburg., 1920 apud Leonardo MR, Leal JM. *Endodontia: tratamento de canais radiculares*. São Paulo: Panamericana, 1991:1-18.
- 17- Holland R, Souza V. Tratamento conservador da polpa dental. In: Leonardo MR, Leal JM. *Endodontia: tratamento de canais radiculares*. São Paulo: Ed Médica Panamericana, 1998:63-75.
- 18- Hook WA, Snyderman R, Mergenhagen SE. Histamine releasing factor generated by the interaction of endotoxin with hamster serum. *Infect Immun* 1970; 2:462-7.
- 19- Horiba N, Maekawa Y, Abe Y, Ito M, Matsumoto T, Nakamura H. Correlations between endotoxin and clinical symptoms or radiolucent areas in infected root canals. *Oral Surg* 1991; 71:492-5.
- 20- Horiba N, Maekawa Y, Abe Y, Ito M, Matsumoto T, Nakamura H, Ozeki M. Cytotoxicity against various cell lines of lipopolysaccharides purified from bacteroides, fusobacterium, and veillonella isolated from infected root canals. *J Endod* 1989; 15:530-4.
- 21- Horiba N, Maekawa Y, Yamauchi Y, Ito M, Matsumoto T, Nakamura H. Complement activation by lipopolysaccharides purified from gram-negative bacteria isolated from infected root canals. *Oral Surg* 1992; 74:648-51.
- 22- Ito HO, Shuto T, Takada H, Koga T, Aida Y, Hirata M, Koga T. Lipopolysaccharides from *Porphyromonas gingivalis*, *Prevotella intermedia* and *Actinobacillus actinomycetemcomitans* promote osteoclastic differentiation *in vitro*. *Archs Oral Biol* 1996; 41:439-44.
- 23- Jiang J, Zuo J, Chen SH, Holiday LS. Calcium hydroxide reduces lipopolysaccharide-stimulated osteoclast formation. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2003; 95:348-54.
- 24- Kakehashi S, Stanley HR, Fitzgerald RJ. The effects of surgical exposures of dental pulps in germ-free and conventional laboratory rats. *Oral Surg Oral Med Oral Pathol* 1965; 20:340-9.
- 25- Katebzadeh N, Hupp J, Trope M. Histological periapical repair after obturation of infected root canals in dogs. *J Endod* 1999; 25:364-8.
- 26- Könönen E, Kanervo A, Takala A, Asikainen S, Jousimies-Somer H. Establishment of oral anaerobes during first year of life. *J Dent Res* 1999; 78:1634-9.
- 27- Kumada H, Haishima Y, Umemoto T, Tanamoto KI. Structural study on the free lipid A isolated from lipopolysaccharide of *Porphyromonas gingivalis*. *J Bacteriol* 1995; 177:2098-106.
- 28- Leonardo MR, Silva LAB, Leonardo RT, Utrilla LS, Assed S. Histological evaluation of therapy using a calcium hydroxide dressing for teeth with incompletely formed apices and periapical lesions. *J Endod* 1993; 19:348-52.
- 29- Leonardo MR, Silva LAB, Leonardo RT. Devemos usar medicação intracanal no tratamento de dentes com necrose pulpar? In: *Odontologia Integrada – atualização multidisciplinar para o clínico e o especialista*. Rio de Janeiro: Editora Pedro Primeiro Ltda, 1999. p.179-95.

- 30- Leonardo MR, Silva LAB, Leonardo RT. Tratamento de canal radicular em sessão única: crença vs. ciência. In: Feller C, Gorab R. Atualização na Clínica Odontológica. São Paulo: Artes Médicas, 2000. p.29-57.
- 31- Matsushita K, Tajima T, Tomita K, Takada H, Nagaoka S, Torii M. Inflammatory cytokine production and specific antibody responses to lipopolysaccharide from endodontopathic black-pigmented bacteria in patients with multilesional periapical periodontitis. *J Endod* 1999; 25:795-9.
- 32- Mattison GD, Haddix JE, Kehoe JC, Progulske-Fox A. The effect of *Eikenella corrodens* endotoxin on periapical bone. *J Endod* 1987; 13:559-65.
- 33- McGee JOD, Isaacson PG, Wright NA. Oxford textbook of pathology. Principles of pathology. Oxford: University Press, 1992.
- 34- Morrison B, Kline L. Activation of the classical and properdin pathways of complement by bacterial lipopolysaccharides (LPS). *J Immunol* 1977; 118:362-8.
- 35- Munford RS, Hall CL. Detoxification of bacterial lipopolysaccharides (endotoxins) by a human neutrophil enzyme. *Science* 1986; 234:203-5.
- 36- Nair BC, Mayberry WR, Dziak R, Chen PB, Levine MJ, Hausmann E. Biological effects of a purified lipopolysaccharides from *Bacteroides gingivalis*. *J. Periodont Res* 1983; 18:40-9.
- 37- Nelson-Filho P, Leonardo ML, Silva LAB., Assed S. Radiographic evaluation of the effect of endotoxin (LPS) plus calcium hydroxide on apical and periapical tissues of dogs. *J Endod* 2002; 28:694-6.
- 38- Nelson-Filho P, Silva LAB, Leonardo MR, Utrilla LS, Figueiredo F. Connective tissue response to calcium hydroxide – based root canal medicaments. *Int Endodon J* 1999; 32:303-11.
- 39- Niwa M, Milner KC, Ribi E, Rudbach JA. Alteration of physical, chemical, and biological properties of endotoxin by treatment with mild alkali. *J Bacteriol* 1969; 97:1069-77.
- 40- Nygreen JA Radgivare angaende basta sattet att varda och levara tandernas friskhet apud Martin DM, Crabb HSM. Calcium hydroxide in root canal therapy. A review. *Br Dent J* 1977; 142:277-83.
- 41- Ohno N, Takada K, Kurasawa T, Liang A, Yadomae T. Detoxification of lipopolysaccharide by lysozyme. Endotoxin and sepsis: molecular mechanism of pathogenesis, host resistance, and therapy. Proceedings of the 4th Conference of the International Endotoxin Society, 1988: p. 170-90.
- 42- Olsen MH, Difiore PM, Dixit SN, Veis A. The effects of calcium hydroxide inhibition on LPS induced release of IL-1b from human monocytes in whole blood. *J Endod* 1999; 25: 289.
- 43- Pitts DL, Williams BL, Morton Jr TH. Investigation of role of endotoxin in periapical inflammation. *J Endod* 1982; 8: 10-8.
- 44- Porro M, Rustici A, Velucchi M, Agnello D, Villa P, Guezzi P. Natural and synthetic polypeptides that recognize the conserved lipid A binding sites of lipopolysaccharides. Endotoxin and sepsis: molecular mechanism of pathogenesis, host resistance, and therapy. Proceedings of the 4th Conference of the International Endotoxin Society, 1998:316-25.
- 45- Raetz CRH. Bacterial endotoxins: extraordinary lipids that activate eucaryotic signal transduction. *J Bacteriol* 1993; 75:5745-53.
- 46- Rietschel ET, Brade H. Bacterial endotoxins. *Scientific American* 1992; 267:26-33.
- 47- Rietschel ET, Kirikae T, Schade FU, Ulmer AJ, Holst O, Brade H, Schmidt G, Mamat U, Grimmecke HD, Kusumoto S, Zahringer U. The chemical structure of bacterial endotoxin in relation to bioactivity. *Immunobiol* 1993; 187:169-90.
- 48- Safavi KE, Nichols FC. Effect of calcium hydroxide on bacterial lipopolysaccharide. *J Endod* 1993; 19:76-8.
- 49- Safavi KE, Nichols FC. Alteration of biological properties of bacterial lipopolysaccharide by calcium hydroxide treatment. *J Endod* 1994; 20:127-9.
- 50- Sant'anna AT, Ramalho LTO, Spolidório DMP. Efeito do formocresol sobre o LPS bacteriano em tecido subcutâneo de camundongos. [In: Anais da 16ª Reunião Anual da Sociedade Brasileira de Pesquisa Odontológica, - SBPqO; 1999 set 8-11; Águas de São Pedro (SP). São Paulo: SBPqO; 1999. p.36
- 51- Seltzer S, Farber PA. Microbiologic factors in endodontology. *Oral Surg* 1994; 78:634-45.
- 52- Schein B, Schilder H. Endotoxin content in endodontically involved teeth. *J Endod* 1975; 1:19-21.
- 53- Shonfeld SE, Greening AB, Glick DD, Frank AL, Simon JH, Herles SM. Endotoxin activity in periapical lesions. *Oral Surg Oral Med Oral Pathol* 1982; 53:82-7.
- 54- Silva LAB. Rizogênese Incompleta - Efeitos de diferentes pastas à base de hidróxido de cálcio na complementação radicular e na reparação periapical em dentes de cães - estudo histológico. Araraquara; 1988 [Dissertação de Mestrado. Faculdade de Odontologia da Universidade Estadual Paulista]
- 55- Silva LAB, Nelson-Filho P, Leonardo MR, Rossi MA, Pansani C.A. Effect of calcium hydroxide on bacterial endotoxin in vivo. *J Endod* 2002; 28:94-8.
- 56- Silva LAB, Assed S, Nelson-Filho P. Proteção Direta da Polpa: como fazer e o que utilizar. In: Atualização na Clínica Odontológica, São Paulo: Artes Médicas; 2002 v.2, p. 267-88.
- 57- Siqueira-Júnior JF, Rôças IN. PCR methodology as a valuable tool for identification of endodontic pathogens. *J Dent* 2003; 31:333-9.

- 58- Soares JA. Avaliação microbiológica, histopatológica e histomicrobiológica de dentes de cães com reação periapical crônica induzida, após preparo biomecânico automatizado e aplicação de curativos de demora à base de hidróxido de cálcio. Araraquara; 2003. [Tese de Doutorado - Faculdade de Odontologia da Universidade Estadual Paulista]
- 59- Stashenko P. The role of immune cytokines in the pathogenesis of periapical lesions. *Endod Dent Traumatol* 1990; 6:89-96.
- 60- Sundqvist G. Ecology of the root canal flora. *J Endod* 1992; 18: 427-30.
- 61- Tanomaru JMG, Leonardo MR, Tanomaru-Filho M, Bonetti-Filho I, Silva LAB. Effect of different irrigation solutions and calcium hydroxide on bacterial LPS. *Int Endod J* 2003; 36:733-9.
- 62- Torabinejad M, Eby WC, Naidorf IJ. Inflammatory and immunological aspects of the pathogenesis of human periapical lesions. *J Endod* 1985; 11:479-88.
- 63- Trope M, Delano EO, Orstavik D. Endodontic treatment of teeth with apical periodontitis: single vs. multivisit treatment. *J Endod* 1999; 25:345-50.
- 64- Wang CY, Stashenko P. Characterization of bone-resorbing activity in human periapical lesions. *J Endod* 1993; 19:107-11.
- 65- Westphal O. Bacterial endotoxins. *Int Archs Allergy Appl Immun* 1975; 49: 1-43.
- 66- Wolff S. Biological effects of bacterial endotoxins in man. *J Infect Dis* 1973; 128: S259-S269.
- 67- Yamasaki M, Nakane A, Kumazawa M, Hashioka K, Horiba N, Nakamura H. Endotoxin and gram-negative bacteria in the rat periapical lesions. *J Endod* 1992: 501-4.
- 68- Zuccolotto CEBG. Detecção de TNF- α , IL-1 e Nitrito produzidos em cultura de monócitos expostos à endotoxina (LPS), associada ou não ao hidróxido de cálcio. Ribeirão Preto; 2003. [Dissertação de Mestrado da Faculdade de Odontologia da Universidade de São Paulo.