

Oxidative stress: therapeutic approaches for cervical cancer treatment

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Silva GA, Nunes RA, Morale MG, Boccardo E, Aguayo F, Termini L. Oxidative stress: therapeutic approaches for cervical cancer treatment. Clinics. 2018;73(suppl 1):e548s

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OBJECTIVES: Oxidative stress results from an imbalance between the generation and elimination of oxidant species. This condition may result in DNA, RNA and protein damage, leading to the accumulation of genetic alterations that can favor malignant transformation. Persistent infection with high-risk human papillomavirus types is associated with inflammatory responses and reactive oxygen species production. In this context, oxidative stress, chronic inflammation and high-risk human papillomavirus can act in a synergistic manner. To counteract the harmful effects of oxidant species, protective molecules, known as antioxidant defenses, are produced by cells to maintain redox homeostasis. In recent years, the use of natural antioxidants as therapeutic strategies for cancer treatment has attracted the attention of the scientific community. This review discusses specific molecules and mechanisms that can act against or together with oxidative stress, presenting alternatives for cervical cancer prevention and treatment.

KEYWORDS: Tumor Markers; Human Papillomavirus; Redox System; Cervical Cancer.

INTRODUCTION

Oxidative stress: general aspects

Oxidative stress (OS) results from an imbalance in the formation and elimination of oxidant species. Accumulation of these molecules may lead to cell dysfunction as a consequence of accumulated oxidative modifications in several biomolecules (1). Free radical generation represents a continuous physiological process that results from biological functions, including metabolism and inflammation. In the mitochondria, cytochrome P450 and peroxisomes are the major endogenous factors leading to reactive oxygen species (ROS) and reactive nitrogen species (RNS) formation. Other exogenous factors, such as radiation, tobacco smoking, chemotherapy and diet, are also important inductors of free radical production (2). Intermediate reactive species that are naturally produced under physiological conditions have a crucial role in metabolic regulation, the cell cycle and intracellular signaling pathways (3).

To maintain redox homeostasis, protective molecules called “antioxidant defenses” act to preserve the balance between formation and removal of ROS and RNS. Cellular antioxidant systems are currently divided into enzymatic and non-enzymatic groups. The enzymatic group comprises catalase, superoxide dismutase (SOD), glutathione peroxidase (Gpx) and glutathione-S-transferase (GST). The non-enzymatic group is composed of molecules such as vitamins C and E, lipoic acid, carotenoids, flavonoids and others (4).

Excess levels of unneutralized free radicals and cellular active intermediates are the major cause of OS (5-7). The presence of high concentrations of oxidized biomolecules is associated with alterations in aerobic metabolism, inflammatory response, exposure to UV radiation, hypoxia, anomalous cell proliferation and viral infections, among others. Therefore, OS is directly associated with several pathological conditions including tumors associated with human papillomavirus (HPV) infection (Figure 1) (8-10).

Human papillomavirus infection and cancer

Persistent infection with high-risk HPV types (HR-HPV) is the main etiological cause for the development of several epithelial tumors at different anatomic locations. The most strongly HPV-associated malignancy is cervical carcinoma, where almost all tumors are positive for HR-HPV DNA (11). Cervical carcinoma is the second most common type of cancer in women worldwide, with approximately 530,000 new cases and 270,000 deaths per year, of which more than 85% occur in developing countries. HPV infection is

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No potential conflict of interest was reported.

Received for publication on February 14, 2018. **Accepted for publication on** September 24, 2018

Commemorative Edition: 10 years of ICESP

DOI: 10.6061/clinics/2018/e548s

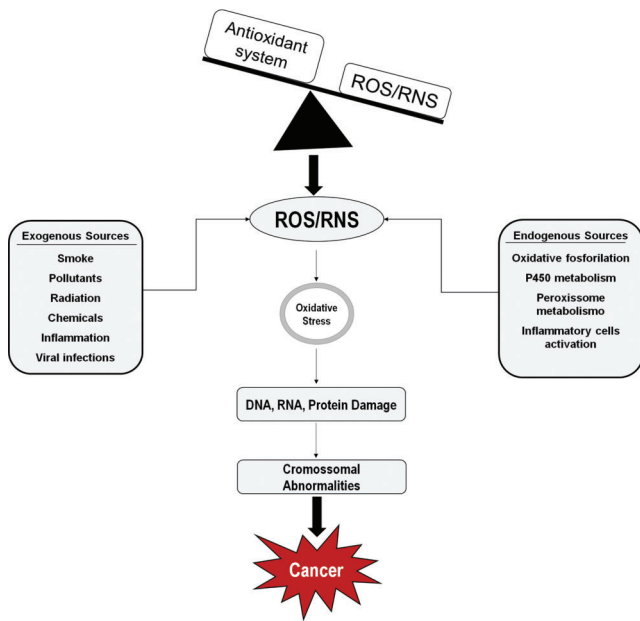


Figure 1 - Several factors, such as environmental pollutants, chemicals, tobacco smoking, chronic inflammation, among others, generate reactive oxygen and nitrogen species (ROS/RNS). An imbalance between oxidant species and the antioxidant system results in DNA, RNA and protein damage, which may lead to the accumulation of genetic alterations and promote malignant transformation. Sources of ROS/RNS.

implicated in a variable, although consistently high, proportion of vaginal, vulvar, anal, penile and head and neck carcinomas (12,13).

In addition to HPV infection, several cofactors contribute to cervical cancer development. These include low socioeconomic status, early sexual initiation, multiple sexual partners, smoking, multiparity, immunosuppression and use of oral contraceptives (14,15). Cervical cancer is mostly a consequence of the continuous evolution of non-invasive precursor lesions called cervical intraepithelial neoplasia (CIN) that are characterized by different degrees of cellular atypia (dysplasia) (16). CIN is divided into the following groups: CIN 1, characterized by mild dysplasia; CIN 2, which represents moderate dysplasia; and CIN 3 or carcinoma in situ, characterized by a severe dysplasia that may progress to an invasive squamous cell carcinoma (SCC). Moreover, changes in the glandular epithelium of the cervix, caused by HPV and other cofactors, are associated with the development of cervical adenocarcinoma (16,17). Furthermore, in persistent viral infection, HPV-induced carcinogenesis involves genetic and epigenetic changes that affect the expression of different cellular proto-oncogenes and tumor suppressor genes. Usually, this process requires an extensive period to accumulate sufficient alterations to trigger and sustain tumor development (18,19). Therefore, immune system evasion and HPV persistence are crucial factors for tumorigenesis (20). This is highlighted by the fact that the great majority of HPV infections are self-limited and spontaneously resolved in few months, and cancer development affects only a small proportion of infected individuals.

E6 and E7 viral oncoproteins have a central role in tumor development. In high-grade lesions and tumors, these proteins are upregulated mainly due to the loss or disruption

of the viral E2 gene (see below), which normally restricts the expression of viral oncogenes (21). The HPV oncoproteins E6 and E7 interact with several tumor suppressor factors and cell cycle regulators, such as p53 and pRB, respectively. These interactions lead to their deregulation and cell immortalization. In addition, E6 and E7 affect many other cellular proteins with several regulatory activities, promoting immune evasion, resistance to apoptosis and epithelial cell proliferation (22). The pleiotropic functions of these oncoproteins deserve to be noted, since they result from a well-adapted evolutionary relationship between virus and host that is essential for HPV tumorigenesis (23). Persistent viral infection, abnormal metabolism of keratinocytes expressing HPV oncogenes and non-effective chronic inflammatory responses lead to ROS production (24) and cause OS, contributing to the cell transformation process (25).

Alterations in expression and activity of some antioxidant proteins, including peroxiredoxins, catalase, quinone oxidoreductase-1 and superoxide dismutase (SOD) family proteins, can be detected in pre-neoplastic and neoplastic tissues associated with HPV infections. For example, expression of SOD2, a crucial antioxidant enzyme responsible for controlling the redox status of normal and tumor cells, is upregulated in several HPV-associated tumors, including penile and cervical carcinomas (26-29). Furthermore, results from different studies have established that the HR-HPV E6 and E7 oncoproteins can modulate OS to favor the accumulation of mutations, a fact that is directly related to cell transformation (23,30,31).

Oxidative Stress and inflammasomes

Inflammasomes are cytosolic multiprotein complexes that assemble after exposure to pathogens or danger-associated molecular patterns, leading to caspase activation and, consequently, secretion of inflammatory cytokines and cell death. Although different inflammasome complexes have been described, with unique activation triggers, they typically consist of a cytosolic pattern recognition receptor (PRR) such as RIG-I-like receptor (RLR), AIM2-like receptor (ALR) or nucleotide-binding domain and leucine-rich repeat-containing (NLR) protein, an adaptor protein (ASC) and pro-caspase-1. The OS generated during HPV infections can be modulated by the infected keratinocytes as well as by activated neutrophils and macrophages (32-34). Moreover, ROS regulate inflammasomes in response to oncogenic viral infections, such as HR-HPV (35-37). Inflammasomes are important because they activate caspase-1 in response to invading pathogens. Therefore, caspase-1 cooperates with apoptosis-associated speck-like protein containing a C-terminal caspase-recruitment domain (ACS), processing the pro-inflammatory cytokines interleukin 1 beta (IL-1 β) and interleukin 18 (IL-18) into their mature forms (38).

Furthermore, protein absent in melanoma-2 (AIM2) activates caspase-1 and inflammasomes by binding to foreign cytoplasmic double stranded DNA (dsDNA) and its adaptor ACS (39-41). Interestingly, HPV16 activates AIM2 inflammasomes and promotes upregulation of proinflammatory cytokines such as IL-1 β , IL-1 α , and IL-18, which are increased in lesions that evolve to cervical cancer (36).

Oxidative Stress and Nuclear Factor-kappa B

Nuclear factor-kappa B (NF- κ B) is a pleiotropic transcription factor composed of dimers of five different members of



the Rel transcription factor family that include p105/p50, p100/p52, RelA, RelB and cRel (42). This factor is found in the cytoplasm from the majority of resting cells, and after activation by a wide range of signals, such as viral infections, bacterial lipopolysaccharides (LPS), DNA damage, OS and proinflammatory cytokines, this protein translocates to the nucleus and regulates the expression of hundreds of genes (42-44). Genes encoding cytokines and chemokines, growth factors, cell adhesion molecules, anti-apoptotic proteins and antioxidant enzymes are regulated by NF- κ B (45,46).

NF- κ B plays a central role in inflammation and immune responses but also acts in different cellular processes, including development, cell growth, survival and proliferation. Furthermore, this transcription factor has a role in many pathological conditions, including cancer, where it may be involved in tumor growth and metastasis (42,44).

ROS levels can be affected by NF- κ B activity through its influence on the expression of antioxidant proteins, including SOD1, SOD2, Ferritin Heavy Chain (FHC), Glutathione S-transferase pi (GST-pi), Glutathione peroxidase-1 (Gpx1) and Dihydrodiol dehydrogenase (DDH1). However, ROS may activate or inhibit NF- κ B signaling depending on the signaling pathway step (upstream or downstream) and cell type analyzed (44,47-49). A recent study showed that Pirin, another NF- κ B regulator and OS sensor, is over-expressed in cervical cancer-derived cell lines and in oral mucosal keratinocytes expressing HPV16 oncogenes (50). This observation shows another mechanism for HR-HPV-dependent NF- κ B activation in cervical cancer cells and further supports the potential use of this factor as a therapeutic target.

NF- κ B activation by free radicals or enzymes is a complex event and can cause various biological effects. For example, its activation by SOD2 has been associated with lung adenocarcinoma progression and poor prognosis (51). Additionally, OS generated in tumors can impair NF- κ B translocation in thymic T cells, which become vulnerable to tumor necrosis factor alpha (TNF α)-mediated apoptosis.

Interestingly, curcumin, a natural antioxidant that is discussed below, could prevent tumor-induced thymic atrophy, restoring NF- κ B activity, and thus acted as an immunorestorative compound (43).

Oxidative Stress and DNA damage

A crucial event in HPV-mediated malignant transformation is the integration of HR-HPV DNA into the cell genome. This event results in E6 and E7 oncogene overexpression, which leads to the destabilization of p53 and pRb tumor suppressor proteins, respectively (52). HR-HPV integration is facilitated by generation of DNA damage/double strand breaks (DSBs) (53). Notably, DNA damage causes DNA oxidation, leading to a plethora of genome alterations, including deletions, insertions, point mutations, DSBs and translocations (54,55).

Genome oxidative damage was previously shown to be strongly involved in epigenetic alterations (56). In an elevated oxidative DNA damage environment, the production of 8-oxo-2'-deoxyguanosine (major product of DNA oxidation) coincides with increased HPV infection, viral-host integration and cervical dysplasia (57). Furthermore, HR-HPV integration into the cell genome generally causes the disruption of the HPV E2 ORF, resulting in E2 gene inactivation and upregulation of E6 and E7 transcription (53).

DNA repair pathways can be affected by HPV oncoproteins, resulting in the inactivation of important groups of genes, such as base excision repair (BER), nucleotide excision repair (NER), DNA mismatch repair (MMR), microhomology-mediated end joining (MMEJ), Fanconi anemia (FA), ataxia-telangiectasia mutated (ATM), and the ATM and Rad3-related (ATR) genes (58). These pathways are implicated in HR-HPV-mediated cervical cancer (59,60).

During HR-HPV infection, apoptosis is hindered by disruption of many regulatory pathways, which results in altered cell proliferation associated with accumulation of mutations and alterations in gene expression (24). In HPV16-related invasive cervical cancer, E2 binding sites (E2BSs) at the viral long control region (LCR) are epigenetically regulated by hypermethylation, dampening E2 protein binding and resulting in the upregulation of E6 and E7 expression (61-63). Moreover, DNA methylation functions as a barrier protecting the virus from immune surveillance (64,65). In summary, theoretical considerations suggest that OS, chronic inflammation, epigenetic alterations and HR-HPV can act in a synergistic manner during oncogenesis.

■ THERAPEUTIC APPROACHES

Oxidative stress control: natural antioxidants

Recently, natural compounds with chemopreventive and chemotherapeutic properties, which have antioxidant features, have received increased attention (66). An inverse association was observed between antioxidant factors present in the diet and HPV tumorigenesis, suggesting that natural antioxidants may protect against HPV persistence and tumor development (67,68). Furthermore, studies using natural compounds present in plant extracts, such as curcumin and resveratrol, have been shown to sensitize tumor cells to radio- and chemotherapy. These observations indicate that natural antioxidants have the potential to reduce the establishment and progression of precursor lesions. Molecules with these properties may be used as a complementary approach for cancer treatment (69,70).

Curcumin

Curcumin, also known as diferuloylmethane (Figure 2A), is one of the main compounds present in turmeric (*Curcuma longa* - Zingiberaceae family) rhizome (71). Moreover, curcumin scavenges ROS and RNS due the activity of the OH or CH₂ group of the β -diketone moiety. In other words, curcumin reacts with free radicals by electron transference followed by proton loss or direct H-atom abstraction, mainly due to the phenolic OH group (72-74). The value of curcumin has been widely addressed and discussed in scientific literature. This compound has anti-inflammatory, antitumoral, antioxidant, and cardioprotective activities and several other properties (75-82). Curcumin can inhibit NF- κ B activation through restriction of I κ B α kinase and Akt activation (83-89). This change results in the inhibition of NF- κ B-regulated gene products that control apoptosis, proliferation, invasion and angiogenesis (90,91).

Moreover, curcumin downregulates other crucial transcription factors responsible for controlling cell growth and survival pathways, including signal transducer and activator of transcription 3 (STAT3), cyclooxygenase 2 (COX2), Akt, antiapoptotic proteins and activator protein 1 (AP1) (69,92,93). In addition, this compound can downregulate angiogenesis *in vivo*, inhibiting the activity of proangiogenic

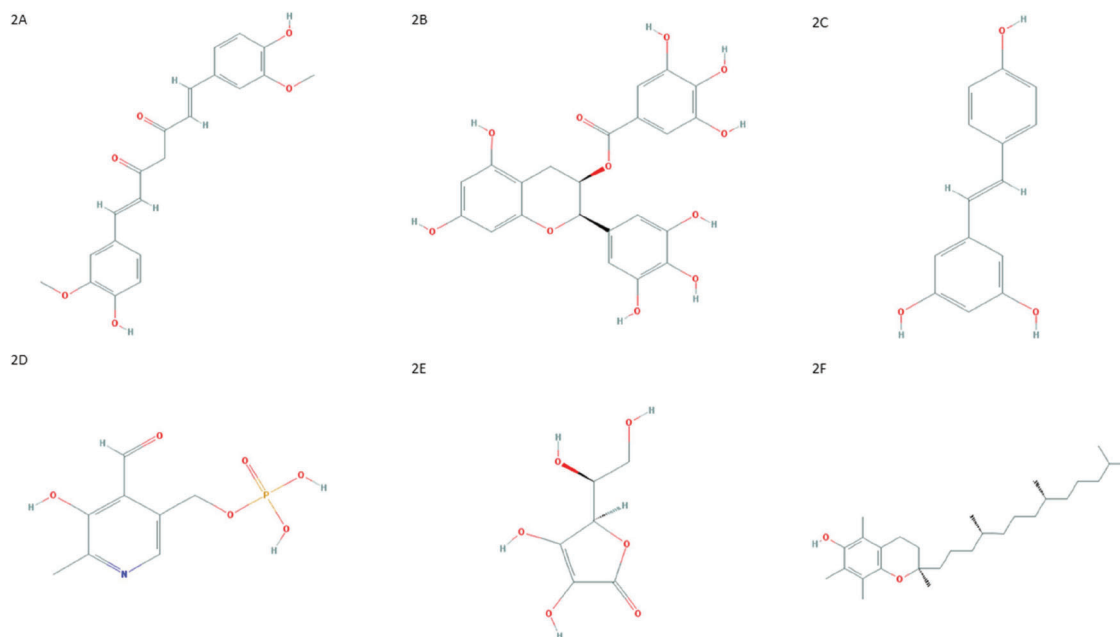


Figure 2 - (A) Chemical structure of curcumin. PubChem CID 969516; **(B)** Chemical structure of epigallocatechin-3-gallate. PubChem CID 65064; **(C)** Chemical structure of resveratrol. PubChem CID 445154; **(D)** Chemical structure of pyridoxal-5'-phosphate. PubChem CID 1051; **(E)** Chemical structure of ascorbic acid. PubChem CID 54670067; **(F)** Chemical structure of α -tocopherol. PubChem CID 14985.

molecules, such as fibroblast growth factor 2 (FGF-2), matrix metalloproteases and COX2, as noted previously (94,95). Finally, curcumin can induce the expression of antioxidant enzymes through nuclear factor erythroid 2-related factor 2 (NRF2) activation (57).

Due to its biological properties and low toxicity, curcumin is an interesting alternative to therapeutic agents used for the treatment of several types of cancer (96-98). Various studies have addressed the antitumor effects of curcumin in experimental models of cervical cancer. For instance, the HPV16-positive cervical cancer-derived cell line SiHa could be sensitized to cisplatin treatment by curcumin co-administration. This treatment inhibited NF- κ B activity and increased cell death (99). Treatment with curcumin also sensitized cervical cancer-derived cells and HPV-positive tumors to taxol and paclitaxel, respectively. Again, these effects involve the downregulation of survival signals regulated by NF- κ B and Akt, such as phosphorylation of MAPKs and induction of Cyclin-D1, COX2, X-linked inhibitor of apoptosis protein (XIAP) and cellular inhibitor of apoptosis protein-1 (cIAP1) (100-102).

In addition, curcumin was shown to downregulate HPV18 transcription in HeLa cells, inhibiting the AP1 pathway and reversing the expression patterns of the c-fos and fra-1 transcription factors (103). Furthermore, depending on the concentration and duration of treatment with curcumin, a cytotoxic effect could be observed in cervical cancer cells (104). Interestingly, another study showed that thioredoxin reductase plays a role in the radiosensitizing effect of curcumin on cervical cancer cells (105). Due to its chemopreventive activity in several types of cancer, curcumin has been evaluated in preclinical studies and clinical trials (106). A study was developed using curcumin in a specific formulation (Meriva) to investigate its impact on the quality of life of 160 cancer patients. Study participants presented solid or hematological malignancies and were undergoing

radio- or chemotherapy following surgical treatment, with significant side effects and increased oxidative stress. Despite the study limitations, which were recognized by the authors, the results provided the first clinical evidence that curcumin may reduce the side effects of cancer treatment and improve patient quality of life. Notably, curcumin has extremely low oral bioavailability, and that it is necessary to develop formulations that allow its clinical use (107).

Epigallocatechin-3-gallate (EGCG)

Another important polyphenol with anticancer properties is epigallocatechin-3-gallate (EGCG). This compound (Figure 2B) it is the major catechin from *Camellia sinensis*, found in green tea (108). This potent antioxidant scavenges ROS and has antiproliferative, antiangiogenic, antimetastatic and proapoptotic effects in several tumor models (109-119).

EGCG is a potent inhibitor of the Akt/NF- κ B and mammalian target of rapamycin (mTOR) signaling pathways inducing apoptosis and cell survival mechanisms. Moreover, EGCG can cause NRF2-mediated antioxidant induction and reduce inflammation (120). Notably, the activation of NRF2 is a key target of cytoprotective agents, and its upregulation results in increased expression of Heme oxygenase-1 (HO-1 antioxidant enzyme) (121). Furthermore, NRF2 prevention of inflammation by ROS occurs through the inactivation of NF- κ B (122). The EGCG antiproliferative effect is related to the accumulation of cells in the G1 phase of the cell cycle, followed by apoptosis (123). Interestingly, this compound can also induce apoptosis in cervical cells extracted from fresh tissues (124).

A recent study showed that EGCG exhibits free radical scavenging properties in cells isolated from cervical cancer biopsies, decreasing cell proliferation and increasing the activity of antioxidant enzymes, such as SOD and Gpx (125). In addition to targeting cell growth and apoptosis, this compound can further inhibit telomerase action, impairing



initiation and development of cervical lesions (126,127). As mentioned before, EGCG also displays antiangiogenic features through inhibition of hypoxia-inducible factor 1 alpha (HIF-1 α) and of vascular endothelial growth factor (VEGF), as a result of proteasome activity and blockage of PI3K/Akt and ERK1/2-regulated pathways (128).

Interestingly, EGCG can act synergistically with cisplatin, inhibiting the growth of cervical cancer-derived cell lines. This finding suggests that cisplatin treatment is potentiated with EGCG in HeLa cells, regulating pathways involved in cell survival and apoptosis. This combination may improve cervical cancer treatment, especially in the context of chemoresistance to cisplatin (129).

A clinical trial was carried out to verify the efficacy of two green tea extract-derived compounds, EGCG and polyphenon E, in a study involving ninety women with HPV-related cervical lesions. These patients were divided in four different groups, according to the following regimens: local application of a polyphenon E ointment, daily oral dose (200 mg) of polyphenon E, both local and oral polyphenon E and 200 mg daily oral dose of EGCG. Patients treated with EGCG or polyphenon E responded positively to the treatment, with a reduction in HPV DNA copies and regression of virus-associated lesions, while the majority of the untreated group showed no improvement (130).

Resveratrol

Resveratrol or 3,5,4-trihydroxystilbene (Figure 2C), a phytoalexin found in grapes, peanuts, blueberries and other food products, shows antitumor effects by interacting with several molecules involved in tumor development, such as first apoptosis signal receptor (Fas), MAPK, NF- κ B and AP1 (131,132).

Resveratrol exhibits antiproliferative effects on cervical cancer-derived cell lines. This effect was characterized by accumulation of cells in the S-phase of the cell cycle (70). The result was further confirmed by another study, using a double dosage of resveratrol, which induced a transient and reversible accumulation of HeLa cells in the S-phase (133). Recently, the glycoprotein cyclooxygenase pathway was shown to be a crucial target of resveratrol. However, it apparently does not directly regulate COX expression. Instead, it may regulate other enzymes related to prostaglandin synthesis acting downstream of the COX pathway (134).

The chemopreventive efficacy of resveratrol has already been demonstrated in hepatocellular, skin, prostate and lung cancers, through several regulatory pathways (135,136). Nevertheless, its exact mechanism in cervical cancer remains to be investigated, which limits its therapeutic applications. Recent data demonstrated that resveratrol inhibits HeLa cell growth and induces apoptosis in a dose- and time-dependent manner. In addition, these cells presented apoptotic characteristics, such as cell shrinkage, formation of apoptotic bodies and DNA fragmentation (137). Interestingly, resveratrol can reduce the active form of Akt, leading to autophagy, cell cycle arrest and apoptosis of SiHa and HeLa cells. Moreover, it can destabilize lysosomes, resulting in cathepsin L (cat L) translocation to the cytosol (131,134,138,139). When the cat L enzymatic activity increases in the cytosol, cytochrome c is released from the mitochondria, leading to cell death by apoptosis. Triggering apoptosis through cat L causes lysosome membrane permeabilization and release of its proteases to the cytosol. (138). Studies conducted by Rezk

et al. (140) showed that resveratrol improves the effectiveness of cisplatin and doxorubicin chemotherapy, suggesting it can be used in cervical cancer treatment.

Pyridoxal-5'-phosphate (Vitamin B6)

Pyridoxal-5'-phosphate (PLP) is the bioactive form of vitamin B6 (Figure 2D). There are three non-phosphorylated vitamin B6 precursors, catalyzed by pyridoxal-kinase (PDXK) into their phosphorylated forms, including PLP (141).

Vitamin B6 is required as a co-enzyme for many biochemical reactions, including amino acid synthesis and catabolism, bioactive amine synthesis (histamine, serotonin, dopamine), hemoglobin synthesis, and glycogenolysis (142-144). Its antioxidant properties include direct effects, due to reaction of its hydroxyl and amine groups with peroxy radicals, or indirect effects, due to its role in homocysteine to cysteine conversion. Cysteine is a necessary substrate for glutathione synthesis and is essential for an ideal redox balance (145).

Vitamin B6 could sensitize several cancer cell lines to apoptosis after cisplatin-mediated DNA damage by depleting intracellular glutathione. Furthermore, low PDXK expression was associated with poor prognosis in two independent cohorts of non-small cell lung cancer patients (146).

In a cohort of thirty-three hepatocellular carcinoma patients that randomly received placebo or vitamin B6 (50 mg/d), the results indicated that vitamin B6 improved the antioxidant capacity by reducing homocysteine levels in patient plasma (145).

Ascorbic acid (vitamin C)

Recent data have shown that vitamin C, also known as ascorbic acid (Figure 2E), is another antioxidant molecule and can increase the effect of chemotherapeutic agents without augmenting their toxicity in normal cells.

In SiHa cells, the association of vitamin C and cisplatin enhanced cisplatin-mediated apoptosis induction through a p53-mediated pathway. Therefore, low concentrations of cisplatin were required to induce cancer cell death. Hence, it is tempting to speculate that, in combination with vitamin C, low amounts of cisplatin could be used in cancer patients, reducing its side effects (147).

Interestingly, ascorbyl stearate (ASC-S), a fatty acid ester derivative from ascorbic acid, exhibits a potent proapoptotic activity. Recently, Mane et al. (148) described the proapoptotic effect of this molecule on HeLa cells due to the induction of changes in the mitochondrial membrane permeability, cytochrome c release and caspase-3 and NF- κ B activation.

Finally, the direct impact of ascorbic acid on cervical carcinogenesis has been suggested. A cross-sectional study published by Hwang et al. (149) showed that a dietary supplementation including vitamin C might reduce the risk of CIN in women with HR-HPV infection.

α -Tocopherol (vitamin E)

Results from previous studies suggest the existence of an association between plasma levels of vitamin E, also known as α -tocopherol or 5,7,8-trimethyltolcol (Figure 2F), and HPV infection status. For example, women with CIN were shown to exhibit decreased plasma levels of this antioxidant compound, which may reflect the existence of elevated OS (150,151). Similar results showed that patients with cervical cancer had low levels of vitamin E (152). Additionally,



Hu and co-workers (153) suggested that vitamin E intake or high circulating levels may reduce the risk of CIN and cervical cancer. Furthermore, a study published by Palan et al. (154) supports previous data suggesting that α -tocopherol and other antioxidants have an important protective role *in vivo*. In addition, the maintenance and regeneration of α -tocopheryl quinone (a non-oxidized form of α -tocopherol) may also have an important role in CIN and in cervical cancer. However, further investigations of α -tocopheryl quinone as a potential marker of OS in precancerous lesions and cervical cancer are strongly needed (154).

Oxidative stress induction: traditional chemotherapy

Despite the positive results with the use of the antioxidant compounds described above, traditional cancer chemotherapy still relies on the induction of OS triggered by ROS levels higher than tumor elimination capacity, which contributes to tumor cell death (155). Nevertheless, evidence has shown that upregulation of antioxidant enzymes by cancer cells has a relevant role in OS control and possibly in drug resistance (156).

One of the most important systems involved in cell protection against elevated levels of free radicals, including those induced by chemotherapy and radiotherapy, is composed of glutathione (GSH) and GSH-related enzymes (157). Regarding this issue, a study analyzed the role of the GSH redox system in drug-resistant glioblastoma multiforme cells. Temozolomide (TMZ)-resistant glioma cells showed low levels of ROS and high GSH and glutathione reductase levels, exhibiting greater antioxidant capacity than sensitive cells, suggesting that this redox system may be an important therapeutic target (158). Interestingly, HPV proteins can also modulate ROS by regulating GSH and SOD levels (159). In fact, a relationship between GSH levels in cervical cancer cells and resistance to doxorubicin treatment, an anticancer drug that induces DNA damage and ROS production, was observed (160).

An alternative treatment for cervical cancer involves molecules capable of inhibiting antioxidant pathways. For example, exposure of HeLa cells to pinostrobin, a dietary bioflavonoid, was associated to reduced cell viability and downregulated GSH and NO²⁻ levels. Pinostrobin-associated cell death involves upregulation of apoptotic extrinsic and intrinsic pathways components, as well as DNA and mitochondrial damage, probably as a consequence of ROS accumulation (161).

Another compound, auranofin, an inhibitor of thioredoxin reductase, can also trigger apoptosis while depleting GSH and increasing ROS intracellular levels. In addition, its effect on cervical cancer cell lines can be intensified in combination with a GSH synthesis inhibitor, L-buthionine sulfoximine (162). By inhibiting antioxidant effects, these treatments overcome HPV resistance to ROS induction, leading to cell death. Finally, therapeutic approaches using ROS inductors or antioxidant molecules such as SOD and SOD-mimetics combined with radiotherapy or chemotherapy have been investigated and may constitute viable alternatives for cancer treatment in the future (163).

Enzyme inhibitors

The use of enzyme inhibitors such as tyrosine kinase inhibitors (TKIs) to modulate ROS effects constitutes an

important alternative to cancer therapies currently under investigation. Sunitinib, for instance, is the most common TKI administered in clinical medicine, and it effectively blocks VEGF receptors (164-166), platelet-derived growth factor receptor (PDGFR) *alfa* and *beta*, and c-Kit (167).

According to some published data, sunitinib could have antioxidant activity due to the improved lipid peroxidation and increased GSH levels observed after cisplatin treatment, reducing OS-triggered side effects and improving chemotherapeutic efficacy (168). When combined with chloroquine, sunitinib augments NOS production, causing an accumulation of RNS and triggering apoptosis; it also increases the levels of GSH, which in turn can interrupt apoptosis (169). In addition, a combination of chloroquine and sunitinib induces increased toxicity in breast, cervical, colorectal, hepatocellular, laryngeal and prostate cancer cell lines as well as in mouse tumor models (169). However, based on the current knowledge, the potential impact of these effects on patients' outcome are still unclear.

Another approach involving enzyme inhibitors was applied in a study that developed a new radiosensitization technique using a hydrogen peroxide solution (Oxydol) named KORTUC (Kochi Oxydol-Radiation Therapy for Unresectable Carcinomas). This approach aimed to treat patients with local advanced unresectable neoplasms, through tumor antioxidative enzyme blockage, leading to tumor sensitization to radiotherapy (170). The same group published further clinical studies showing that this method may be a safe, well-tolerated, and efficient approach (170-173). However, it is also a broad field of study.

Oxidative stress is a consequence of an imbalance between pro- and antioxidant factors. The antioxidant system maintains the oxidative process within physiological limits, preventing injuries that can culminate in irreparable systemic damage and diseases.

Signaling pathways related to redox control in HPV-mediated carcinogenesis represent promising targets for cancer treatment. Some antioxidant molecules, including natural compounds, may be useful preventive or therapeutic alternatives. Despite the positive results of antioxidant compounds, OS induction is still an effective therapeutic approach used in traditional chemotherapy. This approach aims to enhance reactive species production to levels that overcome the tumor elimination capacity, leading to tumor cell death. However, evidence has shown that an increase in antioxidant enzymes by cancer cells has an important role in OS control and possibly in drug resistance.

Current knowledge indicates that several molecules and enzyme inhibitors act to modulate ROS and that they are some of the most significant therapeutic targets for anticancer compound development and should be extensively studied. Despite several lines of evidence based on *in vitro* or animal model studies, relatively few clinical trials have been carried out. These trials are essential for the establishment of new therapies targeting oxidative stress, including those using natural compounds.

■ AUTHOR CONTRIBUTIONS

Silva GA contributed to the abstract, introduction, figures, therapeutic approaches and conclusion and revised the text and references. Nunes RA contributed to preparation and revision of the text. Morale MG contributed to the introduction and revised the text and references. Aguayo F and Boccardo E revised the text. Termini L prepared and revised the text.



■ REFERENCES

- Devasagayam TP, Tilak JC, Bloor KK, Sane KS, Ghaskadbi SS, Lele RD. Free radicals and antioxidants in human health: current status and future prospects. *J Assoc Physicians India*. 2004;52:794-804.
- Bhattacharyya A, Chattopadhyay R, Mitra S, Crowe SE. Oxidative stress: an essential factor in the pathogenesis of gastrointestinal mucosal diseases. *Physiol Rev*. 2014;94(2):329-54, <http://dx.doi.org/10.1152/physrev.00040.2012>.
- Nathan C. Specificity of a third kind: reactive oxygen and nitrogen intermediates in cell signaling. *J Clin Invest*. 2003;111(6):769-78, <http://dx.doi.org/10.1172/JCI200318174>.
- Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. *Int J Biochem Cell Biol*. 2007;39(1):44-84, <http://dx.doi.org/10.1016/j.biocel.2006.07.001>.
- Sies H. Oxidative Stress: Introductory Remarks. In: *Oxidative Stress*. London: Academic Press; 1985.
- Sies H, Cadenas E. Oxidative stress: damage to intact cells and organs. *Philos Trans R Soc Lond B Biol Sci*. 1985;311(1152):617-31, <http://dx.doi.org/10.1098/rstb.1985.0168>.
- Sies H. Biochemistry of oxidative stress. *Angew Chem Int Ed Engl*. 1986;25:1058-71, <http://dx.doi.org/10.1002/anie.198610581>.
- Halliwell B, Whiteman M. Measuring reactive species and oxidative damage in vivo and in cell culture: how should you do it and what do the results mean? *Br J Pharmacol*. 2004;142(2):231-55, <http://dx.doi.org/10.1038/sj.bjp.0705776>.
- Xu Y, Fang F, Dhar SK, Bosch A, St Clair WH, Kasarskis EJ, et al. Mutations in the SOD2 promoter reveal a molecular basis for an activating protein 2-dependent dysregulation of manganese superoxide dismutase expression in cancer cells. *Mol Cancer Res*. 2008;6(12):1881-93, <http://dx.doi.org/10.1158/1541-7786.MCR-08-0253>.
- Gallagher CJ, Ahn K, Knipe AL, Dyer AM, Richie JP Jr, Lazarus P, et al. Association between haplotypes of manganese superoxide dismutase (SOD2), smoking, and lung cancer risk. *Free Radic Biol Med*. 2009;46(1):20-4, <http://dx.doi.org/10.1016/j.freeradbiomed.2008.09.018>.
- Bosch FX, Broker TR, Forman D, Moscicki AB, Gillison ML, Doorbar J. Comprehensive Control of HPV Infections and Related Diseases Vaccine. 2013;31 Suppl 6:G1-31.
- Woodman CB, Collins SL, Young LS. The natural history of cervical HPV infection: unresolved issues. *Nat Rev Cancer*. 2007;7(1):11-22, <http://dx.doi.org/10.1038/nrc2050>.
- Ferlay J, Soerjomataram I, Ervik M, Dikshit R, Eser S, Mathers C, et al. GLOBOCAN 2012 v1.0. Cancer Incidence and Mortality Worldwide: IARC CancerBase No. 11. Worldwide: IARC CancerBase No. 11 [Internet]. Lyon, France: International Agency for Research on Cancer; 2013. Available from: <http://globocon.iarc.fr>.
- Bosch FX, Lorincz A, Muñoz N, Meijer CJ, Shah KV. The causal relation between human papillomavirus and cervical cancer. *J Clin Pathol*. 2002;55(4):244-65, <http://dx.doi.org/10.1136/jcp.55.4.244>.
- Cotton SC, Sharp L, Seth R, Masson LF, Little J, Cruickshank ME, et al. Lifestyle and socio-demographic factors associated with high-risk HPV infection in UK women. *Br J Cancer*. 2007;97(1):133-9, <http://dx.doi.org/10.1038/sj.bjc.6603822>.
- zur Hausen H. Papillomaviruses and cancer: from basic studies to clinical application. *Nat Rev Cancer*. 2002;2(5):342-50, <http://dx.doi.org/10.1038/nrc798>.
- Bruni L, Barrionuevo-Rosas L, Albero G, Serrano B, Mena M, Gómez D, et al. ICO/IARC Information Centre on HPV and Cancer (HPV Information Centre). *Human Papillomavirus and Related Diseases in Albania*; 2014.
- Doorbar J. Molecular biology of human papillomavirus infection and cervical cancer. *Clin Sci (Lond)*. 2006;110(5):525-41.
- Bodily J, Laimins LA. Persistence of human papillomavirus infection: keys to malignant progression. *Trends Microbiol*. 2011;19(1):33-9, <http://dx.doi.org/10.1016/j.tim.2010.10.002>.
- Crosbie EJ, Einstein MH, Franceschi S, Kitchener HC. Human papillomavirus and cervical cancer. *Lancet*. 2013;382(9895):889-99, [http://dx.doi.org/10.1016/S0140-6736\(13\)60022-7](http://dx.doi.org/10.1016/S0140-6736(13)60022-7).
- von Knebel Doeberitz M. New markers for cervical dysplasia to visualise the genomic chaos created by aberrant oncogenic papillomavirus infections. *Eur J Cancer*. 2002;38(17):2229-42, [http://dx.doi.org/10.1016/S0959-8049\(02\)00462-8](http://dx.doi.org/10.1016/S0959-8049(02)00462-8).
- Doorbar J, Quint W, Banks L, Bravo IG, Stoler M, Broker TR, et al. The biology and life-cycle of human papillomaviruses. *Vaccine*. 2012;30 Suppl 5:F55-70, <http://dx.doi.org/10.1016/j.vaccine.2012.06.083>.
- Moody CA, Laimins LA. Human papillomavirus oncoproteins: pathways to transformation. *Nat Rev Cancer*. 2010;10(8):550-60, <http://dx.doi.org/10.1038/nrc2886>.
- Kgatle MM, Spearman CW, Kalla AA, Hairwadzi HN. DNA Oncogenic Virus-Induced Oxidative Stress, Genomic Damage, and Aberrant Epigenetic Alterations. *Oxid Med Cell Longev*. 2017;2017:3179421, <http://dx.doi.org/10.1155/2017/3179421>.
- Williams VM, Filippova M, Soto U, Duerksen-Hughes PJ. HPV-DNA integration and carcinogenesis: putative roles for inflammation and oxidative stress. *Future Virol*. 2011;6(1):45-57, <http://dx.doi.org/10.2217/fvl.10.73>.
- Holley AK, Dhar SK, Xu Y, St Clair DK. Manganese superoxide dismutase: beyond life and death. *Amino Acids*. 2012;42(1):139-58, <http://dx.doi.org/10.1007/s00726-010-0600-9>.
- Kinnula VL, Crapo JD. Superoxide dismutases in malignant cells and human tumors. *Free Radic Biol Med*. 2004;36(6):718-44, <http://dx.doi.org/10.1016/j.freeradbiomed.2003.12.010>.
- Termini L, Filho AL, Maciag PC, Etlinger D, Alves VA, Nonogaki S, et al. Deregulated expression of superoxide dismutase-2 correlates with different stages of cervical neoplasia. *Dis Markers*. 2011;30(6):275-81, <http://dx.doi.org/10.1155/2011/178475>.
- Termini L, Fregani JH, Boccardo E, da Costa WH, Longatto-Filho A, Andreoli MA, et al. SOD2 immunoeexpression predicts lymph node metastasis in penile cancer. *BMC Clin Pathol*. 2015;15:3, <http://dx.doi.org/10.1186/s12907-015-0003-7>.
- Kaina B. DNA damage-triggered apoptosis: critical role of DNA repair, double-strand breaks, cell proliferation and signaling. *Biochem Pharmacol*. 2003;66(8):1547-54, [http://dx.doi.org/10.1016/S0006-2952\(03\)00510-0](http://dx.doi.org/10.1016/S0006-2952(03)00510-0).
- Williams VM, Filippova M, Filippov V, Payne KJ, Duerksen-Hughes P. Human papillomavirus type 16 E6* induces oxidative stress and DNA damage. *J Virol*. 2014;88(12):6751-61, <http://dx.doi.org/10.1128/JVI.03355-13>.
- Levitsky V, Masucci MG. Manipulation of immune responses by Epstein-Barr virus. *Virus Res*. 2002;88(1-2):71-86, [http://dx.doi.org/10.1016/S0168-1702\(02\)00121-1](http://dx.doi.org/10.1016/S0168-1702(02)00121-1).
- Tindle RW. Immune evasion in human papillomavirus-associated cervical cancer. *Nat Rev Cancer*. 2002;2(1):59-65, <http://dx.doi.org/10.1038/nrc700>.
- Wieland SF, Chisari FV. Stealth and cunning: hepatitis B and hepatitis C viruses. *J Virol*. 2005;79(15):9369-80, <http://dx.doi.org/10.1128/JVI.79.15.9369-9380.2005>.
- Torres MA, Jones JD, Dangel JL. Reactive oxygen species signaling in response to pathogens. *Plant Physiol*. 2006;141(2):373-8, <http://dx.doi.org/10.1104/pp.106.079467>.
- Reinholz M, Kawakami Y, Salzer S, Kreuter A, Dombrowski Y, Koglin S, et al. HPV16 activates the AIM2 inflammasome in keratinocytes. *Arch Dermatol Res*. 2013;305(8):723-32, <http://dx.doi.org/10.1007/s00403-013-1375-0>.
- Pontillo A, Bricher P, Leal VN, Lima S, Souza PR, Crovella S. Role of inflammasome genetics in susceptibility to HPV infection and cervical cancer development. *J Med Virol*. 2016;88(9):1646-51, <http://dx.doi.org/10.1002/jmv.24514>.
- Lamkanfi M, Dixit VM. Mechanisms and functions of inflammasomes. *Cell*. 2014;157(5):1013-22, <http://dx.doi.org/10.1016/j.cell.2014.04.007>.
- Du W, Zhen J, Zheng Z, Ma S, Chen S. Expression of AIM2 is high and correlated with inflammation in hepatitis B virus associated glomerulonephritis. *J Inflamm (Lond)*. 2013;10(1):37, <http://dx.doi.org/10.1186/1476-9255-10-37>.
- Han Y, Chen Z, Hou R, Yan D, Liu C, Chen S, et al. Expression of AIM2 is correlated with increased inflammation in chronic hepatitis B patients. *Virol J*. 2015;12:129, <http://dx.doi.org/10.1186/s12985-015-0360-y>.
- Zhen J, Zhang L, Pan J, Ma S, Yu X, Li X, et al. AIM2 mediates inflammation-associated renal damage in hepatitis B virus-associated glomerulonephritis by regulating caspase-1, IL-1 β , and IL-18. *Mediators Inflamm*. 2014;2014:190860, <http://dx.doi.org/10.1155/2014/190860>.
- Mercurio F, Manning AM. NF-kappaB as a primary regulator of the stress response. *Oncogene*. 1999;18(45):6163-71, <http://dx.doi.org/10.1038/sj.onc.1203174>.
- Bhattacharyya S, Mandal D, Sen GS, Pal S, Banerjee S, Lahiry L, et al. Tumor-induced oxidative stress perturbs nuclear factor-kappaB activity-augmenting tumor necrosis factor-alpha-mediated T-cell death: protection by curcumin. *Cancer Res*. 2007;67(1):362-70, <http://dx.doi.org/10.1158/0008-5472.CAN-06-2583>.
- Morgan MJ, Liu ZG. Crosstalk of reactive oxygen species and NF-kB signaling. *Cell Res*. 2011;21(1):103-15, <http://dx.doi.org/10.1038/cr.2010.178>.
- Baeuerle PA, Baichwal VR. NF-kappa B as a frequent target for immunosuppressive and anti-inflammatory molecules. *Adv Immunol*. 1997;65:111-37, [http://dx.doi.org/10.1016/S0065-2776\(08\)60742-7](http://dx.doi.org/10.1016/S0065-2776(08)60742-7).
- May MJ, Ghosh S. Signal transduction through NF-kappa B. *Immunol Today*. 1998;19(2):80-8, [http://dx.doi.org/10.1016/S0167-5699\(97\)01197-3](http://dx.doi.org/10.1016/S0167-5699(97)01197-3).
- Kabe Y, Ando K, Hirao S, Yoshida M, Handa H. Redox regulation of NF-kappaB activation: distinct redox regulation between the cytoplasm and the nucleus. *Antioxid Redox Signal*. 2005;7(3-4):395-403, <http://dx.doi.org/10.1089/ars.2005.7.395>.
- Meyer M, Schreck R, Baeuerle PA. H2O2 and antioxidants have opposite effects on activation of NF-kappa B and AP-1 in intact cells: AP-1 as secondary antioxidant-responsive factor. *EMBO J*. 1993;12(5):2005-15, <http://dx.doi.org/10.1002/j.1460-2075.1993.tb05850.x>.



49. Hirota K, Murata M, Sachi Y, Nakamura H, Takeuchi J, Mori K, et al. Distinct roles of thioredoxin in the cytoplasm and in the nucleus. A two-step mechanism of redox regulation of transcription factor NF-kappaB. *J Biol Chem.* 1999;274(39):27891-7, <http://dx.doi.org/10.1074/jbc.274.39.27891>.
50. Carrillo D, Muñoz JP, Huerta H, Leal G, Corvalán A, León O, et al. Upregulation of PIR gene expression induced by human papillomavirus E6 and E7 in epithelial oral and cervical cells. *Open Biol.* 2017;7(11):pii: 170111, <http://dx.doi.org/10.1098/rsob.170111>.
51. Chen PM, Wu TC, Wang YC, Cheng YW, Sheu GT, Chen CY, et al. Activation of NF-kB by SOD2 promotes the aggressiveness of lung adenocarcinoma by modulating NKX2-1-mediated IKKβ expression. *Carcinogenesis.* 2013;34(11):2655-63, <http://dx.doi.org/10.1093/carcin/bgt220>.
52. Hu Z, Zhu D, Wang W, Li W, Jia W, Zeng X, et al. Genome-wide profiling of HPV integration in cervical cancer identifies clustered genomic hot spots and a potential microhomology-mediated integration mechanism. *Nat Genet.* 2015;47(2):158-63, <http://dx.doi.org/10.1038/ng.3178>.
53. Chiang CM, Ustav M, Stenlund A, Ho TF, Broker TR, Chow LT. Viral E1 and E2 proteins support replication of homologous and heterologous papillomaviral origins. *Proc Natl Acad Sci U S A.* 1992;89(13):5799-803, <http://dx.doi.org/10.1073/pnas.89.13.5799>.
54. Karanjawala ZE, Murphy N, Hinton DR, Hsieh CL, Lieber MR. Oxygen metabolism causes chromosome breaks and is associated with the neuronal apoptosis observed in DNA double-strand break repair mutants. *Curr Biol.* 2002;12(5):397-402, [http://dx.doi.org/10.1016/S0960-9822\(02\)00684-X](http://dx.doi.org/10.1016/S0960-9822(02)00684-X).
55. Cooke MS, Evans MD, Dizdaroglu M, Lunec J. Oxidative DNA damage: mechanisms, mutation, and disease. *FASEB J.* 2003;17(10):1195-214, <http://dx.doi.org/10.1096/fj.02-0752rev>.
56. O'Hagan HM, Mohammad HP, Baylin SB. Double strand breaks can initiate gene silencing and SIRT1-dependent onset of DNA methylation in an exogenous promoter CpG island. *PLoS Genet.* 2008;4(8):e1000155, <http://dx.doi.org/10.1371/journal.pgen.1000155>.
57. Visalli G, Riso R, Facciola A, Mondello P, Caruso C, Picerno I, et al. Higher levels of oxidative DNA damage in cervical cells are correlated with the grade of dysplasia and HPV infection. *J Med Virol.* 2016;88(2): 336-44, <http://dx.doi.org/10.1002/jmv.24327>.
58. Boccardo E. HPV-mediated genome instability: at the roots of cervical carcinogenesis. *Cytogenet Genome Res.* 2010;128(1-3):57-65, <http://dx.doi.org/10.1159/000290657>.
59. Peitsaro P, Hietanen S, Johansson B, Lakkala T, Syrjänen S. Single copy heterozygote integration of HPV 33 in chromosome band 5p14 is found in an epithelial cell clone with selective growth advantage. *Carcinogenesis.* 2002;23(6):1057-64, <http://dx.doi.org/10.1093/carcin/23.6.1057>.
60. Pett MR, Herdman MT, Palmer RD, Yeo GS, Shivji MK, Stanley MA, et al. Selection of cervical keratinocytes containing integrated HPV16 associates with episome loss and an endogenous antiviral response. *Proc Natl Acad Sci U S A.* 2006;103(10):3822-7, <http://dx.doi.org/10.1073/pnas.0600078103>.
61. Kim K, Garner-Hamrick PA, Fisher C, Lee D, Lambert PF. Methylation patterns of papillomavirus DNA, its influence on E2 function, and implications in viral infection. *J Virol.* 2003;77(23):12450-9, <http://dx.doi.org/10.1128/JVI.77.23.12450-12459.2003>.
62. Stucki M, Clapperton JA, Mohammad D, Yaffe MB, Smerdon SJ, Jackson SP. MDC1 directly binds phosphorylated histone H2AX to regulate cellular responses to DNA double-strand breaks. *Cell.* 2005;123(7):1213-26, <http://dx.doi.org/10.1016/j.cell.2005.09.038>.
63. Bhattacharjee B, Sengupta S. CpG methylation of HPV 16 LCR at E2 binding site proximal to P97 is associated with cervical cancer in presence of intact E2. *Virology.* 2006;354(2):280-5, <http://dx.doi.org/10.1016/j.viro.2006.06.018>.
64. Fernandez AF, Rosales C, Lopez-Nieva P, Graña O, Ballestar E, Roperio S, et al. The dynamic DNA methylomes of double-stranded DNA viruses associated with human cancer. *Genome Res.* 2009;19(3):438-51, <http://dx.doi.org/10.1101/gr.083550.108>.
65. Fernandez AF, Esteller M. Viral epigenomes in human tumorigenesis. *Oncogene.* 2010;29(10):1405-20, <http://dx.doi.org/10.1038/onc.2009.517>.
66. Thomasset SC, Berry DP, Garcea G, Marczylo T, Steward WP, Gescher AJ. Dietary polyphenolic phytochemicals—promising cancer chemopreventive agents in humans? A review of their clinical properties. *Int J Cancer.* 2007;120(3):451-8, <http://dx.doi.org/10.1002/ijc.22419>.
67. Siegel EM, Salemi JL, Villa LL, Ferenczy A, Franco EL, Giuliano AR. Dietary consumption of antioxidant nutrients and risk of incident cervical intraepithelial neoplasia. *Gynecol Oncol.* 2010;118(3):289-94, <http://dx.doi.org/10.1016/j.ygyno.2010.05.022>.
68. Tomita LY, Longatto Filho A, Costa MC, Andreoli MA, Villa LL, Franco EL, et al; Brazilian Investigation into Nutrition and Cervical Cancer Prevention (BRINCA) Study Team. Diet and serum micronutrients in relation to cervical neoplasia and cancer among low-income Brazilian women. *Int J Cancer.* 2010;126(3):703-14, <http://dx.doi.org/10.1002/ijc.24793>.
69. Goel A, Aggarwal BB. Curcumin, the golden spice from Indian saffron, is a chemosensitizer and radiosensitizer for tumors and chemoprotector and radioprotector for normal organs. *Nutr Cancer.* 2010;62(7):919-30, <http://dx.doi.org/10.1080/01635581.2010.509835>.
70. Zoberi I, Bradbury CM, Curry HA, Bisht KS, Goswami PC, Roti Roti JL, et al. Radiosensitizing and anti-proliferative effects of resveratrol in two human cervical tumor cell lines. *Cancer Lett.* 2002;175(2):165-73, [http://dx.doi.org/10.1016/S0304-3835\(01\)00719-4](http://dx.doi.org/10.1016/S0304-3835(01)00719-4).
71. Ammon HP, Wahl MA. Pharmacology of Curcuma longa. *Planta Med.* 1991;57(1):1-7, <http://dx.doi.org/10.1055/s-2006-960004>.
72. Vajragupta O, Boonchoong P, Watanabe H, Tohda M, Kummasud N, Sumanont Y. Manganese complexes of curcumin and its derivatives: evaluation for the radical scavenging ability and neuroprotective activity. *Free Radic Biol Med.* 2003;35(12):1632-44, <http://dx.doi.org/10.1016/j.freeradbiomed.2003.09.011>.
73. Das KC, Das CK. Curcumin (diferuloylmethane), a singlet oxygen ((1)O(2)) quencher. *Biochem Biophys Res Commun.* 2002;295(1):62-6, [http://dx.doi.org/10.1016/S0006-291X\(02\)00633-2](http://dx.doi.org/10.1016/S0006-291X(02)00633-2).
74. Khopde SM, Priyadarsini KI, Venkatesan P, Rao MN. Free radical scavenging ability and antioxidant efficiency of curcumin and its substituted analogue. *Biophys Chem.* 1999;80(2):83-9, [http://dx.doi.org/10.1016/S0301-4622\(99\)00070-8](http://dx.doi.org/10.1016/S0301-4622(99)00070-8).
75. Brouet I, Ohshima H. Curcumin, an anti-tumour promoter and anti-inflammatory agent, inhibits induction of nitric oxide synthase in activated macrophages. *Biochem Biophys Res Commun.* 1995;206(2):533-40, <http://dx.doi.org/10.1006/bbrc.1995.1076>.
76. Chan MM, Adapala NS, Fong D. Curcumin overcomes the inhibitory effect of nitric oxide on Leishmania. *Parasitol Res.* 2005;96(1):49-56, <http://dx.doi.org/10.1007/s00436-005-1323-9>.
77. Deodhar SD, Sethi R, Srimal RC. Preliminary study on antirheumatic activity of curcumin (diferuloyl methane). *Indian J Med Res.* 1980;71: 632-4.
78. Khaffi A, Lev-Ari S, Vexler A, Barnea I, Starr A, Karaush V, et al. Curcumin: a potential radio-enhancer in head and neck cancer. *Laryngoscope.* 2009;119(10):2019-26, <http://dx.doi.org/10.1002/lary.20582>.
79. Lin JK, Lin-Shiau SY. Mechanisms of cancer chemoprevention by curcumin. *Proc Natl Sci Coun Repub China B.* 2001;25(2):59-66.
80. Sreejayan, Rao MN. Nitric oxide scavenging by curcuminoids. *J Pharm Pharmacol.* 1997;49(1):105-7, <http://dx.doi.org/10.1111/j.2042-7158.1997.tb06761.x>.
81. Srivastava R, Dikshit M, Srimal RC, Dhawan BN. Anti-thrombotic effect of curcumin. *Thromb Res.* 1985;40(3):413-7, [http://dx.doi.org/10.1016/0049-3848\(85\)90276-2](http://dx.doi.org/10.1016/0049-3848(85)90276-2).
82. Venkatesan N. Curcumin attenuation of acute adriamycin myocardial toxicity in rats. *Br J Pharmacol.* 1998;124(3):425-7, <http://dx.doi.org/10.1038/sj.bjp.0701877>.
83. Aggarwal BB, Shishodia S, Takada Y, Banerjee S, Newman RA, Bueso-Ramos CE, et al. Curcumin suppresses the paclitaxel-induced nuclear factor-kappaB pathway in breast cancer cells and inhibits lung metastasis of human breast cancer in nude mice. *Clin Cancer Res.* 2005;11(20): 7490-8, <http://dx.doi.org/10.1158/1078-0432.CCR-05-1192>.
84. Shishodia S, Amin HM, Lai R, Aggarwal BB. Curcumin (diferuloylmethane) inhibits constitutive NF-kappaB activation, induces G1/S arrest, suppresses proliferation, and induces apoptosis in mantle cell lymphoma. *Biochem Pharmacol.* 2005;70(5):700-13, <http://dx.doi.org/10.1016/j.bcp.2005.04.043>.
85. Siwak DR, Shishodia S, Aggarwal BB, Kurzrock R. Curcumin-induced antiproliferative and proapoptotic effects in melanoma cells are associated with suppression of IkkappaB kinase and nuclear factor kappaB activity and are independent of the B-Raf/mitogen-activated/extracellular signal-regulated protein kinase pathway and the Akt pathway. *Cancer.* 2005;104(4):879-90, <http://dx.doi.org/10.1002/cncr.21216>.
86. Aggarwal S, Ichikawa H, Takada Y, Sandur SK, Shishodia S, Aggarwal BB. Curcumin (diferuloylmethane) down-regulates expression of cell proliferation and antiapoptotic and metastatic gene products through suppression of IkkappaB kinase and Akt activation. *Mol Pharmacol.* 2006;69(1):195-206.
87. Kamat AM, Sethi G, Aggarwal BB. Curcumin potentiates the apoptotic effects of chemotherapeutic agents and cytokines through down-regulation of nuclear factor-kappaB and nuclear factor-kappaB-regulated gene products in IFN-alpha-sensitive and IFN-alpha-resistant human bladder cancer cells. *Mol Cancer Ther.* 2007;6(3):1022-30, <http://dx.doi.org/10.1158/1535-7163.MCT-06-0545>.
88. Deeb D, Jiang H, Gao X, Al-Holou S, Danyluk AL, Dulchavsky SA, et al. Curcumin [1,7-bis(4-hydroxy-3-methoxyphenyl)-1,6-heptadine-3,5-dione; C21H20O6] sensitizes human prostate cancer cells to tumor necrosis factor-related apoptosis-inducing ligand/Apo2L-induced apoptosis by suppressing nuclear factor-kappaB via inhibition of the prosurvival Akt signaling pathway. *J Pharmacol Exp Ther.* 2007;321(2):616-25, <http://dx.doi.org/10.1124/jpet.106.117721>.
89. Aoki H, Takada Y, Kondo S, Sawaya R, Aggarwal BB, Kondo Y. Evidence that curcumin suppresses the growth of malignant gliomas in vitro and in vivo through induction of autophagy: role of Akt and extracellular signal-regulated kinase signaling pathways. *Mol Pharmacol.* 2007;72(1): 29-39, <http://dx.doi.org/10.1124/mol.106.033167>.



90. Aggarwal S, Takada Y, Singh S, Myers JN, Aggarwal BB. Inhibition of growth and survival of human head and neck squamous cell carcinoma cells by curcumin via modulation of nuclear factor-kappaB signaling. *Int J Cancer*. 2004;111(5):679-92, <http://dx.doi.org/10.1002/ijc.20333>.
91. Kunnumakkara AB, Guha S, Krishnan S, Diagaradjane P, Gelovani J, Aggarwal BB. Curcumin potentiates antitumor activity of gemcitabine in an orthotopic model of pancreatic cancer through suppression of proliferation, angiogenesis, and inhibition of nuclear factor-kappaB-regulated gene products. *Cancer Res*. 2007;67(8):3853-61, <http://dx.doi.org/10.1158/0008-5472.CAN-06-4257>.
92. Han SS, Keum YS, Seo HJ, Surh YJ. Curcumin suppresses activation of NF-kappaB and AP-1 induced by phorbol ester in cultured human promyelocytic leukemia cells. *J Biochem Mol Biol*. 2002;35(3):337-42, <http://dx.doi.org/10.5483/BMBRep.2002.35.3.337>.
93. Kang G, Kong PJ, Yuh YJ, Lim SY, Yim SV, Chun W, et al. Curcumin suppresses lipopolysaccharide-induced cyclooxygenase-2 expression by inhibiting activator protein 1 and nuclear factor kappaB bindings in BV2 microglial cells. *J Pharmacol Sci*. 2004;94(3):325-8, <http://dx.doi.org/10.1254/jphs.94.325>.
94. Plummer SM, Holloway KA, Manson MM, Munks RJ, Kaptein A, Farrow S, et al. Inhibition of cyclo-oxygenase 2 expression in colon cells by the chemopreventive agent curcumin involves inhibition of NF-kappaB activation via the NIK/IKK signalling complex. *Oncogene*. 1999;18(44):6013-20, <http://dx.doi.org/10.1038/sj.onc.1202980>.
95. Plummer SM, Hill KA, Festing MF, Steward WP, Gescher AJ, Sharma RA. Clinical development of leukocyte cyclooxygenase 2 activity as a systemic biomarker for cancer chemopreventive agents. *Cancer Epidemiol Biomarkers Prev*. 2001;10(12):1295-9.
96. Cheng AL, Hsu CH, Lin JK, Hsu MM, Ho YF, Shen TS, et al. Phase I clinical trial of curcumin, a chemopreventive agent, in patients with high-risk or pre-malignant lesions. *Anticancer Res*. 2001;21(4B):2895-900.
97. Kawamori T, Lubet R, Steele VE, Kelloff GJ, Kaskey RB, Rao CV, et al. Chemopreventive effect of curcumin, a naturally occurring anti-inflammatory agent, during the promotion/progression stages of colon cancer. *Cancer Res*. 1999;59(3):597-601.
98. Rao CV, Rivenson A, Simi B, Reddy BS. Chemoprevention of colon carcinogenesis by dietary curcumin, a naturally occurring plant phenolic compound. *Cancer Res*. 1995;55(2):259-66.
99. Venkatraman M, Anto RJ, Nair A, Varghese M, Karunakaran D. Biological and chemical inhibitors of NF-kappaB sensitize SiHa cells to cisplatin-induced apoptosis. *Mol Carcinog*. 2005;44(1):51-9, <http://dx.doi.org/10.1002/mc.20116>.
100. Bava SV, Puliappadamba VT, Deepti A, Nair A, Karunakaran D, Anto RJ. Sensitization of taxol-induced apoptosis by curcumin involves downregulation of nuclear factor-kappaB and the serine/threonine kinase Akt and is independent of tubulin polymerization. *J Biol Chem*. 2005;280(8):6301-8, <http://dx.doi.org/10.1074/jbc.M410647200>.
101. Sreekanth CN, Bava SV, Sreekumar E, Anto RJ. Molecular evidences for the chemosensitizing efficacy of liposomal curcumin in paclitaxel chemotherapy in mouse models of cervical cancer. *Oncogene*. 2011;30(28):3139-52, <http://dx.doi.org/10.1038/onc.2011.23>.
102. Bava SV, Sreekanth CN, Thulasidasan AK, Anto NP, Cheriyan VT, Puliappadamba VT, et al. Akt is upstream and MAPKs are downstream of NF-kB in paclitaxel-induced survival signaling events, which are down-regulated by curcumin contributing to their synergism. *Int J Biochem Cell Biol*. 2011;43(3):331-41, <http://dx.doi.org/10.1016/j.biocel.2010.09.011>.
103. Prusty BK, Das BC. Constitutive activation of transcription factor AP-1 in cervical cancer and suppression of human papillomavirus (HPV) transcription and AP-1 activity in HeLa cells by curcumin. *Int J Cancer*. 2005;113(6):951-60, <http://dx.doi.org/10.1002/ijc.20668>.
104. Divya CS, Pillai MR. Antitumor action of curcumin in human papillomavirus associated cells involves downregulation of viral oncogenes, prevention of NFkB and AP-1 translocation, and modulation of apoptosis. *Mol Carcinog*. 2006;45(5):320-32, <http://dx.doi.org/10.1002/mc.20170>.
105. Javvadi P, Hertan L, Kosoff R, Datta T, Kolev J, Mick R, et al. Thioredoxin reductase-1 mediates curcumin-induced radiosensitization of squamous carcinoma cells. *Cancer Res*. 2010;70(5):1941-50, <http://dx.doi.org/10.1158/0008-5472.CAN-09-3025>.
106. Goel A, Kunnumakkara AB, Aggarwal BB. Curcumin as "Curcumin": from kitchen to clinic. *Biochem Pharmacol*. 2008;75(4):787-809, <http://dx.doi.org/10.1016/j.bcp.2007.08.016>.
107. Belcaro G, Hosoi M, Pellegrini L, Appendino G, Ippolito E, Ricci A, Ledda A, Dugall M, Cesarone MR, Maione C, Ciammaichella G, Genovesi D, Togni S. A controlled study of a lecithinized delivery system of curcumin (Meriva[®]) to alleviate the adverse effects of cancer treatment. *Phytother Res*. 2014;28(3):444-50, <http://dx.doi.org/10.1002/ptr.5014>.
108. Du GJ, Zhang Z, Wen XD, Yu C, Calway T, Yuan CS, et al. Epigallocatechin Gallate (EGCG) is the most effective cancer chemopreventive polyphenol in green tea. *Nutrients*. 2012;4(11):1679-91, <http://dx.doi.org/10.3390/nu411679>.
109. Nanjo F, Honda M, Okushio K, Matsumoto N, Ishigaki F, Ishigami T, et al. Effects of dietary tea catechins on alpha-tocopherol levels, lipid peroxidation, and erythrocyte deformability in rats fed on high palm oil and perilla oil diets. *Biol Pharm Bull*. 1993;16(11):1156-9, <http://dx.doi.org/10.1248/bpb.16.1156>.
110. Scott BC, Butler J, Halliwell B, Aruoma OI. Evaluation of the antioxidant actions of ferulic acid and catechins. *Free Radic Res Commun*. 1993;19(4):241-53, <http://dx.doi.org/10.3109/10715769309056512>.
111. Haenen GR, Paquay JB, Korthouwer RE, Bast A. Peroxynitrite scavenging by flavonoids. *Biochem Biophys Res Commun*. 1997;236(3):591-3, <http://dx.doi.org/10.1006/bbrc.1997.7016>.
112. Liang YC, Lin-Shiau SY, Chen CF, Lin JK. Inhibition of cyclin-dependent kinases 2 and 4 activities as well as induction of Cdk inhibitors p21 and p27 during growth arrest of human breast carcinoma cells by (-)-epigallocatechin-3-gallate. *J Cell Biochem*. 1999;75(1):1-12, [http://dx.doi.org/10.1002/\(SICI\)1097-4644\(19991001\)75:1<1::AID-JCB1>3.0.CO;2-N](http://dx.doi.org/10.1002/(SICI)1097-4644(19991001)75:1<1::AID-JCB1>3.0.CO;2-N).
113. Ahmad N, Cheng P, Mukhtar H. Cell cycle dysregulation by green tea polyphenol epigallocatechin-3-gallate. *Biochem Biophys Res Commun*. 2000;275(2):328-34, <http://dx.doi.org/10.1006/bbrc.2000.3297>.
114. Kondo T, Ohta T, Igura K, Hara Y, Kaji K. Tea catechins inhibit angiogenesis in vitro, measured by human endothelial cell growth, migration and tube formation, through inhibition of VEGF receptor binding. *Cancer Lett*. 2002;180(2):139-44, [http://dx.doi.org/10.1016/S0304-3835\(02\)00007-1](http://dx.doi.org/10.1016/S0304-3835(02)00007-1).
115. Nakagawa T, Yokozawa T. Direct scavenging of nitric oxide and superoxide by green tea. *Food Chem Toxicol*. 2002;40(12):1745-50, [http://dx.doi.org/10.1016/S0278-6915\(02\)00169-2](http://dx.doi.org/10.1016/S0278-6915(02)00169-2).
116. Baliga MS, Meleth S, Katiyar SK. Growth inhibitory and antimetastatic effect of green tea polyphenols on metastasis-specific mouse mammary carcinoma 4T1 cells in vitro and in vivo systems. *Clin Cancer Res*. 2005;11(5):1918-27, <http://dx.doi.org/10.1158/1078-0432.CCR-04-1976>.
117. Nihal M, Ahmad N, Mukhtar H, Wood GS. Anti-proliferative and proapoptotic effects of (-)-epigallocatechin-3-gallate on human melanoma: possible implications for the chemoprevention of melanoma. *Int J Cancer*. 2005;114(4):513-21, <http://dx.doi.org/10.1002/ijc.20785>.
118. Qanungo S, Das M, Haldar S, Basu A. Epigallocatechin-3-gallate induces mitochondrial membrane depolarization and caspase-dependent apoptosis in pancreatic cancer cells. *Carcinogenesis*. 2005;26(5):958-67, <http://dx.doi.org/10.1093/carcin/bgi040>.
119. Fassina G, Venè R, Morini M, Minghelli S, Benelli R, Noonan DM, et al. Mechanisms of inhibition of tumor angiogenesis and vascular tumor growth by epigallocatechin-3-gallate. *Clin Cancer Res*. 2004;10(14):4865-73, <http://dx.doi.org/10.1158/1078-0432.CCR-03-0672>.
120. Sahin K, Tuzcu M, Gencoglu H, Dogukan A, Timurkan M, Sahin N, et al. Epigallocatechin-3-gallate activates Nrf2/HO-1 signaling pathway in cisplatin-induced nephrotoxicity in rats. *Life Sci*. 2010;87(7-8):240-5, <http://dx.doi.org/10.1016/j.lfs.2010.06.014>.
121. Na HK, Surh YJ. Modulation of Nrf2-mediated antioxidant and detoxifying enzyme induction by the green tea polyphenol EGCG. *Food Chem Toxicol*. 2008;46(4):1271-8, <http://dx.doi.org/10.1016/j.fct.2007.10.006>.
122. Rahman I, Biswas SK, Kirkham PA. Regulation of inflammation and redox signaling by dietary polyphenols. *Biochem Pharmacol*. 2006;72(11):1439-52, <http://dx.doi.org/10.1016/j.bcp.2006.07.004>.
123. Ahn WS, Huh SW, Bae SM, Lee IP, Lee JM, Namkoong SE, et al. A major constituent of green tea, EGCG, inhibits the growth of a human cervical cancer cell line, CaSki cells, through apoptosis, G(1) arrest, and regulation of gene expression. *DNA Cell Biol*. 2003;22(3):217-24, <http://dx.doi.org/10.1089/104454903321655846>.
124. Asif Siddiqui F, Naim M, Islam N. Apoptotic effect of green tea polyphenol(EGCG) on cervical carcinoma cells. *Diagn Cytopathol*. 2011;39(7):482-8, <http://dx.doi.org/10.1002/dc.21434>.
125. Hussain S. Comparative efficacy of epigallocatechin-3-gallate against H(2)O(2)-induced ROS in cervical cancer biopsies and HeLa cell lines. *Contemp Oncol (Pozn)*. 2017;21(3):209-212, <http://dx.doi.org/10.5114/wo.2017.70110>.
126. Noguchi M, Yokoyama M, Watanabe S, Uchiyama M, Nakao Y, Hara K, et al. Inhibitory effect of the tea polyphenol, (-)-epigallocatechin gallate, on growth of cervical adenocarcinoma cell lines. *Cancer Lett*. 2006;234(2):135-42, <http://dx.doi.org/10.1016/j.canlet.2005.03.053>.
127. Yokoyama M, Noguchi M, Nakao Y, Pater A, Iwasaka T. The tea polyphenol, (-)-epigallocatechin gallate effects on growth, apoptosis, and telomerase activity in cervical cell lines. *Gynecol Oncol*. 2004;92(1):197-204, <http://dx.doi.org/10.1016/j.ygyno.2003.09.023>.
128. Zhang Q, Tang X, Lu Q, Zhang Z, Rao J, Le AD. Green tea extract and (-)-epigallocatechin-3-gallate inhibit hypoxia- and serum-induced HIF-1alpha protein accumulation and VEGF expression in human cervical carcinoma and hepatoma cells. *Mol Cancer Ther*. 2006;5(5):1227-38, <http://dx.doi.org/10.1158/1535-7163.MCT-05-0490>.
129. Kilic U, Sahin K, Tuzcu M, Basak N, Orhan C, Elibol-Can B, et al. Enhancement of Cisplatin sensitivity in human cervical cancer: epigallocatechin-3-gallate. *Front Nutr*. 2015;1:28, <http://dx.doi.org/10.3389/fnut.2014.00028>.
130. Ahn WS, Yoo J, Huh SW, Kim CK, Lee JM, Namkoong SE, et al. Protective effects of green tea extracts (polyphenon E and EGCG) on human cervical lesions. *Eur J Cancer Prev*. 2003;12(5):383-90, <http://dx.doi.org/10.1097/00008469-200310000-00007>.



131. Aggarwal BB, Bhardwaj A, Aggarwal RS, Seeram NP, Shishodia S, Takada Y. Role of resveratrol in prevention and therapy of cancer: pre-clinical and clinical studies. *Anticancer Res.* 2004;24(5A):2783-840.
132. Elshaer M, Chen Y, Wang XJ, Tang X. Resveratrol: An overview of its anti-cancer mechanisms. *Life Sci.* 2018;207:340-9, <http://dx.doi.org/10.1016/j.lfs.2018.06.028>.
133. Kramer MP, Wesierska-Gadek J. Monitoring of long-term effects of resveratrol on cell cycle progression of human HeLa cells after administration of a single dose. *Ann N Y Acad Sci.* 2009;1171:257-63, <http://dx.doi.org/10.1111/j.1749-6632.2009.04884.x>.
134. Harikumar KB, Aggarwal BB. Resveratrol: a multitargeted agent for age-associated chronic diseases. *Cell Cycle.* 2008;7(8):1020-35, <http://dx.doi.org/10.4161/cc.7.8.5740>.
135. Seeni A, Takahashi S, Takeshita K, Tang M, Sugiura S, Sato SY, et al. Suppression of prostate cancer growth by resveratrol in the transgenic rat for adenocarcinoma of prostate (TRAP) model. *Asian Pac J Cancer Prev.* 2008;9(1):7-14.
136. Roy P, Kalra N, Prasad S, George J, Shukla Y. Chemopreventive potential of resveratrol in mouse skin tumors through regulation of mitochondrial and PI3K/AKT signaling pathways. *Pharm Res.* 2009;26(1):211-7, <http://dx.doi.org/10.1007/s11095-008-9723-z>.
137. Li L, Qiu RL, Lin Y, Cai Y, Bian Y, Fan Y, et al. Resveratrol suppresses human cervical carcinoma cell proliferation and elevates apoptosis via the mitochondrial and p53 signaling pathways. *Oncol Lett.* 2018;15(6):9845-51, <http://dx.doi.org/10.3892/ol.2018.8571>.
138. Hsu KF, Wu CL, Huang SC, Wu CM, Hsiao JR, Yo YT, et al. Cathepsin L mediates resveratrol-induced autophagy and apoptotic cell death in cervical cancer cells. *Autophagy.* 2009;5(4):451-60, <http://dx.doi.org/10.4161/auto.5.4.7666>.
139. Sexton E, Van Themsche C, LeBlanc K, Parent S, Lemoine P, Asselin E. Resveratrol interferes with AKT activity and triggers apoptosis in human uterine cancer cells. *Mol Cancer.* 2006;5:45, <http://dx.doi.org/10.1186/1476-4598-5-45>.
140. Rezk YA, Balulad SS, Keller RS, Bennett JA. Use of resveratrol to improve the effectiveness of cisplatin and doxorubicin: study in human gynecologic cancer cell lines and in rodent heart. *Am J Obstet Gynecol.* 2006;194(5):e23-6, <http://dx.doi.org/10.1016/j.ajog.2005.11.030>.
141. Galluzzi L, Vacchelli E, Michels J, Garcia P, Kepp O, Senovilla L, et al. Effects of vitamin B6 metabolism on oncogenesis, tumor progression and therapeutic responses. *Oncogene.* 2013;32(42):4995-5004, <http://dx.doi.org/10.1038/onc.2012.623>.
142. Chang YC, Scott RD, Graves DJ. Function of pyridoxal 5'-phosphate in glycogen phosphorylase: a model study using 6-fluoro-5'-deoxy-pyridoxal- and 5'-deoxy-pyridoxal-reconstituted enzymes. *Biochemistry.* 1987;26(2):360-7, <http://dx.doi.org/10.1021/bi00376a005>.
143. Selhub J. Homocysteine metabolism. *Annu Rev Nutr.* 1999;19:217-46, <http://dx.doi.org/10.1146/annurev.nutr.19.1.217>.
144. Toney MD. Reaction specificity in pyridoxal phosphate enzymes. *Arch Biochem Biophys.* 2005;433(1):279-87, <http://dx.doi.org/10.1016/j.abb.2004.09.037>.
145. Cheng SB, Lin PT, Liu HT, Peng YS, Huang SC, Huang YC. Vitamin B-6 Supplementation Could Mediate Antioxidant Capacity by Reducing Plasma Homocysteine Concentration in Patients with Hepatocellular Carcinoma after Tumor Resection. *Biomed Res Int.* 2016;2016:7658981.
146. Galluzzi L, Vitale I, Senovilla L, Olaussen KA, Pinna G, Eisenberg T, et al. Prognostic impact of vitamin B6 metabolism in lung cancer. *Cell Rep.* 2012;2(2):257-69, <http://dx.doi.org/10.1016/j.celrep.2012.06.017>.
147. Leekha A, Gurjar BS, Tyagi A, Rizvi MA, Verma AK. Vitamin C in synergism with cisplatin induces cell death in cervical cancer cells through altered redox cycling and p53 upregulation. *J Cancer Res Clin Oncol.* 2016;142(12):2503-14, <http://dx.doi.org/10.1007/s00432-016-2235-z>.
148. Mane SD, Thoh M, Sharma D, Sandur SK, Naidu KA. Ascorbyl Stearate Promotes Apoptosis Through Intrinsic Mitochondrial Pathway in HeLa Cancer Cells. *Anticancer Res.* 2016;36(12):6409-17, <http://dx.doi.org/10.21873/anticancer.11238>.
149. Hwang JH, Kim MK, Lee JK. Dietary supplements reduce the risk of cervical intraepithelial neoplasia. *Int J Gynecol Cancer.* 2010;20(3):398-403, <http://dx.doi.org/10.1111/IGC.0b013e3181d02ff2>.
150. Palan PR, Mikhail MS, Goldberg GL, Basu J, Runowicz CD, Romney SL. Plasma levels of beta-carotene, lycopene, canthaxanthin, retinol, and alpha- and tau-tocopherol in cervical intraepithelial neoplasia and cancer. *Clin Cancer Res.* 1996;2(1):181-5.
151. Palan PR, Mikhail MS, Shaban DW, Romney SL. Plasma concentrations of coenzyme Q10 and tocopherols in cervical intraepithelial neoplasia and cervical cancer. *Eur J Cancer Prev.* 2003;12(4):321-6, <http://dx.doi.org/10.1097/00008469-200308000-00013>.
152. Srivastava S, Natu SM, Gupta A, Pal KA, Singh U, Agarwal GG, et al. Lipid peroxidation and antioxidants in different stages of cervical cancer: Prognostic significance. *Indian J Cancer.* 2009;46(4):297-302, <http://dx.doi.org/10.4103/0019-509X.55549>.
153. Hu X, Li S, Zhou L, Zhao M, Zhu X. Effect of vitamin E supplementation on uterine cervical neoplasm: A meta-analysis of case-control studies. *PLoS One.* 2017;12(8):e0183395, <http://dx.doi.org/10.1371/journal.pone.0183395>.
154. Palan PR, Woodall AL, Anderson PS, Mikhail MS. Alpha-tocopherol and alpha-tocopherol quinone levels in cervical intraepithelial neoplasia and cervical cancer. *Am J Obstet Gynecol.* 2004;190(5):1407-10.
155. Castaldo SA, Freitas JR, Concinha NV, Madureira PA. The Tumorigenic Roles of the Cellular REDOX Regulatory Systems. *Oxid Med Cell Longev.* 2016;2016:8413032, <http://dx.doi.org/10.1155/2016/8413032>.
156. Teppo HR, Soini Y, Karihtala P. Reactive Oxygen Species-Mediated Mechanisms of Action of Targeted Cancer Therapy. *Oxid Med Cell Longev.* 2017;2017:1485283, <http://dx.doi.org/10.1155/2017/1485283>.
157. Jiao Y, Wang Y, Guo S, Wang G. Glutathione peroxidases as oncotargets. *Oncotarget.* 2017;8(45):80093-102, <http://dx.doi.org/10.18632/oncotarget.20278>.
158. Yang J, Zhao X, Tang M, Li L, Lei Y, Cheng P, et al. The role of ROS and subsequent DNA-damage response in PUMA-induced apoptosis of ovarian cancer cells. *Oncotarget.* 2017;8(14):23492-506, <http://dx.doi.org/10.18632/oncotarget.15626>.
159. Cruz-Gregorio A, Manzo-Merino J, González-García MC, Pedraza-Chaverri J, Medina-Campos ON, Valverde M, et al. Human Papillomavirus Types 16 and 18 Early-expressed Proteins Differentially Modulate the Cellular Redox State and DNA Damage. *Int J Biol Sci.* 2018;14(1):21-35, <http://dx.doi.org/10.7150/ijbs.21547>.
160. Drozd E, Krzysztos-Russjan J, Marczewska J, Drozd J, Bubko I, Bielak M, et al. Up-regulation of glutathione-related genes, enzyme activities and transport proteins in human cervical cancer cells treated with doxorubicin. *Biomed Pharmacother.* 2016;83:397-406, <http://dx.doi.org/10.1016/j.biopha.2016.06.051>.
161. Jaudan A, Sharma S, Malek SNA, Dixit A. Induction of apoptosis by pinostrobin in human cervical cancer cells: Possible mechanism of action. *PLoS One.* 2018;13(2):e0191523, <http://dx.doi.org/10.1371/journal.pone.0191523>.
162. You BR, Shin HR, Han BR, Kim SH, Park WH. Auranofin induces apoptosis and necrosis in HeLa cells via oxidative stress and glutathione depletion. *Mol Med Rep.* 2015;11(2):1428-34, <http://dx.doi.org/10.3892/mmr.2014.2830>.
163. Bonetta R. Potential therapeutic applications of MnSODs and SOD-mimetics. *Chemistry.* 2018;24(20):5032-41, <http://dx.doi.org/10.1002/chem.201704561>.
164. Bauer S, Joensuu H. Emerging Agents for the Treatment of Advanced, Imatinib-Resistant Gastrointestinal Stromal Tumors: Current Status and Future Directions. *Drugs.* 2015;75(12):1323-34, <http://dx.doi.org/10.1007/s40265-015-0440-8>.
165. Brown C. Targeted therapy: An elusive cancer target. *Nature.* 2016;537(7620):S106-8, <http://dx.doi.org/10.1038/537S106a>.
166. Swain SM, Baselga J, Kim SB, Ro J, Semiglazov V, Campone M, et al. Pertuzumab, trastuzumab, and docetaxel in HER2-positive metastatic breast cancer. *N Engl J Med.* 2015;372(8):724-34, <http://dx.doi.org/10.1056/NEJMoa1413513>.
167. Mendel DB, Laird AD, Xin X, Louie SG, Christensen JG, Li G, et al. In vivo antitumor activity of SU11248, a novel tyrosine kinase inhibitor targeting vascular endothelial growth factor and platelet-derived growth factor receptors: determination of a pharmacokinetic/pharmacodynamics relationship. *Clin Cancer Res.* 2003;9(1):327-37.
168. Suddek GM. Sunitinib improves chemotherapeutic efficacy and ameliorates cisplatin-induced nephrotoxicity in experimental animals. *Cancer Chemother Pharmacol.* 2011;67(5):1035-44, <http://dx.doi.org/10.1007/s00280-010-1402-1>.
169. Abdel-Aziz AK, Shouman S, El-Demerdash E, Elgendy M, Abdel-Naim AB. Chloroquine synergizes sunitinib cytotoxicity via modulating autophagic, apoptotic and angiogenic machineries. *Chem Biol Interact.* 2014;217:28-40, <http://dx.doi.org/10.1016/j.cbi.2014.04.007>.
170. Ogawa Y, Ue H, Tsuzuki K, Tadokoro M, Miyatake K, Sasaki T, et al. New radiosensitization treatment (KORTUC I) using hydrogen peroxide solution-soaked gauze bolus for unresectable and superficially exposed neoplasms. *Oncol Rep.* 2008;19(6):1389-94, <http://dx.doi.org/10.3892/or.19.6.1389>.
171. Ogawa Y, Kubota K, Ue H, Tadokoro M, Matsui R, Yamanishi T, et al. Safety and effectiveness of a new enzyme-targeting radiosensitization treatment (KORTUC II) for intratumoral injection for low-LET radioresistant tumors. *Int J Oncol.* 2011;39(3):553-60, <http://dx.doi.org/10.3892/ijo.2011.1069>.
172. Aoyama N, Ogawa Y, Yasuoka M, Ohgi K, Iwasa H, Miyatake K, et al. Therapeutic results of a novel enzyme-targeting radiosensitization treatment, Kochi oxydol-radiation therapy for unresectable carcinomas II, in patients with stage I primary breast cancer. *Oncol Lett.* 2017;13(6):4741-7, <http://dx.doi.org/10.3892/ol.2017.6074>.
173. Aoyama N, Ogawa Y, Yasuoka M, Takahashi M, Iwasa H, Miyatake K, et al. Therapeutic response to a novel enzyme-targeting radiosensitization treatment (Kochi Oxydol-Radiation Therapy for Unresectable Carcinomas) in patients with recurrent breast cancer. *Oncol Lett.* 2016;12(1):29-34, <http://dx.doi.org/10.3892/ol.2016.4589>.