

**REVIEW ARTICLE** 

# Improving adenoviral vectors and strategies for prostate cancer gene therapy

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Gene therapy has been evaluated for the treatment of prostate cancer and includes the application of adenoviral vectors encoding a suicide gene or oncolytic adenoviruses that may be armed with a functional transgene. In parallel, versions of adenoviral vector expressing the p53 gene (Ad-p53) have been tested as treatments for head and neck squamous cell carcinoma and non-small cell lung cancer. Although Ad-p53 gene therapy has yielded some interesting results when applied to prostate cancer, it has not been widely explored, perhaps due to current limitations of the approach. To achieve better functionality, improvements in the gene transfer system and the therapeutic regimen may be required. We have developed adenoviral vectors whose transgene expression is controlled by a p53-responsive promoter, which creates a positive feedback mechanism when used to drive the expression of p53. Together with improvements that permit efficient transduction, this new approach was more effective than the use of traditional versions of Ad-p53 in killing prostate cancer cell lines and inhibiting tumor progression. Even so, gene therapy is not expected to replace traditional chemotherapy but should complement the standard of care. In fact, chemotherapy has been shown to assist in viral transduction and transgene expression. The cooperation between gene therapy and chemotherapy is expected to effectively kill tumor cells while permitting the use of reduced chemotherapy drug concentrations and, thus, lowering side effects. Therefore, the combination of gene therapy and chemotherapy may prove essential for the success of both approaches.

KEYWORDS: Prostate Cancer; Adenovirus; p53; Chemotherapy; Gene Therapy.

#### Prostate cancer

In the United States, prostate carcinoma is the most frequent cancer among men, accounting for 19% of cancers in 2017. Though incidence rates vary by region and are higher in developed countries, an estimated 1.1 million men were diagnosed with prostate cancer in 2012, making it the second most common cancer in men worldwide. The highest rates are in Australia, North America, and Northern and Western Europe, which are regions where testing for prostate-specific antigen (PSA) has become commonplace. In South America, the rate is 60.1 per 100,000 (age-standardized rate). In a global estimate, prostate cancer is the fifth leading cause of death from cancer in men. The prediction for 2020 is 1,392,727 new cases worldwide (1,2).

If detected early while locally confined, prostate cancer is largely curable by radical prostatectomy or radiotherapy (3). However, initial diagnosis of up to 15% of patients include

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metastatic lesions, and recurrence after conventional radical therapy occurs in up to 40% of patients (4,5). Typically, androgen deprivation therapy (ADT) is given to patients with recurrent disease and is often effective, though most of these patients will relapse after 2-3 years due to the development of castration-resistant prostate cancer (CRPC) (6). The most important driver of resistance is the androgen receptor (AR), whose hyperactivity may arise through multiple mechanisms, such as AR amplification and hypersensitivity, AR mutation leading to promiscuity, androgen-independent AR activation and intratumoral alternative androgen production (7). Docetaxel, a cytotoxic antimicrotubule agent that binds to the β-tubulin subunit of microtubulin, is the first line treatment for CRPC. Although docetaxel may be effective, only approximately 48% of patients responded to combined docetaxel and prednisone treatment, with a median survival of 2.5 months over the control group (8). Other current chemotherapies include abiraterone and enzalutamide (second-generation antiandrogens), which are used as second-line treatments for CRPC, and cabazitaxel (second-generation tubulin-binding taxane) has proven to be beneficial even as a third-line treatment (9,10). Even with these advances, the 5-year survival rate of CRPC is only 31% (11); therefore, continued study of alternative therapies is warranted.

# Gene therapy for prostate cancer

Gene therapy using adenoviral vectors has been evaluated in 17 clinical trials for the treatment of prostate cancer (clinical trials.gov). Here, we will address some of these approaches,

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emphasizing the modifications made to the vector. The use of a prostate-specific promoter to drive viral expression or control oncolytic viral replication assures specificity through transcriptional control. Alternatively, modifications in the fiber protein are expected to alter vector tropism at the level of transduction.

Most of the prostate-specific promoters are derived from the PSA and prostate-specific membrane antigen (PSMA) enhancers (12,13). A chimera composed of the PSA enhancer and the probasin (PB) promoter showed 20% less activity than the cytomegalovirus (CMV) promoter but high prostate cancer cell specificity (14). Promoters of tyrosinase (15), human glandular kallikrein (hKLK2) (16), PB and the mouse mammary tumor virus long terminal repeat (MMTV LTR) (17) have also been tested for their usefulness in treating prostate cancer.

The serotype 5 adenovirus (Ad5) initially makes contact with its cellular receptor (coxsackievirus and adenovirus receptor, CAR) through the fiber protein; thus, its modification can direct viral tropism. For example, compared to an unmodified Ad5 vector, a shortened chimeric Ad5/35 hybrid fiber protein had an increased transgene insert size as well as increased transduction efficiency in different prostate cancer cell lines (18). A fiber knob chimera of Ad5/3 facilitated transduction in a CAR-independent manner (19). Incorporation of the Arg-Gly-Asp (RGD) tripeptide motif into the fiber protein increased transduction, even in CAR-negative prostate cancer cell lines (20,21). Additionally, tissue-specific expression was enhanced in high-capacity adenoviral vectors compared to first-generation adenoviral vectors (15). To avoid destruction of the virus particles by the immune system, use of other adenovirus serotypes (Ad6), shielding of the virus particle with PEG and use of mesenchymal stem cells (MSCs) and dendritic cells as carriers have been tested (22-24).

Conditionally replicating viruses were tested in phase I/II clinical trials. These adenoviral vectors encode the E1 gene under control of a tissue-specific promoter, thus limiting viral replication. In a phase I clinical trial with 20 patients, different doses of the CG7060 virus (Cell Genesys, South San Francisco, CA), which contains the PSA enhancer, were well tolerated, and there was a correlation between viral dose and effect: the patients with the greatest reduction in PSA levels received the highest dose (25). Another vector, CG7870 (Cell Genesys), has the E1 gene under control of the rat PB promoter and the E1B gene under control of the PSA promoter, providing two degrees of specificity. The CG7870 vector was administered in 23 patients with CRPC, and 5 patients showed a 25-49% decrease in PSA levels (26). The combination of CG7870 with taxanes resulted in a synergistic response in vitro (27). Another promoter used is a chimera composed of the PSA and PSMA enhancers, as well as the T-cell receptor y-chain alternate reading frame protein promoter, Ad[I/PPT-E1A]. While not yet tested clinically, the Ad[I/PPT-E1A] oncolytic vector showed prostate-specific activity in both a hormone-dependent and hormone-independent manner, suggesting its usefulness even in patients treated with androgen withdrawal (28). A vaccination protocol using an adenoviral vector expressing PSA was used in a phase I clinical trial and was shown to be safe and to increase survival in CRPC patients (29). Phase II clinical trials are underway (30).

Oncolytic adenoviral vectors have also been armed with functional transgenes. A prostate-restricted replicative adenovirus (PRRA) using a prostate-specific enhancer (PSES) to control the expression of E1A, E1B and E4 was armed with the FasL gene. This conditionally replicating adenoviral vector induced apoptosis of PSA/PMSA-positive cells and was less toxic compared to an AdCMVFasL vector, which killed all mice in 16 hours due to multivisceral failure (31). PRRAs have been armed with other transgenes such as a fusion of the endostatin and angiostatin genes (18), reduced expression in immortalized cells (REIC) (32), glioma pathogenesis-related protein 1 (GLIPR1) (33) and a fusion protein of PSA and CD40L (34).

First-generation adenoviral vectors armed with a suicide gene were safe and well tolerated in phase I clinical trials for patients with local recurrent or metastatic prostate cancer. Only one patient who received the highest viral dose suffered grade 4 thrombocytopenia and grade 3 hepatotoxicity; some patients showed evidence of reduced PSA levels. Suicide gene therapy has also been combined with conditionally replicating adenoviruses in prostate cancer patients with locally recurrent disease, newly diagnosed and locally aggressive disease or metastatic disease. Even though it was shown to be safe and an initial PSA decline was observed, patients relapsed (35-39). Long-term follow up of the patients with local recurrence who were treated with a combination of suicide gene therapy and radiotherapy showed that the patients benefited in terms of the PSA doubling time (PSADT) (40). Introducing suicide therapy into a second-generation adenovirus in combination with intensity-modulated radiotherapy (IMRT) in patients with newly diagnosed prostate cancer resulted in low toxicity and a reduction in the percentage of patients who were positive for adenocarcinoma (41). Ultrasound-directed intraprostatic injection of an adenoviral vector expressing thymidine kinase (Ad-TK) was well tolerated and none of the 10 patients with adenocarcinoma developed metastases (42). Strategies involving suicide gene therapy in prostate cancer are ongoing (clinicaltrials.gov).

# ADENOVIRUS-P53 IN PROSTATE CANCER

#### P53 in prostate cancer

The p53 protein is an important tumor suppressor involved in a variety of cellular responses to stress. P53 degradation is mediated by murine double minute 2 (MDM2), and disruption of the p53/MDM2 complex frees p53 to promote the transcription of specific target genes that, in turn, direct cellular responses such as apoptosis (43,44). In prostate cancer, p53 alterations occur in approximately 5% of cases, while this number rises to 65% in metastatic disease (45,46). The detection of p53 in prostate cancer was analyzed in more than 50 studies showing that immunohistochemical staining of p53 increases in high-grade carcinomas, advanced stage cancer and carcinomas of peripheral zone origin (47). However, there are fewer studies mapping the alterations in p53 in prostate cancer, and such studies show divergent frequencies ranging from 3-40% in p53 gene mutation and 10-60% deletions or loss of heterozygosity (48-50). A more recent study that combines tissue microarray (TMA) and DNA analysis found that different types of p53 alterations characterize subgroups of prostate cancer with distinctively different prognoses; strong p53 immunostaining is rare but represents an independent and worse prognostic event in prostate cancer (47).

#### Ad-p53 clinical trials

Viral vectors expressing p53 have been in development for the last 25 years, and even though retroviral vectors were the first to be tested in patients, adenoviral vectors have been

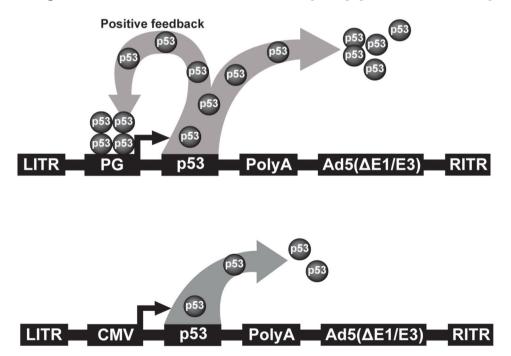


more broadly used, and different versions have been employed to express the tumor suppressor p53 (51-54). Three slightly different first-generation adenoviral vectors reached clinical trials; all with deletions in the E1 gene and expression of the p53 gene under control of constitutive promoters (CMV or Rous sarcoma virus [RSV]). These Ad-p53 vectors known as Advexin (Introgen Therapeutics, Multivir, Inc, both of Houston, TX), SCH58500 (Merck & Co; Schering-Plough, Kenilworth, NJ) and Gendicine (Shenzhen SiBiono GeneTech, Guangdong, China) have been tested for treating different types of cancers, including non-small cell lung cancer (NSCLC), head and neck squamous cell carcinoma (HNSCC), colorectal, bladder and several other cancers (51,55-63). Even with the publication of promising clinical results, only Gendicine has been approved for commercialization and is currently being used in China for the treatment of head and neck cancer. A phase I clinical trial using Advexin for prostate cancer showed that the vector is safe, with no grade 3 or 4 side effects, and that the vector induced the expression of p53 and apoptosis of the tumor cells (64). Even so, no further trials testing Advexin for the treatment of prostate cancer were performed.

## Improvement of Ad-p53 for prostate cancer

In the 1990s, Ad-p53 was tested in prostate carcinoma cell lines and xenograft mouse models, showing varied results. Some groups have demonstrated that Ad-p53 can induce apoptosis and reduce tumor volume (65-67), while another group did not observe any advantage of Ad-p53 compared to the control, revealing instead that Ad-p21 was more effective for reducing tumor volume and increasing survival (68). Most of these studies were conducted 20 years ago, and it seems that these investigators have discontinued such efforts. For the treatment of prostate cancer with gene therapy, improvements in the design of the viral vector as well as the gene transfer approach may increase efficacy, especially with respect to transduction efficiency and transgene expression.

We have developed an improved Ad-p53. Instead of using a constitutive promoter, we have developed a p53-responsive promoter (PG), which was initially incorporated in a retroviral vector. This modified expression system could surpass the parental unmodified vector by up to seven-fold (69). When the p53 gene was placed under control of this PG promoter, an autoregulated positive feedback mechanism was established, leading to more robust inhibition of tumor cell proliferation (70). Next, we transferred this expression system to an adenoviral vector and observed that this promoter was 5-fold stronger than the CMV promoter (71). This vector, named Ad-PGp53, provides higher levels of p53 expression than Ad-CMVp53. A schematic representation of these two vectors is depicted in Figure 1, which are similar to the commercial vectors tested in several clinical trials. We showed that Ad-PGp53 was better able to induce cell death in vitro and in vivo than Ad-CMVp53, and in situ gene therapy resulted in reduced tumor volume and increased overall survival only with Ad-PGp53. In this same work, we observed that the PC3 prostate carcinoma cell line was not efficiently transduced by Ad5 (72). Therefore, we made an additional improvement, incorporating the RGD motif in the fiber protein, thus creating AdRGD-PGp53, which offers both enhanced transduction efficiency in PC3 cells and a high level of p53 expression due to the positive feedback mechanism. This vector showed strong antitumor activity in vitro and in vivo, inducing high levels of reactive oxygen species (ROS), DNA damage and alteration of mitochondrial membrane permeability and resulting in apoptosis (21). Even though this improved



**Figure 1** - Schematic representation of non-replicating serotype 5 adenoviral vectors. Top, Ad-PGp53 featuring the p53 responsive promoter. Leaky expression of the p53 cDNA initiates binding of p53 to the PG promoter, leading to high level p53 expression due to the positive feedback mechanism. In this way, p53 serves to both drive expression as well as act as a tumor suppressor. Bottom, typical Ad-p53 vector where a constitutive promoter is used to drive expression of the p53 cDNA. LITR, left inverted terminal repeat; PG, PGTx $\beta$  chimeric p53-responsive promoter; CMV, cytomegalovirus immediate early enhancer/promoter; p53, wild-type cDNA, PolyA, poly-adenylation signal; Ad5( $\Delta$ E1/E3), adenoviral genome deficient in the E1 and E3 genes; RITR, right inverted terminal repeat.



adenoviral vector has strong antitumor activity against prostate carcinoma cell lines, even better than the versions of Ad-p53 tested in clinical trials, additional benefit may be seen if combined with chemotherapeutic agents.

# Role of p53 in the response to prostate cancer chemotherapeutic agents

For prostate cancer, the most commonly used chemotherapy agents are mitoxantrone, docetaxel and cabazitaxel. One of the first drugs used for treatment of CRPC was mitoxantrone, a synthetic anthracenedione derivative with immunomodulatory and antineoplastic activity that was approved by the FDA in 1987 for treatment of different cancers and in 1996 for prostate cancer. Clinical trials indicated that the combination of mitoxantrone plus prednisone and corticosteroid improved quality of life without affecting survival (8). Its mechanism of action involves cytotoxic activity through intercalation with DNA and inhibition of topoisomerase II (73), resulting in inhibition of replication and transcription (74,75). It also induces double-strand breaks, leading to activation of p53 and its accumulation in the nucleus (76), indicating that p53 status may be important to determine drug sensitivity. Mitoxantrone resistance can be induced by alterations in topoisomerase II or by P-glycoprotein (P-gp, MDR1, ABCB1) overexpression, which results in reduced drug accumulation inside the cell (77).

Docetaxel is the standard chemotherapeutic agent used to treat patients with prostate cancer (78). Docetaxel was synthesized from a precursor (10-deacetylbaccatin III) originally isolated from the needles of the European yew, Taxus baccata (79). It has an antimitotic effect by binding to free tubulin, promoting the formation of stable microtubules, preventing depolymerization and therefore inhibiting mitosis and inducing apoptosis (80). Collapse of microtubules results in the induction of p53, activation or inactivation of a variety of protein kinases and inhibition of cyclin-dependent kinases, resulting in cell cycle arrest in the G2/M phase. Other proapoptotic activities of docetaxel include downregulation of Bcl-2, upregulation of p53 and/or  $p21^{WAF-1}$  and induction of the phosphorylation of Bcl-X(L)/Bcl-2 members (81). Compared to wild-type p53 cells, prostate cancer cells expressing mutant p53 demonstrated reduced sensitivity in response to docetaxel, indicating that functional p53 is essential for sensitivity to docetaxel in prostate cancer cells (82,83). Docetaxel is an effective therapy against prostate cancer, but in some cases, it fails, requiring the use of alternative drugs, including cabazitaxel. Drug resistance is a major barrier for the use of docetaxel, and overcoming this impediment has been a challenge (84). The mechanisms of resistance include overexpression of P-gp (85) and altered beta-tubulin isotypes, as well as deregulation of cell survival and transcription factors (85,86).

Cabazitaxel is a taxane approved by FDA in June 2010 for treatment of prostate cancer subsequent to the use of docetaxel. Like docetaxel, cabazitaxel suppresses the dynamics of microtubules, resulting in inhibition of proliferation and cellular arrest by inducing mitotic spindle deformity. However, cabazitaxel is more efficient (87,88) since it has lower affinity for P-gp and remains inside the cell for more time (89). Resistance to cabazitaxel has been noted, though little is known about the mechanism (90). Even so, the ETS-related gene is overexpressed in prostate cells and leads to resistance to cabazitaxel treatment (91) by aborting p53 function, deregulating apoptosis, overexpressing HER2 and inhibiting tumor cell permeability (92).

# Combined therapy

The combination of Ad-p53 with chemotherapy may benefit both approaches. In a phase II clinical trial, NSCLC patients were treated with Ad-p53, docetaxel or a combination of both, and the median survival time was 7.7 months for patients who received both therapies and 5.9 months for patients who received only docetaxel (93). Patients with stage III or IV oral carcinoma were treated with Ad-p53 and chemotherapy (carboplatin, bleomycin and methotrexate), and the patients with stage III disease treated with the combined therapy had increased survival (94). Clinical trials combining Ad-p53 and chemotherapy provided a synergistic effect (93-98) of reducing side effects and increasing the quality of life and disease control compared to patients treated with only chemotherapy (99). The combination of Ad-p53 with chemotherapy may provide a therapeutic advantage in prostate carcinoma.

There is an intimate relationship between chemotherapy and p53, where the cellular p53 status is important for the prediction of drug efficacy. In ovarian cancer cells, the combination of Ad-p53 and docetaxel was positive only in cells expressing mutant p53, while the treatment was ineffective in cells with wild-type p53 status (100). In NSCLC, the combination of radiotherapy, docetaxel and Ad-p53 reduced tumor growth (101,102) regardless of the cell p53 status (103). Interestingly, Ad-p53 may be especially advantageous in chemoresistant cells since breast cancer cell lines resistant to mitoxantrone were shown to be more sensitive to Ad-p53 compared to drug-sensitive cells (104).

The combination of Ad-p53 and docetaxel resulted in enhanced antitumor effects in a murine model of HNSCC (105). Docetaxel was shown to upregulate CAR in HNSCC cells and cooperate with Ad-p53 to increase the expression of bax and the cleavage of PARP and caspase-3 (106). At the same time, Ad-p53 also favors chemotherapeutic activity by suppressing hepatic enzymes and reducing docetaxel clearance (107). In prostate cancer, the combination of Ad-p53 and cisplatin reduced tumor volume in a xenograft mouse model (108). In vivo, the combination of antisense clusterin oligodeoxynucleotides, mitoxantrone and Ad-p53 eradicated subcutaneous and orthotopic PC3 tumors (109). Docetaxel combined with CV787 (PSA + conditionally replicating adenovirus) synergistically reduced prostate cancer in a xenograft mouse model, where the combinatorial treatment increased the expression of p53 (27). In prostate cancer cell lines, docetaxel and paclitaxel were shown to increase adenoviral transgene expression (110). The combination of chemotherapeutic drugs (cisplatin, docetaxel, mitoxantrone, paclitaxel and etoposide) with an oncolytic adenovirus showed a synergistic response in prostate cancer cell lines; in particular, docetaxel and mitoxantrone were shown to increase viral uptake, exhibiting a trend of increased levels of integrin  $\alpha v\beta 3/\beta 5$  after treatment of DU145 and LNCaP cells with either drug, as well as a trend of increased CAR expression in PC3 cells. The combination of oncolytic adenovirus and docetaxel prolonged overall survival and reduced tumor volume (111).

Prostate cancer is one of the most important tumors in men. Development of new treatments has shown promising results, including gene transfer approaches as an appealing alternative.



Thus, gene therapy is slowly regaining lost territory in the treatment of prostate cancer. The use of prostate cancerspecific oncolytic viruses and suicide gene therapy has reached clinical trials. Adenoviruses expressing the tumor suppressor p53 are employed for HNSCC, but their use has been limited in prostate cancer. We have shown that improvements in the transgene expression system and alteration of viral tropism may improve the suppressor activity of an Ad-p53. Even so, gene therapy may work in cooperation with traditional chemotherapy, benefiting both approaches and bringing about synergistic activity as an effective prostate cancer treatment.

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# AUTHOR CONTRIBUTIONS

Tamura RE, Luna IV, Lana MG and Strauss BE elaborated and reviewed text. Luna IV designed the figure.

#### REFERENCES

- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2017. CA Cancer J Clin. 1. 2017;67(1):7-30, http://dx.doi.org/10.3322/caac.21387
- 2. Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. Int J Cancer. 2015;136(5):E359-86, http:// dx.doi.org/10.1002/ijc.29210.
- 3. Singh J, Trabulsi EJ, Gomella LG. Is there an optimal management for localized prostate cancer? Clin Interv Aging. 2010;5:187-97.
- 4. Dean JL, Knudsen KE. The role of tumor suppressor dysregulation in prostate cancer progression. Curr Drug Targets. 2013;14(4):460-71, http:// dx.doi.org/10.2174/1389450111314040007.
- Ryan CJ, Elkin EP, Small EJ, Duchane J, Carroll P. Reduced incidence of bony metastasis at initial prostate cancer diagnosis: data from CaPSURE. Urol Oncol. 2006;24(5):396-402, http://dx.doi.org/10.1016/j.urolonc.2005. 09 003
- 6. Debes JD, Tindall DJ. Mechanisms of androgen-refractory prostate cancer. N Engl J Med. 2004;351(15):1488-90, http://dx.doi.org/10.1056/NEJMp 048178
- Chandrasekar T, Yang JC, Gao AC, Evans CP. Mechanisms of resistance in castration-resistant prostate cancer (CRPC). Transl Androl Urol. 2015; (43):365-80, http://dx.doi.org/10.3978/j.issn.2223-4683.2015.05.02. Tannock IF, de Wit R, Berry WR, Horti J, Pluzanska A, Chi KN, et al.
- TAX 327 Investigators. Docetaxel plus prednisone or mitoxantrone plus prednisone for advanced prostate cancer. N Engl J Med. 2004;351(15): 1502-12, http://dx.doi.org/10.1056/NEJMoa040720. Heidenreich A, Porres D. Prostate cancer: treatment sequencing for CRPC–what do we know? Nat Rev Urol. 2014;11(4):189-90, http://dx.
- doi.org/10.1038/nrurol.2014.36.
- 10. de Bono JS, Oudard S, Ozguroglu M, Hansen S, Machiels JP, Kocak I, et al. TROPIC Investigators. Prednisone plus cabazitaxel or mitoxantrone for metastatic castration-resistant prostate cancer progressing after docetaxel treatment: a randomised open-label trial Lancet. 2010;376(9747): 1147-54, http://dx.doi.org/10.1016/S0140-6736(10)61389-X.
- Debruyne F. Hormonal therapy of prostate cancer. Semin Urol Oncol. 11. 2002;20(3 Suppl 1):4-9.
- Lee SJ, Kim HS, Yu R, Lee K, Gardner TA, Jung C, et al. Novel prostate-specific promoter derived from PSA and PSMA enhancers. Mol Ther. 2002;6(3):415-21, http://dx.doi.org/10.1006/mthe.2002.0682
- 13. Wu L, Matherly J, Smallwood A, Adams JY, Billick E, Belldegrun A, et al. Chimeric PSA enhancers exhibit augmented activity in prostate cancer gene therapy vectors. Gene Ther. 2001;8(18):1416-26, http://dx.doi.org/ 10.1038/sj.gt.3301549.
- 14. Kraaij R, van der Weel L, de Ridder CM, van der Korput HA, Zweistra JL, van Rijswijk AL, et al. A small chimeric promoter for high prostatespecific transgene expression from adenoviral vectors. Prostate. 2007;67(8): 829-39, http://dx.doi.org/10.1002/pros.20560.
- 15. Shi CX, Hitt M, Ng P, Graham FL. Superior tissue-specific expression from tyrosinase and prostate-specific antigen promoters/enhancers in helper dependente compared with first-generation adenoviral vectors. Hum Gene Ther. 2002;13(2):211-24, http://dx.doi.org/10.1089/104303 40252769743.

- 16. Latham JP, Searle PF, Mautner V, James ND. Prostate-specific antigen promoter/enhancer driven gene therapy for prostate cancer: construction and testing of a tissue-specific adenovirus vector. Cancer Res. 2000;60(2):334-41
- 17. Lu Y, Zhang Y, Chang G, Zhang J. Comparison of prostate-specific promoters and the use of PSP-driven virotherapy for prostate cancer. Biomed Res Int. 2013;2013:624632, http://dx.doi.org/10.1155/2013/ 624632
- 18. Li X, Liu YH, Lee SJ, Gardner TA, Jeng MH, Kao C. Prostate-restricted replicative adenovirus expressing human endostatin-angiostatin fusion gene exhibiting dramatic antitumor efficacy. Clin Cancer Res. 2008;14(1): 291-9, http://dx.doi.org/10.1158/1078-0432.CCR-07-0867.
- 19. Azab BM, Dash R, Das SK, Bhutia SK, Sarkar S, Shen XN, et al. Enhanced prostate cancer gene transfer and therapy using a novel serotype chi-mera cancer terminator virus (Ad.5/3-CTV). J Cell Physiol. 2014;229(1): 34-43, http://dx.doi.org/10.1002/jcp.24408.
- 20. Shen YH, Yang F, Wang H, Cai ZJ, Xu YP, Zhao A, et al. Arg-Gly-Asp (RGD)-Modified E1A/E1B Double Mutant Adenovirus Enhances Antitumor Activity in Prostate Cancer Cells In Vitro and in Mice. PLoS One. 2016;11(1):e0147173, http://dx.doi.org/10.1371/journal.pone.0147173.
- Tamura RE, Hunger A, Fernandes DC, Laurindo FR, Costanzi-Strauss E, 21. Strauss BE. Induction of Oxidants Distinguishes Susceptibility of Prostate Carcinoma Cell Lines to p53 Gene Transfer Mediated by an Improved Adenoviral Vector. Hum Gene Ther. 2017;28(8):639-53, http://dx.doi.org/ 10.1089/hum.2016.139
- Li ZL, Liang X, Li HC, Wang ZM, Chong T. Dendritic cells serve as a "Trojan horse" for oncolytic adenovirus delivery in the treatment of 22. mouse prostate cancer. Acta Pharmacol Sin. 2016;37(8):1121-8, http://dx. doi.org/10.1038/aps.2016.59.
- 23. Nguyen TV, Heller GJ, Barry ME, Crosby CM, Turner MA, Barry MA. Evaluation of polymer shielding for adenovirus serotype 6 (Ad6) for systemic virotherapy against human prostate cancers. Mol Ther Onco-
- Jytics. 2016;3:15021, http://dx.doi.org/10.1038/mto.2015.21. Wang GX, Zhan YA, Hu HL, Wang Y, Fu B. Mesenchymal stem cells modified to express interferon- $\beta$  inhibit the growth of prostate cancer in a mouse model. J Int Med Res. 2012;40(1):317-27, http://dx.doi.org/ 10.1177/147323001204000132.
- DeWeese TL, van der Poel H, Li S, Mikhak B, Drew R, Goemann M, et al. 25 A phase I trial of CV706, a replication-competent, PSA selective oncolytic adenovirus, for the treatment of locally recurrent prostate cancer following radiation therapy. Cancer Res. 2001;61(20):7464-72.
- Small EJ, Carducci MA, Burke JM, Rodriguez R, Fong L, van Ummersen L, et al. A phase I trial of intravenous CG7870, a replication-selective, prostate-specific antigen targeted oncolytic adenovirus, for the treatment of hormone-refractory, metastatic prostate cancer. Mol Ther. 2006;14(1): 107-17, http://dx.doi.org/10.1016/j.ymthe.2006.02.011.
- 27. Yu DC, Chen Y, Dilley J, Li Y, Embry M, Zhang H, et al. Antitumor synergy of CV787, a prostate cancer-specific adenovirus, and paclitaxel and docetaxel. Cancer Res. 2001;61(2):517-25.
- Cheng WS, Dzojic H, Nilsson B, Tötterman TH, Essand M. An oncolytic conditionally replicating adenovirus for hormone-dependent and hormoneindependent prostate cancer. Cancer Gene Ther. 2006;13(1):13-20, http://dx.doi.org/10.1038/sj.cgt.7700881.
- Lubaroff DM, Konety BR, Link B, Gerstbrein J, Madsen T, Shannon M, 29. et al. Phase I clinical trial of na adenovirus/prostate-specific antigen vaccine for prostate cancer: safety and immunologic results. Clin Cancer Res. 2009;15(23):7375-80, http://dx.doi.org/10.1158/1078-0432.CCR-09-1910.
- 30. Lubaroff DM, Vaena D, Brown JA, Zehr P, Griffith KC, Brown E, et al. Vaccine immunotherapy for prostate cancer: from mice to men. Immunol Res. 2014;59(1-3):229-35, http://dx.doi.org/10.1007/s12026-014-8531-2.
- Li X, Liu YH, Zhang YP, Zhang S, Pu X, Gardner TA, et al. Faz ligand delivery by a prostate-restricted replicative adenovirus enhances safety 31. and antitumor efficacy. Clin Cancer Res. 2007;13(18 Pt 1):5463-73.
- Kumon H, Sasaki K, Ariyoshi Y, Sadahira T, Araki M, Ebara S, et al. 32. Feasibility of Neoadjuvant Ad-REIC Gene Therapy in Patients with High-Risk Localized Prostate Cancer Undergoing Radical Prostatectomy. Clin Transl Sci. 2015;8(6):837-40, http://dx.doi.org/10.1111/cts.12362
- Sonpavde G, Thompson TC, Jain RK, Ayala GE, Kurosaka S, Edamura K, 33. et al. GLIPR1 tumor suppressor gene expressed by adenoviral vector as neoadjuvant intraprostatic injection for localized intermediate or highrisk prostate cancer preceding radical prostatectomy. Clin Cancer Res. 2011;17(22):7174-82, http://dx.doi.org/10.1158/1078-0432.CCR-11-1899.
- 34. Yang YF, Xue SY, Lu ZZ, Xiao FJ, Yin Y, Zhang QW, et al. Antitumor effects of oncolytic adenovirus armed with PSA-IZ-CD40L fusion gene against prostate cancer. Gene Ther. 2014;21(8):723-31, http://dx.doi.org/ 10.1038/gt.2014.46.
- Freytag SO, Khil M, Stricker H, Peabody J, Menon M, DePeralta-Venturina M, et al. Phase I study of replication-competent adenovirusmediated double suicide gene therapy for the treatment of locally recurrent prostate cancer. Cancer Res. 2002;62(17):4968-76.
- Freytag SO, Stricker H, Pegg J, Paielli D, Pradhan DG, Peabody J, et al. 36. Phase I study of replication competent adenovirus-mediated double-suicide



gene therapy in combination with conventional-dose three-dimensional conformal radiation therapy for the treatment of newly diagnosed, intermediate- to high-risk prostate cancer. Cancer Res. 2003;63(21):7497-506.

- Nasu Y, Saika T, Ebara S, Kusaka N, Kaku H, Abarzua F, et al. Suicide gene therapy with adenoviral delivery of HSV-tK gene for patients with local recurrence of prostate cancer after hormonal therapy. Mol Ther. 2007;15(4):834-40, http://dx.doi.org/10.1038/sj.mt.6300096.
- Herman JR, Adler HL, Aguilar-Cordova E, Rojas-Martinez A, Woo S, Timme TL, et al. In situ gene therapy for adenocarcinoma of the prostate: a phase I clinical trial. Hum Gene Ther. 1999;10(7):1239-49, http://dx. doi.org/10.1089/10430349950018229.
- Shirakawa T, Terao S, Hinata N, Tanaka K, Takenaka A, Hara I, et al. Long-term outcome of phase I/II clinical trial of Ad-OC-TK/VAL gene therapy for hormone-refractory metastatic prostate cancer. Hum Gene Ther. 2007;18(12):1225-32, http://dx.doi.org/10.1089/hum.2007.074.
- Freytag SO, Stricker H, Peabody J, Pegg J, Paielli D, Movsas B, et al. Five-year follow-up of trial of replication-competent adenovirus-mediated suicide gene therapy for treatment of prostate cancer. Mol Ther. 2007;15(3): 636-42, http://dx.doi.org/10.1038/sj.mt.6300068.
- Freytag SÖ, Movsas B, Åref I, Stricker H, Peabody J, Pegg J, et al. Phase I trial of replication competente adenovirus-mediated suicide gene therapy combined with IMRT for prostate cancer. Mol Ther. 2007;15(5):1016-23, http://dx.doi.org/10.1038/mt.sj.6300120.
- Rojas-Martínez A, Manzanera ÁG, Sukin SW, Esteban-María J, González-Guerrero JF, Gomez-Guerra L, et al. Intraprostatic distribution and long term follow-up after AdV-tk immunotherapy as neoadjuvant to surgery in patients with prostate cancer. Cancer Gene Ther. 2013;20(11):642-9, http:// dx.doi.org/10.1038/cgt.2013.56.
- Bunz F, Dutriaux A, Lengauer C, Waldman T, Zhou S, Brown JP, et al. Requirement for p53 and p21 to sustain G2 arrest after DNA damage. Science. 1998;282(5393):1497-501, http://dx.doi.org/10.1126/science.282. 5393.1497.
- Nakano K, Vousden KH. PUMA, a novel proapoptotic gene, is induced by p53. Mol Cell. 2000;7(3):683-94, http://dx.doi.org/10.1016/S1097-2765(01)00214-3.
- Brooks JD, Bova GS, Ewing CM, Piantadosi S, Carter BS, Robinson JC, et al. An uncertain role for p53 gene alterations in human prostate cancers. Cancer Res. 1996;56(16):3814-22.
- Leroy B, Fournier JL, Ishioka C, Monti P, Inga A, Fronza G, et al. The TP53 website: an integrative resource centre for the TP53 mutation database and TP53 mutant analysis. Nucleic Acids Res. 2013;41 (Database issue):D962-9.
- Kluth M, Harasimowicz S, Burkhardt L, Grupp K, Krohn A, Prien K, et al. Clinical significance of different types of p53 gene alteration in surgically treated prostate cancer. Int J Cancer. 2014;135(6):1369-80, http://dx.doi.org/10.1002/ijc.28784.
- Bettendorf O, Schmidt H, Staebler A, Grobholz R, Heinecke A, Boecker W, et al. Chromosomal imbalances, loss of heterozygosity, and immunohistochemical expression of TP53, RB1, and PTEN in intraductal cancer, intraepithelial neoplasia, and invasive adenocarcinoma of the prostate. Genes Chromosomes Cancer. 2008;47(7):565-72, http://dx.doi.org/10.1002/ gcc.20560.
- Agell L, Hernández S, de Muga S, Lorente JA, Juanpere N, Esgueva R, et al. KLF6 and TP53 mutations are a rare event in prostate cancer: distinguishing between Taq polymerase artifacts and true mutations. Mod Pathol. 2008;21(12):1470-8, http://dx.doi.org/10.1038/modpathol. 2008.145.
- Das K, Lau W, Sivaswaren C, Ph T, Fook-Chong S, Sl T, et al. Chromosomal changes in prostate cancer: a fluorescence in situ hybridization study. Clin Genet. 2005;68(1):40-7, http://dx.doi.org/10.1111/j.1399-0004. 2005.00452.x.
- Roth JA, Nguyen D, Lawrence DD, Kemp BL, Carrasco CH, Ferson DZ, et al. Retrovirus-mediated wild-type p53 gene transfer to tumors of patients with lung cancer. Nat Med. 1996;2(9):985-91, http://dx.doi.org/ 10.1038/nm0996-985.
- Lane DP, Cheok CF, Lain S. p53-based cancer therapy. Cold Spring Harb Perspect Biol. 2010;2(9):a001222, http://dx.doi.org/10.1101/cshperspect. a001222.
- Zhang WW, Fang X, Mazur W, French BA, Georges RN, Roth JA. High efficiency gene transfer and high-level expression of wild-type p53 in human lung câncer cells mediated by recombinant adenovirus. Cancer Gene Ther. 1994;1(1):5-13.
- Gene Ther. 1994;1(1):5-13.
  54. INGN 201: Ad-p53, Ad5CMV-p53, Adenoviral p53, INGN 101, p53 gene therapy--Introgen, RPR/INGN 201. BioDrugs. 2003;17(3):216-22.
- Wills KN, Maneval DC, Menzel P, Harris MP, Sutjipto S, Vaillancourt MT, et al. Development and characterization of recombinant adenoviruses encoding human p53 for gene therapy of cancer. Hum Gene Ther. 1994; 5(9):1079-88, http://dx.doi.org/10.1089/hum.1994.5.9-1079.
- Schuler M, Herrmann R, De Greve JL, Stewart AK, Gatzemeier U, Stewart DJ, et al. Adenovirus-mediated wild-type p53 gene transfer in patients receiving chemotherapy for advanced non-small-cell lung cancer: results of a multicenter phase II study. J Clin Oncol. 2001;19(6):1750-8, http://dx.doi.org/10.1200/JCO.2001.19.6.1750.

- Schuler M, Rochlitz C, Horowitz JA, Schlegel J, Perruchoud AP, Kommoss F, et al. A phase I study of adenovirus-mediated wild-type p53 gene transfer in patients with advanced non-small cell lung cancer. Hum Gene Ther. 1998;9(14):2075-82, http://dx.doi.org/10.1089/hum.1998.9.14-2075.
- Buller RE, Shahin MS, Horowitz JA, Runnebaum IB, Mahavni V, Petrauskas S, et al. Long term follow-up of patients with recurrent ovarian cancer after Ad p53 gene replacement with SCH 58500. Cancer Gene Ther. 2002;9(7): 567-72, http://dx.doi.org/10.1038/sj.cgt.7700473.
- Buller RE, Runnebaum IB, Karlan BY, Horowitz JA, Shahin M, Buekers T, et al. A phase I/II trial of rAd/p53 (SCH 58500) gene replacement in recurrent ovarian cancer. Cancer Gene Ther. 2002;9(7):553-66, http://dx.doi. org/10.1038/sj.cgt.7700472.
- Kuball J, Wen SF, Leissner J, Atkins D, Meinhardt P, Quijano E, et al. Successful adenovirus-mediated wild-type p53 gene transfer in patients with bladder cancer by intravesical vector instillation. J Clin Oncol. 2002;20(4):957-65, http://dx.doi.org/10.1200/JCO.2002.20.4.957.
- Atencio IA, Grace M, Bordens R, Fritz M, Horowitz JA, Hutchins B, et al. Biological activities of a recombinant adenovirus p53 (SCH 58500) administered by hepatic arterial infusion in a Phase 1 colorectal cancer trial. Cancer Gene Ther. 2006;13(2):169-81, http://dx.doi.org/10.1038/ sj.cgt.7700870.
- Peng Z. Current status of gendicine in China: recombinant human Ad-p53 agent for treatment of cancers. Hum Gene Ther. 2005;16(9):1016-27, http://dx.doi.org/10.1089/hum.2005.16.1016.
- Poeta ML, Manola J, Goldwasser MA, Forastiere A, Benoit N, Califano JA, et al. TP53 mutations and survival in squamous-cell carcinoma of the head and neck. N Engl J Med. 2007;357(25):2552-61, http://dx.doi.org/ 10.1056/NEJMoa073770.
- Pisters LL, Pettaway CA, Troncoso P, McDonnell TJ, Stephens LC, Wood CG, et al. Evidence that transfer of functional p53 protein results in increased apoptosis in prostate cancer. Clin Cancer Res. 2004;10(8): 2587-93, http://dx.doi.org/10.1158/1078-0432.CCR-03-0388.
- Ko SC, Gotoh A, Thalmann GN, Zhau HE, Johnston DA, Zhang WW, et al. Molecular therapy with recombinant p53 adenovirus in na androgenindependent, metastatic human prostate cancer model. Hum Gene Ther. 1996;7(14):1683-91, http://dx.doi.org/10.1089/hum.1996.7.14-1683.
- 66. Asgari K, Sesterhenn IA, McLeod ĎG, Cowan K, Moul JW, Seth P, et al. Inhibition of the growth of pre-established subcutaneous tumor nodules of human prostate cancer cells by single injection of the recombinant adenovirus p53 expression vector. Int J Cancer. 1997;71(3):377-82, http:// dx.doi.org/10.1002/(SICI)1097-0215(19970502)71:3 < 377::AID-IJC13 > 3.0. CO;2-D.
- Yang C, Cirielli C, Capogrossi MC, Passaniti A. Adenovirus-mediated wild type p53 expression induces apoptosis and suppresses tumorigenesis of prostatic tumor cells. Cancer Res. 1995;55(19):4210-3.
- Eastham JA, Hall SJ, Sehgal I, Wang J, Timme TL, Yang G, et al. In vivo gene therapy with p53 or p21 adenovirus for prostate cancer. Cancer Res. 1995;55(22):5151-5.
- Strauss BE, Costanzi-Strauss E. pCLPG: a p53-driven retroviral system. Virology. 2004;321(2):165-72, http://dx.doi.org/10.1016/j.virol.2003.12.021.
- Strauss BE, Bajgelman MC, Costanzi-Strauss E. A novel gene transfer strategy that combines promoter and transgene activities for improved tumor cell inhibition. Cancer Gene Ther. 2005;12(12):935-46, http://dx. doi.org/10.1038/sj.cgt.7700846.
- Bajgelman MC, Strauss BE. Development of an adenoviral vector with robust expression driven by p53. Virology. 2008;371(1):8-13, http://dx. doi.org/10.1016/j.virol.2007.11.015.
- Tamura RE, da Silva Soares RB, Costanzi-Strauss E, Strauss BE. Autoregulated expression of p53 from an adenoviral vector confers superior tumor inhibition in a model of prostate carcinoma gene therapy. Cancer Biol Ther. 2016;17(12):1221-30, http://dx.doi.org/10.1080/15384047. 2016.1235655.
- Teply BA, Hauke RJ. Chemotherapy options in castration-resistant prostate cancer. Indian J Urol. 2016;32(4):262-70, http://dx.doi.org/ 10.4103/0970-1591.191239.
- Kapuscinski J, Darzynkiewicz Z. Interactions of antitumor agentes Ametantrone and Mitoxantrone (Novatrone) with double-stranded DNA. Biochem Pharmacol. 1985;34(24):4203-13, http://dx.doi.org/10.1016/0006-2952(85)90275-8.
- Kapuscinski J, Darzynkiewicz Z. Relationship between the pharmacological activity of antitumor drugs Ametantrone and mitoxantrone (Novatrone) and their ability to condense nucleic acids. Proc Natl Acad Sci U S A. 1986;83(17):6302-6, http://dx.doi.org/10.1073/pnas.83.17.6302.
   Ć Ćmielová J, Lesná M, Řezáčová M. Subcellular Localization of Proteins
- Ć Ćmielová J, Lesná M, Řezáčová M. Subcellular Localization of Proteins Responding to Mitoxantrone-Induced DNA Damage in Leukaemic Cells. Folia Biol (Praha). 2015;61(2):60-5.
- Hazlehurst LA, Foley NE, Gleason-Guzman MC, Hacker MP, Cress AE, Greenberger LW, et al. Multiple mechanisms confer drug resistance to mitoxantrone in the human 8226 myeloma cell line. Cancer Res. 1999;59(5): 1021-8.
- Yvon AM, Wadsworth P, Jordan MA. Taxol suppresses dynamics of individual microtubules in living human tumor cells. Mol Biol Cell. 1999;10(4):947-59, http://dx.doi.org/10.1091/mbc.10.4.947.



- Wani MC, Taylor HL, Wall ME, Coggon P, McPhail AT. Plant antitumor agents. VI. The isolation and structure of taxol, a novel antileukemic and antitumor agent from Taxus brevifolia. J Am Chem Soc. 1971;93(9):2325-7, http://dx.doi.org/10.1021/ja00738a045.
- Dagher R, Li N, Abraham S, Rahman A, Sridhara R, Pazdur R. Approval summary: Docetaxel in combination with prednisone for the treatment of androgen-independent hormone-refractory prostate cancer. Clin Cancer Res. 2004;10(24):8147-51, http://dx.doi.org/10.1158/1078-0432.CCR-04-1402.
- Ganansia-Leymarie V, Bischoff P, Bergerat JP, Holl V. Signal transduction pathways of taxanes-induced apoptosis. Curr Med Chem Anticancer Agents. 2003;3(4):291-306, http://dx.doi.org/10.2174/1568011033482422.
- Liu C, Zhu Y, Lou W, Nadiminty N, Chen X, Zhou Q, et al. Functional p53 determines docetaxel sensitivity in prostate cancer cells. Prostate. 2013;73(4):418-27, http://dx.doi.org/10.1002/pros.22583.
- Chappell WH, Lehmann BD, Terrian DM, Abrams SL, Steelman LS, McCubrey JA. p53 expression controls prostate cancer sensitivity to chemotherapy and the MDM2 inhibitor Nutlin-3. Cell Cycle. 2012;11(24): 4579-88, http://dx.doi.org/10.4161/cc.22852.
- Hwang C. Overcoming docetaxel resistance in prostate cancer: a perspective review. Ther Adv Med Oncol. 2012;4(6):329-40, http://dx.doi.org/ 10.1177/1758834012449685.
- Galletti E, Magnani M, Renzulli ML, Botta M. Paclitaxel and docetaxel resistance: molecular mechanisms and development of new generation taxanes. ChemMedChem. 2007;2(7):920-42, http://dx.doi.org/10.1002/ cmdc.200600308.
- Kosaka T, Miyajima A, Shirotake S, Suzuki E, Kikuchi E, Oya M. Long-term androgen ablation and docetaxel up-regulate phosphorylated Akt in castration resistant prostate cancer. J Urol. 2011;185(6):2376-81, http://dx.doi.org/10.1016/j.juro.2011.02.016.
- Paller CJ, Antonarakis ES. Cabazitaxel: a novel second-line treatment for metastatic castration-resistant prostate cancer. Drug Des Devel Ther. 2011;5:117-24.
- Azarenko O, Smiyun G, Mah J, Wilson L, Jordan MA. Antiproliferative mechanism of action of the novel taxane cabazitaxel as compared with the parent compound docetaxel in MCF7 breast cancer cells. Mol Cancer Ther. 2014;13(8):2092-103, http://dx.doi.org/10.1158/1535-7163.MCT-14-0265.
- Kartner N, Riordan JR, Ling V. Cell surface P-glycoprotein associated with multidrug resistance in mammalian cell lines. Science. 1983;221(4617): 1285-8, http://dx.doi.org/10.1126/science.6137059.
- Armstrong CM, Gao AC. Drug resistance in castration resistant prostate cancer: resistance mechanisms and emerging treatment strategies. Am J Clin Exp Urol. 2015;3(2):64-76.
- Galletti G, Matov A, Beltran H, Fontugne J, Miguel Mosquera J, Cheung C, et al. ERG induces taxane resistance in castration-resistant prostate cancer. Nat Commun. 2014;5:5548, http://dx.doi.org/10.1038/ncomms6548.
- Vrignaud P, Semiond D, Benning V, Beys E, Bouchard H, Gupta S. Preclinical profile of cabazitaxel. Drug Des Devel Ther. 2014;8:1851-67, http://dx.doi.org/10.2147/DDDT.S64940.
- Ning X, Sun Z, Wang Y, Zhou J, Chen S, Chen D, et al. Docetaxel plus trans-tracheal injection of adenoviral-mediated p53 versus docetaxel alone in patients with previously treated non-small-cell lung cancer. Cancer Gene Ther. 2011;18(6):444-9, http://dx.doi.org/10.1038/cgt.2011.15.
- Li Y, Li LJ, Wang LJ, Zhang Z, Gao N, Liang CY, et al. Selective intraarterial infusion of rAd-p53 with chemotherapy for advanced oral cancer: a randomized clinical trial. BMC Med. 2014;12:16, http://dx.doi.org/ 10.1186/1741-7015-12-16.
- Xiao J, Zhou J, Fu M, Liang L, Deng Q, Liu X, et al. Efficacy of recombinant human adenovirus-p53 combined with chemotherapy for locally advanced cervical cancer: a clinical trial. Oncol Lett. 2017;13(5):3676-80, http://dx.doi.org/10.3892/ol.2017.5901.
- Dong M, Li X, Hong LJ, Xie R, Zhao HL, Li K, et al. Advanced malignant pleural or peritoneal effusion in patients treated with recombinant adenovirus p53 injection plus cisplatin. J Int Med Res. 2008;36(6):1273-8, http://dx.doi.org/10.1177/147323000803600614.

- Guan YS, Liu Y, Zou Q, He Q, La Z, Yang L, et al. Adenovirus-mediated wild type p53 gene transfer in combination with bronchial arterial infusion for treatment of advanced non-small-cell lung cancer, one year follow-up. J Zhejiang Univ Sci B. 2009;10(5):331-40, http://dx.doi.org/ 10.1631/jzus.B0820248.
- Fujiwara T, Tanaka N, Kanazawa S, Ohtani S, Saijo Y, Nukiwa T, et al. Multicenter phase I study of repeated intratumoral delivery of adenoviral p53 in patients with advanced non-small-cell lung cancer. J Clin Oncol. 2006;24(11):1689-99, http://dx.doi.org/10.1200/JCO.2005.03.4116.
- Biaoxue R, Hui P, Wenlong G, Shuanying Y. Evaluation of efficacy and safety for recombinant human adenovirus-p53 in the control of the malignant pleural effusions via thoracic perfusion. Sci Rep. 2016;6:39355, http://dx.doi.org/10.1038/srep39355.
- Miettinen S, Ylikomi T. Concomitant exposure of ovarian cancer cells to docetaxel, CPT-11 or SN-38 and adenovirus-mediated p53 gene therapy. Anticancer Drugs. 2009;20(7):589-600, http://dx.doi.org/10.1097/CAD. 0b013e32832dad3d.
- 101. Nishizaki M, Meyn RE, Levy LB, Atkinson EN, White RA, Roth JÁ, et al. Synergistic inhibition of human lung cancer cell growth by adenovírus mediated wild-type p53 gene transfer in combination with docetaxel and radiation therapeutics in vitro and in vivo. Clin Cancer Res. 2001;7(9): 2887-97.
- 102. Horio Y, Hasegawa Y, Sekido Y, Takahashi M, Roth JA, Shimokata K. Synergistic effects of adenovirus expressing wild-type p53 on chemosensitivity of non-small cell lung cancer cells. Cancer Gene Ther. 2000; 7(4):537-44, http://dx.doi.org/10.1038/sj.cgt.7700148.
- 103. Inoue A, Narumi K, Matsubara N, Sugawara S, Saijo Y, Satoh K, et al. Administration of wild-type p53 adenoviral vector synergistically enhances the cytotoxicity of anti-cancer drugs in human lung cancer cells irrespective of the status of p53 gene. Cancer Lett. 2000;157(1):105-12, http://dx.doi. org/10.1016/S0304-3835(00)00480-8.
- 104. Seth P, Katayose D, Li Z, Kim M, Wersto R, Craig C, et al. A recombinant adenovirus expressing wild type p53 induces apoptosis in drug-resistant human breast cancer cells: a gene therapy approach for drug-resistant cancers. Cancer Gene Ther. 1997;4(6):383-90.
- 105. Yoo GH, Subramanian G, Ezzat WH, Tulunay OE, Tran VR, Lonardo F, et al. Intratumoral delivery of docetaxel enhances antitumor activity of Ad-p53 in murine head and neck cancer xenograft model. Am J Otolaryngol. 2010;31(2):78-83, http://dx.doi.org/10.1016/j.amjoto.2008.10.002.
- Yoo GH, Piechocki MP, Oliver J, Lonardo F, Zumstein L, Lin HS, et al. Enhancement of Ad-p53 therapy with docetaxel in head and neck cancer. Laryngoscope. 2004;114(11):1871-9, http://dx.doi.org/10.1097/01.mlg. 0000147914.51239.ed.
- Wonganan P, Zamboni WC, Strychor S, Dekker JD, Croyle MA. Drug vírus interaction: effect of administration of recombinant adenoviruses on the pharmacokinetics of docetaxel in a rat model. Cancer Gene Ther. 2009;16(5):405-14, http://dx.doi.org/10.1038/cgt.2008.99.
   Gjerset R, Haghighi A, Lebedeva S, Mercola D. Gene therapy approaches
- 108. Gjerset R, Haghighi A, Lebedeva S, Mercola D. Gene therapy approaches to sensitization of human prostate carcinoma to cisplatin by adenoviral expression. of p53 and by antisense jun kinase oligonucleotide methods. Methods Mol Biol. 2001;175:495-520.
- 109. Yamanaka K, Gleave ME, Hara I, Muramaki M, Miyake H. Synergistic antitumor effect of combined use of adenoviral-mediated p53 gene transfer and antisense oligodeoxynucleotide targeting clusterin gene in an androgen independent human prostate cancer model. Mol Cancer Ther. 2005;4(2):187-95.
- 110. Li Y, Okegawa T, Lombardi DP, Frenkel EP, Hsieh JT. Enhanced transgene expression in androgen independent prostate cancer gene therapy by taxane chemotherapeutic agents. J Urol. 2002;167(1):339-46, http:// dx.doi.org/10.1016/S0022-5347(05)65465-1.
- 111. Radhakrishnan S, Miranda E, Ekblad M, Holford A, Pizarro MT, Lemoine NR, et al. Efficacy of oncolytic mutants targeting pRb and p53 pathways is synergistically enhanced when combined with cytotoxic drugs in prostate câncer cells and tumor xenografts. Hum Gene Ther. 2010;21(10): 1311-25, http://dx.doi.org/10.1089/hum.2010.019.