

Hydrogen, a potential safeguard for graft-versus-host disease and graft ischemia-reperfusion injury?

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Post-transplant complications such as graft-versus-host disease and graft ischemia-reperfusion injury are crucial challenges in transplantation. Hydrogen can act as a potential antioxidant, playing a preventive role against post-transplant complications in animal models of multiple organ transplantation. Herein, the authors review the current literature regarding the effects of hydrogen on graft ischemia-reperfusion injury and graft-versus-host disease. Existing data on the effects of hydrogen on ischemia-reperfusion injury related to organ transplantation are specifically reviewed and coupled with further suggestions for future work. The reviewed studies showed that hydrogen (inhaled or dissolved in saline) improved the outcomes of organ transplantation by decreasing oxidative stress and inflammation at both the transplanted organ and the systemic levels. In conclusion, a substantial body of experimental evidence suggests that hydrogen can significantly alleviate transplantation-related ischemia-reperfusion injury and have a therapeutic effect on graft-versus-host disease, mainly via inhibition of inflammatory cytokine secretion and reduction of oxidative stress through several underlying mechanisms. Further animal experiments and preliminary human clinical trials will lay the foundation for hydrogen use as a drug in the clinic.

KEYWORDS: Molecular Hydrogen; Organ Transplantation; Graft-Versus-Host Disease; Ischemia-Reperfusion Injury; Antioxidant.

Yuan L, Shen J. Hydrogen, a potential safeguard for graft-versus-host disease and graft ischemia-reperfusion injury? *Clinics*. 2016;71(9):544-549

Received for publication on April 29, 2016; First review completed on May 27, 2016; Accepted for publication on June 2, 2016

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INTRODUCTION

Transplantation is a therapeutic modality in which healthy cells, tissues, or organs (an autograft or allograft) are transplanted to restore the anatomical structure and function of damaged organs or tissues. This approach is the final treatment choice for untreatable diseases and end-stage organ diseases (1). Organ transplantation leads to benefits such as functional recovery and prolonged survival, but post-transplantation complications such as ischemia-reperfusion (I/R) injury and acute graft-versus-host disease (aGVHD) remain major challenges (2-4). These complications reduce patients' quality of life, increase medical costs, and worsen prognosis.

Graft ischemia-reperfusion injury

During the process of transplantation, blood flow to the organ to be transplanted is interrupted, leading to ischemia that can damage the organ. In addition, restoration of blood

flow to the transplanted organ may result in local and systemic inflammatory responses that can increase tissue injury. Graft I/R injury is characterized by reactive oxygen species (ROS) production, complement activation, leukocyte infiltration, platelet-leukocyte aggregation, increased microvascular permeability, and decreased endothelium-dependent relaxation (5,6). In the process of prolonged ischemia, adenosine triphosphate (ATP) levels and intracellular pH decrease because of anaerobic metabolism, leading to lactate accumulation. In addition, increased intracellular and mitochondrial calcium levels (calcium overload) are observed because certain ATPase-dependent ion transport mechanisms become dysfunctional (7). This calcium overload leads to cell swelling and rupture as well as cell death by necrotic, necroptotic, apoptotic and autophagic mechanisms.

Graft I/R injury is manifested by increased inflammation mediated by the complement system and cytokines. Once activated, the complement pathway damages the transplanted organ's cells by attacking the plasma membrane or recruiting/activating neutrophils (8,9). Cytokines may play either pro- or anti-inflammatory roles. Among others, tumor necrosis factor (TNF)- α is central in graft I/R injury (10). Increased TNF- α in the graft will lead to increased neutrophil recruitment, increased ROS production and activation of the NF- κ B and JNK pathways (11). Other cytokines are also involved: interleukin (IL)-1 β , IL-18, and interferon (IFN)- γ increase damage, while IL-6, IL-10, and IL-13 attempt to

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No potential conflict of interest was reported.

DOI: 10.6061/clinics/2016(09)10



control the damage (10). In its severest form, I/R injury can lead to dysfunction and possibly death of the transplanted organ (5,6,12).

Graft-versus-host disease

GVHD is a severe complication of organ transplantation, resulting in morbidity and mortality. GVHD consists of three phases, and several of the involved mechanisms are shared with the classical mechanisms of I/R injury. During the first phase, tissues are damaged by the recipient's conditioning regimen, which leads to the release of inflammatory cytokines such as TNF- α , IL-1, and IL-7 (13). These cytokines induce activation of host antigen-presenting cells (APCs). During the second phase of GVHD, the host APCs activate the donors' cells through IL-12 and IL-23 release, resulting in the production of Th1-related cytokines such as IL-2, IL-6, and TNF- γ . IL-10 downregulates the synthesis of these cytokines, but it is usually itself downregulated in the inflammatory context. The activated Th1 cells from the donor secrete IFN- γ to induce secretion of indoleamine 2,3-dioxygenase by the host APCs, thus stimulating immunotolerizing Tregs. IFN- γ also stimulates mononuclear cells to secrete IL-1 and TNF- α , which are inflammatory cytokines (13). Finally, in the third phase, the Th1 cells promote the proliferation and differentiation of cytotoxic T lymphocytes (CTLs) and stimulate natural killer (NK) cells, which in turn induce apoptosis of the cells of the transplanted organ (14). Cellular and inflammatory cytokines such as TNF- γ , IL-1, and IL-6 then directly assault various host tissues, leading to the clinical manifestations of GVHD (15). The activated cells also produce ROS, resulting in severe cell damage and the development of GVHD (16). Several studies have shown that GVHD is characterized by increased oxidative stress (17-19), and it has thus been suggested that antioxidants could be used to prevent GVHD (20).

Hydrogen as an antioxidant

Hydrogen is an inert gas that was long considered to have no effect on higher living organisms. Interestingly, however, in 2007, Ohsawa et al. (21) observed that hydrogen could reduce the levels of hydroxyl radicals (the most cytotoxic of all ROS), effectively protecting cells (22). Subsequently, many studies showed that hydrogen acts as an antioxidant and it has been used broadly in the prevention and treatment of many illnesses in experimental animal models (23-28). The hydrogen used in these studies mainly consisted of two types (hydrogen gas and hydrogen-rich saline) delivered through a number of methods, such as ventilation with mixed gas containing hydrogen (29), oral administration of hydrogen-rich saline (30), intraperitoneal injection of hydrogen gas or hydrogen-rich saline (31), and intravenous injection of hydrogen-rich saline (32).

Hydrogen as a therapeutic modality against transplantation-related I/R injury and GVHD

Increasing evidence has shown that molecular hydrogen could play an important role in the prevention and treatment of GVHD and graft I/R injury. Hence, the current literature regarding the effects of hydrogen on different animal models mimicking GVHD and I/R injury will be reviewed. The mechanisms of hydrogen's effects on I/R injury and GVHD are summarized in Table 1, but although many studies were performed in models of I/R injury, these were not necessarily

Table 1 - Mechanisms of the therapeutic effects of hydrogen on ischemia-reperfusion injury and graft-versus-host disease.

Effects	Mechanism
Antioxidation	<ul style="list-style-type: none"> *Inhibition of increased myeloperoxidase (MPO) activity (56) *Elimination of toxic reactive oxygen species (35,46) *Decreased levels of 8-hydroxydeoxyguanosine (36,37,66) *Decreased tissue malondialdehyde levels (36,42,52,56,64,65,67) *Decreased lipid peroxidation (42,49) *Increased expression of heme oxygenase-1 (43) *Decreased MPO activity (56) *Decreased levels of 8-iso-prostaglandin F₂α (64,65) *Decreased levels of 4-hydroxynonenal (66) *Improved superoxide dismutase activity (67) *Decreased hypoxia-inducible factor-1 levels (40)
Anti-inflammation	<ul style="list-style-type: none"> *Inhibition of the secretion of a variety of inflammatory cytokines (31,42,46,56) *Decreased inflammatory index and oxidative stress (41) *Reduced macrophage infiltration and sequestration (42) *Reduced recruitment of neutrophils (46) *Inhibition of the gene expression of proinflammatory factors (49)
Anti-apoptosis	<ul style="list-style-type: none"> *Increased levels of the B-cell lymphoma-2 and Bcl-extra-large proteins (42) *Reduced number of NF-κB-positive cells (71)
Other	<ul style="list-style-type: none"> *Inhibition of the release of serum alanine aminotransferase (58) *Improved suppression of the graft muscle contractility induced by transplantation (49) *Increased release of brain-derived neurotrophic factor (64,65) *Regulation of signaling pathways (54,64,70,71)

models of graft I/R injury. Therefore, the results may provide clues about the use of hydrogen for the treatment of graft I/R injury, but caution must be taken when examining these results. The studies mainly examined the use of hydrogen for organ pre-conditioning before harvesting, during organ preservation, and just before or during transplantation. Indeed, prolonged hypothermic preservation prior to transplantation is a challenge in the process of transplantation. In addition, graft I/R injury is common during transplantation, wherein multiple factors are involved and contribute to ROS production and ultrastructural injury. A number of studies have shown that hydrogen can decrease inflammation and apoptosis in graft organs, as detailed below.

Heart

Nakao et al. (33) showed that hydrogen could significantly reduce heart I/R injury induced by prolonged hypothermic preservation prior to transplantation through hydrogen's anti-inflammatory and antioxidant properties, as revealed by decreased levels of malondialdehyde (MDA) (an oxidation marker) and levels of troponin I and creatine phosphokinase (markers of heart injury). Similarly, Noda et al. (34) documented that a novel hydrogen-supplemented preservation solution efficiently improved myocardial injury due to cold I/R in a rat heterotopic transplantation model. In this study, the hydrogen-rich preservation solution led to decreased levels of IL-6, IL-1 β , TNF- α , ICAM-1, iNOS, and CCL2,



which are all markers of inflammation. Another study by Noda et al. showed that drinking hydrogen-rich water after heart transplantation could enhance cardiac allograft survival due to hydrogen's antioxidant properties, eliminating toxic ROS and reducing chronic intimal hyperplasia of the aortic artery after heart and artery transplantation (35). Indeed, this study showed that hydrogen led to increased ATP levels and more efficacious mitochondrial respiratory chain function as well as decreased IL-2 and IFN- γ levels and intimal hyperplasia (35). Therefore, the use of hydrogen in heart transplantation seems to be associated with reduced oxidative stress and inflammation. However, the available studies on heart grafts are limited in number and scope.

Kidneys

Hydrogen-rich saline from the University of Wisconsin has been used during hypothermic preservation of renal grafts and has been shown to decrease oxidative stress, as represented by lower MDA and serum 8-hydroxydeoxyguanosine (8-OHdG) levels as well as by prolonged graft survival (36). This preservation solution also decreased macrophage infiltration of this type of graft (36). In animal models of kidney transplantation, Shingu et al. (37) showed that treatment with hydrogen-rich saline could significantly attenuate renal graft I/R injury by reducing the levels of 8-OHdG, therefore improving renal transplant function and maintaining normal tissue structure after transplantation. Cardinal et al. (38) found that oral administration of hydrogen-rich saline could improve kidney function and increase overall survival after allotransplantation through reduction of oxidative stress and limited activation of inflammatory pathways such as MAPK pathways. Taken together, these results suggest that hydrogen improves kidney graft outcomes by decreasing inflammation and oxidative stress.

Lungs

One study showed that lung inflation with 3% hydrogen during the cold ischemia phase lowered graft myeloperoxidase (MPO) activity and serum IL-8 and TNF- α levels, resulting in alleviated lung graft injury and improved function (39). A study by Noda et al. (40) showed similar results in a rat model of lung transplantation using hydrogen preconditioning during the *ex vivo* period. The study also showed that the levels of hypoxia-inducible factor-1 were decreased in hydrogen-treated lungs, leading to decreased levels of the inflammatory cytokines IL-6, IL-1 β , and TNF- α (40). In rat models of brain-dead donor/recipient lung transplantation, it was demonstrated that hydrogen inhalation by the donors and the recipients could improve both lung function and graft histology by decreasing the inflammatory index (higher IL-8 and lower TNF- α levels), oxidative stress (increased superoxide dismutase (SOD) activity and lower MDA levels), and apoptosis (41). In another rat lung transplantation model, inhalation of mixed gas (98% oxygen and 2% hydrogen) alleviated lung graft I/R injury (42) by reducing inflammatory mediator upregulation as well as macrophage infiltration and sequestration; lowering tissue MDA levels 2 hours after reperfusion; and increasing the levels of the B-cell lymphoma (Bcl)-2 and Bcl-extra-large proteins, two proteins involved in apoptosis. A study by the same group showed that inhalation of mixed gas (2% hydrogen and 98% oxygen) by the organ donor could reduce the severity of I/R injury by reducing tissue

edema and the number of apoptotic pulmonary epithelial cells and especially by increasing the expression of heme oxygenase-1 (HO-1), which is a potent, inducible transcription factor with antioxidant, anti-inflammatory, and anti-apoptotic properties, therefore playing important roles in lung graft protection (43). Additionally, a recent study in pigs showed that hydrogen gas inhalation during *ex vivo* lung perfusion improved lung function after donation following cardiac death; the hydrogen group also had lower expression of IL-1 β , IL-6, IL-8, and TNF- α as well as lower scores for lung injury severity (44). Taken together, these studies all suggest that the use of hydrogen in lung grafts reduces inflammation, oxidative stress, and apoptosis.

Liver

One study examined the outcomes of perfusing the donor liver with hydrogen-saturated lactate Ringer's solution just before reperfusion and showed significantly lower aspartate aminotransferase and lactate dehydrogenase levels in animals with hydrogen-perfused livers, suggesting better graft function than in untreated grafts (45). In a rat model of small intestinal transplantation wherein both donors and recipients received 2% hydrogen inhalation, Buchholz et al. (46) found that hydrogen treatment significantly decreased the levels of CCL2, IL-1 β , IL-6, and TNF- α , leading to improved gastrointestinal transit and decreased lipid peroxidation as well as attenuated post-transplant breakdown of mucosal barrier function. Shigeta et al. (47) showed that luminal injection of hydrogen-rich solution attenuated I/R injury in a rat model of intestine transplantation by reducing oxidative stress. In a rat model of pancreas transplantation, hydrogen-rich saline was shown to protect against I/R injury, as demonstrated by better histopathological damage scores (based on edema, inflammation and necrosis) and better pancreatic function as well as by reduced levels of TNF- α , IL-1 β , and IL-6 (48). In a rat model of small-bowel transplantation, rats suffered from symptoms of gastroparesis, and Buchholz et al. (49) showed that hydrogen could alleviate this transplantation-related gastroparesis by improving the suppression of graft muscle contractility, inhibiting the gene expression of proinflammatory factors, and reducing the systemic inflammatory response.

Bone marrow and GVHD

Few studies have specifically examined the effects of hydrogen on GVHD. Qian et al. (50) studied the effects of hydrogen-rich saline treatment in a mouse model of haploidentical allogeneic bone marrow transplantation and found that the hydrogen-rich saline group had significantly reduced GVHD, significantly higher survival and faster recovery of peripheral blood leukocytes compared with the control group. This study further expanded the application range of hydrogen and introduced a new method for treating GVHD. However, the study did not examine the exact mechanisms involved in the results and suggested that reduced TNF- α , IL-2, and/or ROS levels may play roles in the benefits of hydrogen against aGVHD after bone marrow transplantation (50). In a mouse model consisting of lethal irradiation followed by allogeneic hematopoietic stem cell transplantation, hydrogen-rich saline was shown to improve the survival rate, to lower the rate of GVHD and the serum levels of inflammatory cytokines, and to reduce tissue damage (51). Again, the exact mechanisms involved have not been explored.



Hydrogen as a treatment for non-graft I/R injury

A number of studies have shown that hydrogen could be used to prevent I/R injury. In an experimental model of bilateral renal pedicle occlusion for 45 minutes, Wang et al. (31) showed that intraperitoneal injection of hydrogen-rich saline five minutes before reperfusion could alleviate I/R injury by inhibiting the secretion of a variety of inflammatory cytokines. In a rat model of I/R injury, inhalation of 2.5% hydrogen initiated 10 minutes before reperfusion and continued for 120 minutes could attenuated renal I/R injury by decreasing MDA levels (52). Sun et al. (53,54) also showed that hydrogen could reduce myocardial damage in a rat heart with regional myocardial I/R through antioxidative and anti-inflammatory effects.

In a New Zealand white rabbit model of lung I/R injury, hydrogen-rich saline treatment protected the lung from I/R injury by increasing the PaO₂/FiO₂ ratio and reducing the lung wet/dry ratio (55); in particular, the hydrogen-rich saline group displayed a significantly lower proportion of alveolar hemorrhage and pathologic lesions compared with the control group. In a rat model of lung injury induced by intestinal I/R injury, Mao et al. (56) showed that hydrogen-rich saline could reduce lung injury by decreasing MDA levels and MPO activity in the lung tissues.

In a liver injury mouse model, Sun et al. (57) showed that hydrogen-rich saline treatment could have protective effects on the liver. Similarly, in a mouse model of liver I/R injury, Fukuda et al. (58) found that hydrogen inhalation could significantly reduce liver I/R injury by inhibiting the release of serum alanine aminotransferase and MDA production.

Spinal cord injuries can be divided into two phases: i.e., direct mechanical tissue disruption, followed by cell damage by a cascade that includes oxidative stress, calcium mobilization, glutamate toxicity, and inflammation (59). Increased ROS production during spinal cord injuries plays a role in neuronal death and subsequent neuronal deficits (60,61). In addition to causing direct insults to macromolecules, these ROS act as intracellular messengers of neuronal death (62,63). Neurons are among the cells most sensitive to ROS (59). It was found that hydrogen reduced acute spinal cord contusion injury by increasing the release of brain-derived neurotrophic factor and decreasing the levels of oxidative products such as 8-iso-prostaglandin F_{2α} and MDA (64,65).

Retinal I/R injuries are often observed in conditions such as acute angle-closure glaucoma, retinal artery occlusion, and amaurosis fugax. In animal models, retinal I/R injuries are often induced by transient elevation of intraocular pressure. In a model of retinal I/R injury, hydroxyl radicals caused irreversible cellular damage by affecting lipids, proteins and nucleic acids (66). Hydrogen-loaded eye drops were used in these animals and markers such as 4-hydroxynonenal and 8-hydroxy-2-deoxyguanosine were used to evaluate I/R injury. The hydrogen-loaded eye drops dramatically decreased 4-hydroxynonenal and 8-hydroxy-2-deoxyguanosine levels and reduced subsequent retinal cell death after I/R injury (66).

Testicular torsion occurs when the spermatic cord twists, thereby cutting off the testicle's blood supply. This urological condition usually affects children and adolescents and inflammatory cytokines and free radicals play important roles. One study assessed the effect of hydrogen-rich saline on testicular I/R injury after testicular torsion and showed that the injury score in the hydrogen treatment group was

the lowest among all tested groups. Moreover, compared with the other groups, in the hydrogen treatment group, MDA levels were significantly lowered and SOD activity was significantly improved (67).

In a rat model of *in utero* I/R injury, Mano et al. (68) studied the effects of hydrogen on rat fetal hippocampal damage caused by I/R on day 16 of pregnancy. The results indicated that oral administration of hydrogen-saturated water could reduce placental oxidative damage, alleviate neonatal growth retardation and improve the rat fetal hippocampal damage caused by *in utero* I/R (68). Similar effects were observed in another study (69).

Therefore, a number of animal experiments over the last few years have shown that hydrogen can obviously reduce the damage caused by organ transplantation. From the initial simple effect of antioxidative activity (23) to anti-inflammatory and anti-apoptotic activity and regulation of signaling pathways (31,54,64,70), the effects of hydrogen treatment have been demonstrated in many experiments.

A substantial body of experimental evidence suggests that hydrogen can significantly alleviate I/R injury related to transplantation and has a therapeutic effect on complications of transplantation (including GVHD), mainly *via* inhibition of inflammatory cytokine secretion and reduction of oxidative stress. However, the exact mechanisms leading to these effects are currently ill known. In addition, many studies on the effects of hydrogen were performed in models of I/R injury, but not in models of graft I/R injury. Nevertheless, the results may provide clues about the use of hydrogen for the treatment of graft I/R injury, although caution must be taken when examining these results.

With the advantages of being easily available, having a low price, and being a nontoxic small molecule that is easily absorbed, hydrogen has a strong prospect of clinical applications. Further animal experiments and preliminary human clinical trials are needed to lay the foundation for hydrogen use as a drug in the clinic in the near future.

■ AUTHOR CONTRIBUTIONS

All authors contributed to the review of the literature, the data analysis, and the manuscript writing and approved the final version of the manuscript.

■ REFERENCES

- White SL, Hirth R, Mahillo B, Dominguez-Gil B, Delmonico FL, Noel L, et al. The global diffusion of organ transplantation: trends, drivers and policy implications. *Bull World Health Organ.* 2014;92(11):826-35, <http://dx.doi.org/10.2471/BLT.14.137653>.
- Chaib E, Silva FD, Figueira ER, Lima FR, Andraus W, D'Albuquerque LA. Graft-versus-host disease after liver transplantation. *Clinics.* 2011;66(6):1115-8, <http://dx.doi.org/10.1590/S1807-59322011000600035>.
- Kosieradzki M, Rowinski W. Ischemia/reperfusion injury in kidney transplantation: mechanisms and prevention. *Transplant Proc.* 2008; 40(10):3279-88, <http://dx.doi.org/10.1016/j.transproceed.2008.10.004>.
- Foley DP, Chari RS. Ischemia-reperfusion injury in transplantation: novel mechanisms and protective strategies. *Transplant Rev.* 2007;21(1):43-53, <http://dx.doi.org/10.1016/j.trre.2007.01.004>.
- Eltzschig HK, Collard CD. Vascular ischaemia and reperfusion injury. *Br Med Bull.* 2004;70:71-86, <http://dx.doi.org/10.1093/bmb/ldh025>.
- Ricca L, Lemoine A, Cauchy F, Hamelin J, Sebagh M, Esposti DD, et al. Ischemic Postconditioning of the Liver Graft in Adult Liver Transplantation. *Transplantation.* 2015;99(8):1633-43, <http://dx.doi.org/10.1097/TP.0000000000000685>.
- Kalogeris T, Baines CP, Krenz M, Korhuis RJ. Cell biology of ischemia/reperfusion injury. *Int Rev Cell Mol Biol.* 2012;298:229-317, <http://dx.doi.org/10.1016/B978-0-12-394309-5.00006-7>.
- Montalvo-Jave EE, Escalante-Tattersfield T, Ortega-Salgado JA, Pina E, Geller DA. Factors in the pathophysiology of the liver ischemia-reperfusion



- injury. *J Surg Res.* 2008;147(1):153-9, <http://dx.doi.org/10.1016/j.jss.2007.06.015>.
9. Fondevila C, Shen XD, Tsuchihashi S, Uchida Y, Freitas MC, Ke B, et al. The membrane attack complex (C5b-9) in liver cold ischemia and reperfusion injury. *Liver Transpl.* 2008;14(8):1133-41, <http://dx.doi.org/10.1002/lt.21496>.
 10. Abu-Amara M, Yang SY, Tapuria N, Fuller B, Davidson B, Seifalian A. Liver ischemia/reperfusion injury: processes in inflammatory networks—a review. *Liver Transpl.* 2010;16(9):1016-32, <http://dx.doi.org/10.1002/lt.22117>.
 11. Schwabe RF, Brenner DA. Mechanisms of Liver Injury. I. TNF-alpha-induced liver injury: role of IKK, JNK, and ROS pathways. *Am J Physiol Gastrointest Liver Physiol.* 2006;290(4):G583-9, <http://dx.doi.org/10.1152/ajpgi.00422.2005>.
 12. Peralta C, Jimenez-Castro MB, Gracia-Sancho J. Hepatic ischemia and reperfusion injury: effects on the liver sinusoidal milieu. *J Hepatol.* 2013;59(5):1094-106, <http://dx.doi.org/10.1016/j.jhep.2013.06.017>.
 13. Hill GR, Teshima T, Gerbitz A, Pan L, Cooke KR, Brinson YS, et al. Differential roles of IL-1 and TNF-alpha on graft-versus-host disease and graft versus leukemia. *J Clin Invest.* 1999;104(4):459-67, <http://dx.doi.org/10.1172/JCI6896>.
 14. Harris AC, Ferrara JL, Levine JE. Advances in predicting acute GVHD. *Br J Haematol.* 2013;160(3):288-302, <http://dx.doi.org/10.1111/bjh.12142>.
 15. Hill GR, Crawford JM, Cooke KR, Brinson YS, Pan L, Ferrara JL. Total body irradiation and acute graft-versus-host disease: the role of gastrointestinal damage and inflammatory cytokines. *Blood.* 1997;90(8):3204-13.
 16. Ferrara JL, Deeg HJ. Graft-versus-host disease. *N Engl J Med.* 1991;324(10):667-74, <http://dx.doi.org/10.1056/NEJM199103073241005>.
 17. Amer J, Weiss L, Reich S, Shapira MY, Slavin S, Fibach E. The oxidative status of blood cells in a murine model of graft-versus-host disease. *Ann Hematol.* 2007;86(10):753-8, <http://dx.doi.org/10.1007/s00277-007-0321-7>.
 18. Schwab L, Goroncy L, Palaniyandi S, Gautam S, Triantafyllou A, Mocsai A, et al. Neutrophil granulocytes recruited upon translocation of intestinal bacteria enhance graft-versus-host disease via tissue damage. *Nat Med.* 2014;20(6):648-54, <http://dx.doi.org/10.1038/nm.3517>.
 19. Klambt V, Wohlfeil SA, Schwab L, Hulsdunker J, Ayata K, Apostolova P, et al. A Novel Function for P2Y2 in Myeloid Recipient-Derived Cells during Graft-versus-Host Disease. *J Immunol.* 2015;195(12):5795-804, <http://dx.doi.org/10.4049/jimmunol.1501357>.
 20. Im KI, Kim N, Lim JY, Nam YS, Lee ES, Kim EJ, et al. The Free Radical Scavenger NecroX-7 Attenuates Acute Graft-versus-Host Disease via Reciprocal Regulation of Th1/Regulatory T Cells and Inhibition of HMGB1 Release. *J Immunol.* 2015;194(11):5223-32, <http://dx.doi.org/10.4049/jimmunol.1402609>.
 21. Ohsawa I, Ishikawa M, Takahashi K, Watanabe M, Nishimaki K, Yamagata K, et al. Hydrogen acts as a therapeutic antioxidant by selectively reducing cytotoxic oxygen radicals. *Nat Med.* 2007;13(6):688-94, <http://dx.doi.org/10.1038/nm1577>.
 22. Zhang W, Wang M, Xie HY, Zhou L, Meng XQ, Shi J, et al. Role of reactive oxygen species in mediating hepatic ischemia-reperfusion injury and its therapeutic applications in liver transplantation. *Transplant Proc.* 2007;39(5):1332-7, <http://dx.doi.org/10.1016/j.transproceed.2006.11.021>.
 23. Hong Y, Chen S, Zhang JM. Hydrogen as a selective antioxidant: a review of clinical and experimental studies. *J Int Med Res.* 2010;38(6):1893-903, <http://dx.doi.org/10.1177/147323001003800602>.
 24. Feng Y, Wang R, Xu J, Sun J, Xu T, Gu Q, et al. Hydrogen-rich saline prevents early neurovascular dysfunction resulting from inhibition of oxidative stress in STZ-diabetic rats. *Curr Eye Res.* 2013;38(3):396-404, <http://dx.doi.org/10.3109/02713683.2012.748919>.
 25. Chuai Y, Gao F, Li B, Zhao L, Qian L, Cao F, et al. Hydrogen-rich saline attenuates radiation-induced male germ cell loss in mice through reducing hydroxyl radicals. *Biochem J.* 2012;442(1):49-56, <http://dx.doi.org/10.1042/BJ20111786>.
 26. Ji X, Liu W, Xie K, Liu W, Qu Y, Chao X, et al. Beneficial effects of hydrogen gas in a rat model of traumatic brain injury via reducing oxidative stress. *Brain Res.* 2010;1354:196-205, <http://dx.doi.org/10.1016/j.brainres.2010.07.038>.
 27. Liu S, Liu K, Sun Q, Liu W, Xu W, Denoble P, et al. Consumption of hydrogen water reduces paraquat-induced acute lung injury in rats. *J Biomed Biotechnol.* 2011;2011:305086.
 28. Sun Q, Cai J, Liu S, Liu Y, Xu W, Tao H, et al. Hydrogen-rich saline provides protection against hyperoxic lung injury. *J Surg Res.* 2011;165(1):e43-9, <http://dx.doi.org/10.1016/j.jss.2010.09.024>.
 29. Hayashi T, Yoshioka T, Hasegawa K, Miyamura M, Mori T, Ukimura A, et al. Inhalation of hydrogen gas attenuates left ventricular remodeling induced by intermittent hypoxia in mice. *Am J Physiol Heart Circ Physiol.* 2011;301(3):H1062-9, <http://dx.doi.org/10.1152/ajpheart.00150.2011>.
 30. Fang Y, Fu XJ, Gu C, Xu P, Wang Y, Yu WR, et al. Hydrogen-rich saline protects against acute lung injury induced by extensive burn in rat model. *J Burn Care Res.* 2011;32(3):e82-91, <http://dx.doi.org/10.1097/BCR.0b013e318217f84f>.
 31. Wang F, Yu G, Liu SY, Li JB, Wang JF, Bo LL, et al. Hydrogen-rich saline protects against renal ischemia/reperfusion injury in rats. *J Surg Res.* 2011;167(2):e339-44, <http://dx.doi.org/10.1016/j.jss.2010.11.005>.
 32. Zhang DQ, Feng H, Chen WC. Effects of hydrogen-rich saline on taurocholate-induced acute pancreatitis in rat. *Evid Based Complement Alternat Med.* 2013;2013:731932, <http://dx.doi.org/10.1155/2013/731932>.
 33. Nakao A, Kaczorowski DJ, Wang Y, Cardinal JS, Buchholz BM, Sugimoto R, et al. Amelioration of rat cardiac cold ischemia/reperfusion injury with inhaled hydrogen or carbon monoxide, or both. *J Heart Lung Transplant.* 2010;29(5):544-53, <http://dx.doi.org/10.1016/j.jhealun.2009.10.011>.
 34. Noda K, Shigemura N, Tanaka Y, Kawamura T, Hyun Lim S, Kokubo K, et al. A novel method of preserving cardiac grafts using a hydrogen-rich water bath. *J Heart Lung Transplant.* 2013;32(2):241-50, <http://dx.doi.org/10.1016/j.jhealun.2012.11.004>.
 35. Noda K, Tanaka Y, Shigemura N, Kawamura T, Wang Y, Masutani K, et al. Hydrogen-supplemented drinking water protects cardiac allografts from inflammation-associated deterioration. *Transpl Int.* 2012;25(12):1213-22, <http://dx.doi.org/10.1111/j.1432-2277.2012.01542.x>.
 36. Abe T, Li XK, Yazawa K, Hatayama N, Xie L, Sato B, et al. Hydrogen-rich University of Wisconsin solution attenuates renal cold ischemia-reperfusion injury. *Transplantation.* 2012;94(1):14-21, <http://dx.doi.org/10.1097/TP.0b013e318255f8be>.
 37. Shingu C, Koga H, Hagiwara S, Matsumoto S, Goto K, Yokoi I, et al. Hydrogen-rich saline solution attenuates renal ischemia-reperfusion injury. *J Anesth.* 2010;24(4):569-74, <http://dx.doi.org/10.1007/s00540-010-0942-1>.
 38. Cardinal JS, Zhan J, Wang Y, Sugimoto R, Tsung A, McCurry KR, et al. Oral hydrogen water prevents chronic allograft nephropathy in rats. *Kidney Int.* 2010;77(2):101-9, <http://dx.doi.org/10.1038/ki.2009.421>.
 39. Liu R, Fang X, Meng C, Xing J, Liu J, Yang W, et al. Lung inflation with hydrogen during the cold ischemia phase decreases lung graft injury in rats. *Exp Biol Med (Maywood).* 2015;240(9):1214-22, <http://dx.doi.org/10.1177/1535370214563895>.
 40. Noda K, Shigemura N, Tanaka Y, Bhamra J, D'Cunha J, Kobayashi H, et al. Hydrogen preconditioning during ex vivo lung perfusion improves the quality of lung grafts in rats. *Transplantation.* 2014;98(5):499-506, <http://dx.doi.org/10.1097/TP.0000000000000254>.
 41. Zhou H, Fu Z, Wei Y, Liu J, Cui X, Yang W, et al. Hydrogen inhalation decreases lung graft injury in brain-dead donor rats. *J Heart Lung Transplant.* 2013;32(2):251-8, <http://dx.doi.org/10.1016/j.jhealun.2012.11.007>.
 42. Kawamura T, Huang CS, Tochigi N, Lee S, Shigemura N, Billiar TR, et al. Inhaled hydrogen gas therapy for prevention of lung transplant-induced ischemia/reperfusion injury in rats. *Transplantation.* 2010;90(12):1344-51, <http://dx.doi.org/10.1097/TP.0b013e3181fe1357>.
 43. Kawamura T, Huang CS, Peng X, Masutani K, Shigemura N, Billiar TR, et al. The effect of donor treatment with hydrogen on lung allograft function in rats. *Surgery.* 2011;150(2):240-9, <http://dx.doi.org/10.1016/j.surg.2011.05.019>.
 44. Haam S, Lee S, Paik HC, Park MS, Song JH, Lim BJ, et al. The effects of hydrogen gas inhalation during ex vivo lung perfusion on donor lungs obtained after cardiac death. *Eur J Cardiothorac Surg.* 2015;48(4):542-7, <http://dx.doi.org/10.1093/ejcts/ezv057>.
 45. Matsuno N, Watanabe R, Kimura M, Iwata S, Fujiyama M, Kono S, et al. Beneficial effects of hydrogen gas on porcine liver reperfusion injury with use of total vascular exclusion and active venous bypass. *Transplant Proc.* 2014;46(4):1104-6, <http://dx.doi.org/10.1016/j.transproceed.2013.11.134>.
 46. Buchholz BM, Kaczorowski DJ, Sugimoto R, Yang R, Wang Y, Billiar TR, et al. Hydrogen inhalation ameliorates oxidative stress in transplantation induced intestinal graft injury. *Am J Transplant.* 2008;8(10):2015-24, <http://dx.doi.org/10.1111/j.1600-6143.2008.02359.x>.
 47. Shigetani T, Sakamoto S, Li XK, Cai S, Liu C, Kurokawa R, et al. Luminal injection of hydrogen-rich solution attenuates intestinal ischemia-reperfusion injury in rats. *Transplantation.* 2015;99(3):500-7, <http://dx.doi.org/10.1097/TP.0000000000000510>.
 48. Luo ZL, Cheng L, Ren JD, Fang C, Xiang K, Xu HT, et al. Hydrogen-rich saline protects against ischemia/reperfusion injury in grafts after pancreas transplantations by reducing oxidative stress in rats. *Mediators Inflamm.* 2015;2015:281985, <http://dx.doi.org/10.1155/2015/281985>.
 49. Buchholz BM, Masutani K, Kawamura T, Peng X, Toyoda Y, Billiar TR, et al. Hydrogen-enriched preservation protects the isogeneic intestinal graft and amends recipient gastric function during transplantation. *Transplantation.* 2011;92(9):985-92, <http://dx.doi.org/10.1097/TP.0b013e318230159d>.
 50. Qian L, Mei K, Shen J, Cai J. Administration of hydrogen-rich saline protects mice from lethal acute graft-versus-host disease (aGVHD). *Transplantation.* 2013;95(5):658-62, <http://dx.doi.org/10.1097/TP.0b013e31827e6b23>.
 51. Yuan L, Chen X, Qian L, Shen J, Cai J. Administration of hydrogen-rich saline in mice with allogeneic hematopoietic stem-cell transplantation. *Med Sci Monit.* 2015;21:749-54, <http://dx.doi.org/10.12659/MSM.891338>.
 52. Zeng K, Huang H, Jiang XQ, Chen XJ, Huang W. [Protective effects of hydrogen on renal ischemia/reperfusion injury in rats]. *Sichuan Da Xue Xue Bao Yi Xue Ban.* 2014;45(1):39-42.
 53. Sun Q, Kang Z, Cai J, Liu W, Liu Y, Zhang JH, et al. Hydrogen-rich saline protects myocardium against ischemia/reperfusion injury in rats.



- Exp Biol Med (Maywood). 2009;234(10):1212-9, <http://dx.doi.org/10.3181/0812-RM-349>.
54. Zhang Y, Sun Q, He B, Xiao J, Wang Z, Sun X. Anti-inflammatory effect of hydrogen-rich saline in a rat model of regional myocardial ischemia and reperfusion. *Int J Cardiol*. 2011;148(1):91-5, <http://dx.doi.org/10.1016/j.ijcard.2010.08.058>.
 55. Li H, Zhou R, Liu J, Li Q, Zhang J, Mu J, et al. Hydrogen-rich saline attenuates lung ischemia-reperfusion injury in rabbits. *J Surg Res*. 2012;174(1):e11-6, <http://dx.doi.org/10.1016/j.jss.2011.10.001>.
 56. Mao YF, Zheng XF, Cai JM, You XM, Deng XM, Zhang JH, et al. Hydrogen-rich saline reduces lung injury induced by intestinal ischemia/reperfusion in rats. *Biochem Biophys Res Commun*. 2009;381(4):602-5, <http://dx.doi.org/10.1016/j.bbrc.2009.02.105>.
 57. Sun H, Chen L, Zhou W, Hu L, Li L, Tu Q, et al. The protective role of hydrogen-rich saline in experimental liver injury in mice. *J Hepatol*. 2011;54(3):471-80, <http://dx.doi.org/10.1016/j.jhep.2010.08.011>.
 58. Fukuda K, Asoh S, Ishikawa M, Yamamoto Y, Ohsawa I, Ohta S. Inhalation of hydrogen gas suppresses hepatic injury caused by ischemia/reperfusion through reducing oxidative stress. *Biochem Biophys Res Commun*. 2007;361(3):670-4, <http://dx.doi.org/10.1016/j.bbrc.2007.07.088>.
 59. Xu W, Chi L, Xu R, Ke Y, Luo C, Cai J, et al. Increased production of reactive oxygen species contributes to motor neuron death in a compression mouse model of spinal cord injury. *Spinal Cord*. 2005;43(4):204-13, <http://dx.doi.org/10.1038/sj.sc.3101674>.
 60. Anderson DK, Hall ED. Pathophysiology of spinal cord trauma. *Ann Emerg Med*. 1993;22(6):987-92, [http://dx.doi.org/10.1016/S0196-0644\(05\)82739-8](http://dx.doi.org/10.1016/S0196-0644(05)82739-8).
 61. Hall ED. Pathophysiology of spinal cord injury. Current and future therapies. *Minerva Anesthesiol*. 1989;55(3):63-6.
 62. Maher P, Schubert D. Signaling by reactive oxygen species in the nervous system. *Cell Mol Life Sci*. 2000;57(8-9):1287-305, <http://dx.doi.org/10.1007/PL00000766>.
 63. Rhee SG. Redox signaling: hydrogen peroxide as intracellular messenger. *Exp Mol Med*. 1999;31(2):53-9, <http://dx.doi.org/10.1038/emmm.1999.9>.
 64. Chen C, Chen Q, Mao Y, Xu S, Xia C, Shi X, et al. Hydrogen-rich saline protects against spinal cord injury in rats. *Neurochem Res*. 2010;35(7):1111-8, <http://dx.doi.org/10.1007/s11064-010-0162-y>.
 65. Huang Y, Xie K, Li J, Xu N, Gong G, Wang G, et al. Beneficial effects of hydrogen gas against spinal cord ischemia-reperfusion injury in rabbits. *Brain Res*. 2011;1378:125-36, <http://dx.doi.org/10.1016/j.brainres.2010.12.071>.
 66. Oharazawa H, Igarashi T, Yokota T, Fujii H, Suzuki H, Machide M, et al. Protection of the retina by rapid diffusion of hydrogen: administration of hydrogen-loaded eye drops in retinal ischemia-reperfusion injury. *Invest Ophthalmol Vis Sci*. 2010;51(1):487-92, <http://dx.doi.org/10.1167/iovs.09-4089>.
 67. Jiang D, Wu D, Zhang Y, Xu B, Sun X, Li Z. Protective effects of hydrogen rich saline solution on experimental testicular ischemia-reperfusion injury in rats. *J Urol*. 2012;187(6):2249-53, <http://dx.doi.org/10.1016/j.juro.2012.01.029>.
 68. Mano Y, Kotani T, Ito M, Nagai T, Ichinohashi Y, Yamada K, et al. Maternal molecular hydrogen administration ameliorates rat fetal hippocampal damage caused by in utero ischemia-reperfusion. *Free Radic Biol Med*. 2014;69:324-30, <http://dx.doi.org/10.1016/j.freeradbiomed.2014.01.037>.
 69. Liu W, Chen O, Chen C, Wu B, Tang J, Zhang JH. Protective effects of hydrogen on fetal brain injury during maternal hypoxia. *Acta Neurochir Suppl*. 2011;111:307-11, <http://dx.doi.org/10.1007/978-3-7091-0693-8>.
 70. Liu Q, Shen WF, Sun HY, Fan DF, Nakao A, Cai JM, et al. Hydrogen-rich saline protects against liver injury in rats with obstructive jaundice. *Liver Int*. 2010;30(7):958-68, <http://dx.doi.org/10.1111/j.1478-3231.2010.02254.x>.
 71. Wang C, Li J, Liu Q, Yang R, Zhang JH, Cao YP, et al. Hydrogen-rich saline reduces oxidative stress and inflammation by inhibit of JNK and NF-kappaB activation in a rat model of amyloid-beta-induced Alzheimer's disease. *Neurosci Lett*. 2011;491(2):127-32, <http://dx.doi.org/10.1016/j.neulet.2011.01.022>.