BRAZILIAN JOURNAL OF OCEANOGRAPHY, 62(3):167-177, 2014

TEMPORAL VARIATIONS OF SHALLOW SUBTIDAL MEIOFAUNA IN LOS CRISTIANOS BAY (TENERIFE, CANARY ISLANDS, NE ATLANTIC OCEAN)

Rodrigo Riera^{1,*}, Jorge Nunez² and Maria del Carmen Brito²

¹Centro de Investigaciones Medioambientales del Atlántico (CIMA SL) (Arzobispo Elías Yanes, 44, 38206 La Laguna, Tenerife, Canary Islands, Spain)

²University of La Laguna - Benthos Lab, Department of Animal Biology, Faculty of Biology (La Laguna, 38206Tenerife, Canary Islands, Spain)

*Corresponding author: rodrigo@cimacanarias.com

http://dx.doi.org/10.1590/S1679-87592014027406203

ABSTRACT

A subtidal meiofaunal assemblage in Los Cristianos Bay, Tenerife, Canary Islands was sampled from May 2000 to April 2001, at 3 m depth. Nematodes dominated overwhelmingly during the study period, ranging from 84.52% in May 2000 to 95.93% in October 2000. Copepods and polychaetes were the second and the third most abundant groups, respectively. Meiofaunal densities showed significant differences throughout the study period, with minimum abundances during the spring-summer months (May-July) and highest densities in winter (January and February). This seasonality is mainly due to the temporal variations of the most abundant species (nematodes Daptonema hirsutum and Pomponema sedecima), with differences in meiofauna species composition and abundance during May and June 2000 as compared to the remaining months of the study period. Environmental variables partly explained meiofaunal community structure, being the sedimentary type of very fine sands the most important, jointly with other variables, such as nitrogen and organic matter content.

RESUMO

As assembleias da meiofauna de Los Cristianos Bay, Tenerife, Ilhas Canárias, foi amostrada de maio de 2000 a abril de 2001 a 3 m de profundidade. Nematoda dominou durante todo o período de estudo, variando entre 84.52% em maio de 2000 a 95.93% em outubro do mesmo ano. Copepoda e Polychaeta foram o segundo e terceiro grupos mais abundantes, respectivamente. A densidade apresentou diferenças significativas ao longo do período, com valores mínimos na primavera-verão (maio-julho) e máximos no inverno (janeiro e fevereiro). Essa sazonalidade está sendo atribuída principalmente às variações temporais das espécies mais abundantes de nemátodes (Daptonema hirsutum e Pomponema sedecima). A meiofauna apresentou ainda diferenças na composição e abundância em maio e junho de 2000 quando comparado com os meses restantes do estudo. As variáveis ambientais explicaram parcialmente as variações na estrutura da comunidade e o tipo sedimentar areia muito fina destacou-se entre os mais importantes, juntamente com outras variáveis, tais como nitrogênio e conteúdo de matéria orgânica.

Descriptors: Subtidal, Meiofauna, Nematodes, Temporal distribution, Canary Islands, Atlantic

Descritores: Sublitoral, Meiofauna, Nematodes, Distribuição temporal, Ilhas Canárias, Oceano Atlântico.

Introduction

Shallow marine meiobenthos are known to vary seasonally with physico-chemical regimes and environmental trophic dynamics (GUIDI-**GUILVARD** AND BUSCAIL, 1995; DANOVARO, 1996; GRÉMARE ET AL, 1997; ÓLAFSSON AND ELMGREN, 1997). However,

one of the main drawbacks in the field is to know how to separate the effects of the different parameters, e.g. temperature and food availability (FLEEGER ET AL, 1989), temperature and salinity (Santos et al, 1996) or muds and organic matter content, because they are generally closely linked.

There is an extensive literature on the temporal variations of meiofauna (mobile metazoan benthic invertebrates that pass through a 0.5 mm mesh sieve but are retained on a 0.063 or 0.042 mm mesh sieve) over a study period, normally of one year. However, most of these studies are limited to estuarine (e.g. NOZAIS ET AL, 2005) or intertidal environments (e.g. SCHIZAS AND SHIRLEY, 1996) or temperate (e.g. VINCX, 1989; ÓLAFSSON AND ELMGREN, 1997) or polar regions (e.g. VANHOVE ET AL, 2000) but seasonal studies on marine subtropical or tropical regions are rather scarce and limited to intertidal habitats (e.g. SANTOS ET AL, 1996).

In temperate regions, meiofaunal abundances reach maximum abundances during winter (COULL 1988, PALACÍN 1990), though some inconsistencies have also been reported (MAZZOLA ET AL. 2000, MIRTO ET AL. 2000). In subtropical regions, several studies have documented an increase in meiofaunal abundances during spring and summer (HICKS AND COULL 1983, COULL 1985, RUDNICK ET AL. 1985). In tropical latitudes, meiofauna has shown different temporal patterns, although the highest abundances are found during the wet season, when temperatures are usually higher (ALBUQUERQUE ET AL.

The Canary archipelago can he characterized as a subtropical area of the Macaronesian (Azores, Madeira, Canary Islands and Cape Verde islands) biogeographic region. The Canary Islands are bathed by oligotrophic waters (0.0-2.5 µ atom-g 1-1 of phosphates, nitrates and silicates) (BRAUN, 1981), implying the presence of low concentrations of particulated organic matter and thus of infaunal densities (macro- and meiofauna). The meiofauna assemblage structure is thus directly affected and low abundances are to be expected in this region. Moreover, the subtropical oceanic climate is almost constantly present throughout the year, with little variation in temperature (18-25°C). Hence a lack of seasonality in marine assemblages is to be expected and this has, in fact, been observed in intertidal meiofauna communities (RIERA ET AL, 2011b).

In the present study, patterns in the assemblage structure and abundance of meiofauna assemblages inhabiting a shallow subtidal site (3 m deep) were investigated on the Canary Islands over a one-year cycle (May 2000-April 2001). It was sought: (i) to determine whether patterns in the assemblage structure, total meiofaunal abundance and the abundance of the most common species followed a seasonal trend during the study period, and (ii) to ascertain whether three sedimentary variables (grain size, organic content and total nitrogen) affected meiofaunal assemblage structure during this annual cycle.

MATERIAL AND METHODS

Study Area

This study was conducted from May 2000 to April 2001 in Los Cristianos Bay, a locality on the south coast of Tenerife (Canary Islands, NE Atlantic Ocean; Lat. N. 28°02′59′′ Long. W. 16°42′54′′). The study site is located in a sheltered bay, with a recreational harbour inside it (Fig. 1).

Sediment samples were collected on a monthly basis manually by SCUBA divers at one shallow-subtidal station (3 m deep), with an average silt-clay fraction of 0.34%. Sediment corers (4.5 cm inner diameter; 15.9 cm²) were pushed into the sediment to a depth of 30 cm. Five replicates were collected for faunistic analysis and one for analysis of abiotic factors (organic matter, total nitrogen and granulometry).

Analysis of Meiofauna

Samples were preserved in a 10% seawater formaldehyde solution and decanted through a 500 and a 63 µm mesh sieve. The fraction remaining on the 63 µm mesh sieve was separated into different taxonomical groups under a binocular microscope and preserved in 70% ethanol (SOMERFIELD AND WARWICK, Meiofaunal specimens were mounted in jelly glycerine and examined using a LEICA DMLB microscope equipped with Nomarski interference contrast and identified to species level, whenever possible. All meiofaunal specimens were identified and a subsample of 200 nematode individuals taken, following SOMERFIELD AND WARWICK (1996). Some taxonomic groups (i.e. harpacticoid copepods, turbellarians) required dissection, and/or careful inspection of taxonomic characters of their internal anatomy. The remaining taxonomic groups were determined to the lowest taxonomic level using current scientific literature.

Statistical Analysis

Biological descriptors of the assemblage (abundance, species richness, n° of taxonomical groups) were estimated for each sample. Differences in species richness and abundance patterns throughout the study period (May 2000-April 2001) were tested by one-way ANOVA, after verifying normality using the Kolmogorov-Smirnov test and Levene's test for homogeneity of variances. When the requirements for normality and/or homogeneity of variances were not met, the Kruskal-Wallis ANOVA (KW ANOVA) test was used instead.

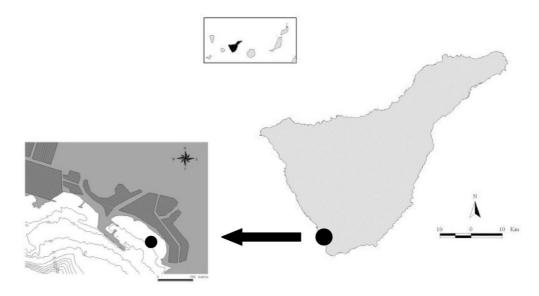


Fig. 1. Map of the study area, showing sampling station.

To visualize affinities in assemblage structure throughout the study period, a n-MDS (non-metric multidimensional scaling) was carried out on square rooted-transformed abundance data via the Bray-Curtis similarity index. ANOSIM analysis (CLARKE, 1993) was used to detect the significance of differences in meiofauna assemblage structure between months. A comparative test of similarity matrices (RELATE routine) was conducted to detect temporal differences in meiofauna assemblage structure between the months throughout the study period.

The relationship between the entire set of environmental variables and the meiofaunal assemblage structure was investigated using distance-based redundancy analysis (db-RDA, Legendre AND Anderson 1999). Multivariate multiple regression, using the DISTLM routine via 4999 permutations of the data (ANDERSON, 2001), tested the significance of these relationships by fitting a linear model based on Bray-Curtis dissimilarities to squared-root transformed abundance data.

RESULTS

Sedimentary Variables

The sampling site was dominated by fine sands (0.125-0.25 mm grain size diameter) throughout the study period (May 2000-April 2001),

ranging from a minimum of 55.79% (February 2001) to a maximum of 82.86% (October 2000). Medium sands were also a well-represented sedimentary fraction, ranging from 7.67% (November 2000) to 34.32% (May 2000). The remaining sediment types were scarce (average < 5%). The mean percentage of organic matter content was 0.54% throughout the study period, with a maximum content of 1.01% (September 2000) and a minimum content of 0.006% (November 2000). The mean percentage of nitrogen remained almost constant over the study period, with a mean of 0.013%, ranging from 0.012% to 0.014% (Table 1). For more details see RIERA ET AL. (2011).

Meiofauna

A total of 33,262 specimens were collected during the study period (May 2000-April 2001). Free-living nematodes were the most abundant group with 31,125 individuals (92.9% of the overall abundance), ranging from 84.52% in May 2000 to 95.93% in October 2000. Harpacticoid copepods and polychaetes were the second and third taxonomic groups in order of importance, representing 8.86% and 5.58% of the overall abundance, respectively. The remaining taxonomic groups (Turbellarians, Amphipods, Tanaids, Oligochaetes, Acari, Ostracods, Misids and Nemerteans) were scarce during the study period.

	May-00	Jun-00	Jul-00	Aug-00	Sep-00	Oct-00	Nov-00	Dec-00	Jan-01	Feb-01	Mar-01	Apr-01
Nitrogen (%)	0.014	0.013	0.014	0.012	0.013	0.014	0.014	0.013	0.013	0.013	0.013	0.012
Organic matter												
(%)	0.71	0.86	0.63	0.02	1.01	0.64	0.006	0.73	0.17	0.33	0.81	0.53
Gravels (%)	0.39	0.04	0.27	6.19	0.03	0.15	1.93	0.05	0.13	8.59	0.13	0.03
Very coarse												
sands (%)	0.82	0.46	0.69	4.59	0.25	0.13	0.68	0.33	0.64	4.49	0.79	0.61
Coarse sands												
(%)	3.14	1.61	0.95	4.14	0.64	0.17	0.77	0.81	0.8	5.86	2.18	0.63
Medium sands												
(%)	34.32	20.44	19.18	20.27	26.59	9.61	7.67	24.92	22.47	19.55	22.76	17.87
Fine sands (%)	59.61	73.22	75.85	56.26	67.67	82.86	78.31	68.8	71.2	55.79	69.99	76.45
Very fine sands												
(%)	1.49	3.83	2.86	8.01	4.57	6.78	9.87	4.78	4.53	5.35	3.9	4.17
Silt/clay (%)	0.22	0.4	0.2	0.53	0.25	0.31	0.78	0.31	0.24	0.36	0.24	0.25

Table 1. Values of sedimentary variables throughout the study period in Los Abrigos subtidal.

In terms of species richness, 90 taxa were identified at the sampling station during the study basis. Nematodes were the most diverse group with 48 species, followed by polychaetes (17 taxa) and turbellarians (7 taxa). Species richness varied throughout the study period, with maximum mean values in March 2001 (21.20 taxa) and minimum in July 2000 (14.80). Significant differences were found among the months studied (One-way ANOVA, F = 3.271, p = 0.002) (Fig. 2E).

Meiofaunal abundances ranged from 301 ind $10~\rm cm^{-2}$ in May 2000 and July 2000 to 760 ind $10~\rm cm^{-2}$ (December 2000) and 730 ind $10~\rm cm^{-2}$ (January 2001). Meiofaunal densities varied over the study period, showing minimum abundances during the summer months (May, June and July 2000) and highest densities in winter (January and February 2001) (Fig. 2A). As expected, meiofaunal abundances showed significant differences during the study period (One-way ANOVA, F=25.53, p <<0.0001) (Table 3).

Nematodes

Nematode densities varied from 242.4 ind. $10~\text{cm}^{-2}$ in May 2000 to 725.8 ind. $10~\text{cm}^{-2}$ in December 2000 (Fig. 2B). As was expected, nematodes presented significant differences over the study period (May 2000-April 2001) (One-way ANOVA, F = 27.53; p << 0.0001) (Table 3).

The most abundant nematodes were Daptonema hirsutum and *Pomponema sedecima*, with overall abundances of 9.230 ind. and 9.007 ind., respectively, throughout the study period. The remaining nematode species attained densities lower than 4,500 individuals. The species *Acanthopharynx aff. denticulata*, *Actarjania* sp.1, *Ceramonema*

yunfengi and Scaptrella cf. cincta were rather scarce (3 ind.) during the study period (Table 2).

Copepods

Harpacticoid copepods reached their maximum abundances in January and February 2001, with 41 and 39 ind. 10 cm⁻², respectively. The lowest densities were encountered in April 2001 (14 ind. 10 cm⁻²) and June 2000 (15 ind. 10 cm⁻²).

A total of five species of copepods were identified at the sampling station, *Canuella aff, perplexa* and *Halectinosoma* sp1 being the most abundant with 901 and 565 individuals, respectively. The remaining copepod species attained densities lower than 30 specimens, the species *Asellopsis* sp being represented by one single individual (Table 2).

Polychaetes

Polychaetes attained their highest abundances in June 2000 (20 ind. 10 cm⁻²) and May 2000 (16 ind. 10 cm⁻²), and the lowest densities in July 2000 (3 ind. 10 cm⁻²) and November 2000 (4 ind. 10 cm⁻²).

A total of 16 polychaete species were identified at the sampling station during the study period, the spionids Spio filicornis (269 ind.) and Rhynchospio glutaea (144 ind.) being the most abundant ones. The remaining polychaete taxa were scarce, with densities lower than 60 specimens. The polychaetes Aricidea assimilis, Cirrophorus armatus. Dispio uncinata, Microphthalmus pseudoaberrans, Platynereis dumerilii, Scolelepis squamata and Erinaceusyllis criptica were represented by one single individual (Table 2).

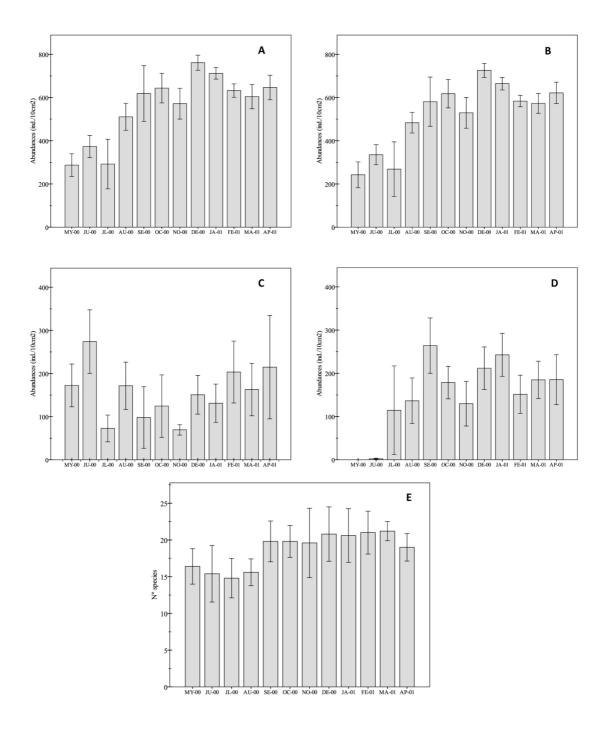


Fig. 2. Mean abundances (\pm SE) throughout the study period (May 2000-April 2001). A. Overall meiofauna. B. Nematodes. C. *Daptonema hirsutum*. D. *Pomponema sedecima*. E. Species richness.

Table 2. Abundances of meiofaunal species (mean \pm SD) throughout the study period (May 2000-April 2001).

	A	May-00	Jun-00	Jul-00	Aug-00	Sep-00	Oct-00	Nov-00	Dec-00	Jan-01	Feb-01	Mar-01	Apr-01
Nematoda	Acanthopharynx aff. denticulata	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.6 ± 1.34	0 ± 0	0 ± 0
Nematoda	Actarjania sp1	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.6 ± 1.34
Nematoda	Actinonema sp	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.8 ± 1.79	0 ± 0
Nematoda	Amphimonhystera sp	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	1 ± 2.24	0 ± 0	0.8 ± 1.79	2.6 ± 3.97	5 ± 5.57	3.6 ± 4.51	0.6 ± 1.34
Nematoda	Sp Catanema sp	1.8 ± 2.49	1 ± 1.41	1 ± 1.41	1 ± 2.24	5.6 ± 8.17	6.8 ± 4.44	1.6 ± 1.67	3.8 ± 4.55	7.2 ± 12.13	6 ± 6.96	4.6 ± 2.79	1.6 ± 2.19
	Ceramonema aff												
Nematoda	yunfengi Metadasynemella	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.6 ± 1.34	0 ± 0
Nematoda	sp	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.8 ± 1.79	0 ± 0	0 ± 0
Nematoda	Choniolaimus sp	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	1 ± 2.24	1.6 ± 3.58	0.6 ± 1.34	0.8 ± 1.79	0 ± 0	2.2 ± 2.05	0 ± 0
Nematoda	Chromadorita sp	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	1.4 ± 1.95
Nematoda	Daptonema hirsutum	172.4 ± 49.57	274 ± 73.56	72.8 ± 31.06	171.6 ± 54.83	98 ± 71.39	124.6 ± 72.26	69.4 ± 12.05	150.8 ± 44.82	131.2 ± 44.46	203.4 ± 71.76	163 ± 61	214.8 ± 119.77
Nematoda	Dasynemoides sp Eleutherolaimus	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	1 ± 2.24	0 ± 0	4.2 ± 4.27	2.6	3.8 ± 2.86	2.6 ± 2.88	1.4 ± 3.13
Nematoda	sp	0 ± 0	0 ± 0	0.4 ± 0.89	0 ± 0	5.2 ± 7.26	2.8 ± 2.59	1.4 ± 3.13	0 ± 0	0 ± 0	0 ± 0	0 ± 0 9.4 ±	0.6 ± 1.34
Nematoda	Enoploides gryphus	3.2 ± 3.56	2.8 ± 3.03	0.8 ± 1.09	16.8 ± 11.48	6.4 ± 8.79	3.8 ± 4.49	4.6 ± 1.82	5.8 ± 3.90	3.4 ± 3.43	3.4 ± 3.71	9.4 ± 10.41	11.6 ± 13.22
Nematoda	Enoplolaimus aff propinquus	0 ± 0	0 ± 0	0 ± 0	1.4 ± 3.13	0 ± 0	0 ± 0	0.6 ± 1.34	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Nematoda	Filitonchus filiformis	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.8 ± 1.79	0 ± 0	0 ± 0
Nematoda	Laimella sp Mesacanthion	0 ± 0	0 ± 0	0 ± 0	0 ± 0 26 ±	0 ± 0 10 ±	0 ± 0	0 ± 0	0 ± 0 10.2 ±	1 ± 2.24	0 ± 0	0 ± 0	0 ± 0
Nematoda	diplechma Microlaimus aff	9.8 ± 2.68	7.4 ± 7.06	5.8 ± 3.03	12.75	11.49	5.4 ± 3.65	1.8 ± 1.79	4.44	7.6 ± 5.73	4.8 ± 7.46	3.4 ± 4.10	3 ± 3.09
Nematoda	acinaces	0 ± 0	0 ± 0	0 ± 0	0 ± 0	4.2 ± 2.77	0 ± 0	1 ± 1.41	2.2 ± 4.92	3.6 ± 3.51	4.6 ± 6.54	0.8 ± 1.79	0.8 ± 1.79
Nematoda	Microlaimus sp3 Monoposthia aff	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.6 ± 1.34	1 ± 2.24	0 ± 0	0 ± 0	0 ± 0
Nematoda	mirabilis	0 ± 0	0 ± 0	0.6 ± 1.34	0 ± 0	0 ± 0	0.8 ± 1.79	6.8 ± 5.89	3.4 ± 4.67	10 ± 9.41	3.8 ± 2.86	0 ± 0	0.8 ± 1.79
Nematoda	Monoposthia sp	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	1.8 ± 4.02	2.8 ± 2.95	2 ± 4.47
Nematoda	Odontophora aff longisetosa	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	5 ± 9.59	0 ± 0	0 ± 0	0 ± 0	3 ± 5.20	0 ± 0
Nematoda	Odontophora aff wieseri	2 ± 2	1.6 ± 2.61	6.4 ± 5.59	16.2 ± 17.51	12.8 ± 10.78	32 ± 17.42	40 ± 14.35	38.4 ± 9.07	52 ± 17.62	43.6 ± 12.72	34.2 ± 20.92	64.6 ± 40.79
Nematoda	Oncholaimellus calvadosicus	9.6 ± 7.06	11.8 ± 5.26	28.8 ± 16.21	62.6 ± 22.87	102 ± 44.06	83 ± 31.92	191.8 ± 24.47	154 ± 31.98	92.6 ± 37.96	29.4 ± 12.05	94.2 ± 9.31	39.2 ± 18.20
Nematoda	Oncholaimus aff skawensis	0 + 0	0 ± 0	0 + 0	0 + 0	0 + 0	0 + 0	2 + 3.08	0 ± 0	1 + 2.24	0.8 ± 1.79	0 + 0	0 + 0
	Oncholaimus	34.4 ±	20.6 ±	19.4 ±	31 ±	24.6 ±	25.8 ±	24.6 ±			67 ±		31 ±
Nematoda Nematoda	campylocercoides Paracomesoma sp	14.26 0 ± 0	10.50 0 ± 0	10.53 0 ± 0	12.25 0 ± 0	4.16 0 ± 0	13.33 0 ± 0	23.86 1.2 ± 1.79	17 ± 4.53 0 ± 0	12 ± 5.57 0 ± 0	33.88 0 ± 0	18 ± 9.27 0 ± 0	22.68 0.6 ± 1.34
Nematoda	Paralinhomoeus sp	1.6 ± 2.19	0.6 ± 1.34	0 ± 0	0 ± 0	6.4 ± 10.06	1.8 ± 2.49	0 ± 0	5.6 ± 5.32	2 ± 2.74	0.8 ± 1.79	0.8 ± 1.79	1.6 ± 2.19
Nematoda	Paralongicyathola imus sp	0 ± 0	0.0 ± 1.54	0.8 ± 1.79	0 ± 0	0 ± 0	1 ± 2.24	0 ± 0	0+0	0+0	0.0 ± 1.79	0.0 ± 1.77	0+0
Nematoda	nnus sp Paramesonchium	0 ± 0	0 ± 0	0.8 ± 1.79	10.4 ±	30.8 ±	1 ± 2.24 96 ±	26.8 ±	89.8 ±	57.8 ±	21.4 ±	20.8 ±	40.8 ±
Nematoda	sp D	0 ± 0	0 ± 0	5.8 ± 4.32 114.6 ±	6.77	18.20	65.67	17.02	30.39	28.77	8.50	9.12 184.8 ±	34.48 185.4 ±
Nematoda	Pomponema sedecima	0 ± 0	2.2 ± 1.48	114.6 ± 102.47	136.4 ± 52.74	264 ± 63.92	178.6 ± 37.84	129.6 ± 51.64	211.8 ± 49.20	242.6 ± 49.62	151.4 ± 44.11	184.8 ± 43.11	185.4 ± 57.70
Nematoda	Pomponema aff reducta	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	2.4 ± 5.37	0 ± 0	0 ± 0	0 ± 0
Nematoda	Pseudochromador a sp1	0 ± 0	0 ± 0	0.6 ± 1.34	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	2.6 ± 3.97	0 ± 0	0 ± 0	0 ± 0
Nematoda	Pseudochromador a sp2	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	4.2 ± 6.94	0.4 ± 0.89	1 ± 2.24	5.2 ± 4.76	2.2 ± 2.04	2.2 ± 2.05	1.6 ± 2.19
Nematoda	Richtersia sp	0 ± 0	0 ± 0	0 ± 0	0.8 ± 1.79	0 ± 0	0 ± 0	3.6 ± 8.05	0 ± 0	0.8 ± 1.79	0 ± 0	0 ± 0	0 ± 0
Nematoda	Sabatieria aff celtica	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.8	0 ± 0
Nematoda	Sabatieria aff elongata	1.2 ± 1.79	1.6 ± 3.58	0 ± 0	0 ± 0	1.4 ± 3.13	0 ± 0	1 ± 1.41	4.2 ± 5.76	3.6 ± 5.68	2.2 ± 3.19	3.6 ± 6.50	1.2 ± 1.64
Nematoda	Sabatieria aff longisetosa	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.6 ± 1.34	0 ± 0	0 ± 0	0.8 ± 1.79	0 ± 0	0 ± 0
Nematoda	Scaptrella cf. cinctap	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.6 ± 1.34	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Nematoda	Setosabatieria aff hilarula	3.4 ± 3.71	1.4 ± 1.95	1.8 ± 4.02	0.4 ± 0.89	2.8 ± 4.76	3.4 ± 3.43	9.6 ± 8.08	10 ± 5.96	3.6 ± 2.07	8 ± 5.70	6 ± 2.45	3.4 ± 3.51
Nematoda	Siphonolaimus aff niger	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.4 ± 0.89	1 ± 2.24	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Nematoda	Siphonolaimus aff pelllucidus	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0±0	0 ± 0	12 ± 26.83	0 ± 0	0.8 ± 1.79	0 ± 0
	-										0 ± 0 0 ± 0		0 ± 0 0.6 ± 1.34
Nematoda	Siphonolaimus sp2 Spirinia	0 ± 0	0 ± 0 9.8 ±	0 ± 0	0.8 ± 1.09	0 ± 0	9 ± 15.26 33.8 ±	0.4 ± 0.89	0 ± 0	0 ± 0	17.8	1.6 ± 2.19	12.2 ±
Nematoda	parasitifera	2.2 ± 2.86	12.38	9.2 ± 4.44	5.8 ± 4.09	5.2 ± 5.45	16.51	1.4 ± 1.95	9 ± 8.46	2.8 ± 2.59	±11.88	8.4 ± 3.91	7.43
Nematoda	Thalassironus sp1	0 ± 0	0.4 ± 0.89	0 ± 0	0.8 ± 1.79	0.6 ± 1.34	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Nematoda	Trefusia sp Trileptium aff	0 ± 0	0 ± 0	0 ± 0	1.4 ± 3.13	0 ± 0	1 ± 2.24	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Nematoda	parisetum	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.8 ± 1.79	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0

Table 2. Continuation.

		May-00	Jun-00	Jul-00	Aug-00	Sep-00	Oct-00	Nov-00	Dec-00	Jan-01	Feb-01	Mar-01	Apr-01
Nematoda	Viscosia glabra	0.8 ± 1.09	0 ± 0	0 ± 0 12.4 ±	0 ± 0	0 ± 0	0.8 ± 1.79	1.4 ± 1.95	1.6 ± 2.19	0 ± 0 21.6 ±	0 ± 0 14.2 ±	0 ± 0 12.2 ±	0 ± 0
Copepoda	Halectinosoma sp	5 ± 5.96	3.4 ± 1.52	10.97	6.2 ± 6.38	8.4 ± 6.07	4 ± 1	3.4 ± 3.13	6.8 ± 3.77	7.47	3.03	4.55	9.6 ± 9.99
Copepoda	Tryphonema sp	0.6 ± 0.89	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Copepoda	Asellopsis sp Canuella aff.	0 17.6 ±	0 ± 0 13.4 ±	0 ± 0	0.2 ± 0.45 10.6 ±	0 ± 0 17.2 ±	0 ± 0 13.2 ±	0 ± 0 32.8 ±	0 ± 0 15.6 ±	0 ± 0 18.2 ±	0 ± 0 23 ±	0 ± 0 11.8 ±	0 ± 0
Copepoda	perplexa Harpacticus aff.	12.15	10.88	6 ± 5.48	4.72	11.26	5.02	21.98	12.11	6.02	11.25	11.88	6.2 ± 5.02
Copepoda	flexus	2.2 ± 3.90	0 ± 0	0 ± 0	0.2 ± 0.45	0.2 ± 0.45	0.2 ± 0.45	0.4 ± 0.89	0 ± 0	0.8 ± 1.79	1.6 ± 1.52	0.4 ± 0.89	0 ± 0
Polychaeta	Aricidea assimilis Capitomastus	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Polychaeta	minimus Cirrophorus	0 ± 0	0 ± 0	0 ± 0	0.4 ± 0.89	0 ± 0	0 ± 0	0.2 ± 0.45	0 ± 0	0 ± 0	0.2 ± 0.45	0 ± 0	0 ± 0
Polychaeta	armata	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.2 ± 0.45	0 ± 0
Polychaeta	Dispio uncinata	0 ± 0	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Polychaeta	Erinaceusyllis cryptica	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Polychaeta	Exogone breviantennata Microphthalmus	1 ± 1	0 ± 0	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.2 ± 0.45	0 ± 0	0.2 ± 0.45	0 ± 0	0.2 ± 0.45
Polychaeta	pseudoaberrans	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Polychaeta	Pararicia sp Pionosyllis	0.2 ± 0.45	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Polychaeta	spinisetosa Platynereis	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.2 ± 0.45	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0.2 ± 0.45	0.2 ± 0.45	0 ± 0
Polychaeta	dumerilii Pseudopolydora	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Polychaeta	sp Rhynchospio	0 ± 0	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Polychaeta	glutaea	2.8 ± 2.68	3.4 ± 1.52	0.4 ± 0.89	2.6 ± 1.82	2.6 ± 2.97	1.8 ± 2.39	1.4 ± 1.52	3.8 ± 4.21	1 ± 1	1 ± 1.73	0.8 ± 0.84	1.2 ± 0.45
Polychaeta	Scoloplos armiger Schoeredella	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.6 ± 0.55	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Polychaeta	laubieri Scolelepis	0.8 ± 1.30	1 ± 0.71	0 ± 0	0 ± 0	0.8 ± 0.45	1 ± 1.41	0.6 ± 0.55	0.8 ± 0.84	0 ± 0	0 ± 0	0 ± 0	0.4 ± 0.55
Polychaeta	squamata	0 ± 0	0 ± 0 12.4 ±	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0 ± 0 5.8 ±
Polychaeta	Spio filicornis Streptosyllis	9 ± 5.24	3.85	1.2 ± 0.84	4.6 ± 3.78	1.6 ± 2.07	3.4 ± 2.19	1.6 ± 1.52	5.6 ± 4.39	2 ± 1.73	3.2 ± 2.59	3.2 ± 0.84	4.76
Polychaeta	bidentata	2.2 ± 1.92	2.6 ± 1.67	0.8 ± 0.84	0.4 ± 0.55	1.6 ± 1.14	1.2 ± 1.09	0.2 ± 0.45	1.2 ± 2.17	0.2 ± 0.45	0.4 ± 0.55	0.2 ± 0.45	0.2 ± 0.45
Turbellaria	Acoela sp 1	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.4 ± 0.55	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Turbellaria	Catenulida sp 1	0.6 ± 0.89	0 ± 0	0.6 ± 0.89	0.4 ± 0.89	1.8 ± 2.39	0.2 ± 0.45	0 ± 0	0 ± 0	3.2 ± 3.03	1.2 ± 1.09	0.4 ± 0.55	0.2 ± 0.45
Turbellaria	Catenulida sp 2	0 ± 0	0 ± 0	0.2 ± 0.45	0 ± 0	0.6 ± 0.89	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0	0 ± 0
Turbellaria	Catenulida sp 3 Catenulida aff	0.2 ± 0.45	0.2 ± 0.45	0.6 ± 1.34	0 ± 0	0.4 ± 0.55	0.4 ± 0.55	0.2 ± 0.45	0 ± 0	0.4 ± 0.89	1.6 ± 2.30	1.4 ± 1.14	0.6 ± 0.55
Turbellaria	catenulidae Haplopharyngida	0 ± 0	0 ± 0	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Turbellaria	sp 1	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Turbellaria	Proseriata sp 1	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.2 ± 0.45	0 ± 0	0 ± 0	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Acari	Acaridae Ampelisca	0 ± 0	0.4 ± 0.55	0.2 ± 0.45	0 ± 0	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Amphipoda	brevicornis	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0.2 ± 0.45	0 ± 0
Amphipoda	Bathyporeia sp	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Amphipoda	Lysianassidae Pontocrates	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.2 ± 0.45	0 ± 0	0 ± 0
Amphipoda	arenarius	0.4 ± 0.89	0.8 ± 1.30	0 ± 0	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.2 ± 0.45	1.4 ± 2.61	0 ± 0	0.2 ± 0.45
Amphipoda Nemertea	Stenothoidae Ototyphlonemertes	0.2 ± 0.45 0 ± 0	0 ± 0 0 ± 0	0 ± 0	0 ± 0 0 ± 0	0 ± 0	0.2 ± 0.45 0 ± 0	0 ± 0	0 ± 0 0 ± 0	0 ± 0			
	sp 2 Gastrosaccus						0 ± 0		0 ± 0		0 ± 0		0.2 ± 0.45
Mysidacea	sanctus	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.2 ± 0.45	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Oligochaeta	Aktedrilus sp	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.6 ± 0.55	0 ± 0	0 ± 0	0 ± 0	0.2 ± 0.45
Oligochaeta	Grania sp Cypridina aff	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0.8 ± 0.84	0.2 ± 0.45	0 ± 0	0.4 ± 0.89	0 ± 0	0 ± 0	0 ± 0
Ostracoda	mediterranea	0.4 ± 0.89	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Ostracoda	Leptocythere sp	0 ± 0	0 ± 0	0 ± 0	0.2 ± 0.45	0 ± 0 1.4 ±	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0
Tanaidacea	Apseudes talpa	1 ± 1.73	0.6 ± 0.55	0.2 ± 0.45	0 ± 0	0.89	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0

Minor Taxa

Minor taxonomic groups comprised turbellarians, tanaids, amphipods, oligochaetes, acari, ostracods, misids and nemerteans. A total of 20 species were identified during the study period (7 turbellarians, 5 amphipods, 2 oligochaetes, 2 ostracods, 1 tanaid, 1 acari, 1 misid and 1 nemertean). The most abundant species were the

turbellarians Catenulida sp.1 (43 ind.) and Catenulida sp.3 (30 ind.), the remaining minor taxa being scarce (< 20 ind.). The least abundant species were the amphipod *Bathyporeia* sp., the misid *Gastrosaccus sanctus*, the turbellarian Haplopharyngida sp.1, the ostracod *Leptocythere* sp.1, the amphipod Lysianassidae and the nemertean *Ototyphlonemertes* sp.2, with one single specimen (Table 2).

Table 3. Results of univariate ANOVA testing for differences in overall meiofauna, nematode, *Daptonema hirsutum* and *Pomponema sedecima* abundance throughout the study period (May 2000-April 2001). Significant differences are highlighted in bold.

Overall meiofaunal abundance			Nematode abundances			Daptonema hirsutum abundances			Pomponema sedecima abundances				
Source of variation	df	MS	F	p	MS	F	p	MS	F	p	MS	F	p
Time	11	123,606.28	25.53	<<0.0001	122,373.20	27.53	<<0.0001	34033.22	12.20	<<0.0001	17,975.92	4.357	0.00016

Species Assemblage

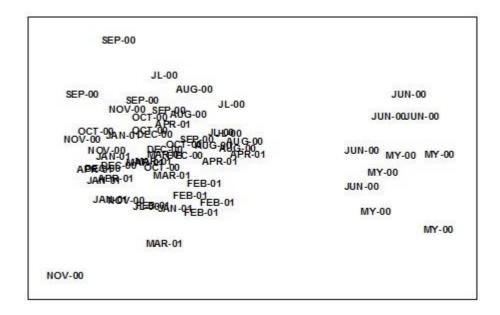
The most abundant species were the nematodes *Daptonema hirsutum* and *Pomponema sedecima*, that made up 54.82% of the overall meiofaunal abundance. *Daptonema hirsutum* was more abundant in June $2000~(274~{\rm ind}~10{\rm cm}^{-2})$ and April $2001~(214.8~{\rm ind}~10{\rm cm}^{-2})$, decreasing to 69.4 and $72.8~{\rm ind}~10~{\rm cm}^{-2}$ in July and November 2000, respectively (Fig. 2C). *Daptonema hirsutum* densities varied significantly over the study period (One-way ANOVA; F=4.357, p=0.00016) (Table 3).

The nematode *Pomponema sedecima* reached its maximum densities in September 2000 (264 ind 10cm⁻²) and January 2001 (242.6 ind 10cm⁻²). Its minimum abundances were registered in May 2000 (absent), June 2000 (2.2 ind 10 cm⁻²) and July 2000 (114.6 ind 10 cm⁻²) (Fig. 2D). *Pomponema sedecima* densities showed highly

significant differences during the study period (One-way ANOVA; F = 12.196, $p \ll 0.0001$) (Table 3).

Multivariate Analysis

Differences in the meiofauna assemblage structure during the months studied were observed in the MDS (Fig. 3), with a clear distinction between May and June 2000 and the remaining months of the study year. The former months were characterized by characterized by low abundances of nematodes and high densities of polychaetes. In terms of species, that group (May and June 2000) was dominated by the nematode Daptonema hirsutum and, to a lesser extent, by the nematodes Oncholaimus campylocercoides and Oncholaimellus calvadosicus, as well as, the harpacticoid copepod Canuella aff: perplexa.



 $Fig.\ 3.\ Multidimensional\ Scaling\ (MDS)\ of\ sampling\ months\ during\ the\ study\ period\ (May\ 2000-April\ 2001)\ (stress=0.16).$

The analysis of matrices similarity (RELATE) showed significant differences throughout the sampling year (number of permutations used = 20.000, Rho = 0.234; significance level = 0.1%). There is thus a seasonality in the meiofaunal community structure of Los Cristianos subtidal station during the sampling period (May 2000-April 2001). These results were mainly due to the seasonal variability of the two most abundant nematode species (Daptonema hirsutum and Pomponema sedecima) and, to a lesser extent, of the meiofaunal species composition of May and June 2000.

The multivariate multiple regression showed that none of the environmental variables measured contributed significantly to explain the patterns of the meiofaunal assemblage structure (all p-values > 0.05, Table 4). This is mainly due to the environmental stability at the sampling station throughout the study period (May 2000-April 2001).

DISCUSSION

Meiofauna densities showed intermediate values, with an overall mean of 554.4 ind 10 cm⁻² throughout the sampling period (May 2000-April 2001). These abundances are not consistent with previous data concerning shallow subtidal densities (SANDULLI ET AL, 2002; VEZZULLI AND FABIANO, 2006; LEONARDIS ET AL, 2008), because of the lower meiofaunal ones (ca. 200-400 ind 10 cm⁻²) observed in the western Mediterranean. In this area, low primary production rates which are related to the depletion of nutrients in the euphotic zone, which provides a very low organic matter supply to the benthos due to a restriction of the vertical transport of particles to the seabed (SACHS AND REPETA, 1999). In terms of meiofauna composition, the high dominance of nematodes in fine-sandy subtidal seabeds has already been observed (e.g. SEMPRUCCI ET AL, 2010). In short, meiofaunal and nematode abundances were higher during winter months (December 2000-March 2001), with a sharp decrease in the spring-summer months, especially accentuated in May 2000

Apparently, the meiofaunal assemblages from Los Cristianos bay here studied are not affected by the oligotrophic state of the water masses of the Canary archipelago (BARTON ET AL, 1998). Moreover, low organic matter content (avg. 0.54%) was found in the sediments analyzed, throughout the study period (RIERA ET AL, 2011a); the sedimentary dynamics of Los Cristianos bay could, however, affect meiobenthos community structure. It will be necessary to take this effect into consideration in future studies, giving special emphasis to sediment deposition.

The present study showed a meiofauna community structure typical of a shallow site with several abundant species that are influenced by coastal constraints (winds, waves, currents, freshwater runoff, etc.). The most abundant species (the nematodes Daptonema hirsutum and Pomponema sedecima) showed several peaks during the study period, with no clear temporal pattern. These species dominated the meiofauna assemblage structure throughout the study period, and they have been recorded abundantly in fine sands worldwide (KENNEDY, 1994; BLOME ET AL, 1999; GESHKIERE ET AL, 2002). The xyalid D. hirsutum has been observed to be an active migrator to fine sands in experiments of simulated deposition of dredged material (SCHRATZBERGER ET AL, 2000). Moreover, these species are typical of sandy substrates with low organic sedimentary content, being scarce or absent in enriched muddy sediments or the "thiobios" components (RPD layer) of reduced environments (SEMPRUCCI ET AL., 2010). Both species (D. hirsutum and P. sedecima) showed significant differences in their abundances throughout the study period, directly influenced the overall meiofaunal abundances.

Table 4. Multivariate multiple regression testing the effect of environmental variables on the overall meiofaunal assemblage structure SS(trace) = portion of sum of squares relative to the analysed predictor variable; pseudo-F = statistic; p = significance level; prop = proportion of variation explained.

Variable	SS (trace)	Pseudo-F	р	prop
Total Nitrogen (%)	2314.5	8.633	0.423	0.098
Organic matter (%)	2102.9	10.002	0.401	0.089
Gravels (%)	1788.7	0.876	0.614	0.074
Very coarse sands (%)	1214.5	0.543	0.689	0.069
Coarse sands (%)	3180.9	17.011	0.115	0.156
Medium sands (%)	1103.3	0.465	0.856	0.056
Fine sands (%)	1567	0.834	0.324	0.091
Very fine sands (%)	859.35	0.456	0.917	0.047
Silt/clay (%)	1014.9	0.502	0.826	0.058

The study station was characterized by low variations of sedimentary properties during the year studied, with a dominance of fine sands and low concentrations of organic matter. However, the environmental variables measured were unable to explain the temporal variability of meiofauna assemblages during the study period.

Thus, other environmental factors could be responsible for the meiofaunal variations in the Los Cristianos subtidal location. For areas, no inequivocal example, in subtidal influence of phytoplankton sedimentation on the density and biomass of meiofaunal nematodes was found (HEIP ET AL, 1985, ÓLAFSSON ET AL, 1999). However, SCHRATZBERGER ET AL (2007) showed that populations of the nematode Spilophorella paradoxa that feed on fresh organic material exhibited temporal patterns in their abundance and size distribution. Moreover, the temporality of this species was clearly linked to the quality and quantity of organic matter in the sediment (SCHRATZBERGER ET AL, 2007).

One important ecological factor is predation, the effects of which on soft-substrates have received little attention (GREGG AND FLEEGER, 1997). Meiofauna are potentially an important food source for higher trophic levels, particularly macrofauna, flat-fish, juveniles of larger fish, crabs, shrimps and epibenthic predators (HOYT ET AL, 2000; FELLER, 2006). Numerous individuals of sand steenbras (*Lithognathus mormyrus*) were observed at the sampling station throughout the study period (RIERA, unpubl. data). This species is an important predator on interstitial species, especially epibenthic taxa (e.g. some harpacticoid species) (COULL, 1990).

A more detailed and complete ecological study of the study location is necessary in order to understand meiofauna dynamics and community structure; this study should include other environmental factors (e.g. sedimentary dynamics) and biological interactions (e.g. predation).

ACKNOWLEDGEMENTS

We are grateful to Dr. T. Gheskiere and Dr. M. Vincx (Department of Marine Biology, University of Ghent, Belgium) for the bibliographical facilities offered and their encouragement during the stay of the first author (R.R.), as also to Dr. F. Tuya (University of Las Palmas de Gran Canaria) for his continuous encouragement and interchange of ideas.

REFERENCES

- ALBUQUERQUE, E. F.; PINTO, A. P. ALCÂNTARA, A. GOMES, V. Spatial and temporal changes in interstitial meiofauna on a sandy ocean beach of South America. Braz. J. of Oceanogr., 55(2):121-131. 2007.
- BLOME, D.; SCHELIER, U. VAN BERNEM, K. H. Analysis of the small-scale spatial patterns of freeliving marine nematodes from tidal flats in the East Frisian Wadden Sea. Mar. Biol., 133(4): 717-726. 1999.
- COULL, B. C. Long-term variability of estuarine meiobenthos: an 11 year study. Mar. Ecol. Prog. Ser., 24:205-218. 1985.
- COULL, B. C. Ecology of the marine meiofauna. In: Higgins RP and Thiel H (ed.) Introduction to the study of Meiofauna. Washington, D.C. Smithsonian Institution Press, 18-38 pp. 1988.
- COULL, B. C. Are members of the meiofauna food for higher trophic levels? Trans. Am. Micros. Soc., 109(3): 233-246. 1990.
- DANOVARO, R.; FABIANO, M. VINCX, M. Meiofauna response to the Agrip Abruzzo oil spill in subtidal sediments of the Ligurian Sea. **Mar. Pollut. Bull.**, 30: 133-145, 1995.
- DANOVARO, R. Detritus-bacteria-meiofauna interactions in a seagrass bed (Posidonia oceanica) of the NW Mediterranean. **Mar. Biol.**, 127: 1-13. 1996.
- FELLER, R. J. Weak meiofaunal tropic linkages in Crangon crangon and Carcinus maenas. J. Exp. Mar. Biol. Ecology, 330: 274-283. 2006.
- FLEEGER, J. W.; SHIRLEY, T. C. ZIEMANN, D. A. Meiofaunal responses to sedimentation from an Alaskan spring bloom. I. Major taxa. Mar. Ecol. Prog. Ser., 57: 137-145. 1989.
- GHESKIERE, T.; HOSTE, E. KOTWICKI, L. DEGRAER, S. VANAVERBEKE, J. VINCX, M. The sandy beach meiofauna and free-living nematodes from De Panne (Belgium). Bulletin de L'Institut Royal des Sciences Naturelles de Belgique, 72(S): 43-49, 2004.
- GREGG, J. C.; FLEEGER, J. W. Importance of emerged and suspended meiofauna to the diet of the darter goby (Gobionella bolesoma Jordan and Gilbert). J. Exp. Mar. Biol. Ecol., 209: 103-142. 1997.
- GRÉMARE, A.; AMOROUX, J. M. CHARLES, F. DINET, A. RIAUX-GOBIN, C. BAUDART, J. MEDERNACH, L. BODIOU, J. Y. VETION, G. COLOMINES, J. C. ALBERT, P. Temporal changes in the biochemical composition and nutritional value of the particulate organic matter available to surface deposit-feeders: a two year study. Mar. Ecol. Progr. Ser., 150: 195-206. 1997.
- GUIDI-GUILVARD, L. D.; BUSCAIL, R. Seasonal survey of metazoan méiofaune and surface sediment organics in a non-tidal turbulent sublittoral prodelta (northwestern Mediterranean). Cont. Shelf Res.,15(6): 633-653. 1995.
- HEIP, C.; VINCX, M. VRANKEN, G. The ecology of marine nematodes. Oceanography and Marine Biology: An Annual Review 30: 149-190. 1985.

- HICKS, G. R.; COULL, B. C. The ecology of marine meiobenthic harpacticoid copepods. Oceanography and Marine Biology Annual Review 21:67-175. 1983.
- HOYT, M.; FLEEGER, J. W. SEIBELING, R. FELLER, R. J. Serological estimation of prey-protein gutresidence time and quantification of meal size for grass shrimp consuming meiofaunal copepods. J. Exp. Mar. Biol. Ecol., 248: 105-119. 2000.
- KENNEDY, A. D. Carbon partitioning within meiobenthic nematode communities in the Exe Estuary, UK. Mar. Ecol. Prog. Ser., 105: 71-78. 1994.
- DE LEONARDIS, C.; SANDULLI, R. VANAVERBEKE, J. VINCX, M. DE ZIO, S. Meiofauna and nematode diversity in some Mediterranean areas of the Adriatic and Ionian Sea. Scientia Marina, 72(1): 5-13. 2008.
- MAZZOLA, A.; MIRTO, S. LA ROSA, T. FABIANO, M. DANOVARO, R. Fish-farming effects on benthic community structure in coastal sediments: analysis of meiofaunal recovery. ICES Journal of Marine Science 57:1454-1461. 2000.
- MIRTO, S.; LA ROSA, T. DANOVARO, R. MAZZOLA, A. Microbial and meiofaunal response to intensive mussel-farm biodeposition in coastal sediments of the Western Mediterranean. Mar. Pollut. Bull., 40(3):244-252. 2000.
- NOZAIS, C.; PERISSINOTTO, R. TITA, G. Seasonal dynamics of meiofauna in a South African temporarily open/closed estuary (Mdloti Estuary, Indian Ocean). **Estuar. Coast., Shelf Sci.**, 62: 325-338. 2005.
- ÓLAFSSON, E.; ELMGREN, R. Seasonal dynamics of sublittoral meiobenthos in relation to phytoplankton sedimentation in the Baltic Sea. Estuar. Coast., Shelf Sci., 45: 149-164. 1997.
- ÓLAFSSON, E.; MODIG, H. VAN DEN BUND, W. J. Species-specific uptake or radiolabelled phytodetritus by benthic meiofauna from the Baltic Sea. Mar. Ecol. Prog. Ser., 177: 63-72. 1999.
- PALACÍN, C. Estudio ecológico de la meiofauna bentónica de la Bahía de Els Alfacs (Delta del Ebro). Ecología y sistemática de las poblaciones de nematodos. Ph-D Thesis. University of Barcelona, 406 pp. 1990.
- RIERA, R.; NÚÑEZ, J. BRITO, M. C. Short-term variability in sedimentary and compositional properties of two beaches in Tenerife (Canary Islands, NE Atlantic Ocean). **Revista de la Sociedad Geológica de España**, 23(3-4): 81-87. 2011a.
- RIERA, R.; NÚÑEZ, J. BRITO, M. C. Temporal variability of a subtropical intertidal meiofauna assemblage: contrasting effects at the species and assemblage-level. **Vie et Milieu**, 61(3): 1-9. 2011b.

- RUDNICK, D.; ELMGREN, R. FRITHSEN, J. B. Meiofaunal prominence and benthic seasonality in a coastal marine ecosystem. Oecologia, 67: 157-168. 1985
- SACHS, J. P., REPETA, D. J. Oligotrophy and nitrogen fixation during eastern Mediterranean Sapropel events. Science, 286, 2485. 1999.
- SANTOS, P. J. P.; CASTEL, J. SOUZA-SANTOS, L. P. Seasonal variability of meiofauna abundance in the oligo-mesohaline area of the Gironde estuary, France. Estuar. Coast. Shelf Sci., 43: 549-563. 1996.
- SCHIZAS, N. V.; SHIRLEY, T. C. Seasonal changes in structure to an Alaskan intertidal meiofaunal assemblage. Mar. Ecol. Progr. Ser., 133: 115-124. 1996
- SCHRATZBERGER, M.; FORSTER, R. M. GOODSIR, F. JENNINGS, S. Nematode community dynamics over an annual production cycle in the central North Sea. Mar. Environ. Res., 66: 508-519. 2008.
- SCHRATZBERGER, M.; REES, H. L. BOYDS, S. E. Effects of simulated deposition of dredged material on structure of nematode assemblages – the role of burial. Mar. Biol., 136: 519-530. 2000.
- SEMPRUCCI, F.; COLANTONI, P. BALDELLI, G. ROCCHI, M. BALSAMO, M. The distribution of meiofauna on back-reef sandy platform in the Maldives (Indian Ocean). Mar. Ecol., 31(4): 592-607.
- SOMERFIELD P.; WARWICK, R. M. Meiofauna in marine pollution programmes. A laboratory manual. Ministry of Agriculture, Fisheries and Food, Directorate of Fisheries Research, Lowestoft, 71 pp. 1996
- VANHOVE, S.; BEGHYN, M. VAN GANSBEKE, D. BULLOUGH, L. W. VINCX, M. A seasonally varying biotope at Signy Island, Antarctica: implications for meiofaunal structure. **Mar. Ecol. Progr. Ser.**, 202: 13-25. 2000.
- VEZZULLI, L. FABIANO, M. Sediment biochemical and microbial variables for the evaluation of trophic status along the Italian and Albanian Continental Shelves. J. Mar. Biol. Assoc. UK., 86, 27-37. 2006.
- VINCX, M. Seasonal fluctuations and production of nematode communities in the Belgian coastal zone of the North Sea. Belgian Journal of Zoology, 15: 57-66. 1989.

(Manuscript received 30 April 2011; revised 16 June 2014; accepted 14 June 2014)