

Fall 10-7-2013

Mechanism of amyloid β -protein dimerization determined using single-molecule AFM force spectroscopy.

Zhengjian Lv

University of Nebraska Medical Center, zhengjian.lv@unmc.edu

Robin Roychaudhuri

David Geffen School of Medicine at UCLA

Margaret M Condrón

David Geffen School of Medicine at UCLA

David B. Teplow

David Geffen School of Medicine at UCLA

Yuri L. Lyubchenko

University of Nebraska Medical Center, ylyubchenko@unmc.edu

Follow this and additional works at: https://digitalcommons.unmc.edu/cop_pharmsci_articles

 Part of the [Pharmacy and Pharmaceutical Sciences Commons](#)

Recommended Citation

Lv, Zhengjian; Roychaudhuri, Robin; Condrón, Margaret M; Teplow, David B.; and Lyubchenko, Yuri L., "Mechanism of amyloid β -protein dimerization determined using single-molecule AFM force spectroscopy." (2013). *Journal Articles: Pharmaceutical Sciences*. 16.
https://digitalcommons.unmc.edu/cop_pharmsci_articles/16

This Article is brought to you for free and open access by the Pharmaceutical Sciences at DigitalCommons@UNMC. It has been accepted for inclusion in Journal Articles: Pharmaceutical Sciences by an authorized administrator of DigitalCommons@UNMC. For more information, please contact digitalcommons@unmc.edu.



OPEN

Mechanism of amyloid β -protein dimerization determined using single-molecule AFM force spectroscopy

SUBJECT AREAS:

SINGLE-MOLECULE
BIOPHYSICS

ALZHEIMER'S DISEASE

Received
26 July 2013Accepted
20 September 2013Published
7 October 2013Correspondence and
requests for materials
should be addressed to
Y.L.L. (lyubchenko@
unmc.edu)Zhengjian Lv¹, Robin Roychoudhuri², Margaret M. Condrón², David B. Teplow^{2,3} & Yuri L. Lyubchenko¹

¹Department of Pharmaceutical Sciences, University of Nebraska Medical Center, 986025 Nebraska Medical Center, Omaha, NE 68198, United States, ²Department of Neurology, ³Brain Research Institute and Molecular Biology Institute, David Geffen School of Medicine at UCLA, Los Angeles, CA 90095, United States.

A β 42 and A β 40 are the two primary alloforms of human amyloid β -protein (A β). The two additional C-terminal residues of A β 42 result in elevated neurotoxicity compared with A β 40, but the molecular mechanism underlying this effect remains unclear. Here, we used single-molecule force microscopy to characterize interpeptide interactions for A β 42 and A β 40 and corresponding mutants. We discovered a dramatic difference in the interaction patterns of A β 42 and A β 40 monomers within dimers. Although the sequence difference between the two peptides is at the C-termini, the N-terminal segment plays a key role in the peptide interaction in the dimers. This is an unexpected finding as N-terminal was considered as disordered segment with no effect on the A β peptide aggregation. These novel properties of A β proteins suggests that the stabilization of N-terminal interactions is a switch in redirecting of amyloids from the neurotoxic aggregation pathway, opening a novel avenue for the disease preventions and treatments.

A aberrant folding (misfolding) and aggregation of amyloid proteins are pathological hallmarks of a large class of neurodegenerative diseases, including Alzheimer's disease (AD) (reviewed in^{1,2}). In AD, amyloid precursor proteins are sequentially cleaved by β -secretase and γ -secretase, producing the amyloid β -protein (A β)³. A β peptides contain several alloforms with varying sequences of amino acids, dependent on the cleavage sites of γ -secretase. While A β 40 is the most abundant species among these alloforms, A β 42 is the most amyloidogenic species, and also appears to be the most toxic⁴⁻⁶. It was shown that the two extra C-terminal residues of A β 42 dramatically change the peptide's oligomerization properties compared to A β 40^{7,8}. A β 42 prefers to form a pentamer/hexamer structure (paranucleus) that can nucleate fibril formation⁷. By contrast, A β 40 reaches rapid equilibrium with coexisting monomers, dimers, trimers, and tetramers⁷. According to computer simulations, A β 42 has a turn-like fold at the C-terminus that involves Met35-Gly38 (referred to as a C-turn), suggesting that the more structured C-terminus of A β 42 generates greater intramolecular contacts than A β 40⁹. The hypothesis that the C-terminal residues play an important role in the A β aggregation process was recently tested by¹⁰, in which a set of A β peptides with defined alterations in the C-terminus were studied. Specifically, it was shown that VPV substitutions (Gly33Val-Val36Pro-Gly38Val) stabilized the β -hairpin structure by reducing the flexibility of the peptide backbone and by strengthening hydrophobic interactions between the putative β -strands. A similar VPV motif in an A β 40 mutant changed the oligomerization pattern to one similar to A β 42, resulting in increased toxicity of the oligomers compared with the toxicity of wild type (WT) A β 40¹⁰. *pP* substitutions (Val36/D-Pro)-(Gly37/L-Pro) destabilized the putative C-turn structure and thereby made [*pP*]A β 42 "A β 40-like"¹⁰.

We have developed a single-molecule atomic force microscopy (SMFS) approach enabling us to probe and characterize the interactions of the peptides and proteins in transiently misfolded conformations¹¹⁻¹⁵. In this approach, the protein is end-immobilized on the AFM tip and substrate, allowing us to measure the interprotein interactions at the single-molecule level. The dynamics force spectroscopy (DFS) approach has been quite informative with respect to the lifetimes of transiently formed misfolded dimers (reviewed in¹⁶). These studies, applied to A β 40¹², A β 42¹⁴, α -synuclein¹³ and Sup35¹⁵, revealed that misfolded dimers have an unexpectedly high stability. Their lifetimes exceeded the characteristic lifetimes of monomers by orders of magnitudes, suggesting



that dimerization stabilized the misfolded protein state. Recent molecular dynamic (MD) simulations of A β (13–23) peptide directly support this view and show that the monomers adopt antiparallel β -sheet structures resulting in high dimer stability¹⁷. In addition to measuring the stability of dimers, SMFS allowed us to characterize the structure of the dimers and to identify the protein segments involved in dimer formation^{11–14}. Using this approach, we showed that Cu²⁺ cations change the conformation of A β 42 peptide, and dimer formation involves the N-terminal segments¹⁴. In the absence of the cations, A β 42 peptides formed dimers via the C-terminal segments.

Here, we applied both SMFS approaches to characterize the effects of C-terminal substitutions on the structure of transiently formed dimers. The DFS approach revealed that VPV mutations in both A β 40 and A β 42 peptides resulted in stabilization. Additionally, the patterns of formation of misfolded A β dimers were studied by contour length analysis. These data provided an explanation of the stabilizing effect of the peptide C-terminus. We demonstrated that the interaction pattern of A β 40 involving N-termini reduces dimers stability. These results are discussed in the context of the role of the N-terminus on the global structural reorganization of the misfolded A β dimers and their aggregation propensity.

Results

Single-molecule force spectroscopy. The schematic of the SMFS experiments is shown in Fig. 1a. The cysteinyl-peptides were covalently immobilized on the tip and the surface via flexible tethers terminated with maleimide groups. Based on NMR and hydrogen-deuterium exchange experiments, Cys residues placed at the N-termini of the A β peptides should not significantly affect peptide structure, or aggregation^{18,19}. Peptide aggregation was minimized by using a very low peptide concentration (20 nM) during the coupling reaction with the functionalized mica surface and the tips. Probing was performed by multiple approach-retraction cycles over various spots of the mica surface.

Figures 1–b1 and 1–b2 show typical force curves for non-specific and specific interactions, respectively. Specific rupture events were characterized by the appearance of the event indicated with an arrow in Fig. 1–b2. The rupture peak appears at the distance defined by the extension of the flexible linkers (polyethylene glycol, PEG and maleimide silatrane, MAS) and the segments of the peptides not involved in dimer formation (see Supplementary Methods for specifics). The force–distance curves, as shown in Fig. 1–b2, were analyzed with worm-like chain (WLC) fitting to extract parameters including the rupture force and the contour length. Rupture force distributions at specified loading rates were plotted and fitted with a probability density function, as demonstrated by the histogram in Fig. 1c obtained for A β 40. The measurements for the [VPV]A β 40 mutant performed at similar conditions (ionic strength, loading rate, etc.) are shown in Fig. 1d. Both histograms (1c and 1d) comprise narrow distributions of rupture forces, but the histogram 1d is shifted towards significantly higher rupture forces. The maxima of Gaussians were 83.6 pN and 57.0 pN for [VPV]A β 40 and A β 40, respectively, suggesting that the dimers formed by [VPV]A β 40 are more stable than those formed by the WT A β 40. The rupture forces increase with increasing apparent loading rates (Supplementary Fig. S1). To quantitatively characterize the properties of the dimers formed by A β 40 and A β 42 peptides, we employed the DFS approach, as described below.

Dynamic force spectroscopy of misfolded A β dimers. In the DFS approach, multiple force measurements were performed at different pulling rates and the data were grouped into several clusters according to the selected loading rates. According to equation (3) (see Methods), the force data were linearly dependent on the logarithm of the loading rate. After extrapolating the data to the

zero pulling rate, the intercept and the slope of the linear plots (DFS spectrum) produced characteristics of the complexes, including their stability (off-rate constant k_{off}) and the energy barrier position (x_{β})^{20,21}.

Figure 2 shows a DFS spectrum for A β 40 peptide obtained from one experiment. The experiments were repeated three times, producing similar results. The k_{off} and x_{β} values determined from each separate experiment are shown in Supplementary Table S1. The mean values were calculated from this dataset (Supplementary Table S1) and equation (4) (see Methods) was used to extract the barrier height (ΔG), along with x_{β} values, to generate the energy profile for the A β 40 dissociation process. The profile is shown in the inset of Fig. 2.

Similar DFS spectra were obtained for [VPV]A β 40 (Supplementary Fig. S2a), A β 42 (Supplementary Fig. S2b), [VPV]A β 42 (Supplementary Fig. S2c), and [pP]A β 42 (Supplementary Fig. S2d) and the representative DFS plots for each mutant are shown in Supplementary Fig. S2. Each DFS was approximated by one linear plot, suggesting that the dissociation energy profiles have one barrier. The energy profiles were reconstructed from the mean values for x_{β} and k_{off} obtained from independent experiments for each peptide, shown in Supplementary Table S1. The corresponding energy profiles for each peptide are shown in the insets of Supplementary Fig. S2. The barrier heights were 27.3, 28.8, 27.7, 29.0 and 27.3 $k_B T$ for A β 40, [VPV]A β 40, A β 42, [VPV]A β 42 and [pP]A β 42, respectively.

The stabilities of the dimers formed by the peptides were characterized by their lifetimes, (Supplementary Table S1). These data show that the initial stability of [VPV]A β 42 dimers was more than three times higher than the wild type dimer (0.18 ± 0.01 s and 0.63 ± 0.12 s for A β 42 and [VPV]A β 42, respectively). A similar stabilization effect was observed for the A β 40 peptide. The VPV mutation increased the lifetime from 0.11 ± 0.03 s for A β 40 to 0.53 ± 0.03 s for [VPV]A β 40. The pP mutation decreased the stability of [pP]A β 42 dimers when compared to WT A β 42 dimers (0.18 ± 0.01 s for A β 42 vs. 0.11 ± 0.01 s for [pP]A β 42). Overall, the stability of all five dimers decreased in the following order: [VPV]A β 42 (0.63 s) > [VPV]A β 40 (0.53 s) > A β 42 (0.18 s) > [pP]A β 42 (0.11 s) = A β 40 (0.11 s). The data obtained for WT A β 40 were similar to the recent data obtained at higher ionic strength (150 mM vs 10 mM)¹², suggesting that changes in ionic strength in this broad range do not have pronounced effects on A β 40 interactions.

Characterization of misfolded states of A β dimers. We used the contour length measurements to identify segments of the peptides involved in dimer formation^{12–14}. The measured contour length is comprised of the length of the flexible tethers used for peptide immobilization and the length of the segment of the peptide from the anchoring point to the rupture position. The latter, according to the schematic in Fig. 1, was obtained by subtracting the lengths of all linkers from the measured contour lengths.

Figure 3a shows the contour length distribution for A β 40 peptide obtained from 170 rupture events at loading rates of 5000–7000 pN/s. The distribution was broad but not smooth. The data primarily clustered around 33 nm, indicating that complexes dissociating from this pathway were the predominant species of A β 40 dimers. The interactions via central and middle parts of the peptide exist, but their contribution is minor. The distribution of rupture forces from this dataset is shown in Fig. 3b. This histogram was rather narrow, with a most probable rupture force of 63.4 ± 3.2 pN, suggesting that the interaction strengths for dimers with different contour lengths are rather close, although minor events with forces beyond the main distribution also exist. The combined plot of rupture force versus contour length is shown in Fig. 3c. The data primarily aligned along a horizontal line at ~ 60 pN, with a limited number of rupture events with forces above this line appearing.

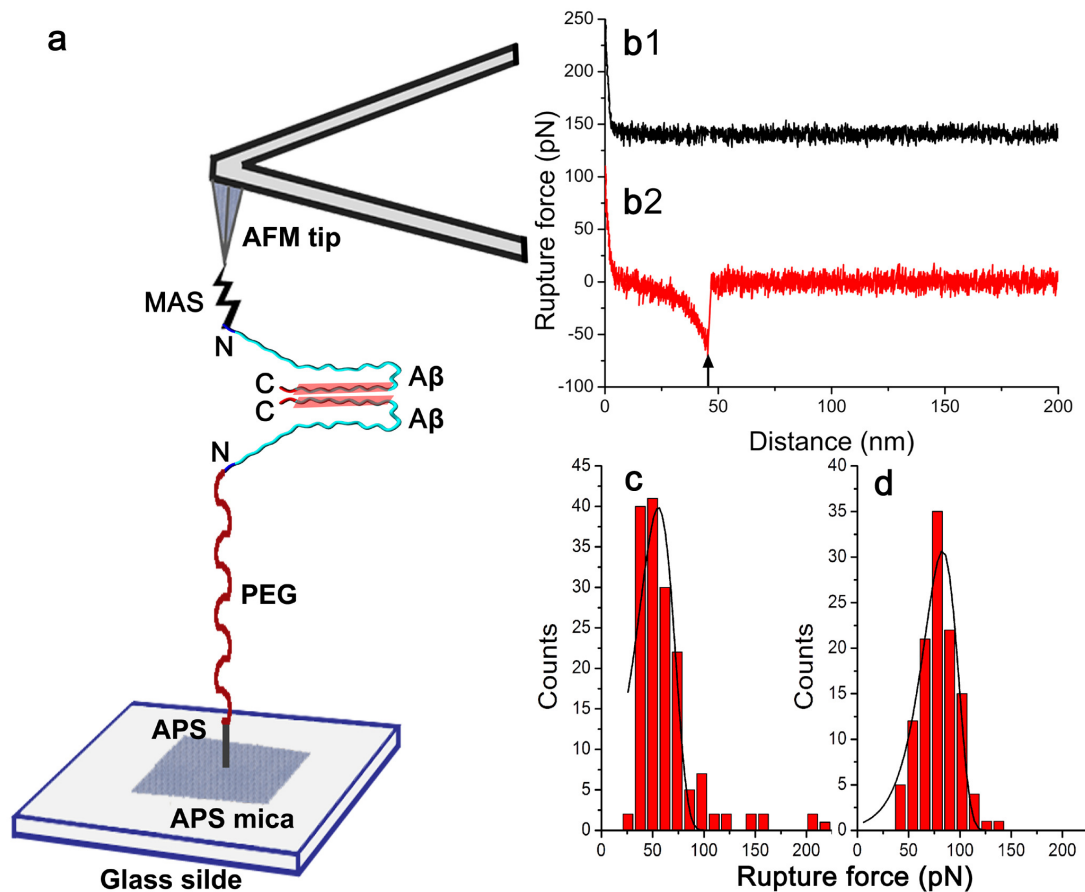


Figure 1 | Schematic of the experimental setup. (a) One A β molecule immobilized on the mica substrate is picked up by another A β molecule functionalized on the AFM tip. Putative interaction segments within a dimer are highlighted in pink bands. A rupture event occurs when a misfolded dimer is torn apart. For clarity, only one linker molecule is drawn on AFM tip and substrate, and the sizes of objects in the scheme are not to scale. N and C denote N-terminus and C-terminus, respectively. (b) Typical force–distance curve that shows either no interactions (black) or specific interactions (red). In the case of specific interactions, breaking a misfolded A β dimer resulted in a rupture distance at ~ 48 nm and a rupture force at ~ 73 pN. A vertical black arrow indicates the rupture position. For clarity, only the retraction curves are shown. Y offset was performed with the red line but not with the black line. A typical histogram of rupture force distribution at a loading rate of 6215 pN/s for A β 40 (c). The most probable rupture force for A β 40 was 57.0 pN. A typical histogram of rupture force distribution at a loading rate of 6068 pN/s for [VPV]A β 40 (d). The most probable rupture force for [VPV]A β 40 was 83.6 pN. The solid lines represent the fit of probability density function.

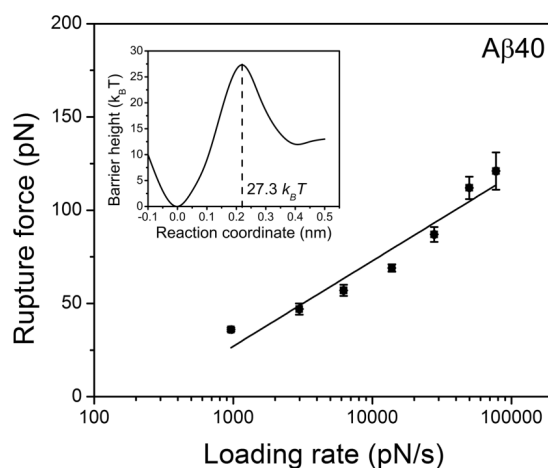


Figure 2 | Representative dynamic force spectra of A β 40. The solid red line represents the results fit with the Bell–Evans model. The obtained energy profile parameters are summarized in Table S1. The inset shows the reconstructed energy landscapes of misfolded A β dimers. Error bars represent \pm SEM.

A similar analysis performed for [VPV]A β 40 is shown in Fig. 4. The contour length distribution of [VPV]A β 40 (Fig. 4a) is broad, but entirely different from the one for the wild type A β 40 (Fig. 3a). The distribution for [VPV]A β 40 dimers has three clusters at ~ 33 nm, ~ 42 nm and ~ 57 nm that correspond to the interactions of distant C-terminus region of the peptide. Thus the major effect of VPV mutation is that the major data cluster at ~ 33 nm of A β 40 dimers became the minor one of [VPV]A β 40 dimers, whereas the other two data clusters of [VPV]A β 40 dimers increased in size with approximately the same rate. The most probable rupture force of [VPV]A β 40 dimers was 25% higher than that of A β 40 dimers (79.3 vs 63.4 pN; Fig. 4b). Similar to A β 40 dimers, the distribution of rupture forces of [VPV]A β 40 dimers showed concentrated data points at low force levels (Fig. 4c), but had some doubled forces at contour lengths of ~ 32 nm (see arrow).

A similar contour length analysis was performed for A β 42 and two mutants. The contour length distribution for A β 42 is broad with the major contributions to the dimer stability from large contour length values ~ 56 nm (Supplementary Fig. S3a). VPV mutation shifts the distribution to the central region forming a major data cluster at ~ 42 nm with a substantial drop of distant rupture events (Supplementary Fig. S4a). The contour length distribution of the rupture of [pP]A β 42 is entirely different from the above described distributions (cf. Supplementary Fig. S5a with Supplementary Figs.

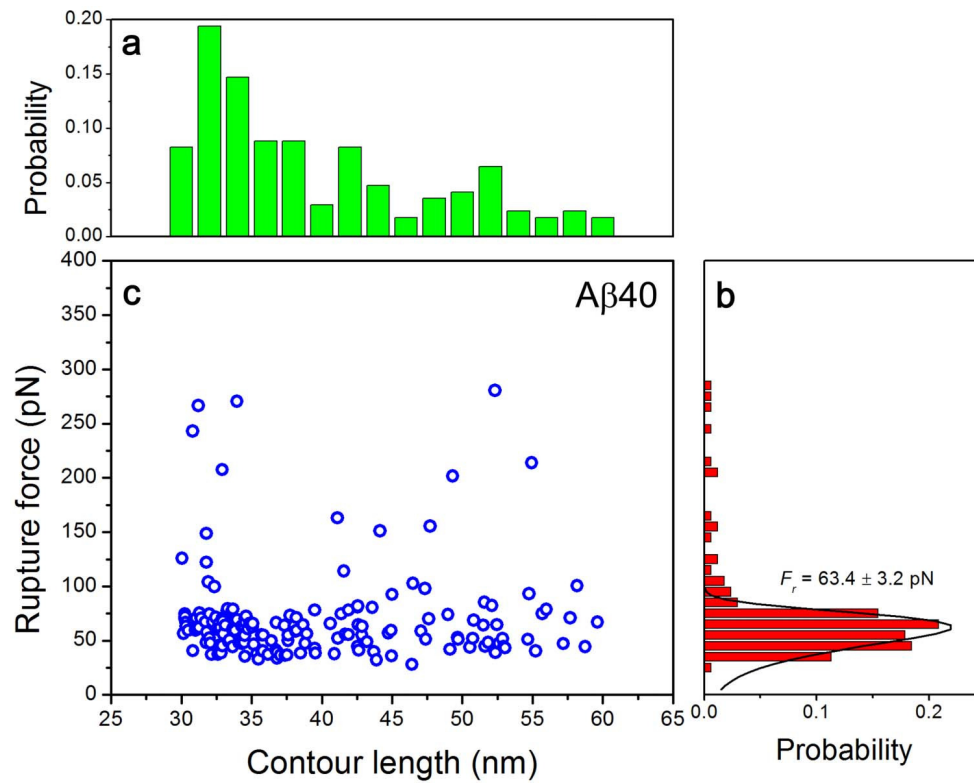


Figure 3 | The distributions of contour length (a) and rupture force (b) at loading rates of 5000–7000 pN/s for Aβ40. The contour length distribution showed a major data cluster at ~33 nm. The most probable rupture force was 63.4 ± 3.2 pN. The scatter distribution of rupture forces with varying contour lengths is shown in (c). The total number of data was 170. Error is shown by \pm SEM.

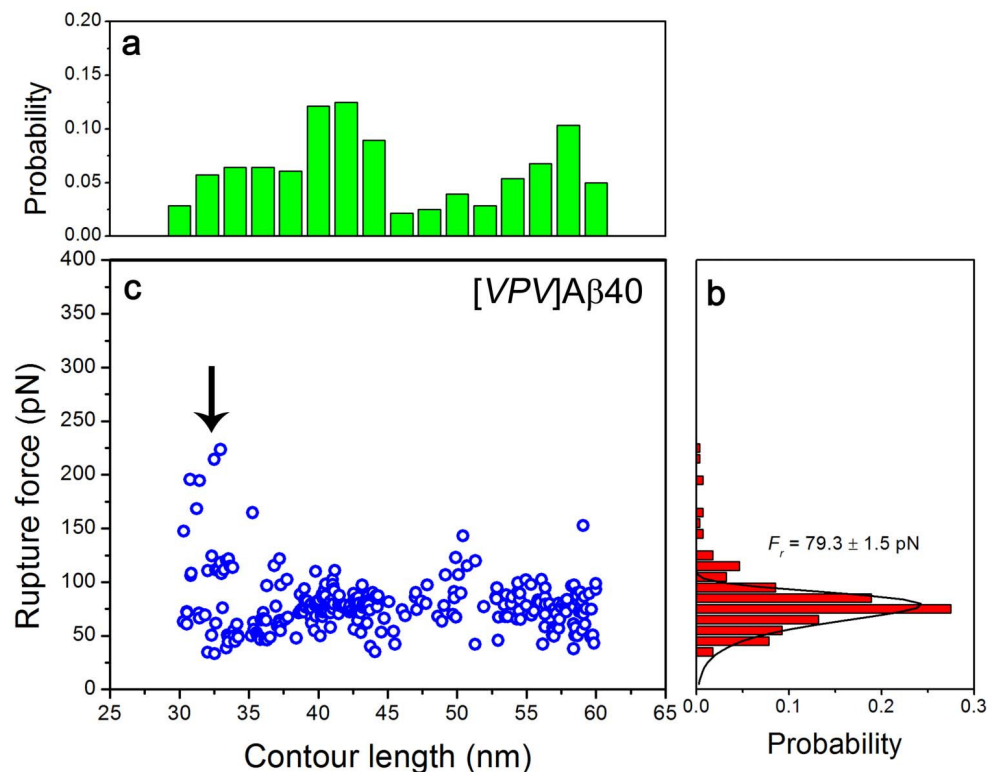


Figure 4 | The distributions of contour lengths (a) and rupture forces (b) at loading rates of 5000–7000 pN/s for [VPV]Aβ40. The contour length distribution showed a major data cluster at ~42 and a comparable cluster at ~57 nm. The most probable rupture force was 79.3 ± 1.5 pN. The scatter distribution of rupture forces with varying contour lengths is shown in (c). The total number of data was 280. Error is shown by \pm SEM.



S3a and S4a). It has a maximum corresponding to small contour lengths values (~ 33 nm) and the entire distribution is very similar to the one for A β 40 (Fig. 3a). Overall, the contour length distributions for A β 40 and A β 42 and their mutants are broad suggesting multiple populations of dimer structures. This finding is consistent with published MD simulation results^{22–26}. Compared to the variability of contour lengths profiles for A β 42 and the two mutants, the rupture force values are relatively similar. The most probable rupture force was 66.2 ± 0.8 pN for A β 42 dimers (Supplementary Fig. S3b), 67.9 ± 1.0 pN for [VPV]A β 42 dimers (Supplementary Fig. S4b), 68.0 ± 1.4 pN for [pP]A β 42 dimers (Supplementary Fig. S5b), close to that of A β 42.

Discussion

The strong effect of the A β C-terminus on self-assembly is an intriguing property. The extra two amino acids of A β 42 peptide are responsible for elevated neurotoxicity and a higher aggregation propensity than the A β 40 peptide²⁷. Here, we applied SMFS approach to investigate the interactions of inter-A β peptides. The SMFS analysis of A β peptides revealed that mutations of the very C-terminal regions caused significant differences in the propensity of the peptides to form transient dimeric species. The mutations: (1) change the stability of the dimeric species, and (2) alter the interaction pattern of the monomers within the dimers, leading to interactions a significant distance away from their C-terminal location.

The rupture force is a straightforward characteristic of interpeptide interactions, but this parameter is not very sensitive to C-terminal mutations of A β peptides. The DFS approach is a more sensitive technique than standard approaches and provides the ability to quantitatively characterize the effect of mutations. Our DFS results reveal that both [VPV]A β 40 and [VPV]A β 42 dimers are more stable than their WT counterparts (Fig. 2, Supplementary Fig. S2 and Table S1). These mutations increased the lifetimes of corresponding dimers by three fold: 0.53 s vs. 0.11 s for [VPV]A β 40 vs. A β 40 and 0.63 s vs. 0.18 s for [VPV]A β 42 vs. A β 42 dimers. By contrast, the [pP]A β 42 dimers (0.11 s lifetime) are less stable than the WT A β 42 dimers (0.18 s lifetime), which is also longer than the lifetime of A β 40 dimers (0.11 s). Additionally, the energy profile of [pP]A β 42 is analogous to that of A β 40 dimers, as shown by the same lifetime (0.11 s for both [pP]A β 42 and A β 40).

According to previous simulation results¹⁰, the stabilization effect of VPV substitutions can be attributed to the constrained flexibility of the peptide backbone and the strengthened hydrophobic interactions at the C-terminus. The finding that A β 42 dimers are more stable than A β 40 dimers is consistent with the fact that A β 42 is more prone to aggregation than A β 40. The most probable rupture force of [VPV]A β 40 is significantly higher than that of A β 40 (Figs. 3 and 4). Combined with the longer lifetime of [VPV]A β 40 dimers, these data indicate that VPV substitutions strengthen the interactions between A β 40 peptides. However, this correlation is not true of A β 42 peptides. The most probable rupture forces of WT A β 42 and its mutants are similar (Supplementary Figs. S3–S5), but their lifetimes are different. These findings suggest that dimer characteristics other than rupture forces contribute to the stabilization effect of VPV mutations. The contour length measurements, as described below, were instrumental in unraveling these interaction patterns.

Probing interactions of A β peptides within a dimer by SMFS provides a pattern of peptide interactions within the dimers. According to the immobilization procedure, the peptide is covalently bound at its N-terminal cysteine. The experimentally measured contour length consists of two major components, the lengths of the flexible tethers used for peptide immobilization and the length of the peptide segment between the N-terminus and the segment of the peptide involved in interpeptide interactions. Therefore, the position of the peak on the force–distance curve is defined by the peptide length between the N-terminus and the interacting segment. If A β

peptide adopts various conformations in these transient states, the positions of interacting segments, and hence the length of the non-interacting segments, will vary. The closer the interacting segment is to the N-terminus, the shorter the contour length will be when measured by AFM. The variable contour length measurements for all A β peptides in this study suggest that the monomers in the dimer adopt various conformations. The contour length of PEG 3400 was estimated to be 22.0 ± 0.9 nm²⁸. Lengths of both aminopropyl silatrane (APS) and MAS were ~ 2.5 nm¹³. Therefore, the total length of all linkers was 24.5 ± 0.9 nm. In addition, the distance between N-termini of two peptides was estimated to be 1.0 nm.

Figure 3 shows the contour length distribution for A β 40 peptide. After subtracting the contour lengths of the tethers, the lengths of the stretchable peptide segments are obtained. The length distribution of the A β 40 N-terminal segments is shown in Fig. 5a. This value was converted into the length in number of residues and is shown as the upper X axis on the graph. Our conversion values of 0.40 for A β 42 and 0.41 for A β 40 are close to the 0.38 nm value previously reported^{29,30}. On this distribution, the predominant peak 1 corresponds to the segment of 1–10 residues, peak 2 corresponds to the longer segment of ~ 20 residues, and the third peak comprises almost 35 residues of the peptide from the N-terminus. This distribution suggests that A β 40 can adopt at least three primary conformations within the dimers, which differ by the overall size of the peptide segments involved in the interpeptide interactions. The predominant conformation defined by peak 1 corresponds to the peptide conformation of a relatively short N-terminal segment. The rest of the peptide is involved with dimer formation. However, the interaction pattern changes dramatically if the VPV mutation is introduced into A β 40 (Fig. 5b). Peak 1 exists, but it is not the predominant one, and peak 2 becomes larger. Additionally, there is a substantial increase in peak 3 that also moves to the very C-terminus of the peptide. Notably, this mutation leads to an increase in the rupture forces and the lifetime of the dimer. Combined with the change of the interaction pattern, these data suggest that the N-terminal sequence of the A β 40 peptide dramatically alters the interaction pattern of the peptide in dimers and could modulate the toxicity of oligomers formed by the peptide.

A similar analysis was applied to the A β 42 peptide and its mutants (Figs. 5c, 5d and 5e). First, we compared the contour length profiles of A β 42 and A β 40 peptides (Supplementary Figs. 5a and 5c). The extra two amino acids present in A β 42 dramatically alter its interaction pattern compared to A β 40. The primary interactions that occur via the N-termini for A β 40 are replaced in A β 42 by interactions within the C-terminus (Peak 3, Fig. 5c). This is consistent with a recent paper by Gu et al.³¹, in which A β 42 oligomers were found to be tightly packed at C-terminal region. Additional C-terminal modifications with triple substitutions (VPV mutations; Fig. 5d) result in further alterations in A β 42 interaction. The most dramatic change observed in this mutant was the increase in interactions within the central part of the protein (observed by the growth of central peak 2) combined with decreased interactions of peptides via the C-termini (observed by the drop of peak 3). As mentioned above, the difference in dimer stability between A β 42 and [VPV]A β 42 as detected by the SMFS analysis was not due to the rupture forces because they were essentially similar. The contour length analysis explained the difference in the stabilities of A β 42 and [VPV]A β 42 dimers. The data in Figs. 5c and 5d suggest that a conformational change in the peptides occurs. As a result, the interpeptide interactions within the C-terminal region of A β 42 (peak 3) are supplanted by interactions within the central segments of [VPV]A β 42 (peak 2).

The A β 42 double substitution of residues 36 and 37 with proline (pP mutations) results in very dramatic changes in the interpeptide interaction profile (*cf.* Figs. 5c and 5e). Overall, the interpeptide interaction is shifted to the N-terminus, as evidenced by the growth

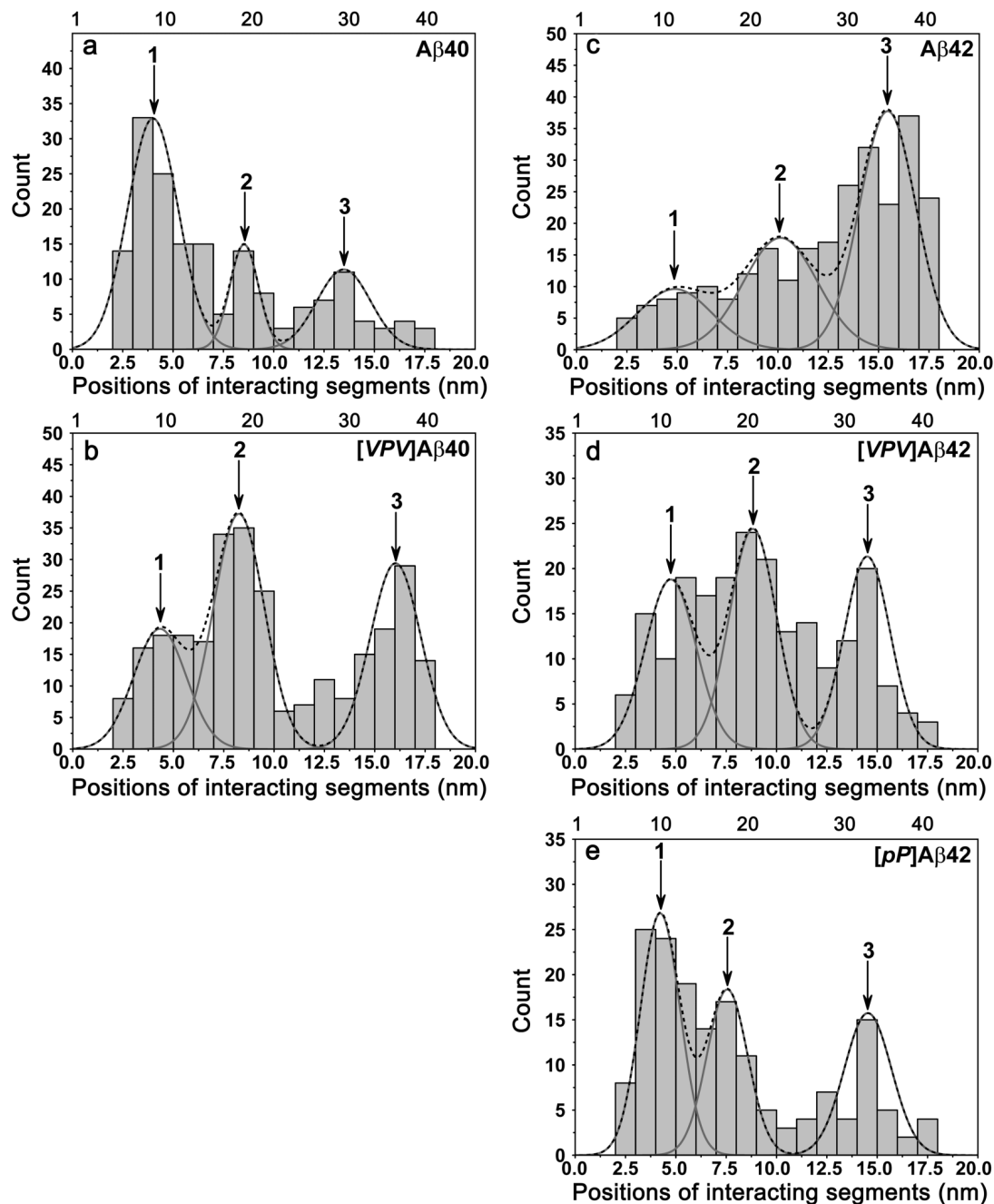


Figure 5 | The contour length distributions for A β 40, [VPV]A β 40, A β 42, [VPV]A β 42, and [pP]A β 42 dimers are shown in (a), (b), (c), (d) and (e), respectively. The interaction segments are aligned according to the contour length distribution. The dotted lines denote the distribution profiles and peaks are approximated according to the profiles. Notably, there are some uncertainties in length estimation. The arrows indicate interaction segments.

of peak 1. Surprisingly, the peptide interaction profile becomes similar to the profile for A β 40 peptide (Fig. 5a). Notably, according to the DFS study (Supplementary Table S1), the stability of [pP]A β 42 dimers as defined by their lifetime becomes as low as the lifetime for A β 40 (0.11 s). Therefore, we hypothesize that the pP substitution causes [pP]A β 42 monomer structures within the dimers to become similar to those for A β 40 peptides.

It is important to compare our data with the aggregation studies of similar A β peptide samples. The studies performed by Roychaudhuri R et al¹⁰ showed that the aggregation properties of A β 40 and [pP]A β 42 are similar. Our analysis suggests that the similarity in aggregation is due to almost identical interaction profiles for both peptides. Similarly, mutating A β 40 by VPV substitutions alters its aggregation propensity, causing it to be A β 42-like. This effect is due

to increased interactions between the central and C-terminal segments of [VPV]A β 40 peptide dimers, compared to the predominate contribution of N-terminal interactions in A β 40 dimers. This change in the interaction pattern also could explain the elevated neurotoxicity of [VPV]A β 40 compared with the neurotoxicity of the WT peptide described in¹⁰. The model that the modulation of N-terminal segments of A β regulates toxicity is supported by the low toxicity of [pP]A β 42 peptide aggregates¹⁰. V36p-G37P replacement makes [pP]A β 42 interaction similar to A β 40 interaction, with predominant interactions via N-terminal segments (cf. Figs. 5a and 5e). These data lead to a model of A β interaction suggesting that the N-terminal segments destabilize the dimer and thus decrease the aggregation propensity of the peptide and the cytotoxicity of the oligomers. *In silico* studies¹⁰ have suggested that the formation of



the Val36–Gly37 turn is a key feature controlling A β 42 aggregation and elevated neurotoxicity. Furthermore, VPV substitutions stabilize this structure. Our data show that stabilization of the turn is not a local structural change. Rather, interactions with other segments of the peptide are involved with both the stabilization of dimers and possibly higher order oligomers.

Our model on the effect of the N–terminal on the aggregation propensity of A β peptides is in line with the results of a recent paper³² in which the interaction of A β 40 and A β 42 monomers in solution was studied. They found that A β 40 monomer forms a type of anti-parallel β –hairpin involving the central hydrophobic cluster (residues 16–21) with the N–terminal residues 9–13, while A β 42 monomer forms a major antiparallel β –hairpin involving the central hydrophobic cluster with the C–terminus. The contribution of N–terminal residues to the peptide interaction also reported in paper by Maji et al.³³, in which effects of Tyr substitutions on the aggregation were studied. They report that Tyr substitution at Asp1 for both A β 40 and A β 42 leads to the slowest aggregation kinetics as opposed to Tyr substitutions at any other segments of the peptides.

A number of novel features of the interaction and self–assembly process of A β 40 and A β 42 peptides were determined using single–molecule force spectroscopy. The dynamic force spectroscopy analysis revealed stronger interpeptide interactions for A β 42 than for A β 40. The change in the peptide sequence at the C–terminus can increase or decrease dimer stability. VPV substitutions stabilize the Val36–Gly37 turn in A β 42, whereas a *pP* dipeptide substitution destabilizes the dimer. A complementary single–molecule force spectroscopy approach, in which contour lengths were analyzed, revealed an unexpected long–range interaction effect in peptide dimerization. The N–terminal region of A β 40 peptide cause its low interpeptide interaction, but the extra two residues of A β 42 eliminate this effect. However, *pP* substitution in A β 42 restores the effect of the N–terminus, therefore the stability of [*pP*]A β 42 and A β 40 become equal. The significance of this new finding on long–distance interactions within A β is two–fold. First, it demonstrates that A β peptide interaction patterns are complex and the seemingly neutral N–terminal region plays an important role in interpeptide interactions. Second, the involvement of the N–terminal region of A β in the interaction process suggests that this region may be a potential target for intervening into the A β misfolding process. These are novel avenues in the amyloid misfolding and aggregation area.

Methods

All cysteinyl–A β peptides were synthesized using 9–fluorenylmethoxycarbonyl (Fmoc) chemistry and purified by reverse phase high performance liquid chromatography (RP–HPLC). The identity and purity (usually > 97%) of the peptides were confirmed by amino acid analysis followed by mass spectrometry and RP–HPLC. Functionalization of AFM tips and mica substrates were prepared according to previously published procedures^{11,12,14}. Briefly, 20 nM A β monomers were immobilized on mica functionalized with aminopropyl silatrane (APS) via PEG 3400 tethers and on AFM tips through maleimide silatrane (MAS). For DFS study, the tip retraction speeds ranged from 100 nm/s to 4000 nm/s. The dwell time was set at 0.3 s when retraction speed reached 1000 nm/s and above. Force curves with rupture event were subject to WLC analysis^{12,14}, as shown below.

$$F(x) = \frac{k_B T}{L_p} \left[\frac{1}{4} \left(1 - \frac{x}{L_c} \right)^{-2} - \frac{1}{4} + \frac{x}{L_c} \right] \quad (1)$$

Rupture force distributions were fitted with a probability density function:

$$p(F) = k_{off} \exp\left(\frac{Fx_\beta}{k_B T}\right) \frac{1}{r} \exp\left(-k_{off} \int_0^F \exp\left(\frac{Fx_\beta}{k_B T}\right) \frac{1}{r} df\right) \quad (2)$$

Dynamic force spectra were fitted with Bell–Evans model²¹:

$$F = \frac{k_B T}{x_\beta} \ln\left(\frac{r \cdot x_\beta}{k_{off} k_B T}\right) \quad (3)$$

The energy profiles were reconstructed according to previous papers^{20,34}, using the equation below.

$$\Delta G^\ddagger = \ln\left(\frac{k_B T}{k_{off} \cdot h}\right) k_B T \quad (4)$$

Positions of interacting segments of A β monomers within dimers were estimated by subtracting from the measured contour lengths the lengths of all linkers and the distance between N–termini of monomers (see Fig. 1a). The contour length of PEG was estimated with the following equation²⁸:

$$L_c(F) = N_s \cdot \left(\frac{L_{planar}}{e^{\Delta G(F)/k_B T} + 1} + \frac{L_{helical}}{e^{-\Delta G(F)/k_B T} + 1} \right) \quad (5)$$

Where $L_c(F)$ is the contour length, N_s is the average number of monomers, L_{planar} is the length of monomers with planar conformation, $L_{helical}$ is the length of monomers with helical conformation, $\Delta G(F)$ is the free energy difference at zero applying force. The N_s is 77 ± 10 for 3400 Da PEG. The L_{planar} and $L_{helical}$ are 3.58 Å and 2.8 Å, respectively. The $\Delta G(F)$ is fixed at $3 k_B T$. The contour length of PEG was thus estimated to be 22.0 ± 0.9 nm. All error bars represent SEM. Detailed methods can be found in Supplementary Information.

- Chiti, F. & Dobson, C. M. Protein misfolding, functional amyloid, and human disease. *Annu Rev Biochem* **75**, 333–366 (2006).
- Roychaudhuri, R., Yang, M., Hoshi, M. M. & Teplow, D. B. Amyloid beta-protein assembly and Alzheimer disease. *J Biol Chem* **284**, 4749–4753 (2009).
- Benilova, I., Karran, E. & De Strooper, B. The toxic Abeta oligomer and Alzheimer's disease: an emperor in need of clothes. *Nat Neurosci* **15**, 349–357 (2012).
- De Strooper, B., Vassar, R. & Golde, T. The secretases: enzymes with therapeutic potential in Alzheimer disease. *Nat Rev Neural* **6**, 99–107 (2010).
- Karran, E., Mercken, M. & De Strooper, B. The amyloid cascade hypothesis for Alzheimer's disease: an appraisal for the development of therapeutics. *Nat Rev Drug Discov* **10**, 698–712 (2011).
- Pauwels, K. et al. Structural basis for increased toxicity of pathological abeta42:abeta40 ratios in Alzheimer disease. *J Biol Chem* **287**, 5650–5660 (2012).
- Bitan, G. et al. Amyloid beta-protein (Abeta) assembly: Abeta 40 and Abeta 42 oligomerize through distinct pathways. *Proc Natl Acad Sci U S A* **100**, 330–335 (2003).
- Bernstein, S. L. et al. Amyloid-beta protein oligomerization and the importance of tetramers and dodecamers in the aetiology of Alzheimer's disease. *Nat Chem* **1**, 326–331 (2009).
- Yang, M. & Teplow, D. B. Amyloid beta-protein monomer folding: free-energy surfaces reveal alloform-specific differences. *J Mol Biol* **384**, 450–464 (2008).
- Roychaudhuri, R. et al. C-terminal turn stability determines assembly differences between Abeta40 and Abeta42. *J Mol Biol* **425**, 292–308 (2013).
- Yu, J., Warnke, J. & Lyubchenko, Y. L. Nanoprobeing of alpha-synuclein misfolding and aggregation with atomic force microscopy. *Nanomedicine* **7**, 146–152 (2011).
- Kim, B. H. et al. Single-molecule atomic force microscopy force spectroscopy study of Abeta-40 interactions. *Biochemistry* **50**, 5154–5162 (2011).
- Krasnoslobodtsev, A. V. et al. Effect of spermidine on misfolding and interactions of alpha-synuclein. *PLoS One* **7**, e38099 (2012).
- Lv, Z., Condrum, M. M., Teplow, D. B. & Lyubchenko, Y. L. Nanoprobeing of the Effect of Cu(2+) Cations on Misfolding, Interaction and Aggregation of Amyloid beta Peptide. *J Neuroimmune Pharmacol* **8**, 262–273 (2013).
- Portillo, A. M., Krasnoslobodtsev, A. V. & Lyubchenko, Y. L. Effect of electrostatics on aggregation of prion protein Sup35 peptide. *J Phys Condens Matter* **24**, 164205 (2012).
- Lyubchenko, Y. L., Kim, B. H., Krasnoslobodtsev, A. V. & Yu, J. Nanoimaging for protein misfolding diseases. *Wiley Interdiscip Rev Nanomed Nanobiotechnol* **2**, 526–543 (2010).
- Lovas, S., Zhang, Y., Yu, J. & Lyubchenko, Y. L. Molecular mechanism of misfolding and aggregation of abeta(13–23). *J Phys Chem B* **117**, 6175–6186 (2013).
- Petkova, A. T. et al. A structural model for Alzheimer's beta-amyloid fibrils based on experimental constraints from solid state NMR. *Proc Natl Acad Sci U S A* **99**, 16742–16747 (2002).
- Luhns, T. et al. 3D structure of Alzheimer's amyloid-beta(1–42) fibrils. *Proc Natl Acad Sci U S A* **102**, 17342–17347 (2005).
- Krasnoslobodtsev, A. V., Shlyakhtenko, L. S. & Lyubchenko, Y. L. Probing Interactions within the synaptic DNA-Sfil complex by AFM force spectroscopy. *J Mol Biol* **365**, 1407–1416 (2007).
- Merkel, R., Nassoy, P., Leung, A., Ritchie, K. & Evans, E. Energy landscapes of receptor–ligand bonds explored with dynamic force spectroscopy. *Nature* **397**, 50–53 (1999).
- Mitternacht, S., Staneva, I., Hard, T. & Irbach, A. Monte Carlo study of the formation and conformational properties of dimers of Abeta42 variants. *J Mol Biol* **410**, 357–367 (2011).
- Zhu, X., Bora, R. P., Barman, A., Singh, R. & Prabhakar, R. Dimerization of the full-length Alzheimer amyloid beta-peptide (Abeta42) in explicit



- aqueous solution: a molecular dynamics study. *J Phys Chem B* **116**, 4405–4416 (2012).
24. Barz, B. & Urbanc, B. Dimer formation enhances structural differences between amyloid beta-protein (1–40) and (1–42): an explicit-solvent molecular dynamics study. *PLoS One* **7**, e34345 (2012).
 25. Melquiond, A., Dong, X., Mousseau, N. & Derreumaux, P. Role of the region 23–28 in Abeta fibril formation: insights from simulations of the monomers and dimers of Alzheimer's peptides Abeta40 and Abeta42. *Curr Alzheimer Res* **5**, 244–250 (2008).
 26. Cote, S., Laghaei, R., Derreumaux, P. & Mousseau, N. Distinct dimerization for various alloforms of the amyloid-beta protein: Abeta(1–40), Abeta(1–42), and Abeta(1–40)(D23N). *J Phys Chem B* **116**, 4043–4055 (2012).
 27. Walsh, D. M. & Teplow, D. B. Alzheimer's disease and the amyloid beta-protein. *Prog Mol Biol Transl Sci* **107**, 101–124 (2012).
 28. Oesterhelt, F., Rief, M. & Gaub, H. E. Single molecule force spectroscopy by AFM indicates helical structure of poly(ethylene-glycol) in water. *New Journal of Physics* **1**, 6 (1999).
 29. Sarkar, A., Caamano, S. & Fernandez, J. M. The elasticity of individual titin PEVK exons measured by single molecule atomic force microscopy. *J Biol Chem* **280**, 6261–6264 (2005).
 30. Zhao, J. M. *et al.* Single-molecule detection of structural changes during Per-Arnt-Sim (PAS) domain activation. *Proc Natl Acad Sci U S A* **103**, 11561–11566 (2006).
 31. Gu, L., Liu, C. & Guo, Z. Structural insights into Abeta42 oligomers using site-directed spin labeling. *J Biol Chem* **288**, 18673–18683 (2013).
 32. Ball, K. A., Phillips, A. H., Wemmer, D. E. & Head-Gordon, T. Differences in beta-strand Populations of Monomeric Abeta40 and Abeta42. *Biophys J* **104**, 2714–2724 (2013).
 33. Maji, S. K. *et al.* Amino acid position-specific contributions to amyloid beta-protein oligomerization. *J Biol Chem* **284**, 23580–23591 (2009).
 34. Tinoco Jr, I. & Bustamante, C. The effect of force on thermodynamics and kinetics of single molecule reactions. *Biophys Chem* **101–102**, 513–533 (2002).

Acknowledgements

The work was supported by grants to Y.L.L. from the National Institutes of Health (NIH: 5R01 GM096039-02) and National Science Foundation (EPS-1004094) and grants to D.B.T. from NIH (NS038328, AG041295) and the Jim Easton Consortium for Drug Development and Biomarkers.

Author contributions

Z.L., D.B.T. and Y.L.L. designed the experiments. R.R. and M.M.C. prepared initial samples. Z.L. performed the experiments and analyzed the data. Z.L., D.B.T. and Y.L.L. discussed the results and wrote the manuscript. All authors reviewed the manuscript.

Additional information

Supplementary information accompanies this paper at <http://www.nature.com/scientificreports>

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Lv, Z., Roychaudhuri, R., Condrón, M.M., Teplow, D.B. & Lyubchenko, Y.L. Mechanism of amyloid β -protein dimerization determined using single-molecule AFM force spectroscopy. *Sci. Rep.* **3**, 2880; DOI:10.1038/srep02880 (2013).



This work is licensed under a Creative Commons Attribution 3.0 Unported license. To view a copy of this license, visit <http://creativecommons.org/licenses/by/3.0>