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The Chromosomes of three species of the *Nasonia* complex (Hymenoptera, Pteromalidae)

With 1 figure and 2 tables

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Summary

The karyotypes of three sibling species, *Nasonia vitripennis* (WALKER, 1836), *N. longicornis* DARLING, 1990 and *N. giraulti* DARLING, 1990 were examined, the latter two species for the first time. All species have chromosome numbers of $n=5$ and $2n=10$, their chromosomes being metacentric. The distribution of constitutive heterochromatin also appears to be similar in the genus *Nasonia* ASHMEAD, 1904. However, statistical karyotypic differences between studied species were revealed using chromosome morphometrics. These data are consistent with the order of divergence in the *Nasonia* complex found on the basis of molecular studies.

Zusammenfassung

Die Ergebnisse der Untersuchung des Karyotyps der Schwesterarten *Nasonia vitripennis* (WALKER, 1836), *N. longicornis* DARLING, 1990 und *N. giraulti* DARLING, 1990 werden mitgeteilt. Für die beiden letztgenannten Arten liegen bisher keine karyologischen Daten vor. Die Karyotypen der drei untersuchten Arten mit $n=5$ und $2n=10$ bestehen aus metacentrischen Chromosomen. Die Verteilung des konstitutiven Chromatins scheint in der Gattung *Nasonia* ASHMEAD, 1904 ebenfalls sehr einheitlich zu sein. Demgegenüber ergab die Morphometrie der Chromosomen statistisch signifikante Unterschiede zwischen den drei Arten. Die vorliegenden Daten entsprechen der Abfolge der Artenspaltungen im *Nasonia*-Komplex, die auf der Basis molekulargenetischer Untersuchungen festgestellt wurde.

Резюме

Исследованы кариотипы трех видов-двойников, *Nasonia vitripennis* (WALKER, 1836), *N. longicornis* DARLING, 1990 и *N. giraulti*, 1990, причем два последних вида изучены впервые. У всех трех наездников $n=5$ и $2n=10$, и в их кариотипах присутствуют только метацентрические хромосомы. В пределах рода *Nasonia* ASHMEAD, 1904 распределение конститутивного гетерохроматина также является весьма сходным. Однако, с помощью морфометрических методов между хромосомными наборами рассматриваемых видов выявлены статистически достоверные отличия. Эти результаты соответствуют порядку дивергенции в комплексе *Nasonia*, определенному по данным молекулярно-генетических исследований.

Nasonia vitripennis (WALKER, 1836) is a well-known cosmopolitan parasite of cyclorrhaphous diptera (BOUČEK & RASPLUS, 1991). Various aspects of its biology have been extensively investigated all over the world (e.g. WHITING, 1967; VAN DEN ASSEM, 1976; WERREN, 1983). This species was widely used in genetic research, a limited genomic map based on visible

markers is available (SAUL, 1990). First data on its chromosome number, $n=5$, were obtained by GERSHENZON (1946) and later confirmed by other studies (PENNYPACKER, 1958; GERSHENZON, 1968; WHITING, 1968; NUR et al., 1988). The only detailed karyotypic description of *N. vitripennis* was made by REED (1993) who provided data on chromosome morphometrics as well as on C- and Ag-NOR-bandings in this species.

The genus *Nasonia* ASHMEAD, 1904 was considered to be monotypic for many years. Surprisingly enough, two new sibling species, *N. longicornis* DARLING, 1990 and *N. giraulti* DARLING, 1990 were found in this genus in North America about ten years ago (DARLING & WERREN, 1990). An extensive study of this complex revealed a number of morphological characters which could be used as distinction features for these taxa. In addition, many other characters could be used for separating these species with 0.95–0.999% certainty. Furthermore, *N. vitripennis*, *N. longicornis* and *N. giraulti* substantially differ in their courtship behaviour (VAN DEN ASSEM & WERREN, 1994).

Chromosome number and other karyotypic features of the Pteromalidae appear to be highly conservative. Specifically, eight studied species of the family have $n=5$ (GOKHMAN & QUICKE, 1995; GOKHMAN, *in press*). Except for *N. vitripennis*, neither morphometric studies of karyotypes nor differential chromosome staining in these species, however, were performed to date. Moreover, $n=6$ was found in a local population of *N. vitripennis* from California (GOODPASTURE, 1974). Even among laboratory stocks of this species normally having $n=5$, some populations carrying a particular B chromosome were revealed (NUR et al., 1988; WERREN, 1991). The present paper deals with the results of a karyological study of *N. vitripennis*, *N. longicornis* and *N. giraulti* using routine and differential (C-) chromosome staining.

Material and methods

Chromosomes of all three *Nasonia* species were studied in wasps from laboratory cultures maintained at the Institute of Evolutionary and Ecological Sciences, University of Leiden, namely: Lab II, a laboratory stock of *Nasonia vitripennis* originally collected in Leiden more than 20 years ago; strain IV7 R2 of *N. longicornis* derived from the strain IV7 (nest UT007) which had been collected in Utah, USA, in July 1988; and strain R16A of *N. giraulti* derived from a particular strain (nest VA002) which had been collected in Virginia, USA, in September 1988. Air-drying chromosome preparations were made from cerebral ganglia of male and female prepupae, according to the routine procedure described by IMAI et al. (1988). Differential chromosome staining (C-banding) was performed according to the method developed by SUMNER (1972) and modified by GOKHMAN (1997). Chromosomes were studied under microscopes MBR-15 and Zeiss NU-2. For making chromosome measurements, ten diploid metaphase plates for each species were scanned using static TV camera equipped with the image analysis program ImageExpert version 1.00. Scanned images were measured using Adobe Photoshop version 3.0.5. Statistical data analysis was performed with the help of STATISTICA version 4.3. The *t* test for independent samples was used as a statistical criterion.

Results

Numbers of studied individuals and metaphase plates for each species are presented in Table 1. Relative lengths and centromeric indices of all chromosomes of the three species are shown in Table 2. Chromosomes of *N. vitripennis*, *N. longicornis* and *N. giraulti* are very similar, especially for their centromeric indices. Due to this similarity, a unified karyotypic description is given for all species, with interspecific differences being added.

Tab. 1: Number of studied individuals of the *Nasonia* complex and their metaphase plates.

Species	No. of individuals		No of metaphase plates	
	males	females	haploid	diploid
<i>N. vitripennis</i> (WALKER, 1836)	1	9	20	15
<i>N. longicornis</i> DARLING, 1990	1	9	2	37
<i>N. giraulti</i> DARLING, 1990	1	8	10	41

Tab. 2: Relative length (RL) and centromeric index (CI) of chromosomes of the *Nasonia* complex. Values having a common superscript significantly differ: ^a at $p < 0.001$, ^{bc} at $p < 0.05$.

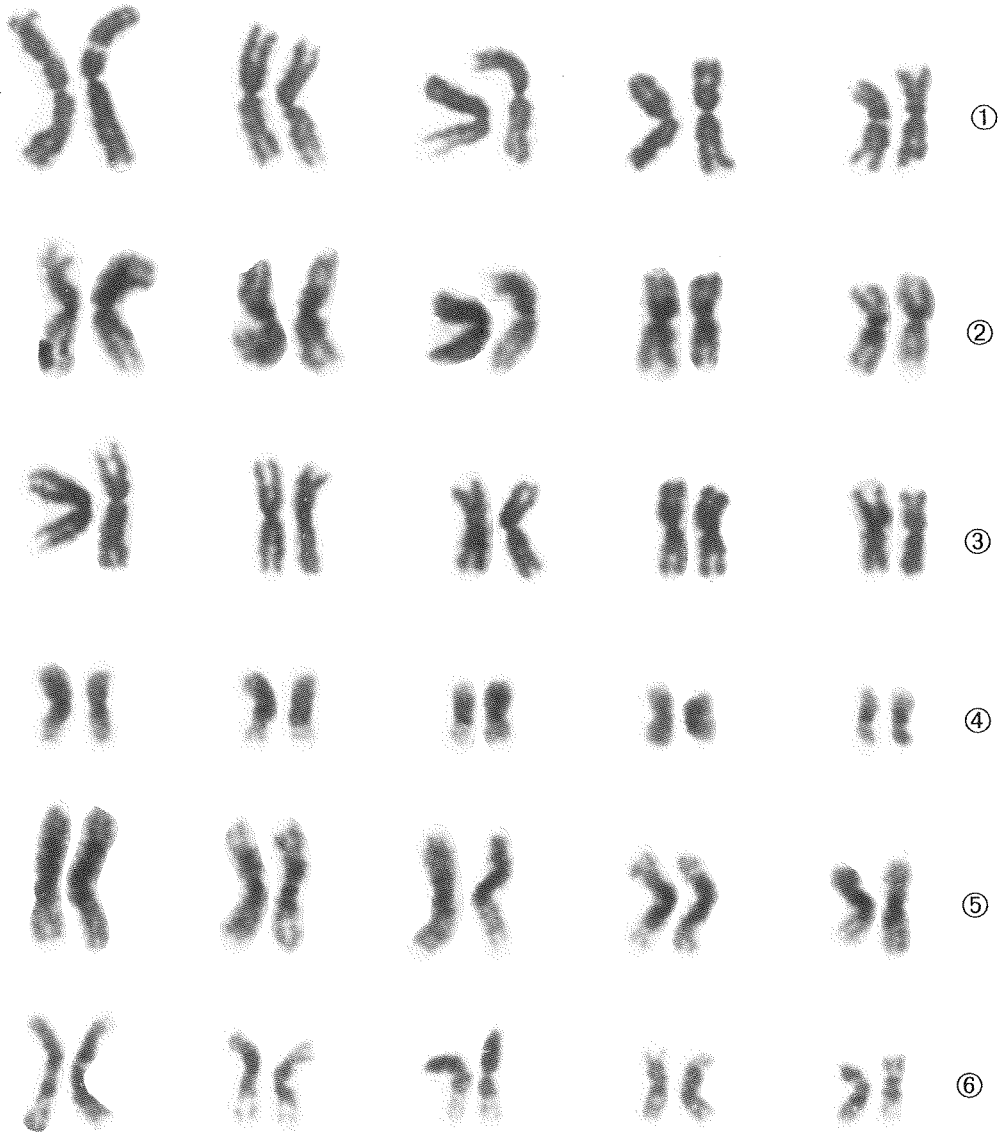
Chr. no.	Species					
	<i>N. vitripennis</i>		<i>N. longicornis</i>		<i>N. giraulti</i>	
	RL	CI	RL	CI	RL	CI
1	25.97±1.44	46.93±2.64	25.48±0.82	46.82±2.68	25.58±1.28	46.89±1.83
2	21.03±0.63 ^{ab}	47.52±2.29	21.84±0.63 ^a	46.65±2.27	21.59±0.85 ^b	46.27±2.01
3	19.59±0.74	47.37±2.68	19.80±0.70 ^c	46.66±2.94	19.34±0.72 ^c	46.92±2.15
4	17.71±0.81	46.88±2.47	17.57±0.49	45.22±3.21	17.78±0.64	46.84±2.51
5	15.70±0.67	46.97±2.20	15.32±0.97	46.20±3.46	15.71±1.18	46.07±2.50

The same chromosome numbers, $n=5$ and $2n=10$, were found in *N. vitripennis*, *N. longicornis* and *N. giraulti*, all chromosomes being obviously metacentric and gradually decreasing in size (Figs. 1-3). The first chromosome pair is slightly larger than the others. Chromosome 2 is significantly shorter in *N. vitripennis* than in *N. giraulti* or *N. longicornis*. Similarly, chromosome 3 is longer in the latter species than in *N. giraulti*.

The distribution of the constitutive heterochromatin also appeared quite similar in the three *Nasonia* species. All chromosomes have characteristic large segments of centromeric heterochromatin. Moreover, the shorter arm of the chromosome 3 is also heterochromatic, at least in most of the metaphase plates (Figs. 4-6).

Discussion

All karyotypes of the *Nasonia* complex are symmetrical and very much similar to each other. However, they differ from chromosome sets of some other Pteromalidae also having $n=5$. Unfortunately, these differences can usually be revealed only after making detailed chromosome measurements. For example, one of the two species belonging to the *Anisopteromalus calandrae* (HOWARD, 1881) species complex also has $n=5$ (GOKHMAN et al., 1998), but its haploid karyotype is more asymmetrical, with two smaller chromosomes being visibly shorter than the third one. Moreover, the smallest chromosome of the latter species is submetacentric, whereas the others are clearly metacentric.



Figs 1-6. Diploid karyotypes of *Nasonia* species. Figs 1-3. Routine staining. 1 - *N. vitripennis*, 2 - *N. longicornis*, 3 - *N. giraulti*. Figs 4-6. Differential C-banding. 4 - *N. vitripennis*, 5 - *N. longicornis*, 6 - *N. giraulti*. Bar equals 10 μ m.

Our data on relative chromosome lengths in *N. vitripennis* are very close to those calculated by REED (1993), namely: 25.10, 21.36, 19.74, 17.89 and 15.91 (transformed from absolute chromosome lengths). The latter author was also able to demonstrate the presence of pericentromeric heterochromatin in all chromosomes of this wasp species.

Evidence obtained from the study of an rDNA internal transcribed spacer (ITS2) (CAMPBELL et al., 1993) and mitochondrial 16S DNA of the *Nasonia* complex (REED & WERREN, *in litt.*) suggests that the time of divergence between *N. vitripennis* and the two remaining species is approximately 200,000 years, and that between *N. longicornis* and *N. giraulti* is about 100,000 years. Our data are consistent with the order of divergence in the *Nasonia* complex found on the basis of molecular studies. Specifically, a comparison of relative lengths of the chromosome 2 demonstrates that *N. vitripennis* is likely to stem out first from the common lineage which split later into *N. longicornis* and *N. giraulti*. The significant difference on the relative lengths of chromosome 3 between the two latter species may reflect an autapomorphous chromosomal rearrangement occurred in one of these taxa.

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