Aristotle Biomedical Journal, Vol 1, No 2 e-ISSN: 2653-9748

Brief Report Towards the identification of miRNAs targeting the translational machinery as novel cancer therapeutics

Christos Papagiannopoulos¹, Konstantinos Kyritsis¹, Ioannis Vizirianakis¹

¹ Department of Pharmacology, School of Pharmacy, Aristotle University of Thessaloniki, Thessaloniki, Greece

Abstract

Increased protein production is a prerequisite for cell proliferation, thus rendering translation and ribosome deregulation a common hallmark of cancer cell biology. A frequently observed mechanism in malignancies is the overactivation of the translational process. As a consequence, strategies that selectively target the ribosomal machinery bear significant promise as cancer therapeutic approaches. To this end, this report provides a workflow for the identification of novel miRNA molecules with the ability to target and suppress ribosomal activity in cancer cells.

Keywords: Cancer, miRNA therapeutics, Bioinformatics, Ribosome, Pharmacology

Corresponding author:

Prof. Dr. Ioannis S. Vizirianakis, Department of Pharmacology, School of Pharmacy, Aristotle University of Thessaloniki, 54124 Thessaloniki, Greece; e-mail: ivizir@pharm.auth.gr

Abbreviations:

RPs: Ribosomal Proteins

rRNAs: Ribosomal RNAs

miRNA: MicroRNA

Ribosomes are the main molecular machines responsible for protein synthesis within the cells of all domains of life. Each ribosome is composed of 80 Ribosomal Proteins (RPs) and 4 Ribosomal RNAs (rRNAs) which assemble into two subunits within the nucleolus. After their export to the cytoplasm, the 40S and 60S subunits work together to convert genetic information into protein with both speed and accuracy.

Cancer cells reside in a permanent and uncontrolled cellular proliferation state. This trait induces a pressing need for protein production, which is crucial in order to support the incremented metabolic needs of the rapidly duplicating cells (Sulima et al., 2017). As a consequence, ribosomal function is often modulated in cancer cells with alterations mainly directing towards one or both of the following pathways: 1) to increase the abundance of the ribosomal machinery (and therefore protein synthesis); and/or 2) to reduce the translational accuracy (fig. 1A) (Topisirovic and Sonenberg, 2015, Tahmasebi et al., 2018, Robichaud et al., 2018). A notable example of the first mechanism is mediated via the oncogene The MYC. latter. is frequently amplified in various human cancers, where it has been shown to promote oncogenesis by acting as a direct transcriptional inducer of RPs genes (Robichaud et al., 2018). Alike MYC, additional factors, such as ΔN and the Epithelial netrin-1 Cell Transforming 2 (ECT2) oncogene, attain aberrant functionality within tumor cells and function as strong inducers of rRNAs. thus driving ribosome biogenesis and vastly increasing protein synthesis (Delloye-Bourgeois et al., 2012, Huh et al., 2003). Interestingly though, lesions mutations. deletions such as or downregulation of specific RPs have,

also, been observed in various types of cancers and those, although not always clearly understood, are presumed to impair accuracy of translation (second mechanism, fig. 1A). Examples include specific mutations in ribosomal proteins, such as uL18/RPL5 which was found mutated in patients' samples of glioblastoma (11%), melanoma (28%) and breast cancer (34%) (Fancello et al., 2017). Similarly, acute lymphocytic patients with leukemia repeatedly bear mutations in uL18/RPL5 and uL16/RPL10 (De Keersmaecker et al., 2013), whereas uS10/RPS20 mutations seem to play a role in the pathogenesis of a hereditary type of colorectal cancer (Nieminen et al., 2014). In summary, the entirety of the changes described above impair the central process of translation, thus inducing chaotic alterations in the cancer cell proteome, and ostensibly promoting cell multiplication, metastasis and resistance to therapy (fig. 1A).

In this direction, targeting the aberrant and uncontrolled ribosomal machinery of the cancer cell may constitute an effective anti-cancer strategy (outlined in fig. 1B). To this end, we have developed a strategy and a workflow pipeline to identify miRNAs with the ability to bind to RPs, and therefore leverage the gene-suppressing nature of the interaction between miRNAs and their target genes. Our goal is to unravel novel molecules of pharmacological interest aiming to selectively suppress the impairedfrequently overactivated- translation process of the tumor cells and thereby render those cells vulnerable to cell death (fig. 1B). Notably, the versatile class of miRNAs may include very potent pharmacological candidates as evidently shown by the multiple clinical trials that investigate such therapies as well as the recent approval





A. Alterations in ribosomal function and oncogenesis. Oncogenic mutations affect ribosomal function via two main mechanisms: a) an increase in ribosome biogenesis (e.g. via the MYC

oncogene); and b) a reduction in translational fidelity (e.g. mutations in uL18/RPL5). Both mechanisms benefit cancer initiation and progression.

B. Targeting the ribosomal machinery with miRNA molecules. The suggested approach includes the identification of miRNAs that target RPs, thus suppressing ribosomal function in cancer cells and inducing cell death.

C. List of the miRNAs identified and number of RPs, out of the 80 expressed by the human genome, which are targeted by each miRNA. The results shown were generated using the online platforms miRWalk (Dweep et al., 2014) and miRNet (Fan et al., 2016).

D-I. miR-129-5p expression in patient samples from 6 cancer types. D. breast cancer, E. colon cancer, F. head & neck cancer, G. rectum cancer, H. hepatocellular cancer, I. prostate cancer. The gene expression data was retrieved from The Cancer Genome Atlas (TCGA) database. Blue boxes denote normal tissue, whereas red refers to tumor sample of the indicated cancer type.

of the first RNAi therapy. Prominent examples of such onco-suppressor miRNAs include miR-34a, let-7 and miR-16-5p, shown to inhibit the oncogene RAS (Yamakuchi et al., 2008), activate the onco-suppressor p53 (Cho, 2007), and block oncogenic signaling via VEGFA (Qu et al., 2017), respectively.

To identify our candidate miRNAs, we undertook an in-silico approach with the aim to analyze all the potential between interactions the known miRNAs (as annotated in miRBase) and the 80 RPs expressed by the The human genome. available databases that contain miRNA-mRNA interactions incorporate either predicted (usually generated by an algorithm) or validated interactions (generated during wet-lab experiments). For a thorough analysis we opted to include data from both types of datasets, using two platforms: miRNet for validated interactions (Fan et al., 2016) and miRwalk for predicted interactions (Dweep et al., 2014). Surprisingly, we identified a small group of 10 miRNAs whose members are reported to target at least 12 ribosomal proteins. Notably, four of those miRNAs seem to possess the ability to bind to more than 30 RPs

encoding genes. This data suggests that the action of only a few miRNAs is adequate to modulate the expression of most RPs and thereby affect translation. Those miRNAs may represent potential biomolecules with anti-cancer activity. Such oncosuppressor miRNAs are frequently downregulated or deleted in cancer cells, thus removing this barrier in tumor initiation and progression. To investigate whether our miRNAs are subjected to this kind of regulation, we performed an expression analysis of miR-129-5p in patient samples from 6 cancers types (breast cancer, colon cancer, head & neck cancer, rectum cancer, hepatocellular cancer, and prostate cancer) (Fig1. D-I) by using the gene expression data deposited in The Cancer Genome Atlas (TCGA) database (Weinstein et al., 2013). Interestingly, we found that one of our candidate miRNAs, miR-129-5p is downregulated in all tested samples (Fig.1 D.). This is consistent with a hypothetical tumor-suppressor function of this miRNA and, likely, the anticancer activity of miR-129-5p is mediated via targeting ribosome biogenesis. Conclusively, this approach has resulted in the identification of a group of miRNAs with potential modulatory role in ribosome biogenesis and at least one miRNA that seems to possess the capacity to act, upon its pharmacological exploitation, as anticancer therapeutic.

Treating cancer remains a pressing problem worldwide. medical Bv integrating biological knowledge as well as bioinformatic analysis we can only identify potential not "druggable" targets but also discover pharmacological agents to block those targets. In this direction, our approach stems from the thorough understanding of ribosomal (de)regulation in cancer cells and integrates bioinformatic tools to identify miRNA molecules able to reverse the impaired translational activity of cancer cells. Indeed, our analysis suggests that at least 10 miRNAs may have a potential modulatory role in ribosomal function. We hypothesized that some of those miRNAs will act as tumor suppressors and, as a proof of concept we analyzed the expression of miR-129-5p across 6 types. cancer MiR-129-5p is consistently downregulated in cancer patients, suggesting that the miRNA exhibits potential anti-cancer activity. Thus, the delivery of miR-129-5p in patients will likely be associated with a beneficial therapeutic outcome and thereby, its study in cancer cell samples highly interesting. is However, a more thorough molecular analysis has to be carried out to pharmacologically exploit the possibility of successfully developing targeted anticancer miRNAs as therapeutics with increased efficacy and safety profiles in the clinical setting. In conclusion, this brief report provides workflow for a the identification of novel miRNA molecules with highly interesting properties in modulating ribosome function in cancer cells.

Acknowledgements

This research is co-financed by Greece and the European Union (European Social Fund - ESF) through the Operational Programme "Human Resources Development, Education and Lifelong Learning 2014-2020" in the context of the project "Strengthening Human Resources Research Potential via Doctorate Research – 2nd Cycle" (MIS 5000432) awarded to CP.

References

Cho, W. C. (2007). Oncomirs: The Discovery And Progress Of Micrornas In Cancers. Mol Cancer, 6, 60.

De Keersmaecker, K., Atak, Z. K., Li, N., Vicente, C., Patchett, S., Girardi, T., Gianfelici, V., Geerdens, E., Clappier, E., Porcu, M., Lahortiga, I., Luca, R., Yan, J., Hulselmans, G., Vranckx, H., Vandepoel, R., Sweron, B., Jacobs. K., Mentens, N., Wlodarska, I., Cauwelier, B., Cloos, J., Soulier, J., Uyttebroeck, A., Bagni, C., Hassan, B. A., Vandenberghe, P., Johnson, A. W., Aerts, S. & Cools, J. (2013). Exome Sequencing Identifies Mutation In Cnot3 And Ribosomal Genes Rpl5 And Rpl10 In T-Cell Acute Lymphoblastic Leukemia. Nat Genet, 45, 186-90.

Delloye-Bourgeois, C., Goldschneider, D., Paradisi, A., Therizols, G., Belin, S., Hacot, S., Rosa-Calatrava, M., Scoazec, J. Y., Diaz, J. J., Bernet, A. & Mehlen, P. (2012). Nucleolar Localization Of A Netrin-1 Isoform Enhances Tumor Cell Proliferation. Sci Signal, 5, Ra57.

Dweep, H., Gretz, N. & Sticht, C. (2014). Mirwalk Database For Mirna-Target Interactions. Methods Mol Biol, 1182, 289-305. Fan, Y., Siklenka, K., Arora, S. K., Ribeiro, P., Kimmins, S. & Xia, J. (2016). Mirnet - Dissecting Mirna-Target Interactions And Functional Associations Through Network-Based Visual Analysis. Nucleic Acids Res, 44, W135-41.

Fancello, L., Kampen, K. R., Hofman, I. J., Verbeeck, J. & De Keersmaecker, K. (2017). The Ribosomal Protein Gene Rpl5 Is A Haploinsufficient Tumor Suppressor In Multiple Cancer Types. Oncotarget, 8, 14462-14478.

Huh, W. K., Falvo, J. V., Gerke, L. C., Carroll, A. S., Howson, R. W., Weissman, J. S. & O'shea, E. K. (2003). Global Analysis Of Protein Localization In Budding Yeast. Nature, 425, 686-91.

Nieminen, T. T., O'donohue, M. F., Wu, Y., Lohi, H., Scherer, S. W., Paterson, A. D., Ellonen, P., Abdel-Rahman, W. M., Valo, S., Mecklin, J. P., Jarvinen, H. J., Gleizes, P. E. & Peltomaki. P. (2014). Germline Mutation Of Rps20, Encoding A Ribosomal Protein, Causes Predisposition То Hereditary Nonpolyposis Colorectal Carcinoma Without Dna Mismatch Repair Deficiency. Gastroenterology, 147, 595-598.E5.

Qu, Y., Liu, H., Lv, X., Liu, Y., Wang, X., Zhang, M., Zhang, X., Li, Y., Lou, Q., Li, S. & Li, H. (2017). Microrna-16-5p Overexpression **Suppresses** Proliferation And Invasion As Well As Apoptosis By Targeting Triggers Vegfa Expression In Breast Carcinoma. Oncotarget, 8, 72400-72410.

Robichaud, N., Sonenberg, N., Ruggero, D. & Schneider, R. J. (2018). Translational Control In Cancer. Cold Spring Harb Perspect Biol. Sulima, S. O., Hofman, I. J. F., De Keersmaecker, K. & Dinman, J. D. (2017). How Ribosomes Translate Cancer. Cancer Discov, 7, 1069-1087.

Tahmasebi, S., Khoutorsky, A., Mathews, M. B. & Sonenberg, N. (2018). Translation Deregulation In Human Disease. Nat Rev Mol Cell Biol, 19, 791-807.

Topisirovic, I. & Sonenberg, N. (2015). Translation And Cancer. Biochim Biophys Acta, 1849, 751-2.

Weinstein, J. N., Collisson, E. A., Mills, G. B., Shaw, K. R., Ozenberger, B. A., Ellrott, K., Shmulevich, I., Sander, C. & Stuart, J. M. (2013). The Cancer Genome Atlas Pan-Cancer Analysis Project. Nat Genet, 45, 1113-20.

Yamakuchi, M., Ferlito, M. & Lowenstein, C. J. (2008). Mir-34a Repression Of Sirt1 Regulates Apoptosis. Proc Natl Acad Sci U S A, 105, 13421-6.