Final accepted version of

```
Roncoroni, M., Brandani, J., Battin, T. and Lane, S.N., 2019.
Ecosystem engineers: biofilms and the ontogeny of glacier
floodplain ecosystems. WIRES Water, 6, e1390
```

A fully typeset version may be viewed at

https://rdcu.be/bVZOf

Page 1 of 47	WIREs Water
1 2 3 4 5 6 7	
8	
9	
10	
11	
12	
13	
14	
15	
20	
21	
22	
Pia Ci	
	m Development
26 Soil	
27 Stabiliz	ation and coil fertilization
28	
29	



Article Title: Ecosystem engineers: biofilms and the ontogeny of glacier floodplain ecosystems

First author Matteo Roncoroni*1, ORCID iD: 0000-0001-6957-6225 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, matteo.roncoroni@unil.ch, No conflicts of interest. Equal first author Jade Brandani', ORCID iD: 0000-0002-0435-1807 Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, Jade brandani@epfl.ch, No conflicts of interest. Second author Tom I. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, Jade Brandani@epfl.ch, No conflicts of interest. Second author Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom battin@epfl.ch, No conflicts of interest. Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, stuart.lane@unil.ch, No conflicts of interest. "Matteo Roncoroni and Jade Brandani contributed equally to the develonment of and writing of this namer	floodplain ecosystems	, nogony o
First author Matteo Roncoroni*1, ORCID iD: 0000-0001-6957-6225 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, matteo.roncoroni@unil.ch, No conflicts of interest. Equal first author Jade Brandani ¹ , ORCID iD: 0000-0002-0435-1807 Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, jade.brandani@epfl.ch, No conflicts of interest. Second author Tom I. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom.battin@epfl.ch, No conflicts of interest. Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, stuart.lane@unil.ch, No conflicts of interest. 'Matteo Roncoroni and Jade Brandani contributed equally to the develorment of and writing of this paper		
First author Matteo Roncoroni* ¹ , ORCID iD: 0000-0001-6957-6225 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, matteo.roncoroni@unil.ch, No conflicts of interest. Equal first author Jade Brandani ¹ , ORCID iD: 0000-0002-0435-1807 Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, jade.brandani@epfl.ch, No conflicts of interest. Second author Ton I. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, jom. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom.battin@epfl.ch, No conflicts of interest. Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, stuart.lane@unil.ch, No conflicts of interest. 'Matteo Roncoroni and Jade Brandani contributed equally to the develorment of and writing of this paper		
First author Matteo Roncoroni* ¹ , ORCID iD: 0000-0001-6957-6225 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, matteo.roncoroni@unil.ch, No conflicts of interest. Equal first author Jade Brandani ¹ , ORCID iD: 0000-0002-0435-1807 Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, jade.brandani@epfl.ch, No conflicts of interest. Second author Tom I. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, jade.brandani@epfl.ch, No conflicts of interest. Second author Tom I. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom.battin@epfl.ch, No conflicts of interest. Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, stuart.lane@unil.ch, No conflicts of interest. 'Matteo Roncoroni and Jade Brandani contributed equally to the development of and writing of this paper		
First author Matteo Roncoroni* ¹ , ORCID iD: 0000-0001-6957-6225 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, matteo.roncoroni@unil.ch, No conflicts of interest. Equal first author Jade Brandani ¹ , ORCID iD: 0000-0002-0435-1807 Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, jade.brandani@epfl.ch, No conflicts of interest. Second author Tom I. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom.battin@epfl.ch, No conflicts of interest. Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, stuart.lane@unil.ch, No conflicts of interest. 'Matteo Roncoroni and Jade Brandani contributed equally to the development of and writing of this paper		
First author Matteo Roncoroni* ¹ , ORCID iD: 0000-0001-6957-6225 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, matteo.roncoroni@unil.ch, No conflicts of interest. Equal first author Jade Brandani ¹ , ORCID iD: 0000-0002-0435-1807 Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, jade.brandani@epfl.ch, No conflicts of interest. Second author Tom I. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom.battin@epfl.ch, No conflicts of interest. Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, stuart.lane@unil.ch, No conflicts of interest. ¹ Matteo Roncoroni and Jade Brandani contributed equally to the development of and writing of this paper		
First author Matteo Roncoroni*1, ORCID iD: 0000-0001-6957-6225 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, matteo.roncoroni@unil.ch, No conflicts of interest. Equal first author Jade Brandani ¹ , ORCID iD: 0000-0002-0435-1807 Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, jade.brandani@epfl.ch, No conflicts of interest. Second author Tom I. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom.battin@epfl.ch, No conflicts of interest. Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, stuart.lane@unil.ch, No conflicts of interest. 'Matteo Roncoroni and Jade Brandani contributed equally to the development of and writing of this paper		
First author Matteo Roncoroni* ¹ , ORCID iD: 0000-0001-6957-6225 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, matteo.roncoroni@unil.ch, No conflicts of interest. Equal first author Jade Brandani ¹ , ORCID iD: 0000-0002-0435-1807 Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, jade.brandani@epfl.ch, No conflicts of interest. Second author Tom I. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom.battin@epfl.ch, No conflicts of interest. Second author Tom I. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom.battin@epfl.ch, No conflicts of interest. Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, stuart.lane@unil.ch, No conflicts of interest. ¹ Matteo Roncoroni and Jade Brandani contributed equally to the development of and writing of this paper		
First author Matteo Roncoroni* ¹ , ORCID iD: 0000-0001-6957-6225 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, matteo.roncoroni@unil.ch, No conflicts of interest. Equal first author Jade Brandani ¹ , ORCID iD: 0000-0002-0435-1807 Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, jade.brandani@epfl.ch, No conflicts of interest. Second author Ton I. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom.battin@epfl.ch, No conflicts of interest. Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, stuart.lane@unil.ch, No conflicts of interest. ¹ Matteo Roncoroni and Jade Brandani contributed equally to the development of and writing of this paper		
First author Matteo Roncoroni* ¹ , ORCID iD: 0000-0001-6957-6225 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, matteo.roncoroni@unil.ch, No conflicts of interest. Equal first author Jade Brandani ¹ , ORCID iD: 0000-0002-0435-1807 Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, jade.brandani@epfl.ch, No conflicts of interest. Second author Tom I. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom.battin@epfl.ch, No conflicts of interest. Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, stuart.lane@unil.ch, No conflicts of interest. ¹ Matteo Roncoroni and Jade Brandani contributed equally to the development of and writing of this paper		
Matteo Roncoroni*1, ORCID iD: 0000-0001-6957-6225 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, matteo.roncoroni@unil.ch, No conflicts of interest. Equal first author Jade Brandani ¹ , ORCID iD: 0000-0002-0435-1807 Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, jade.brandani@epfl.ch, No conflicts of interest. Second author Tom I. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, iom.battin@epfl.ch, No conflicts of interest. Second author Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom.battin@epfl.ch, No conflicts of interest. Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, stuart.lane@unil.ch, No conflicts of interest. 'Matteo Roncoroni and Jade Brandani contributed equally to the development of and writing of this paper	First author	
Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, matteo.roncoroni@unil.ch, No conflicts of interest. Equal first author Jade Brandani ¹ , ORCID iD: 0000-0002-0435-1807 Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, jade.brandani@epfl.ch, No conflicts of interest. Second author Tom I. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom.battin@epfl.ch, No conflicts of interest. Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, stuart.lane@unil.ch, No conflicts of interest. 'Matteo Roncoroni and Jade Brandani contributed equally to the development of and writing of this paper	Matteo Roncoroni*1, ORCID iD: 0000-0001-6957-6225	
Lausanne, Switzerland, matteo.roncoroni@unil.ch, No conflicts of interest. Equal first author Jade Brandani ¹ , ORCID iD: 0000-0002-0435-1807 Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, jade.brandani@epfl.ch, No conflicts of interest. Second author Tom I. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom.battin@epfl.ch, No conflicts of interest. Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, stuart.lane@unil.ch, No conflicts of interest. ¹ Matteo Roncoroni and Jade Brandani contributed equally to the development of and writing of this paper	Institute of Earth Surface Dynamics (IDYST), University of Lau	sanne, 1015
Jade Brandani ¹ , ORCID iD: 0000-0002-0435-1807 Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, jade.brandani@epfl.ch, No conflicts of interest. Second author Tom I. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom.battin@epfl.ch, No conflicts of interest. Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, stuart.lane@unil.ch, No conflicts of interest. ¹ Matteo Roncoroni and Jade Brandani contributed equally to the development of and writing of this paper	Lausanne, Switzerland, <u>matteo.roncoroni@unil.ch</u> , No conflicts of int	erest.
Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, jade.brandani@epfl.ch, No conflicts of interest. Second author Tom I. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom.battin@epfl.ch, No conflicts of interest. Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, stuart.lane@unil.ch, No conflicts of interest. ¹ Matteo Roncoroni and Jade Brandani contributed equally to the development of and writing of this paper	Jade Brandani ¹ , ORCID iD: 0000-0002-0435-1807	
Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, jade.brandani@epfl.ch, No conflicts of interest. Second author Tom I. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom.battin@epfl.ch, No conflicts of interest. Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, stuart.lane@unil.ch, No conflicts of interest. ¹ Matteo Roncoroni and Jade Brandani contributed equally to the development of and writing of this paper	Stream Biofilm and Ecosystem Research Laboratory (SB	ER), École
Second author Tom I. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom.battin@epfl.ch, No conflicts of interest. Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, stuart.lane@unil.ch, No conflicts of interest. ¹ Matteo Roncoroni and Jade Brandani contributed equally to the development of and writing of this paper	Polytechnique Fédérale de Lausanne, 1015 Lausanne,	Switzerland,
Tom I. Battin Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom.battin@epfl.ch, No conflicts of interest. Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, stuart.lane@unil.ch, No conflicts of interest. ¹ Matteo Roncoroni and Jade Brandani contributed equally to the	Second author	
Stream Biofilm and Ecosystem Research Laboratory (SBER), École Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom.battin@epfl.ch, No conflicts of interest. Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, stuart.lane@unil.ch, No conflicts of interest. ¹ Matteo Roncoroni and Jade Brandani contributed equally to the development of and writing of this paper	Tom I. Battin	
Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland, tom.battin@epfl.ch, No conflicts of interest. Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, stuart.lane@unil.ch, No conflicts of interest. ¹ Matteo Roncoroni and Jade Brandani contributed equally to the development of and writing of this paper	Stream Biofilm and Ecosystem Research Laboratory (SB	ER), École
Second author Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, <u>stuart.lane@unil.ch</u> , No conflicts of interest. ¹ Matteo Roncoroni and Jade Brandani contributed equally to the development of and writing of this paper	Polytechnique Fédérale de Lausanne, 1015 Lausanne,	Switzerland,
Stuart N. Lane, ORCID iD: 0000-0002-6077-6076 Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, <u>stuart.lane@unil.ch</u> , No conflicts of interest. ¹ Matteo Roncoroni and Jade Brandani contributed equally to the development of and writing of this paper	Second author	
Institute of Earth Surface Dynamics (IDYST), University of Lausanne, 1015 Lausanne, Switzerland, <u>stuart.lane@unil.ch</u> , No conflicts of interest. 1Matteo Roncoroni and Jade Brandani contributed equally to the development of and writing of this paper	Stuart N. Lane, ORCID iD: 0000-0002-6077-6076	
Lausanne, Switzerland, <u>stuart.lane@unil.ch</u> , No conflicts of interest. ¹ Matteo Roncoroni and Jade Brandani contributed equally to the development of and writing of this paper	Institute of Earth Surface Dynamics (IDYST), University of Lau	sanne, 1015
development of and writing of this paper	Lausanne, Switzerland, <u>stuart.lane@unil.ch</u> , No conflicts of interest.	ally to the
	development of and writing of this paper	ing to the

6 Abstract

The term 'ecosystem engineering' emerged in the 1990s and is commonly used to refer to the activities of larger organisms like beavers and trees in rivers and streams. The focus on larger organisms may be motivated by their more visible effects on the environment. However, whilst it may be intuitive to suggest that the bigger the organism the bigger its potential engineering effects, there may be micro-scale organisms who through their number rather than their size can act simultaneously to result in significant impacts. This paper considers biofilms as a candidate ecosystem engineer. It is well known that biofilms play an important role in enriching the sediment matrix of nutrients and in stabilizing sediments. Biofilms may be critical in increasing the habitability of the benthic substratum. In this paper, we consider their potential role in the ontogeny of ecosystems in recently deglaciated terrain. We show how by changing sediment stoichiometry, decreasing sediment erodibility and reducing surface sediment permeability they may promote primary succession on lateral, incised terraces, which are less perturbed compared with the main active floodplain.





In streams, biofilms should be depicted as ecosystem engineers as they stabilize and fertilize sediments. Their development is thus critical for river ecosystems, particularly in glacier floodplains, where sediments tend to be unstable and nutrient-limited.

1. Introduction

Some organisms have the capacity to transform ecosystems echoing ways engineers build, create and modify their surrounding environment (Polvi and Sarneel, 2018). The term "ecosystem engineering", commonly used in ecology, describes this phenomenon and takes place in diverse ecosystems involving potentially a multitude of organisms (Jones et al., 1994; 1997). In an introduction to ecosystem engineers in rivers, Polvi and Sarneel (2018) state that while the balance between hydraulic energy and sediment size and quantity influence the shape and function of streams, ecosystem engineers may considerably modify the functioning of aquatic and riparian ecosystems by changing streamflow characteristics. Freshwater ecosystem engineers can be found from headwaters downstream to estuaries, and in channels with different perimeter sedimentology (e.g., gravel, sand, and cohesive or coarse material), patterns (e.g., single-thread, braided) and morphodynamics. A stream's hydrological context may set constraints upon the growth and survival of ecosystem engineers in aquatic and riparian ecosystems (Gurnell, 2014; Bätz et al., 2015) and organisms will generally only engineer positive feedbacks where resources are available (e.g., trapping and stabilizing fine sediments) within those constraints (Moore, 2006).

The floodplains of glacier-fed rivers, characterized by a recent switch to phototrophic conditions and extensive deposits of glaciogenic sediment, are usually slow to revegetate (Hotaling et al., 2017a). Altitude, and hence temperature, may be a factor restricting vegetation succession, but fundamentally, glaciogenic sediments are nutrient-poor and unstable. Although conceptual models of vegetation succession in glacial forefields are well-developed (see review in Miller and Lane, 2019) the early stages of ontogenesis, and notably the role played by microbial life in driving related processes, is very poorly understood. Freshwater biofilms can develop rapidly and are implicated in multiple ecosystem processes (Battin et al., 2003; 2016) and, because of the profound impacts that they can have upon resource availability, they are candidate ecosystem engineers. In this review, we focus upon freshwater stream benthic biofilms, as potential ecosystem engineers (Gerbersdorf et al., 2008a; 2008b; 2009) with a particular focus on the early ontogeny of glacial floodplain ecosystems following glacier retreat. Biofilms have received less focus compared to more visible organisms such as beavers, mussels, and snails (Gurnell, 2014), and thus their engineering capacity is less well documented (Polvi and Sarneel, 2018). The aim of this paper is to review the role of biofilms as ecosystem engineers in streams and to develop a conceptual model for the relevance of microbial biofilms for glacial floodplain ecosystem ontogeny.

- 2. Ecosystem engineering: key concepts

2.1 The concept of ecosystem engineering

Almost three decades ago, two new terms emerged in ecology: 'ecosystem engineering' and 'ecosystem engineers' (Lawton and Jones, 1993; Jones et al., 1994). The former describes the act of engineering an ecosystem, while the latter describes how organisms act as engineers (Jones et al., 1994). The notion of ecosystem engineering and engineers in rivers and streams has been reviewed by Moore (2006) and more recently by Polvi and Sarneel (2018) and so only a short summary is provided here.

Jones et al. (1994) described ecosystem engineers as "organisms that directly or indirectly modulate the availability of resources [...] to other species, by causing physical state changes in biotic and abiotic materials" (Jones et al., 1994, p. 374). Jones et al. (1994; 1997) also introduced a key distinction between ecosystem engineers: autogenic versus allogenic. Autogenic engineers modify the surrounding environment through their own living or dead tissue (Jones et al., 1994; Polvi and Sarneel, 2018). Allogenic engineers change the environment through the transformation of materials from one state to another (Jones et al., 1994; 1997). As an example, vegetation (e.g., willows) plays an important role in the riparian zone (Gurnell and Petts, 2002) by stabilizing sediments and leading to the creation of more stable habitats like islands (Gurnell, 2014), as well as increasing bank stability (Simon and Collison, 2002). Hence, riparian plants are autogenic engineers as it is their own root system that increases the resistance of river banks to erosion. In contrast, caddisfly larvae build nets that reduce sediment erodibility by increasing the critical shear stress and hence resistance to sediment entrainment (Statzner et al., 1999; Cardinale et al., 2004; Johnson et al., 2009). They are allogenic engineers as it is material made by the larvae that increases the resistance of gravels to erosion by water. The distinction between autogenic and allogenic engineers is not fixed; allogenic engineers can also have autogenic effects, and vice versa (Jones et al., 1994), sometimes simultaneously. For instance, Salix autogenically increases the stability of river banks but allogenically changes depositional processes and can influence soil formation (Bätz et al., 2015).

Ecosystem engineering is not concerned with direct provision of resources (e.g., as part of a food chain) (Jones et al., 1994; 1997). For instance, predator-prey relationships between fish, such as salmonids, and macroinvertebrates, such as stoneflies, (Soluk and Richardson, 1997) are not engineering because prey simply provide food to predators. Rather, ecosystem engineers effectively manage resources; either intentionally or unintentionally. The former describes the traits ('extended phenotypes'; see Dawkins, 1982) that an organism possesses that allows intentional transformation of its environment (e.g., when a beaver explicitly builds a dam or the female salmon digs a redd) (Jones et al., 1994; 1997). The latter describes the traces that have been left incidentally by organisms but which "manage" the environment for others (e.g., woody debris in the stream) (Jones et al., 1994; 1997). Whether or not ecosystem engineering is intentional or unintentional, it is thought to occur in all ecosystems (Jones et al., 1994) and the majority of, if not all, organisms are capable of doing it (Jones et al., 1994; 1997; Wright and Jones, 2004).

Jones et al.'s statements about the ubiquity of ecosystem engineering opened a scientific debate about the usefulness of the terms 'ecosystem engineering' and 'ecosystem engineer';

what use is such terms if they apply everywhere and to all things (Reichman and Seabloom, 2002a; 2002b)? Ecosystem engineering needs to be distinguished in terms of scale and magnitude of impacts and not simply based upon a functional definition regarding how an organism engineers. Hence, we consider both functions and scale/magnitude effects. Hastings et al. (2007) noted that ecosystem engineering impacts can be either small or large, both in size and in time, and are related to the size and lifespan of the engineer itself. Impacts should be important and visible at large scales for organisms to be true engineers of ecosystems (Hastings et al., 2007). Intentional engineering may seem to be more important than unintentional, but this is not necessarily the case. For instance, large woody debris, which is unintentional engineering, can substantially impact salmonid populations (House and Boehne, 1986; Zika and Peter, 2002).

2.2 Stream ecosystem engineering: a matter of scale

The physical processes that drive rivers and the ways in which rivers in turn erode and deposit sediment to make landscapes have been identified at scales from the sub-daily to millions of years (Schumm and Lichty, 1965). Such processes not only produce spatio-temporal variation in the landscapes we see but also create the habitat templates upon which life develops and also the constraints to which life must adapt (e.g., flow regime, accommodation space, perturbation frequency, microclimate, etc.). At the same time, and still poorly understood, organisms can modify stream ecosystems (Moore, 2006; Polvi and Sarneel, 2018) through ecosystem engineering. Not all organisms have similar engineering impacts on streams, and these effects depend upon the engineers themselves (Hastings et al., 2007). Ecosystem engineering has tended to focus on visible impacts (e.g., beaver dams) made by large organisms (see Gerbersdorf et al., 2009) but scaling stream ecosystem engineers based on their size is not forcibly correct for three reasons.

First, within-organism size variations may be important and bigger individuals may have bigger engineering effects. For instance, in relation to brown trout (Salmo trutta) redds, bigger females produce bigger redds (Crisp and Carling, 1989). Redd construction, by sorting sediments, decreases streambed erodibility by increasing the critical shear stress for sediment transport (Montgomery et al., 1996). Thus, bigger female trout have bigger engineering effects on the streambed compared with smaller females.

Second, scale depends upon your point of view. Depending on the chosen geomorphological scale (e.g., watershed versus pool-riffle sequence) ecosystem engineering can have substantial differences in apparent impacts. For instance, vegetation might have a greater stabilization potential compared to caddisflies at the overall watershed level, whereas for a given streambed surface caddisflies might have greater potential. Hence, the smaller the scale, the more important smaller organisms become.

- Third, there is an important relationship between engineering magnitude and the number of organisms taking part in the engineering effort, and this association is generally positive and may be more than additive. This is because it is the assemblage of organisms that multiply the engineering potential of any individual organism (Boogert et al., 2006). Consider for example microorganisms (e.g., microbes); individually they are not able to stabilize

1 2		
3 4	152 153	sediments; in communities (biofilms) they may have remarkable stabilization effects (Section 3.2).
5 6	100	
7 8		
9 10		
11 12		
13 14		
15 16		
17 18		
19 20		
21 22		
23 24		
25 26		
27 28		
29 30		
31 32		
33 34 25		
35 36 37		
37 38 20		
40 41		
42 43		
44 45		
46 47		
48 49		
50 51		
52 53		
54 55		
56 57		
58 59		
60		6

3. Stream biofilms as ecosystem engineers

3.1 Biofilms: an introduction

Biofilms dominate microbial life in most aquatic ecosystems (Flemming and Würtz, 2019) and particularly in streams (Battin et al., 2016). Biofilms are surface-attached microbial communities that dwell in a matrix composed of extracellular polymeric substances (EPS). Several properties emerge from the biofilm mode of life that underpin its ecological success (Flemming et al., 2016). The spatial organisation of microbial life within a matrix induces the establishment of physical and chemical gradients that provide habitat diversity and hence increases biodiversity. The biofilm matrix also facilitates the sorption of solutes and resource capture, stimulating biotic interactions and protecting from erosion, grazing and UV radiation, for instance.

19 166

Benthic biofilms in streams are highly diverse, including taxa from all three domains of life (that is, archaea, bacteria and eukaryotes) (Battin et al., 2016; Bengtsson et al., 2018) and even abundant and diverse viruses (Meriem Bekliz, personal communication). Prokaryotes and algae (mostly diatoms) rapidly colonize sedimentary surfaces where, after an initial adhesion phase, growth triggers a succession that often involves bacteria that are typical biofilm formers (Niederdorfer et al., 2016). As this succession proceeds, niches can develop within the nascent biofilm, which fosters its biodiversity (Jackson et al., 2001) (Figure 1).

Depending on local hydraulics and sediment size distribution, benthic biofilms form complex and dynamic physical structures, including ridges or filamentous streamers (in the centimeter range) that float in water (Flemming and Wingender, 2010). The physical structure of biofilms is critical for mass transfer and related microbial processes including the uptake and metabolism of organic compounds (e.g., Battin et al. 2003; Singer et al. 2010). It is the combination of the very large reactive surface, high biodiversity and continuous exposure to the flow of streamwater and associated solutes that make biofilms so important for stream ecosystem processes. For instance, they are the major sites of ecosystem metabolism (that is, gross primary production and respiration) and therefore of nutrient and carbon cycling in streams (Battin et al., 2016; Demars, 2018). As such, they also form the basis of the food web in numerous streams.

42 ¹⁸⁵ 43 186

A critical aspect of biofilms is the matrix, extracellular material in which biofilms cells are embedded, acting as a protection for organisms (Flemming et al., 2016). The matrix is also recognized to store all available material from lysed cells and to capture from the water phase both dissolved and particulate nutrients to make them available as nutrient and energy source (Flemming and Wingender, 2010). EPSs are the principal constituent of this matrix and is responsible for the biofilm's internal structure, surface attachment, and cohesion by maintaining cells in close proximity leading to strong cell-to-cell interactions (Flemming and Wingender, 2010). EPSs differ between biofilms in terms of nutrient availability, community composition, hydrologic forces and temperature. In their review, Flemming and Wingender (2010) describe the functions of EPSs bacterial biofilms. These include functions such as adhesion, aggregation, cohesion of biofilms, sorption of inorganic ions, enzymatic activity, nutrient source and water retention. It is thus possible to understand how the matrix can affect the mode of life of a given biofilm.





Figure 1: Biofilm development stages. From planktonic phase, microbes deposit and attach on the surface and start producing extracellular polymeric substances (EPS). With the secretion of EPS substances, the biofilm enters its growth stage and microcolonies are formed. When the biofilm matures, it may reach a 'mushroom' structure where slough of biofilm will later be affected by erosion forces and dispersal. This dispersal stage will link together local communities and form biofilm metacommunities.

3.2 Biostabilization of sediments by biofilms

The hypothesis that biofilms cause biostabilization was suggested early in the 1930s by Carter (1932; 1933a; 1933b), but it was only quantitatively confirmed in the 1970s (Neumann et al., 1970; Scoffin, 1970; Holland et al., 1974). Traditionally, sediment transport has been seen as a physical process (Le Hir et al., 2007), related to channel hydraulics (e.g., Shields, 1936). However, in natural ecosystems, sediment transport processes can be altered by biotic influences (Flemming, 2002). This process is known as 'biostabilization', where organisms decrease the susceptibility of sediment to erosion (Paterson and Daborn, 1991).

Particle movement in streams starts when bed shear stress τ_o exceeds a certain threshold, the critical bed shear stress τ_{oc} (e.g., Leopold *et al.*, 1964; Richards, 1982; Fang *et al.*, 2014). It is now recognized that biofilms tend to increase τ_{oc} (Neumeier *et al.*, 2006; Fang *et* al., 2014; Pivato et al., 2019). EPSs are the major driver of increased cohesion and stability of sediments (de Boer, 1981; Grant and Gust, 1987; Paterson, 1989; 1994; Dade et al., 1990). Following Tolhurst et al. (2002), EPSs involve sediment stabilization by either

physical binding (apparent cohesion) or molecular electrochemical interactions (chemical cohesion).

It is also known that the biofilms modify bed morphology, so changing bed roughness, influencing near-bed velocities and ultimately impacting sediment transport (van Rijn, 2007). Biofilms may themselves have a morphology and, as a result, they may increase (Flemming et al., 2007a) or reduce (Fang et al., 2014) bed roughness. Biofilms can increase the roughness by developing free-flowing "fluffy" morphologies (e.g., free flowing streamers) that are easily erodible along with the uppermost sediment layers even at low flows (Flemming et al., 2007a). On the other hand, biofilms can also reduce bed roughness by developing smoother mats, which reduce the shear stress available to transport the particles (Fang et al., 2014). However, this can also reduce the energy losses at the surface, change shear stress partitioning and ultimately increase the component of shear stress available to entrain and transport sediment. Bed-smoothing (e.g., by introduction of finer sediments) has been shown to lead to bed destabilization (Wilcock et al., 2001; An et al., 2019) independent of any increases in critical shear stress. Whilst biofilms may increase resistance to motion, their impact on shear stress partitioning may also increase the shear stress available for transport, which results in an increase of the susceptibility of the bed to being eroded. Thus, the bed stabilization provided by biofilms is a function of the balance between the forces that increase sediment cohesion and hence resistance to erosion and the forces that tend to erode them, both external (e.g., shear stress) and internal to the biofilms (e.g., fluffy morphologies, smoothed mats).

Whether or not biofilms stabilize sediments, it appears that this process is particle size dependent, and proportionally more important for finer particles. For instance, Statzner et al. (1999) found that benthic microalgae and their biofilms are able to stabilize sand particles, i.e. those finer than 2 mm. The fact that biofilm stabilization is potentially particle size dependent, and greater for finer sediments, perhaps explains why there has been more focus on marine environments (e.g., tidal flats) than on freshwater environments (Fang et al., 2014). Freshwater environments can have coarser bed sediment but many coastal environments are depositional, composed of terrestrially-derived material that has been sorted by rivers and streams, such that most material is finer.

Biostabilization also appears to be more efficient in saltwater than in freshwater because of ion concentrations related to seawater chemical properties (Spears et al., 2008; Gerbersdorf and Wieprecht, 2015). It has been shown (Flemming and Wingender, 2010) that biostabilization could be enhanced by a surplus of cations, such as in saltwater, that helps to create strong electrochemical bridges between the EPS and the sediments. For instance, Stal (2003) and Flemming et al. (2007a) found that an increase in Ca²⁺ (and in general of all divalent cations) has the consequence of increasing the binding forces between the matrix and the sediments resulting in greater biostabilization.

To investigate the role of biofilms in stabilizing sediments, the analysis of shear stress and/or critical shear stress is common. Grant and Gust (1987) investigated the role of purple sulphur bacterial mats in stabilizing marine sediments, and they tested their hypothesis in laboratory flumes with samples collected directly on a South Florida beach. By increasing flume discharge (i.e., increasing τ_0) until the erosion threshold was reached, they

demonstrated that colonized sediments have a τ_{oc} up to five times higher compared with

sterile sediments (Grant and Gust, 1987). Similar results were recorded by Tolhurst et al. (2002) who found a 3.7 times increase in τ_{oc} . In their experiments, they used cohesive sediments from a Scottish estuary and a variable amount of commercially extracted EPSs (Tolhurst et al., 2002), which mimics the natural increase of EPSs following biofilm growth but without the need to wait for natural biofilm growth.

Neumeier et al. (2006) simulated diatom biofilm growth for a tidal environment in a laboratory flume in a 28-day experiment. They reported a 10 fold increase in τ_{ac} for sediments colonized by a well-developed biofilm (after 28 days of colonization) compared with abiotic sediments (Neumeier et al., 2006). In two 5-week experiments using small glass beads and biofilms sampled in a Scottish estuary, Gerbersdorf et al. (2008b) and Gerbersdorf et al. (2009) found that τ_{oc} was up to 3.6 and 10.8 times higher respectively compared with abiotic controls. The difference between these two increases was explained by the use of bacteria only in the former experiment and by the use of a mixture of bacteria and diatoms in the latter experiment (Gerbersdorf et al., 2009). Gerbersdorf et al. (2009) stated that one possible reason for this increase was synergistic interactions between diatoms and bacteria, with the former profiting from the exudates of the latter and vice versa.

Vignaga (2012) investigated biostabilization by freshwater cyanobacteria using experimental flumes, different particle sizes (1 mm, 1.2 mm and 2.2 mm) and different growth periods (up to 10 weeks of colonization). Results showed the greatest biostabilization after 4 weeks with a maximum increase of τ_{oc} of 43% for particles of 1 mm, 35% for particles of 2.2 mm and 30% for particles of 1.2 mm (Vignaga, 2012). This confirms the above-mentioned hypothesis that the impact of biofilms is likely to be greater for finer sediment. Thom et al. (2015), in 4-to 8-week experiments using flumes, riverine water, and glass beads of 0.1 and 0.2 mm showed a maximum increase of stability of almost 10 times compared with an abiotic reference ($\tau_{oc} = 0.23 \text{ Nm}^{-2}$) (Thom *et al.*, 2015).

Thom et al. (2015) also found that biostabilization is a seasonally driven phenomenon; the increase of τ_{oc} changes through the year. Sediment stability was just 1 to 1.5 times τ_{oc} for late autumn conditions, 3 times τ_{oc} for summer conditions and at maximum (10 times) for spring conditions (Thom et al., 2015). Schmidt et al. (2016) also found a maximum increase of sediment stability with spring conditions and a minimum with autumn conditions. Seasonality has also been recorded in tidal environments (e.g., Widdows et al., 2000; Amos et al., 2004).

Pivato et al. (2019) showed that during winter and early spring biofilm growth was limited by low sediment temperatures and light availability. The low growth rate results in less developed winter biofilms (low biomass) incapable of stabilizing sediment because the mat was not developed enough to significantly increase the shear stress (Pivato et al., 2019). However, when meteorological and environmental conditions (e.g., light availability and sediment temperature) were suitable, the biofilm was able to mature rapidly and to provide effective biostabilization because the mat was developed enough to support higher shear stress (Pivato et al., 2019).

 Other studies have considered the role of disturbance, here referred as an increase in the shear stress that could lead to bed destabilization and subsequent biofilm detachment, in conditioning how biofilms influence biostabilization. Mariotti and Fagherazzi (2012) modeled biofilms impacts under different disturbance scenarios in tidal environments. In their first scenario, strong but infrequent disturbances were applied to the biofilm, in the second, weak and infrequent disturbances were applied, while in the third weak (of the same magnitude of scenario 2) weak and frequent disturbances were applied. In the first case, biofilm biomass and τ_{oc} increased until the occurrence of the disturbance, which systematically destroyed the biofilm and re-started the cycle. In the second case, disturbances were neither large enough nor frequent enough to have a negative impact on the biofilm, which increased in both biomass and τ_{oc} until an equilibrium state was reached, defined here as the point in which biomass and critical shear stress values fluctuate around a steady mean value. In the third case, frequent weak disturbances destroyed the biofilm progressively. From the last two scenarios, it appears that biofilm biostabilization is also a function of biofilm growth history (Mariotti and Fagherazzi, 2012). In similar projects, Yallop et al. (1994), Valentine et al. (2014) and Chen et al. (2017; 2019) for tidal environments and Thom et al. (2015) for riverine systems found that shorter growth periods lead to reduced stabilization effects, whilst long periods allow for stronger stability resulting in an increased ability to support greater shear stresses.

Chen *et al.* (2017) proposed also that the role of EPSs in stabilizing sediment is not only a
 surficial phenomenon, and they argued that EPSs can penetrate into sediment beds allowing
 for a deeper impact of biostabilization. In fact, they showed that after the failure of the
 surficial biofilm layer, the sediment stability still remained in place (Chen *et al.*, 2017).

In summary, biofilms have the capacity to increase the critical shear stress of sediments, in particular by producing viscoelastic EPSs. With increasing biostabilization, biofilms construct their own niche and enhance the habitability of an otherwise unstable sedimentary environment. With better habitability, biofilms proliferate resulting in greater EPS production and increased sediment stability, and so on until they are scoured or disintegrate. Hence, by changing the sediment state from unconsolidated to consolidated, biofilms act as allogenic engineers and this is done intentionally. Therefore, we propose the biostabilization of sedimentary environments as another important emergent property of the biofilm mode of life in streams, but whose importance is likely to be context specific (e.g., more important for fine sediments than coarse sediments).

47 350 **3.3 Fertilization of sediments by biofilms**

The mineral surfaces of stream sediments are typically devoid of significant amounts of organic substances. Dissolved molecules can sorb to the mineral surfaces thereby forming organomineral complexes (Aufdenkampe et al., 2011), which are typically thought to be protected from metabolism and hence of minor relevance for ecological processes (Rothman and Forney, 2007). As microbial biofilms colonize and accumulate on these mineral surfaces, they change the chemical stoichiometry of the sedimentary environment. Several processes are involved in this chemical shift. The adhesion of bacterial cells to minerals enriches their surface in nitrogen (N) and phosphorus (P) relative to carbon (C). This is because the C:N and C:P ratio of bacterial cells typically varies from 4 to 8 and 10 to 80,

respectively (Hall et al., 2010). Microbial in origin, EPS are biopolymers that not only contain polysaccharides, but also DNA and a large variety of proteins as well (Flemming et al., 2007b). As cells start proliferating and producing EPS they add organic carbon to the mineral matrix. Depending on the contribution of extracellular DNA and proteins to the EPS, the build-up of a biofilm matrix further enriches the mineral environment in P and N. As cells decay within the biofilms, their organic constituents are released to the matrix where they are transiently protected from loss through water flow, for instance. The exudation of storage compounds by algae to maintain stoichiometric homeostasis during peak photosynthesis or shortly thereafter further contributes to chemical enrichment with the biofilm matrix. It is the close spatial proximity between the transient storage of reactive solutes and active cells that facilitates the remineralisation and elemental cycling within biofilms (Battin et al., 2003). At the same time, the EPS matrix easily scavenges solutes from the streamwater owing to its very large surface area and chemical surface properties (Freeman and Lock, 1995; Flemming et al., 2016). As the biofilm matrix is enriched by EPS secretion and other chemical processes, more dissolved molecules sorb on mineral surfaces. This process is thus claimed to fertilize the biofilm's surrounding environment (Flemming and Wingender, 2010).

We argue therefore that the enrichment in nutrients by microorganisms and aided by the biofilm matrix acts as a fertilizer of an otherwise nutrient-poor mineral matrix. This is analogous to the soil build-up during primary vegetation succession. In streams, this process may gain relevance with concomitant biostabilization. In fact, it would be the latter that provides a time scale for microbial biofilms to accumulate and to recycle nutrients, and thereby fertilize the sediments. This is why we propose that biofilms act as allogenic engineers that alter the environment through material transformation (Jones et al., 1994). The resources resulting from this engineering become available for other microorganisms, but also for macroinvertebrates and potentially for plant seedlings as well. Fertilization can be understood as an intentional engineering process with a positive feedback on all microorganisms composing the biofilm matrix. In fact, nutrient enrichment within the sedimentary environment facilitates biofilm proliferation and further enhances nutrient stripping from the streamwater.

Key definitions:

> Autogenic: autogenic engineers are organisms that change their surrounding environment through their own living or dead tissues, such as vegetation that increases river bank stability through its own biological structure.

Allogenic: allogenic engineers are organisms that change their surrounding environment through the transformation of materials from one state to another, such as beavers that build wooden dams by cutting living trees.

Intentional engineering: intentional modification of the surrounding environment to benefit from it (e.g., nest digging by salmonids).

Unintentional engineering: unintentional modification of the surrounding environment that does not provide direct feedbacks to the engineer, but it can have positive or negative impacts upon other organisms. Extracellular Polymeric Substances (EPSs): biomolecules binding cells to each other and to solid materials mainly composed of polysaccharides and proteins. **Prokaryotes:** Unicellular organisms lacking internal membrane-bound structure without a distinct nucleus. Lysed cells: Cells affected by the breakdown of membrane due to virial, enzymatic or osmotic processes compromising its integrity. Stoichiometric homeostasis: property of an organism to keep its biomass element ratio relatively stable independently of its substrate. **Roughness:** roughness describes the loss of energy experienced by the flowing water due to the friction against the streambed and/or banks. Related to channel morphology (e.g., grain size, grain angle, etc.), it influences hydraulic components such as flow velocities, turbulence and particle motion, as well as the ecological setting of the benthic substratum. **Critical shear stress:** the critical shear stress (τ_{oc}) is the threshold from which particles can be mobilized. Below this threshold, resisting forces (e.g. gravity) keep grains on the streambed; above this threshold, moving forces initiate particle movement. Hence, particle movement begins only when the shear stress (τ_o) exerted by water on the grains is greater than the critical shear stress defined by properties of the streambed. **Ontogeny:** in biology, ontogeny refers to development of an organism, an anatomical part or a behavioral trait through time. In this paper, we refer to the ecological development of glacial floodplains. In particular, we refer to the way in which life develops in these environments by converting barren, hostile and water-limited soils to fertile and water retaining ones (i.e., morphogenesis of a rocky environment).

4. Biofilms, ecosystem engineering and the ontogeny of glacial floodplains

4.1 Glacial floodplains and streams: highly dynamic environments

Glacial floodplains are highly dynamic environments experiencing continuous morphological change (Marren, 2005; Heckmann et al., 2016). Stream braiding is common in these environments (Maizels, 1995; Marren, 2005), and is controlled by both allogenic and autogenic factors (Ashworth and Ferguson, 1986). The system is externally influenced by discharge and sediment delivery. These combine with the extant morphology to determine spatial patterns of flow velocity and shear stress, as well as the sediment available for transport (Ashworth and Ferguson, 1986). Internally, shear stress partitioning and sediment availability regulate sediment transport. The latter shape channel morphology; and may sort sediment on the streambed, so changing bed roughness and velocity distributions and consequently the shear stresses (Ashworth and Ferguson, 1986). Bed sediment sorting will also change τ_{oc} . Thus, external factors control the internal ones and vice versa.

Discharge in glaciated river basins, and consequently in glacial forefields, has substantial variations both daily and yearly (Maizels, 1995; Marren, 2005), and it is usually charged with sediment in the form of bedload or suspended load (Gurnell, 1987; Milner and Petts, 1994). Daily or diurnal flow changes interact with stream bathymetry to drive rapid increases and then decreases in bed shear stress and hence sediment supply and sediment transport. Runoff can be produced by snowmelt or ice melt. Because the majority of ice melt is produced at the glacier surface (Nienow et al., 1998), ice melt is driven primarily by solar radiation, temperature and wind speed (see Fountain, 1996; Nienow et al., 1998; Swift et al., 2005). Annually, discharge is strongly bound with radiation and temperature variations (Maizels, 1995; Lane and Nienow, 2019). Discharge is low in winter (Maizels, 1995; Malard et al., 1999) because the air temperature is below the melting point of ice. During late spring and summer, solar radiation becomes more intense and temperatures rise, and the daily flow maxima and variability increase due to snowpack melt (Nienow et al., 1998). As snow cover tends both to slow melt and to buffer melt water delivery to the stream, as long as the basin remains significantly snow covered, flow maxima tend to be damped and baseflows maintained (Lane and Nienow, 2019). However, as snow melt occurs and the snow line retreats up a glacier, ice melt begins. As ice has a lower albedo than snow, melt rates are higher and as ice has a much lower porosity that snow, buffering and hence flow attenuation are reduced. Thus, peak flows become bigger and baseflows become smaller (Nienow et al., 1998; Malard et al., 1999; Swift et al., 2005; Lane and Nienow, 2019) through the melt season. By the end of the summer, falling incoming solar radiation causes glacier melt and so peak discharge to decrease but diurnal discharge variation is maintained. By late autumn, discharge approaches the winter minima (Maizels, 1995).

This variability in discharge is also coupled to strong variability in both suspended load and bedload. In glacial forefields, sediment sources may be either glacial or periglacial. The latter can be important, although this does depend on the degree of valley sidewall to proglacial stream coupling, which may be limited by alluvial fan formation (Lane et al., 2017). Over annual to decadal time-scales glacial sediment sources are predominantly related to subglacial sediment evacuation by glacier melt. Measurements close to glacier margins (Mao et al., 2019) show distinct timescales of variation in suspended sediment concentration

(SSC)-discharge relations. At the daily time-scale (Clifford et al., 1995; Perolo et al., 2019), clockwise hysteresis can result because discharge forces shear stress to rise above the critical value needed for bed sediment transport and subglacial bed-accumulated fines are released. These become exhausted during the day, leading to lower concentrations on the falling limb. Superimposed on such daily time-scales there may be a seasonal evolution. As sediment evacuation from glaciers falls to almost zero through winter, but at least some glacial erosion (due to ice deformation) may continue, there may be subglacial sediment accumulation during winter and early spring. This is evacuated and progressively exhausted through the melt season such that the degree of daily clockwise hysteresis may reduce (Clifford et al., 1995; Hodgkins, 1996; Riihimaki et al., 2005; Stott et al., 2014; Mao and Carrillo, 2017). It may also reduce due to the development of more channelized subglacial streams which reduce meltwater access to the bed of the glacier where sediment has accumulated and hence reduces sediment supply (Nienow et al., 1998; Mair et al., 2002; Swift et al., 2002; Kulessa et al., 2008; Gimbert et al., 2016).

Bedload is more difficult to measure and so it is less well known (Mao et al., 2019). Glacial streams are thought to be bedload dominated (Alley et al., 1997). Evidence points to some similarities with suspended sediment dynamics. Bedload transport appears to be controlled by discharge magnitude and variability. Perolo et al. (2019) showed how late season bedload export from an Alpine valley glacier switched on during the day and off during the night in response to variation in discharge and hence shear stress, leading to clockwise bedload transport hysteresis at the daily time-scale. Using a hydraulic model for the whole melt season, they showed that bedload transport capacity was higher when the intensity of diurnal discharge fluctuation was greater because bedload responded as a non-linear function of discharge over a critical value. Such changes in capacity may not be reflected in actual transport because, as with suspended sediment, there may be exhaustion effects at both the daily timescale but also at the seasonal timescale. However, such processes have rarely been studied.

The regular rise and fall of stream discharge, and hence bed shear stress, results in rates of morphodynamic change that are elevated as compared with non-glacial streams. Provided there is enough lateral accommodation space, high rates of sediment supply coupled to discharges that frequently exceed the threshold at which sediments start to move, lead to braided channel patterns. This may be reinforced by relatively low rates of ecosystem engineering due to vegetation, given climate conditions. Channel reworking rates have been shown to be a function of combined discharge and sediment supply variability (Lane et al., 1996) and give rise to a complex template of channels, submerged bedforms and exposed gravel bars (Germanoski and Schumm, 1993). Due to data collection challenges there are no studies that have quantified reworking rates at high frequency over an entire melt season. Bakker et al. (2019) presented data for a three week period, calculating bedload transport from measured erosion and deposition patterns. Figure 2 shows how bedload transport (and hence erosion and deposition) occurs over a number of different time-scales and that during their 3 week study there was no part of the river occupied by water that was not reworked. This is of particular importance for biofilm development as rates of disturbance are so high that it is only likely to be in the early and late summer, when channel reworking rates fall, that there will be enough stability for biofilm development.



Figure 2: Transport frequency in an Alpine braided system showing the frequency of disturbances for 15 surveys of a river over a 21-day period (after Bakker et al., 2019). The x- and y-scales are given in meters.

Away from the morphodynamically active zone, zones that were previously active may become stable and allow vegetation succession (Figure 3). These are commonly on terraces, where incision of the active zone has reduced inundation and hence sediment transport and channel change frequency (Thompson and Jones, 1986; Thompson, 1988; Germanosky and Schumm, 1993; Marren, 2002; Marren and Toomath, 2013; 2014; Roussel et al., 2018). Terrace formation depends upon the balance between sediment supply and sediment transport capacity (and hence glacier melt). With glacier recession, there is evidence (de Winter et al., 2012; Cordier et al., 2017; Roussel et al., 2018) that sediment supply does not keep up with capacity. The result is typically incision in front of retreating glacier margins, which provides sediment necessary for downstream aggradation (incision-aggradation, Marren, 2002; Beylich et al., 2009; Marren and Toomath, 2014; Roussel et al., 2018), although the extent to which this happens depends on valley slope and on whether or not proglacial lakes form that can disconnect downstream sediment delivery (Bogen et al., 2015). But, conditions have been observed where supply can keep up with capacity increases and aggradation is observed throughout the forefield (Curran et al., 2017). In flume experiments, Germanoski and Schumm (1993) showed that channel patterns associated with incision-aggradation are nearly always braided in aggradational zones, but may be braided or straight in incision zones, depending on incision rate; incising reaches invariably produce terraces, but higher incision rates are needed for a straight channel.

Given the apparent importance of more stable zones in supporting biofilm development (see Miller and Lane, 2019 and also Figures 3 and 5), this terrace formation is important. However, it is unlikely to be sufficient for biofilm development and vegetation succession. Glaciogenic deposits are typically extremely well drained and terraces, higher than the morphodynamically active river may be hydrologically stressed. To support ecological processes and succession, including biofilm development, water accessibility remains crucial (Rydgren et al., 2014) which implies that terraces need to be hydrologically connected, whether to surface water or groundwater (see Miller and Lane, 2019). This makes ecosystem processes very dependent upon water sources besides those derived from the morphodynamically active stream itself, especially as incision is likely to have decoupled such terraces from groundwater supply by the mainstream. Such terraces are more likely to be dependent on snow-melt fed or spring-fed streams supplying water from the hillslopes (e.g., Malard et al., 1999; Ward et al., 1999) or larger-scale groundwater transfers. Snow-melt sources also tend to be seasonally-variable (Malard et al., 1999) and evidence (Malard et al., 1999) suggests that in summertime terraces are supported by the hillslope-derived groundwater.





4.2 Proposed model of biofilms in glacier floodplain ecosystem ontogeny

From theory to reality

Previous sections showed how biofilms can act both as biostabilizators and fertilizers. In an ideal ecosystem, with the absence of abiotic disturbances both fertilization and biostabilization effects will increase with time (Figure 4A). The more stable the sediments are, the more biofilms will expand, grow and stabilize (Figure 1), and the more these effects will extend laterally to surrounding areas. The more nutrients captured within the biofilm matrix, the more enzymes will be produced, biofilm growth enhanced, and fertilization increased. These two processes should be complementary to one another and create a positive feedback loop that represents the first of the two types of ecosystem engineering identified above.

In theory, sediment fertilization allows microorganism and small plants to grow, and roots will enhance the stabilisation of these sediments. Biostabilization should enhance fertilization by making biofilms and sediments less sensitive to erosion and by allowing the biofilm to store nutrients within its matrix and in sediments. This is the second type of bioengineering identified above. However, glacier-fed streams are continuously exposed to high-flow induced disturbances that may counter the potential effects of biostabilization. There are also questions that are yet unresolved regarding the extent to which biostabilization can really increase τ_{oc} sufficiently. The typical magnitude of the increases in τ_{oc} reported for the fine sediments typical of tidal environments are much lower than the magnitudes of the τ_{oc} values reported for the sediments, gravels and coarser (e.g., 100 to 150 Nm⁻²) typical of glacial floodplain material. Thus, the biostabilization effects may be variable spatially, more important where there is sandy or silty material, which itself is more easily erodible and less likely to be stable. Licz

John Wiley & Sons



Figure 4: A) Ideal model where disturbances do not occur during biofilm development leading to an exponential increase of habitability; B) Conceptual model of the variation in habitability in relation with stream discharge (Q) in the active floodplain. The black dashed line is the discharge (Q), the solid red line is the habitability, the blue dashed line represents the case in which habitability is increased during the winter time; C) Conceptual model of the variation in habitability in relation with stream discharge (Q) in the abandoned channels.

607 Active floodplain

As highlighted in Section 4.1, stream dynamics continuously shape and re-shape glacial forefields. This reworking may be seasonal, with periods of higher and lower rates of reworking that follow glacial hydrological and sediment supply regimes. Four periods can be identified and related to potential biofilm ecosystem engineering, namely summer, autumn, winter and spring (Figure 4B). These periods don't map onto normal (e.g. astronomical or meteorological) definitions of the seasons and will vary substantially from year to year: for instance, at altitudes of 2000 m or higher, snowmelt in the glacial forefield may start as early as May or as late as early July depending on altitude and orientation (and hence shading); similarly, permanent snowfall in winter may occur as early as late September or as late as early December.

Summer experiences the highest discharges because of ice melt (Nienow et al., 1998). Additionally, summertime has intense diurnal flow variations (Nienow et al., 1998; Swift et al., 2005; Lane and Nienow, 2019). The increase in peak daily discharge and in the intensity of diurnal discharge variation through the melt season, which also drives high rates of sediment delivery (Perolo et al., 2019), leads to the morphodynamically most active period. Not only may such activity erode the sediments to which biofilms are bound, the high sediment transport rate (Milner and Petts, 1994) and flow shear stresses (Cullis et al., 2014) may scour biofilms from bed sediments. Because disturbances are frequent, biofilms cannot fully develop and they cannot impact stream habitability. The time between two subsequent disturbances is too short to allow biofilms to reach a state of maturity (Thom et al., 2015). Thus, during the summer, biofilm development tends to be restricted to morphodynamically inactive zones (e.g., terraces; see below) and then limited by access to other resources, notably water.

- When autumn approaches, from late August onwards, temperatures drop as does the discharge due to reduced melt. Although flow variation is commonly still important (Lane and Nienow, 2019), peak flows are commonly reduced and the rate of morphodynamic change falls. As a result, the size of the morphodynamically active zone contracts (Malard et al., 1999). Less-incised channels receive less discharge compared with well-incised ones where the bulk water flows preferentially, resulting in lower shear stress and greater probability that flow depth is less than the euphotic depth and solar radiation can reach the stream bed. Provided such channels are stable, biofilms may develop and increase habitability because irreversible attachment and EPS secretion is possible (Stoodley et al., 2002). Whether or not this has a positive feedback on stability of such streams is not yet known. The increase in habitability continues until the first significant snow which pauses photosynthesis. The latter may also be reduced by the progressive reduction in solar radiation exacerbated in Alpine valley due to relief-driven effects on shading. We know little about how photosynthetic activity evolves as a function of time in such streams during the transition from late summer through autumn to early winter.
- In winter, and even if there are almost no disturbances, we predict that biofilms cannot increase the habitability of the forefields, mainly because of the absence of light, which precludes phototrophic activity. This impedes the provision of energy for other organisms (Lavandier and Décamps, 1984), and given the nutrient depleted nature of glacier forefields (Bardgett and Walker, 2004; Bardgett et al., 2007), there should be few opportunities for chemotrophs to develop. That said, however, some chemotrophic activity could develop during winter if biofilms have accumulated sufficient nutrients in autumn to sustain their metabolic needs when light is no longer available. Coupled with longer periods of stability and appropriate water supply by groundwater (Malard et al., 1999), this should lead biofilms to reach a sufficient state of maturity, able to increase the habitability to its theoretical maximum. This hypothesis is relevant and is one reason why potential consumers of biofilms, such as macroinvertebrate communities, reach their maximum richness during winter in Alpine streams (Gabbud et al., 2019).
- In spring, solar radiation and temperature rise leading to snowmelt. The presence of snow in the basin may attenuate runoff significantly leading to low diurnal flow fluctuations (Lane and Nienow, 2019) with relatively high baseflows. Runoff is typically lower in terms of suspended

sediment concentrations, until glacier melt starts. Provided discharges are not sufficient enough to cause τ_{oc} to be exceeded, low levels of morphodynamic activity should reduce disturbance which, coupled with high solar radiation intensity once the snowpack disappears, produces ideal conditions for biofilms to increase habitability. We might expect some spatial variability in this process, with the less incised channels being more morphodynamically stable. This "window of opportunity" begins to close as the melt season progresses, diurnal discharge variation becomes more intense, and rates of morphodynamic change increase. Whether or not this increase in habitability has any longer term effect is a moot point as it depends on the extent to which subsequent morphodynamic activity destroys biofilm communities.

Lateral gradients

As noted above, because of the potential for continuous reworking of the active floodplain, it is the abandoned channels located on terraces where it is most likely that biofilms will lead to a long-term increase in habitability. Such channels are likely to be only very rarely (if ever) inundated by the main stream due to the associated elevation differences. For instance, in the Glacier d'Otemma (Valais, Switzerland), Miller and Lane (2019) found that c. 1 m height difference separated the active plain from one terrace on which very developed biofilms were observed (Figure 5) and this surface could not be inundated even at the highest discharges measured. Given their less disturbed nature, abandoned channels on terraces should support biofilm development and so ecosystem engineering (Figures 4C and 5). Moisture cannot be guaranteed, but may be supported by groundwater (Malard et al., 1999), by hillslope-derived flows, or by the positive feedbacks following biofilm development, which reduces vertical infiltration of water (Miller and Lane, 2019).

In summer water sourced from groundwater and hillslopes will not scour biofilm to a significant degree. Disturbances are small in magnitude and the time between two of them may be long. Hence, biofilms can grow, reach a state of maturity and increase the local habitability to greater levels as compared with the active floodplain. The feedbacks triggered by fertilization and biostabilization can propagate into vegetation succession with evidence that certain plants (i.e. debris vegetation), such as Saxifraga Aizodes, Trifolium Badium, Poa Alpina and Festuca Sp., colonize preferentially near channel zones where biofilms create a perched water table (Figure 5B). The only process that can counter this development is lateral erosion of terraces by migration of the morphodynamically active zone.

In autumn, habitability falls progressively as solar radiation reduces until there is a permanent snow cover and with habitability eventually dropping to winter levels. With the onset of permanent snow cover, the environment should become more suitable for chemotrophs that might profit from nutrients that accumulated before snowfall. During winter, predictions made for the active floodplain should apply to channels located on terraces. In spring, the behavior is similar to that explained in the active floodplain, but the habitability reaches greater levels.

In summary, as terraces are sites of low disturbance magnitude and frequency, they may allow for ecosystem engineering by biofilms provided that there is sufficient moisture available, commonly related to non-glacial runoff and groundwater. Here we have identified

an important additional engineering effect of biofilms in glacial floodplains, which is the effect of biofilms on drainage (Figure 5B), crucially important for vegetation establishment (Rydgren et al., 2014) given the ease of drainage of glaciogenic sediments. In that sense, it is plausible that after glacier recession (the surfaces of Figure 5 were still covered by ice in the mid 1990s) and terrace formation, biofilms have played a crucial role in maintaining water at the surface and allowing vegetation to develop progressively in the years after terrace formation. This is of particular interest given that pioneering vegetation has not been able to develop in other proximal zones of the Otemma forefield over the last 30 years, and it has been shown (e.g., Burga et al., 2010) that the first plants can appear as few as ten years after deglaciation. Indeed, it is possible that the effects of biofilms on drainage may be significantly greater than the effects of biofilms on stabilization of sediment, given the size of the glaciogenic sediments that have to be stabilized. These observations emphasize a need to test hypotheses regarding biostabilization and drainage impacts of biofilms in glacial environments, and to include studies of groundwater in such analyses.



Figure 5: A) Biofilm formation in a stream on a terrace in the Val d'Otemma, Valais, Switzerland; B) A biofilm mat that has formed in a stream in the Val d'Otemma supplied with hillslope-sourced groundwater showing successional colonization of the channel margin by vegetation. The stream is on a terrace about 1 m above the morphodynamically active channel. Note to the right there are stable bar surfaces, also on the terrace, but largely void of primary production due to severe moisture limitations related to well-drained glaciogenic sediments.

736 Longitudinal effects

These latitudinal effects need to be put into context. The glacial signals of colder water,
 discharge variation and sediment supply will all attenuate with distance downstream from a

 WIREs Water

glacier (e.g., Milner and Petts, 1994; Füreder, 1999; Burgherr et al., 2002; Uehlinger et al., 2003). Thus, there is likely to be a progressive reduction in the constraints on biofilm development that results in enhanced biofilm presence, greater rates of biofilm development and increasing habitability. This has been demonstrated for macroinvertebrates by showing that moving away from the glacier snout (i.e. downstream) results in the increase of species richness and diversity (Finn et al., 2010).

We might conceptualize this as some kind of cusp model: as these constraints reduce downstream, there may be some sort of threshold at which the positive feedbacks that follow from biofilm development become sufficiently strong that they can support vegetation succession. The ecosystem engineering effects that then follow from this succession then start to create more stable zones, more capable of retaining water, where biofilm-driven habitability can increase further.

How can biofilm promote the ontogeny of glacial floodplains?

The above model suggests a need to think critically about the idea that biofilms are simple agents of habitability in glacial floodplains. As discussed above, there is no evidence of an exponential growth in habitability with time. At the same time, there is no evidence to believe that in the active floodplain biofilms can develop enough to promote habitability. By contrast, it appears that in abandoned and less disturbed channels, notably on terraces, the habitability can increase to a favorable point, which means that biofilms can grow up and increase their ecosystem engineering effects. Visual observations (see Figures 3C and 5) and research (Miller and Lane, 2019) confirm that vegetation develops along channels. One might think that this is a result of the presence of flowing water. However, we argue that this view might be limited by the nature of these creeks that tend to dry out gradually during the summer (e.g., Malard et al., 1999), both by infiltration and exhaustion of solid water sources (i.e. the snow pack); a phenomenon that should reduce the development of vegetation. Thus, biofilms may be important in determining where vegetation can develop (Figure 5; Miller and Lane, 2019). Additionally, biofilms should increase the amount of soil nutrients available for plants to grow (Kaštovská et al., 2005; Schulz et al., 2013; Cicazzo et al., 2016), which further explains why terraces are not fully covered by vegetation (see the upper-right part of Figure 5B). This effect may be a lateral one related to terraces but also a longitudinal one due to the attenuation described above.





Figure 6: Floodplain ontogeny. A) Floodplain at time t_0 biofilm is no longer developed; B) Floodplain at time t_1 biofilm is developing and coalescing in bigger colonies in abandoned channels, commonly on terraces; C) Floodplain at time t_2 . There is vegetation where there were biofilms; D) Floodplain at time t_3 . Biofilms develop in the new abandoned channels; E) Floodplain at time t_4 . New branches are now disconnected; F) Floodplain at time t_5 . There is vegetation where were biofilms; G) Floodplain at time t_6 . Vegetation has colonized more floodplain space.

Figure 6 proposes a model for the ontogeny of glacial floodplain ecosystems in relation to biofilms. At the very beginning, the floodplain has a fully-developed braided system, where biofilms do not develop because of the high rate of disturbances (Figure 6A). As mentioned above, there could be some biofilm development in less incised channels during autumn and spring, but this phenomenon is ephemeral and systematically reset by stream dynamics in summer.

56789As the floodplain evolves, some surfaces are eroded and terraces form (e.g., Thompson and57790Jones, 1986), becoming disconnected from the main active plain (Figure 6B). Because these58791abandoned channels are no longer heavily perturbed by stream dynamics, biofilms are able

60

to develop. The water needed to sustain biofilm development is provided mainly by hillslope groundwater (Malard et al., 1999), which is also thermally less variable (Ward, 1994), less turbid and enriched in nutrients (Brown et al., 2007). These conditions allow biofilms to trigger their engineering feedbacks, in which biostabilization increases fertilization, and vice versa. These feedbacks drive an increase of the organic matter content of sediments (Miller and Lane, 2019), promoting primary succession (Raab et al., 2012; Cicazzo et al., 2016) (Figure 6C). Additionally, biofilms can also promote moisture retention by reducing the permeability of channel beds (Miller and Lane, 2019; Figure 5), which reduces loss of water through drainage. This is of importance due to the low permeability of glaciogenic sediment (Burga et al., 2010). The additional engineering effect can be important for maintaining soil moisture if groundwater availability is reduced. Hence, by maintaining water at the surface of channels, biofilms can self-sustain their growth but also promote vegetation succession. Such a phenomenon has been recorded in Glacier d'Otemma forefield, where vegetation seems to grow preferentially along the impermeabilized channels (Figure 5B, Miller and Lane, 2019).

As the floodplain evolves, new surfaces are created by terrace formation, which increases the space for biofilm engineering and vegetation development (Figure 6D, E, F). Theoretically, these processes lead to a decrease of the accommodation space of the braided system. Incision will then create new terraces, further reducing the accommodation space for active braiding and leading to the formation of a more straight single channel (Germanosky and Schumm, 1993). This process may be aided by glacier recession which increases the distance available for the attenuation of glacier flow, sediment and temperature signals.

This overall ecological process is triggered because microbial biofilms not only have the ability to uptake nutrients but also to store them within the biofilm matrix. Overtime, these available nutrients will also be stored in the surrounding soil environment (Kaštovská et al., 2005; Schmidt et al., 2008; Schulz et al., 2013), leading to an enrichment of soil with phosphorous, carbon and nitrogen. Soil enrichment may take some time to occur (already in the first 4 years after deglaciation; Schmidt et al., 2008), however it is a crucial step for glacier floodplain fertilization and succession. By adding nutrients to the soil, biofilms modify soil nutrient content allowing successional processes (Kaštovská et al., 2005; Zumsteg et al., 2012; Raab et al., 2012; Frey et al., 2013; Miller and Lane, 2019). Again, this process provides positive feedbacks where biofilm growth leads to more nutrient storage in soil, enhanced sediment stabilization and greater moisture retention. With stabilization and soil enrichment, succession and vegetation development will occur (already in the first 10 years after deglaciation; Burga et al., 2010), further reinforcing the stabilization of sediment and biofilm growth. As architects of soils, microbes are responsible for bedrock material weathering, the source of nutrients in vegetation free sites, potential surface biostabilization and important changes in near surface hydrology (Schulz et al., 2013).

As mentioned by Miller and Lane (2019), the roles of biofilms and vegetation should not be seen separately; they must be linked because the former interact with the latter and vice versa. Our model predicts that biofilms have a primordial role in fertilizing the substrate of abandoned channels on fluvial terraces, making other resources (notably water) available and possibly stabilizing sediment. As vegetation develops, the system becomes more stable,

2								
3	839	as well as	more enriched	in organic	matter. Our model	predicts	also that the	habitability
4	840	provided	by	biofilms	reduces	as	winter	comes.
6								
7								
, 8								
9								
10								
11								
12								
13								
14								
15								
16								
17								
18								
19								
20								
21								
23								
24								
25								
26								
27								
28								
29								
30								
31								
32								
33 24								
24 25								
36								
37								
38								
39								
40								
41								
42								
43								
44 45								
45 46								
40 47								
47								
49								
50								
51								
52								
53								
54								
55								
56								
57								
58								
59								
00								26

Conclusion

This review paper has highlighted how biofilms may engineer the environment through fertilization and biostabilization, and there are no reasons to exclude them from a list of potential ecosystem engineers just because their engineering is not as visible as that of other organisms or because of their relative small size compared with other ecosystem engineers. Furthermore, it has been argued that it is not accurate to evaluate biostabilization without accounting for fertilization, and vice versa, because they both interact together; and that the effects of biofilms on the biostabilization of glaciogenic deposits have not been proved. This is particularly important in scientific terms because it brings together scientists from different domains, such as geomorphology, microbiology and geochemistry.

- We also proposed a model for glacial floodplain ecosystem ontogeny that accounts for the engineering effects of biofilms by showing that biofilms are a means of promoting ecological succession in abandoned channels. Their pioneering role allows vegetation to install because conditions become less harsh. As a result, vegetation becomes ecologically relevant. For this reason, it is important to look at the entire successional process instead of looking at a single biological community. It is probably better to consider ecological succession as an extended organism; it is not the single organism (e.g., biofilms and vegetation) that can explain why barren soils become prone to life and then colonized, but the organic whole (i.e. extended organism) acting at each stage. Hence, we argue that it can be erroneous to evaluate the process of ecological succession by investigating one single stage or one single community, because each stage relies on the previous one and each community has synergies with another, and so on. This is the reason why the role of biofilms in ecological succession in deglaciated terrain should be considered as an extended ecosystem engineer that modifies the environment with the ultimate goal to transform it from a hostile one to a livable one.
- That said, this model raises a number of questions that need to be investigated. First, we know very little about the biostabilization effects of biofilms in these kinds of environments. Second, we need to consider the timescales of ecosystem succession in relation to the timescales of biofilm development and their engineering effects. Third, we need to validate the latitudinal and longitudinal impacts of biofilms and how these influence ecosystem succession holistically over decadal timescales. Fourth, we need to consider whether vegetation can develop without biofilm engineering in these environments. Finally, we need to look at biofilms at the annual timescale and not just the spring or summer timescale. We need to understand what happens under snow to habitability, for instance. We need to monitor stream morphodynamics continually in parallel with the continuous monitoring of biofilm development and their ecosystem engineering functions. Through such intensive study, we may develop a much better understanding of the ecosystem engineering of biofilms in glacier floodplain ecosystems.

882 Funding Information

This study is benefited through the financial support from SNF Project CRSII5_180241 *ENSEMBLE* awarded to T. I. Battin, S. N. Lane, M. Lever and P. Wilmes.

885 Acknowledgments

We would like to acknowledge Senior Editor Dr. Jan Seibert, Associate Editor Gemma Harvey and two anonymous reviewers for the detailed and critical but constructive comments and suggestions provided on earlier version of this article. We would like to acknowledge Mr. Alexandre Simard for his contribution to the graphical abstract and Figure 1 and PhD Student Elisa Giaccone for having given us useful information about Otemma's vegetation. This research is supported by Swiss National Science Foundation Synergia project ENSEMBLE (CRSII5-180241/1).

894 References

- Alley, R. B., Cuffey, K. M., Evenson, E. B., Strasser, J. C., Lawson, D. E., & Larson, G. J.
 (1997). How glaciers entrain and transport basal sediment: Physical constraints.
 Quaternary Science Reviews, 16(9), 1017–1038. https://doi.org/10.1016/S0277 3791(97)00034-6
- Amos, C. L., Bergamasco, A., Umgiesser, G., Cappucci, S., Cloutier, D., DeNat, L., ...
 900
 901
 901
 901
 902
 911–241. https://doi.org/10.1016/j.jmarsys.2004.05.013
 924
 - An, C., Parker, G., Hassan, M. A., & Fu, X. (2019). Can magic sand cause massive
 degradation of a gravel-bed river at the decadal scale? Shi-ting River, China.
 Geomorphology, 327, 147–158. https://doi.org/10.1016/j.geomorph.2018.10.026
- Ashworth, P. J., & Ferguson, R. I. (1986). Interrelationships of Channel Processes, Changes and Sediments in a Proglacial Braided River. Geografiska Annaler: Series A, Physical Geography, 68(4). 361-371. https://doi.org/10.1080/04353676.1986.11880186
- 41 910 Aufdenkampe, A. K., Mayorga, E., Raymond, P. A., Melack, J. M., Doney, S. C., Alin, S. R.,
 42 911 ... & Yoo, K. (2011). Riverine coupling of biogeochemical cycles between land,
 43 912 oceans, and atmosphere. *Frontiers in Ecology and the Environment*, 9(1), 53-60.
 45 913 https://doi.org/10.1890/100014
- 914 Bakker, M., Antoniazza, G., Odermatt, E., & Lane, S. N. (2019). Morphological Response of
 915 an Alpine Braided Reach to Sediment-Laden Flow Events. *Journal of Geophysical* 916 *Research: Earth Surface*. https://doi.org/10.1029/2018JF004811
- Bardgett, R. D., & Walker, L. R. (2004). Impact of coloniser plant species on the development of decomposer microbial communities following deglaciation. Soil Bioloav and Biochemistry, 555-559. 36(3), https://doi.org/10.1016/j.soilbio.2003.11.002
- Bardgett Richard D, Richter Andreas, Bol Roland, Garnett Mark H, Bäumler Rupert, Xu
 Xingliang, ... Wanek Wolfgang. (2007). Heterotrophic microbial communities use
 ancient carbon following glacial retreat. *Biology Letters*, 3(5), 487–490.
 https://doi.org/10.1098/rsbl.2007.0242

- Battin, Tom J., Kaplan, L. A., Denis Newbold, J., & Hansen, C. M. E. (2003). Contributions of
 microbial biofilms to ecosystem processes in stream mesocosms. *Nature*, *426*(6965),
 439–442. https://doi.org/10.1038/nature02152
- Battin, T. J., Wille, A., Psenner, R., & Richter, A. (2004). Large-scale environmental controls
 on microbial biofilms in high-alpine streams. *Biogeosciences*, *1*(2), 159–171.
- 9
 930 Battin, Tom J., Sloan, W. T., Kjelleberg, S., Daims, H., Head, I. M., Curtis, T. P., & Eberl, L.
 931 (2007). Opinion: Microbial landscapes: new paths to biofilm research. *Nature Reviews Microbiology*, *5*(1), 76–81. https://doi.org/10.1038/nrmicro1556
- Battin, Tom J., Kaplan, L. A., Findlay, S., Hopkinson, C. S., Marti, E., Packman, A. I., ...
 Sabater, F. (2008). Biophysical controls on organic carbon fluxes in fluvial networks.
 Nature Geoscience, *1*(2), 95–100. https://doi.org/10.1038/ngeo101
- Battin, Tom J., Besemer, K., Bengtsson, M. M., Romani, A. M., & Packmann, A. I. (2016).
 The ecology and biogeochemistry of stream biofilms. *Nature Reviews Microbiology*, 14(4), 251–263. https://doi.org/10.1038/nrmicro.2016.15
- 20 939 Bätz, N., Verrecchia, E. P., & Lane, S. N. (2015). Organic matter processing and soil 21 in а braided river 940 evolution system. CATENA, 126, 86-97. 22 941 https://doi.org/10.1016/j.catena.2014.10.013 23
- P42
 P42
 P43
 P43
 P43
 P44
 P44
 P45
 P44
 P46
 P47
 P48
 P48
 P49
 P49
 P41
 P41
 <

²⁷ 945 https://doi.org/10.1111/mec.14696

1 2 3

4

5

6

- 946 Besemer, K., Singer, G., Limberger, R., Chlup, A.-K., Hochedlinger, G., Hödl, I., ... Battin, T.
 947 J. (2007). Biophysical Controls on Community Succession in Stream Biofilms. *Appl.* 948 *Environ. Microbiol.*, 73(15), 4966–4974. https://doi.org/10.1128/AEM.00588-07
- 949 Besemer Katharina, Singer Gabriel, Quince Christopher, Bertuzzo Enrico, Sloan William, &
 950 Battin Tom J. (2013). Headwaters are critical reservoirs of microbial diversity for
 951 fluvial networks. *Proceedings of the Royal Society B: Biological Sciences*, 280(1771),
 952 20131760. https://doi.org/10.1098/rspb.2013.1760
- 37953Besemer, K. (2015). Biodiversity, community structure and function of biofilms in stream38954ecosystems.Research in Microbiology, 166(10), 774–781.39955https://doi.org/10.1016/j.resmic.2015.05.006
- Beylich, A. A., Laute, K., Liermann, S., Hansen, L., Burki, V., Vatne, G., ... Berthling, I. 956 41 42 957 (2009). Subrecent sediment dynamics and sediment budget of the braided sandur 43 958 system at Sandane, Erdalen (Nordfjord, Western Norway). Norsk Geografisk 44 959 Tidsskrift Norwegian Journal of Geography, 63(2), 123-131. 45 https://doi.org/10.1080/00291950902907934 960 46
- Bogen, J., Xu, M., & Kennie, P. (2015). The impact of pro-glacial lakes on downstream
 sediment delivery in Norway: THE IMPACT OF PRO-GLACIAL LAKES. *Earth Surface Processes and Landforms*, 40(7), 942–952. https://doi.org/10.1002/esp.3669
- 50 964 Boogert, N. J., Paterson, D. M., & Laland, K. N. (2006). The Implications of and Ecosystem 51 965 Engineering for Conservation Biology. BioScience, 56(7), 570-578. 52 966 https://doi.org/10.1641/0006-3568(2006)56[570:TIONCA]2.0.CO:2 53
- 54
 967
 Brown, L. E., Milner, A. M., & Hannah, D. M. (2007). Groundwater influence on alpine stream

 55
 968
 ecosystems. Freshwater Biology, 52(5), 878–890. https://doi.org/10.1111/j.1365

 56
 969
 2427.2007.01739.x
- ⁵⁸ 970 Burga, C. A., Krüsi, B., Egli, M., Wernli, M., Elsener, S., Ziefle, M., ... Mavris, C. (2010).
 ⁵⁹ 971 Plant succession and soil development on the foreland of the Morteratsch glacier
 ⁶⁰ 29

1		
∠ 3	072	(Destroping Switzerland): Stroight forward or chaptic? Flore Marshales
4	972	(Pontresina, Switzerland): Straight forward or chaotic? Flora - Morphology,
5	973	Distribution, Functional Ecology of Plants, 205(9), 561-576.
6	974	nttps://doi.org/10.1016/j.tiora.2009.10.001
7	975	Burgnerr, P., Ward, J. V., & Robinson, C. T. (2002). Seasonal variation in zoobenthos across
8 Q	976	habitat gradients in an alpine glacial floodplain (Val Roseg, Swiss Alps). Journal of
10	977	the North American Benthological Society, 21(4), 561–575.
11	978	https://doi.org/10.2307/1468430
12	979	Cardinale, B. J., Gelmann, E. R., & Palmer, M. A. (2004). Net spinning caddisflies as stream
13	980	ecosystem engineers: the influence of Hydropsyche on benthic substrate stability.
14 15	981	<i>Functional Ecology</i> , 18(3), 381–387. https://doi.org/10.1111/j.0269-
16	982	8463.2004.00865.x
17	983	Carrivick, J. L., Heckmann, T., Turner, A., & Fischer, M. (2018). An assessment of landform
18	984	composition and functioning with the first proglacial systems dataset of the central
19	985	European Alps. <i>Geomorphology</i> , 321, 117–128.
20	986	https://doi.org/10.1016/j.geomorph.2018.08.030
21 22	987	Carter, N. (1932). A Comparative Study of the Alga Flora of Two Salt Marshes. Part I.
23	988	<i>Journal of Ecology</i> , 20(2), 341–370. https://doi.org/10.2307/2256083
24	989	Carter, N. (1933a). A Comparative Study of the Alga Flora of Two Salt Marshes. Part II.
25	990	<i>Journal of Ecology</i> , <i>21</i> (1), 128–208. https://doi.org/10.2307/2255878
26	991	Carter, N. (1933b). A Comparative Study of the Alga Flora of Two Salt Marshes. Part III.
27	992	Journal of Ecology, 21(2), 385–403. https://doi.org/10.2307/2256588
20 29	993	Chapin, F. S., Walker, L. R., Fastie, C. L., & Sharman, L. C. (1994). Mechanisms of Primary
30	994	Succession Following Deglaciation at Glacier Bay, Alaska. Ecological Monographs,
31	995	64(2), 149–175. https://doi.org/10.2307/2937039
32	996	Chen, X. D., Zhang, C. K., Zhou, Z., Gong, Z., Zhou, J. J., Tao, J. F., Feng, Q. (2017).
33 24	997	Stabilizing Effects of Bacterial Biofilms: EPS Penetration and Redistribution of Bed
54 35	998	Stability Down the Sediment Profile: EPS Penetration Down Sediment Profile.
36	999	Journal of Geophysical Research: Biogeosciences, 122(12), 3113–3125.
37	1000	https://doi.org/10.1002/2017JG004050
38	1001	Chen, X., Zhang, C., Paterson, D. M., Townend, I. H., Jin, C., Zhou, Z., Feng, Q. (2019).
39	1002	The effect of cyclic variation of shear stress on non-cohesive sediment stabilization
40 41	1003	by microbial biofilms: the role of 'biofilm precursors', Earth Surface Processes and
42	1004	Landforms, 0(0), https://doi.org/10.1002/esp.4573
43	1005	Cherif M & Loreau M (2009) When microbes and consumers determine the limiting
44	1006	nutrient of autotrophs: a theoretical analysis Proceedings of the Royal Society B.
45	1007	Biological Sciences 276(1656) 487–497 https://doi.org/10.1098/rspb.2008.0560
46 47	1008	Ciccazzo S Esposito A Borruso I & Brusetti I (2016) Microbial communities and
47	1000	primary succession in high altitude mountain environments Annals of
49	1005	microbiology 66(1) 43-60 https://doi.org/10.1007/s13213-015-1130-1
50	1010	Clifford N I Richards K S Brown R A & Lane S N (1995) Scales of Variation of
51	1011	Suspended Sediment Concentration and Turbidity in a Clacial Meltwater Stream
52	1012	Coografiaka Annalor: Sorias A Physical Coography 77(1,2) 45.65
53 54	1013	Geographiska Annaler. Series A, Physical Geography, $77(1-2)$, 43-05.
55	1014	Colling C. L. & Ling C. E. (1993). The Effect of Early Opring Durning on Magdation in
56	1015	Duffele Wellows Bulletin of the Terroy Deterior Club 440(4) 474 404
57	1016	Dullalo vvallows. Bullelin of the Forrey Botanical Club, 110(4), 474–481.
58	1111/	
50	1017	Cordior S Adamson K Dolman M Calvot M & Harmond D (2017) Of ice and water

Quaternary fluvial response to glacial forcing. Quaternary Science Reviews, 166, 57-

1 2 3

4

5

6

7 8

9

1019

73. https://doi.org/10.1016/j.quascirev.2017.02.006 1020 Costerton, J W, Cheng, K. J., Geesey, G. G., Ladd, T. I., Nickel, J. C., Dasgupta, M., & 1021 1022 Marrie, T. J. (1987). Bacterial Biofilms in Nature and Disease. Annual Review of Microbiology, 41(1), 435–464. https://doi.org/10.1146/annurev.mi.41.100187.002251 1023 Costerton, J. William, Lewandowski, Z., Caldwell, D. E., Korber, D. R., & Lappin-Scott, H. M. 1024 10 (1995). Microbial Biofilms. Annual Review of Microbiology, 49(1), 711-745. 1025 11 1026 https://doi.org/10.1146/annurev.mi.49.100195.003431 12 Crisp, D. T., & Carling, P. A. (1989). Observations on siting, dimensions and structure of 1027 13 14 1028 salmonid redds. Journal of Fish Biology, 34(1), 119-134. 15 https://doi.org/10.1111/j.1095-8649.1989.tb02962.x 1029 16 Cullis, J. D. S., Stanish, L. F., & McKnight, D. M. (2014). Diel flow pulses drive particulate 1030 17 1031 organic matter transport from microbial mats in a glacial meltwater stream in the 18 19 1032 McMurdo Dry Valleys. Water Resources Research, 50(1), 86-97. 20 https://doi.org/10.1002/2013WR014061 1033 21 Curran, J. H., Loso, M. G., & Williams, H. B. (2017). Glacial conditioning of stream position 1034 22 1035 and flooding in the braid plain of the Exit Glacier foreland, Alaska. Geomorphology, 23 293, 272-288. https://doi.org/10.1016/j.geomorph.2017.06.004 1036 24 25 Dade, W. B., Davis, J. D., Nichols, P. D., Nowell, A. R. M., Thistle, D., Trexler, M. B., & 1037 26 1038 White, D. C. (1990). Effects of bacterial exopolymer adhesion on the entrainment of 27 1039 sand. Geomicrobiology Journal. 1-16. 8(1), 28 https://doi.org/10.1080/01490459009377874 1040 29 Dawkins, R. (1982). The extended phenotype. Oxford: Oxford University Press. 1041 30 31 1042 de Boer, P. L. de. (1981). Mechanical effects of micro-organisms on intertidal bedform 32 1043 migration*. Sedimentology, 28(1), 129–132. https://doi.org/10.1111/j.1365-33 3091.1981.tb01670.x 1044 34 de Winter, I. L., Storms, J. E. A., & Overeem, I. (2012). Numerical modeling of glacial 1045 35 sediment production and transport during deglaciation. Geomorphology, 167-168, 36 1046 37 1047 102-114. https://doi.org/10.1016/j.geomorph.2012.05.023 38 Demars, B. O. (2018). Hydrological pulses and burning of dissolved organic carbon by 1048 39 respiration. *Limnology* Oceanography, 406-421. 1049 stream and 64(1), 40 1050 doi:10.1002/Ino.11048 41 Elwood, J. W., Newbold, J. D., Trimble, A. F., & Stark, R. W. (1981). The Limiting Role of 42 1051 43 1052 Phosphorus in a Woodland Stream Ecosystem: Effects of P Enrichment on Leaf 44 Ecology, 1053 Decomposition and Primary Producers. 62(1), 146-158. 45 https://doi.org/10.2307/1936678 1054 46 1055 Fang, H., Shang, Q., Chen, M., & He, G. (2014). Changes in the critical erosion velocity for 47 48 1056 sediment colonized by biofilm. Sedimentology, 61(3), 648-659. 49 https://doi.org/10.1111/sed.12065 1057 50 1058 Fastie, C. L. (1995). Causes and Ecosystem Consequences of Multiple Pathways of Primary 51 1059 Succession at Glacier Bay, Alaska. Ecology, 76(6), 1899-1916. 52 1060 https://doi.org/10.2307/1940722 53 54 Finn, D. S., Räsänen, K., & Robinson, C. T. (2010). Physical and biological changes to a 1061 55 lengthening stream gradient following a decade of rapid glacial recession: SMALLER 1062 56 GLACIERS, LONGER STREAMS. Global Change Biology, 16(12), 3314-3326. 1063 57 https://doi.org/10.1111/j.1365-2486.2009.02160.x 1064 58 59 1065 Flemming, H.-C., & Wingender, J. (2001). Relevance of microbial extracellular polymeric 60 31

1		
2		
3	1066	substances (EPSs) - Part I: Structural and ecological aspects. Water Science and
4 5	1067	<i>Technology</i> , <i>43</i> (6), 1–8. https://doi.org/10.2166/wst.2001.0326
6	1068	Flemming, B. W. (2002). Chapter Six Geographic distribution of muddy coasts. In T. Healy,
7	1069	Y. Wang, & JA. Healy (Eds.), Proceedings in Marine Science (pp. 99–201).
8	1070	https://doi.org/10.1016/S1568-2692(02)80080-8
9	1071	Flemming, HC., Strathmann, M., & Morales, C. F. L. (2007a). Microbial Effects. In B.
10	1072	Westrich & U. Förstner (Eds.). Sediment Dynamics and Pollutant Mobility in Rivers:
11	1073	An Interdisciplinary Approach (pp 343–378) https://doi.org/10.1007/978-3-540-
12	1074	34785-9 9
14	1075	Elemming H Neu T R & Wozniak D J (2007b) The EPS Matrix: The "House of Biofilm
15	1075	Cells" Journal of Bacteriology 189(22) 7945-7947 doi:10.1128/ib.00858-07
16	1070	Elemming H & Wingender I (2010) The biofilm matrix Nature Peviews Microbiology
17	1077	2(0) 622 622 doi:10.1028/prmioro2415
18	1078	o(9), 023-033. UOI. 10. 1030/1111110/02413
19 20	1079	Fiemming, H. C., Wingender, J., Szewzyk, U., Steinberg, P., Rice, S. A., & Kjelleberg, S.
20	1080	(2016). Biotilims: an emergent form of bacterial life. Nature Reviews
22	1081	<i>Microbiology</i> , 14(9), 563. https://doi.org/10.1038/nrmicro.2016.94
23	1082	Flemming, H. C., & Würtz, S. (2019). Bacteria and archaea on Earth and their abundance in
24	1083	biofilms. <i>Nature Reviews Microbiology</i> , 17(4), 247. https://doi.org/10.1038/s41579-
25	1084	019-0158-9
26 27	1085	Fountain, A. G. (1996). Effect of Snow and Firn Hydrology on the Physical and Chemical
27	1086	Characteristics of Glacial Runoff. Hydrological Processes, 10(4), 509–521.
29	1087	https://doi.org/10.1002/(SICI)1099-1085(199604)10:4<509::AID-HYP389>3.0.CO;2-3
30	1088	Freeman, C., & Lock, M. A. (1995). The biofilm polysaccharide matrix: A buffer against
31	1089	changing organic substrate supply? Limnology and Oceanography, 40(2), 273–278.
32	1090	https://doi.org/10.4319/lo.1995.40.2.0273
33 24	1091	Frey, B., Bühler, L., Schmutz, S., Zumsteg, A., & Furrer, G. (2013). Molecular
34 35	1092	characterization of phototrophic microorganisms in the forefield of a receding glacier
36	1093	in the Swiss Alps. Environmental Research Letters, 8(1), 015033.
37	1094	https://doi.org/10.1088/1748-9326/8/1/015033
38	1095	Füreder, L. (1999). High alpine streams: cold habitats for insect larvae. In R. Margesin & F.
39	1096	Schinner (Eds.) Cold-Adapted Organisms (nn. 181–196)
40 41	1090	https://doi.org/10.1007/978-3-662-06285-2.10
41	1097	Gabbud C. Robinson C. T. & Lane S. N. (2019) Summer is in winter: Disturbance-driven
43	1000	shifts in macroinvertebrate communities following hydroelectric power exploitation
44	1100	Science of The Total Environment 650 2164 2180
45	1100	$\frac{104-2100}{2104-2100}$
46	1101	Carboradorf S. I.I. Japaka T. Waatrich P. & Dataroon D. M. (2008a) Microbial
47 10	1102	Gerbersdorf, S., U., Jancke, T., Westneri, B., & Paterson, D. M. (2006a). Microbial
40 49	1103	stabilization of riverine sediments by extracellular polymenc substances. <i>Geobiology</i> ,
50	1104	6(1), 57–69. https://doi.org/10.1111/J.1472-4669.2007.00120.x
51	1105	Gerbersdorf, S., U., Manz, W., & Paterson, D. M. (2008b). The engineering potential of
52	1106	natural benthic bacterial assemblages in terms of the erosion resistance of
53	1107	sediments. FEMS Microbiology Ecology, 66(2), 282–294.
54 57	1108	https://doi.org/10.1111/j.1574-6941.2008.00586.x
55 56	1109	Gerbersdorf, S., U., Bittner, R., Lubarsky, H., Manz, W., & Paterson, D. M. (2009). Microbial
57	1110	assemblages as ecosystem engineers of sediment stability. Journal of Soils and
58	1111	Sediments, 9(6), 640–652. https://doi.org/10.1007/s11368-009-0142-5
59	1112	Gerbersdorf, S. U., & Wieprecht, S. (2015). Biostabilization of cohesive sediments: revisiting
60		32

3 1113 the role of abiotic conditions, physiology and diversity of microbes, polymeric 4 secretion, and biofilm architecture. Geobiology, 13(1), 68-97. 1114 5 https://doi.org/10.1111/gbi.12115 1115 6 Germanoski, D., & Schumm, S. A. (1993). Changes in Braided River Morphology Resulting 7 1116 8 from Aggradation and Degradation. The Journal of Geology, 101(4), 451-466. 1117 9 https://doi.org/10.1086/648239 1118 10 Gimbert, F., Tsai, V. C., Amundson, J. M., Bartholomaus, T. C., & Walter, J. I. (2016). 1119 11 1120 Subseasonal changes observed in subglacial channel pressure, size, and sediment 12 transport: SUBGLACIAL HYDROLOGY FROM SEISMIC NOISE. Geophysical 1121 13 14 Research Letters, 43(8), 3786-3794. https://doi.org/10.1002/2016GL068337 1122 15 Godwin, C. M., & Cotner, J. B. (2018). What intrinsic and extrinsic factors explain the 1123 16 stoichiometric diversity of aquatic heterotrophic bacteria? The ISME Journal, 12(2), 1124 17 598-609. https://doi.org/10.1038/ismej.2017.195 1125 18 19 1126 Grant, J., & Gust, G. (1987). Prediction of coastal sediment stability from photopigment 20 content of mats of purple sulphur bacteria. 330(6145), 1127 Nature. 244. 21 https://doi.org/10.1038/330244a0 1128 22 1129 Gurnell, A. M., & Petts, G. E. (2002). Island-dominated landscapes of large floodplain rivers, 23 1130 а European perspective. Freshwater Biology, 47(4), 581-600. 24 25 https://doi.org/10.1046/j.1365-2427.2002.00923.x 1131 26 Gurnell, A. (2014). Plants as river system engineers. Earth Surface Processes and 1132 27 Landforms, 39(1), 4-25. https://doi.org/10.1002/esp.3397 1133 28 Hastings, A., Byers, J. E., Crooks, J. A., Cuddington, K., Jones, C. G., Lambrinos, J. G., ... 1134 29 1135 Wilson, W. G. (2007). Ecosystem engineering in space and time. Ecology Letters, 30 31 1136 10(2), 153–164. https://doi.org/10.1111/j.1461-0248.2006.00997.x 32 1137 Heckmann, T., McColl, S., & Morche, D. (2016). Retreating ice: research in pro-glacial areas 33 1138 matters. Earth Surface Processes and Landforms, 41(2), 271-276. 34 https://doi.org/10.1002/esp.3858 1139 35 Hodgkins, R. (1996). Seasonal trend in suspended-sediment transport from an Arctic glacier, 36 1140 37 1141 and implications for drainage-system structure. Annals of Glaciology, 22, 147-151. 38 https://doi.org/10.3189/1996AoG22-1-147-151 1142 39 Hodkinson, I. D., Webb, N. R., & Coulson, S. J. (2002). Primary community assembly on 1143 40 land - the missing stages: why are the heterotrophic organisms always there first? 1144 41 42 Journal of Ecology. 90(3), 569-577. https://doi.org/10.1046/j.1365-1145 43 1146 2745.2002.00696.x 44 1147 Hodkinson, I. D., Coulson, S. J., & Webb, N. R. (2003). Community assembly along 45 proglacial chronosequences in the high Arctic: vegetation and soil development in 1148 46 north-west Svalbard. Journal of Ecology, 91(4), 651–663. 47 1149 48 1150 https://doi.org/10.1046/j.1365-2745.2003.00786.x 49 Holland, A. F., Zingmark, R. G., & Dean, J. M. (1974). Quantitative evidence concerning the 1151 50 stabilization of sediments by marine benthic diatoms. Marine Biology, 27(3), 191-1152 51 196. https://doi.org/10.1007/BF00391943 1153 52 Hotaling, S., Finn, D. S., Joseph Giersch, J., Weisrock, D. W., & Jacobsen, D. (2017a). 1154 53 54 Climate change and alpine stream biology: progress, challenges, and opportunities 1155 55 for the future: Climate change and alpine stream biology. Biological Reviews, 92(4), 1156 56 2024-2045. https://doi.org/10.1111/brv.12319 1157 57 Hotaling, S., Hood, E., & Hamilton, T. L. (2017b). Microbial ecology of mountain glacier 1158 58 59 1159 ecosystems: biodiversity, ecological connections and implications of a warming 60 33

3	1160	climate: Microbial ecology of mountain glaciers. Environmental Microbiology, 19(8),
4	1161	2935–2948. https://doi.org/10.1111/1462-2920.13766
5 6	1162	House, R. A., & Boehne, P. L. (1986). Effects of Instream Structures on Salmonid Habitat
7	1163	and Populations in Tobe Creek, Oregon. North American Journal of Fisheries
8	1164	Management, 6(1), 38–46. https://doi.org/10.1577/1548-
9	1165	8659(1986)6<38:EOISOS>2.0.CO:2
10	1166	Hurvn, A. D., Slavik, K. A., Lowe, R. L., Parker, S. M., Anderson, D. S., & Peterson, B. J.
11	1167	(2005) Landscape heterogeneity and the biodiversity of Arctic stream communities.
12	1168	a habitat template analysis. Canadian Journal of Eisheries and Aquatic Sciences
14	1169	62(8) 1905–1919 https://doi.org/10.1139/f05-100
15	1170	Jackson E I Rowden A A Attrill M J Bossey S J & Jones M B (2001) The
16	1171	importance of seagrass beds as a habitat for fishery species Oceanography and
17	1172	marine biology 20, 260, 304
18	1172	Interime biology, 59, 209-504.
19 20	11/3	Johnson, M. F., Reid, I., Rice, S. P., & Wood, P. J. (2009). Stabilization of fine gravers by
21	11/4	net-spinning caddisity larvae. Earth Surface Processes and Landiorms, 34(3), 413-
22	11/5	423. https://doi.org/10.1002/esp.1750
23	11/6	Jones, C. G., Lawton, J. H., & Snachak, M. (1994). Organisms as Ecosystem Engineers.
24	1177	Olkos, 69(3), 373–386. https://doi.org/10.2307/3545850
25	1178	Jones, C. G., Lawton, J. H., & Shachak, M. (1997). Positive and Negative Effects of
20 27	1179	Organisms as Physical Ecosystem Engineers. <i>Ecology</i> , 78(7), 1946–1957.
28	1180	https://doi.org/10.1890/0012-9658(1997)078[1946:PANEOO]2.0.CO;2
29	1181	Kaštovská, K., Elster, J., Stibal, M., & Šantrůčková, H. (2005). Microbial Assemblages in Soil
30	1182	Microbial Succession After Glacial Retreat in Svalbard (High Arctic). Microbial
31	1183	<i>Ecology</i> , <i>50</i> (3), 396–407. https://doi.org/10.1007/s00248-005-0246-4
32	1184	Kaufmann, R. (2001). Invertebrate Succession on an Alpine Glacier Foreland. Ecology,
33 34	1185	82(8), 2261–2278. https://doi.org/10.1890/0012-
35	1186	9658(2001)082[2261:ISOAAG]2.0.CO;2
36	1187	Kulessa, B., Booth, A. D., Hobbs, A., & Hubbard, A. L. (2008). Automated monitoring of
37	1188	subglacial hydrological processes with ground-penetrating radar (GPR) at high
38	1189	temporal resolution: scope and potential pitfalls. Geophysical Research Letters,
39 40	1190	35(24). https://doi.org/10.1029/2008GL035855
40 41	1191	Lane, S. N., & Nienow, P. W. (2019). Decadal-scale climate forcing of Alpine glacial
42	1192	hydrological systems. Water Resources Research. 0(ja).
43	1193	https://doi.org/10.1029/2018WR024206
44	1194	Lane, S. N., Richards, K. S., & Chandler, J. H. (1996). Discharge and sediment supply
45	1195	controls on erosion and deposition in a dynamic alluvial channel. <i>Geomorphology</i>
46 47	1196	15(1) 1–15 https://doi.org/10.1016/0169-555X(95)00113-J
48	1197	Lane S. N. Bakker M. Gabbud C. Micheletti N. & Saugy JN. (2017). Sediment export
49	1198	transient landscape response and catchment-scale connectivity following ranid
50	1100	climate warming and Alpine glacier recession. Geomorphology 277 210–227
51	1200	https://doi.org/10.1016/i.geomorph 2016.02.015
52	1200	Langenheder S. Wang J. Kariolainen S. M. Langenheder S. Wang J. Kariolainen S. M. Langenheder S. Wang J. Kariolainen S. M. Langenheder S. M. Langenheder S. Wang J. Kariolainen S. M. Langenheder S. M. Langenhede
55 54	1201	Langernieuer, S., Wang, J., Karjalanen, S. W., Laamanen, T. W., Tolonen, K. T., Viimi, A., &
55	1202	reino, J. (2010). Bacierial metacommunity organization in a myny-connecteu
56	1203	aqualic system. FEMS Microbiology Ecology, IIw225.
57	1204	nups.//doi.org/10.1093/temsec/fiw225
58	1205	Lawton, J. H., & Jones, C. G. (1993). Linking species and ecosystem perspectives. <i>Trends</i>
59 60	1206	In Ecology & Evolution, $\delta(9)$, 311–313. https://doi.org/10.1016/0169-5347(93)90236-I
00		34

1207 Le Hir, P., Monbet, Y., & Orvain, F. (2007). Sediment erodability in sediment transport 4 1208 modelling: Can we account for biota effects? Continental Shelf Research, 27(8), 5 1116-1142. https://doi.org/10.1016/j.csr.2005.11.016 1209 6 Leopold, L. B., Wolman, M. G., & Miller, J. P. (1964). Fluvial processes in geomorphology. 1210 7 8 Retrieved from https://pubs.er.usgs.gov/publication/70185663 1211 9 Lyautey, E., Jackson, C. R., Cayrou, J., Rols, J.-L., & Garabétian, F. (2005). Bacterial 1212 10 Community Succession in Natural River Biofilm Assemblages. *Microbial Ecology*, 1213 11 1214 50(4), 589-601. https://doi.org/10.1007/s00248-005-5032-9 12 1215 Lyon, D. R., & Ziegler, S. E. (2009). Carbon cycling within epilithic biofilm communities 13 14 across a nutrient gradient of headwater streams. Limnology and Oceanography, 1216 15 54(2), 439-449. https://doi.org/10.4319/lo.2009.54.2.0439 1217 16 Mair, D., Nienow, P., Sharp, M. J., Wohlleben, T., & Willis, I. (2002). Influence of subglacial 1218 17 drainage system evolution on glacier surface motion: Haut Glacier d'Arolla, 1219 18 19 1220 Switzerland. Journal of Geophysical Research, 107(B8). 20 https://doi.org/10.1029/2001JB000514 1221 21 Maizels, J. (2002). 9 - Sediments and landforms of modern proglacial terrestrial 1222 22 1223 environments. In J. Menzies (Ed.), Modern and Past Glacial Environments (pp. 279-23 316). https://doi.org/10.1016/B978-075064226-2/50012-X 1224 24 25 Malard, F., Tockner, K., & Ward, J. V. (1999). Shifting Dominance of Subcatchment Water 1225 26 Sources and Flow Paths in a Glacial Floodplain, Val Roseg, Switzerland. Arctic, 1226 27 1227 Antarctic, Alpine Research, 135-150. and 31(2), 28 https://doi.org/10.1080/15230430.1999.12003291 1228 29 1229 Mao, L., & Carrillo, R. (2017). Temporal dynamics of suspended sediment transport in a 30 31 1230 alacierized Andean basin. Geomorphology, 116-125. 287. 32 1231 https://doi.org/10.1016/j.geomorph.2016.02.003 33 1232 Mao, L., Comiti, F., Carrillo, R., & Penna, D. (2019). Sediment Transport in Proglacial Rivers. 34 In T. Heckmann & D. Morche (Eds.), Geomorphology of Proglacial Systems (pp. 1233 35 199–217). https://doi.org/10.1007/978-3-319-94184-4 12 36 1234 37 1235 Mariotti, G., & Fagherazzi, S. (2012). Modeling the effect of tides and waves on benthic 38 Research: 1236 biofilms. Journal of Geophysical Biogeosciences, 117(G4). 39 https://doi.org/10.1029/2012JG002064 1237 40 1238 Marren, P. M. (2002). Glacier margin fluctuations, Skaftafellsjökull, Iceland: implications for 41 sandur evolution. Boreas. 31(1), 75-81. https://doi.org/10.1111/j.1502-42 1239 43 1240 3885.2002.tb01057.x 44 1241 Marren, P. M. (2005). Magnitude and frequency in proglacial rivers: a geomorphological and 45 sedimentological perspective. Earth-Science Reviews, 70(3), 203-251. 1242 46 https://doi.org/10.1016/j.earscirev.2004.12.002 47 1243 48 1244 Marren, P. M., & Toomath, S. C. (2013). Fluvial adjustments in response to glacier retreat: 49 Skaftafellsjökull, Iceland. Boreas, 42(1), 57-70. https://doi.org/10.1111/j.1502-1245 50 1246 3885.2012.00275.x 51 Marren, P. M., & Toomath, S. C. (2014). Channel pattern of proglacial rivers: topographic 1247 52 1248 forcing due to glacier retreat. Earth Surface Processes and Landforms, 39(7), 943-53 54 1249 951. https://doi.org/10.1002/esp.3545 55 1250 McDowell, D. M., & Naiman, R. J. (1986). Structure and function of a benthic invertebrate 56 (Castor canadensis). Oecologia, 68(4), 481stream community as influenced by 1251 57 489. https://doi.org/10.1007/BF00378759 1252 58 59 1253 Miller, H. R., & Lane, S. N. (2019). Biogeomorphic feedbacks and the ecosystem 60 35

2		
3	1254	engineering of recently deglaciated terrain. Progress in Physical Geography: Earth
4	1255	and Environment, 43(1), 24–45. https://doi.org/10.1177/0309133318816536
5 6	1256	Milner, A. M., & Petts, G. E. (1994). Glacial rivers: physical habitat and ecology. Freshwater
7	1257	Biology. 32(2). 295–307. https://doi.org/10.1111/i.1365-2427.1994.tb01127.x
8	1258	Montgomery, D. R., Buffington, J. M., Peterson, N. P., Schuett-Hames, D., & Quinn, T. P.
9	1259	(1996) Stream-bed scour egg burial depths and the influence of salmonid spawning
10	1260	on bed surface mobility and embryo survival. Canadian Journal of Fisheries and
11	1260	Aquatic Sciences 53(5) 1061-1070 https://doi.org/10.1130/f96-028
12	1201	Moore I. W. (2006) Animal Ecosystem Engineers in Streams, $BioScience, 56(3), 237, 246$
13 14	1202	https://doi.org/10.1641/0006.2568/2006/05660227/AEEIS12.0.CO/2
15	1263	https://doi.org/10.1641/0006-3568(2006)056[0237.AEEIS]2.0.CO,2
16	1264	Mosisch, T. D., Bunn, S. E., & Davies, P. M. (2001). The relative importance of shading and
17	1265	nutrients on algal production in subtropical streams. Freshwater Biology, 46(9),
18	1266	1269–1278. https://doi.org/10.1046/j.1365-2427.2001.00747.x
19	1267	Neumann, A. C., Gebelein, C. D., & Scoffin, T. P. (1970). The Composition, Structure and
20	1268	Erodability of Subtidal Mats, Abaco, Bahamas. Journal of Sedimentary Research,
21	1269	40(1). Retrieved from http://archives.datapages.com/data/sepm/journals/v38-
23	1270	41/data/040/040001/0274.htm
24	1271	Neumeier, U., Lucas, C. H., & Collins, M. (2006). Erodibility and erosion patterns of mudflat
25	1272	sediments investigated using an annular flume. Aquatic Ecology, 40(4), 543-554.
26	1273	https://doi.org/10.1007/s10452-004-0189-8
27	1274	Niederdorfer, R., Peter, H., & Battin, T. J. (2016). Attached biofilms and suspended
28	1275	aggregates are distinct microbial lifestyles emanating from differing hydraulics.
30	1276	Nature Microbiology, 1(12), doi:10.1038/nmicrobiol.2016.178
31	1277	Nienow P Sharp M & Willis I (1998) Seasonal changes in the morphology of the
32	1278	subalacial drainage system Haut Glacier d'Arolla Switzerland Farth Surface
33	1270	Processes and Landforms 23(9) 825–843 https://doi.org/10.1002//SICI)1096-
34	12/9	17000000000000000000000000000000000000
35	1200	Baterson D. M. (1980) Short term changes in the predibility of intertidal cohosive sediments
37	1201	raterson, D. W. (1969). Short-term changes in the elouibility of intertidal conesive sediments
38	1282	24(4) 222 224 https://doi.org/40.4240/ls.1000.24.4.0222
39	1283	34(1), 223–234. https://doi.org/10.4319/10.1989.34.1.0223
40	1284	Paterson, D. M., & Daborn, G. R. (1991). Sediment stabilisation by biological action:
41	1285	significance for coastal engineering. <i>Developments in coastal engineering</i> , 111-119.
42	1286	Paterson, D. M. (1994). Microbiological mediation of sediment structure and behaviour. In L.
45 44	1287	J. Stal & P. Caumette (Eds.), Microbial Mats (pp. 97–109). Springer Berlin
45	1288	Heidelberg.
46	1289	Perolo, P., Bakker, M., Gabbud, C., Moradi, G., Rennie, C., & Lane, S. N. (2019). Subglacial
47	1290	sediment production and snout marginal ice uplift during the late ablation season of a
48	1291	temperate valley glacier. Earth Surface Processes and Landforms, 44(5), 1117-
49	1292	1136. https://doi.org/10.1002/esp.4562
50 51	1293	Pivato, M., Carniello, L., Moro, I., & D'Odorico, P. (n.d.). On the feedback between water
52	1294	turbidity and microphytobenthos growth in shallow tidal environments. Earth Surface
53	1295	Processes and Landforms, 0(0). https://doi.org/10.1002/esp.4567
54	1296	Polvi, L. E., & Sarneel, J. M. (2018). Ecosystem engineers in rivers: An introduction to how
55	1297	and where organisms create positive biogeomorphic feedbacks. Wilev
56	1298	Interdisciplinary Reviews: Water. 5(2), e1271, https://doi.org/10.1002/wat2.1271
5/ 50	1299	Raab, T., Krümmelbein, J., Schneider A. Gerwin W. Maurer T. & Naeth M. A. (2012)
59	1300	Initial Ecosystem Processes as Key Factors of Landscape Development_A Review
60	1300	
		50

Physical Geography, 33(4), 305–343. https://doi.org/10.2747/0272-3646.33.4.305 Reichman, O. J., & Seabloom, E. W. (2002a). Ecosystem engineering: a trivialized concept?: Response from Reichman and Seabloom. Trends in Ecology & Evolution, 17(7), 308. https://doi.org/10.1016/S0169-5347(02)02512-0 Reichman, O. J., & Seabloom, E. W. (2002b). The role of pocket gophers as subterranean Trends in Ecology ecosvstem enaineers. & Evolution. 17(1), 44-49. https://doi.org/10.1016/S0169-5347(01)02329-1 Richards, K. S. (1982). Rivers: form and process in alluvial channels. Retrieved from https://trove.nla.gov.au/version/31361403 Riihimaki, C. A., MacGregor, K. R., Anderson, R. S., Anderson, S. P., & Loso, M. G. (2005). Sediment evacuation and glacial erosion rates at a small alpine glacier. Journal of Geophysical Research, 110(F3). https://doi.org/10.1029/2004JF000189 Romaní, Α. Μ. (2010). Freshwater Biofilms. In Biofouling 137-153). (pp. https://doi.org/10.1002/9781444315462.ch10 Roussel, E., Marren, P. M., Cossart, E., Toumazet, J.-P., Chenet, M., Grancher, D., & Jomelli, V. (2018). Incision and aggradation in proglacial rivers: Post-Little Ice Age long-profile adjustments of Southern Iceland outwash plains. Land Degradation & Development, 29(10), 3753-3771. https://doi.org/10.1002/ldr.3127 Rothman, D. H., & Forney, D. C. (2008). Response to comment on" physical model for the decay and preservation of marine organic carbon". Science, 319(5870), 1616-1616. https://doi.org/10.1126/science.1148678 Rydgren, K., Halvorsen, R., Töpper, J. P., & Njøs, J. M. (2014). Glacier foreland succession and the fading effect of terrain age. Journal of Vegetation Science, 25(6), 1367-1380. https://doi.org/10.1111/jvs.12184 Sauer, K., Camper, A. K., Ehrlich, G. D., Costerton, J. W., & Davies, D. G. (2002). Pseudomonas aeruginosa Displays Multiple Phenotypes during Development as a of 1140-1154. Biofilm. Journal Bacteriology. 184(4). https://doi.org/10.1128/jb.184.4.1140-1154.2002 Schmidt, S. K., Reed, S. C., Nemergut, D. R., Stuart Grandy, A., Cleveland, C. C., Weintraub, M. N., ... & Martin, A. M. (2008). The earliest stages of ecosystem succession in high-elevation (5000 metres above sea level), recently deglaciated soils. Proceedings of the Royal Society B: Biological Sciences, 275(1653), 2793-2802. https://doi.org/10.1098/rspb.2008.0808 Schmidt, H., Thom, M., King, L., Wieprecht, S., & Gerbersdorf, S. U. (2016). The effect of seasonality upon the development of lotic biofilms and microbial biostabilisation. Freshwater Biology, 61(6), 963-978. https://doi.org/10.1111/fwb.12760 Schulz, S., Brankatschk, R., Dümig, A., Kögel-Knabner, I., Schloter, M., & Zeyer, J. (2013). The role of microorganisms at different stages of ecosystem development for soil formation. Biogeosciences, 10(6), 3983-3996. https://doi.org/10.5194/bg-10-3983-Schumm, S. A., & Lichty, R. W. (1965). Time, space, and causality in geomorphology. American Journal of Science, 263(2), 110-119. https://doi.org/10.2475/ajs.263.2.110 Scoffin, T. P. (1970). The Trapping and Binding of Subtidal Carbonate Sediments by Marine Vegetation in Bimini Lagoon, Bahamas. Journal of Sedimentary Research, 40(1). http://archives.datapages.com/data/sepm/journals/v38-Retrieved from 41/data/040/040001/0249.htm Shields, A. (1936). Anwendung der Aehnlichkeitsmechanik und der Turbulenzforschung

- 1348 Simon, A., & Collison, A. J. C. (2002). Quantifying the mechanical and hydrologic effects of riparian vegetation on streambank stability. Earth Surface Processes and Landforms, 1349 27(5), 527-546. https://doi.org/10.1002/esp.325 1350
- Singer, G., Besemer, K., Schmitt-Kopplin, P., Hödl, I., & Battin, T. J. (2010). Physical 1351 heterogeneity increases biofilm resource use and its molecular diversity in stream 1352 mesocosms. PLoS One, 5(4), e9988. https://doi.org/10.1371/journal.pone.0009988 1353
- Soluk, D. A., & Richardson, J. S. (1997). The Role of Stoneflies in Enhancing Growth of 1354 1355 Trout: A Test of the Importance of Predator-Predator Facilitation within a Stream 1356 Community. Oikos, 80(2), 214-219. https://doi.org/10.2307/3546588 13
- 14 Spears, B. M., Saunders, J. E., Davidson, I., & Paterson, D. M. (2008). Microalgal sediment 1357 15 biostabilisation along a salinity gradient in the Eden Estuary, Scotland: unravelling a 1358 16 paradox. Marine and Freshwater Research, 313-321. 1359 59(4), 17 https://doi.org/10.1071/MF07164 1360 18
- 19 1361 Statzner, B., Arens, M.-F., Champagne, J.-Y., Morel, R., & Herouin, E. (1999). Silk-20 producing stream insects and gravel erosion: Significant biological effects on critical 1362 21 Water Research, 35(11), 3495-3506. 1363 shear stress. Resources 22 https://doi.org/10.1029/1999WR900196 1364 23
- Stoodley, P., Sauer, K., Davies, D. G., & Costerton, J. W. (2002). Biofilms as Complex 1365 24 25 Differentiated Communities. Annual Review of Microbiology, 56(1), 187-209. 1366 26 https://doi.org/10.1146/annurev.micro.56.012302.160705 1367
- 27 Stott, T., Nuttall, A.-M., & Biggs, E. (2014). Observed run-off and suspended sediment 1368 28 dynamics from a minor glacierized basin in south-west Greenland. Geografisk 1369 29 1370 Tidsskrift-Danish Journal of Geography, 114(2), 93-108. 30 31 1371 https://doi.org/10.1080/00167223.2013.862911
- 32 1372 Sutherland, I. (2001). The biofilm matrix - an immobilized but dynamic microbial 33 1373 environment. Trends in Microbiology, 9(5), 222-227. https://doi.org/10.1016/S0966-34 842X(01)02012-1 1374 35
- Swift, D. A., Nienow, P. W., Spedding, N., & Hoey, T. B. (2002). Geomorphic implications of 36 1375 37 1376 subglacial drainage configuration: rates of basal sediment evacuation controlled by 38 seasonal drainage system evolution. Sedimentary Geology, 149(1-3), 5-19. 1377 39 https://doi.org/10.1016/S0037-0738(01)00241-X 1378 40
- Swift, D. A., Nienow, P. W., Hoey, T. B., & Mair, D. W. F. (2005). Seasonal evolution of 1379 41 42 1380 runoff from Haut Glacier d'Arolla, Switzerland and implications for glacial geomorphic 43 1381 processes. Journal of Hydrology, 309(1), 133–148. 44 1382 https://doi.org/10.1016/j.jhydrol.2004.11.016 45
- Tank, J. L., & Dodds, W. K. (2003). Nutrient limitation of epilithic and epixylic biofilms in ten 1383 46 1384 North American streams. Freshwater Biology, 48(6), 1031-1049. 47 48 1385 https://doi.org/10.1046/j.1365-2427.2003.01067.x
- 49 Thom, M., Schmidt, H., Gerbersdorf, S. U., & Wieprecht, S. (2015). Seasonal biostabilization 1386 50 and erosion behavior of fluvial biofilms under different hydrodynamic and light 1387 51 conditions. International Journal of Sediment Research, 30(4), 273-284. 1388 52 1389 https://doi.org/10.1016/j.ijsrc.2015.03.015 53
- 54 Thompson, A. (1988). Historical development of the proglacial landforms of Svinafellsjokull 1390 55 and Skaftafellsjokull, southeast Iceland. 1391 56
- Thompson, Alan, & Jones, A. (1986). Rates and causes of proglacial river terrace formation 1392 57 in southeast Iceland: an application of lichenometric dating techniques. Boreas, 1393 58 59 1394 15(3), 231–246. https://doi.org/10.1111/j.1502-3885.1986.tb00928.x 60

2 3 1395 Thompson, L., Gray, V., Lindsay, D., & Holy, A. V. (2006). Carbon: nitrogen: phosphorus 4 1396 ratios influence biofilm formation byEnterobacter cloacaeandCitrobacter freundii. 5 Microbiology, 101(5), 1105-1113. doi:10.1111/j.1365-1397 Journal of Applied 6 2672.2006.03003.x 1398 7 8 Tolhurst, T. J., Gust, G., & Paterson, D. M. (2002). The influence of an extracellular 1399 9 polymeric substance (EPS) on cohesive sediment stability. In J. C. Winterwerp & C. 1400 10 Kranenburg Proceedings 1401 (Eds.), in Marine Science (pp. 409-425). 11 1402 https://doi.org/10.1016/S1568-2692(02)80030-4 12 Uehlinger, U., Malard, F., & Ward, J. V. (2003). Thermal patterns in the surface waters of a 1403 13 14 glacial river corridor (Val Roseg, Switzerland). Freshwater Biology, 48(2), 284-300. 1404 15 https://doi.org/10.1046/j.1365-2427.2003.01000.x 1405 16 Valentine, K., Mariotti, G., & Fagherazzi, S. (2014). Repeated erosion of cohesive sediments 1406 17 with biofilms. Advances in Geosciences, 39, 9-14. https://doi.org/10.5194/adgeo-39-1407 18 19 1408 9-2014 20 1409 Van Rijn, L. C. (2007). Unified view of sediment transport by currents and waves. I: Initiation 21 of motion, bed roughness, and bed-load transport. Journal of hydraulic 1410 22 1411 engineering, 133(6), 649-667. https://doi.org/10.1061/(ASCE)0733-23 9429(2007)133:6(649) 1412 24 25 Vignaga, E. (2012). The effect of biofilm colonization on the stability of non-cohesive 1413 26 1414 sediments (Doctoral dissertation, University of Glasgow). 27 Ward, J. V. (1994). Ecology of alpine streams. Freshwater Biology, 32(2), 277-294. 1415 28 https://doi.org/10.1111/j.1365-2427.1994.tb01126.x 1416 29 Ward, J. V., Malard, F., Tockner, K., & Uehlinger, U. (1999). Influence of ground water on 1417 30 31 1418 surface water conditions in a glacial flood plain of the Swiss Alps. Hydrological 32 1419 Processes. 13(3), 277-293. https://doi.org/10.1002/(SICI)1099-33 1420 1085(19990228)13:3<277::AID-HYP738>3.0.CO;2-N 34 Widdows, J., Brinsley, M. D., Salkeld, P. N., & Lucas, C. H. (2000). Influence of biota on 1421 35 spatial and temporal variation in sediment erodability and material flux on a tidal flat 36 1422 37 1423 (Westerschelde, The Netherlands). Marine Ecology Progress Series, 194, 23-37. 38 https://doi.org/10.3354/meps194023 1424 39 Wilcock, P. R., Kenworthy, S. T., & Crowe, J. C. (2001). Experimental study of the transport 1425 40 of mixed sand and gravel. Water Resources Research, 37(12), 3349-3358. 1426 41 42 1427 https://doi.org/10.1029/2001WR000683 43 1428 Wilhelm, L., Singer, G. A., Fasching, C., Battin, T. J., & Besemer, K. (2013). Microbial 44 1429 biodiversity in glacier-fed streams. The ISME Journal, 7(8), 1651-1660. 45 https://doi.org/10.1038/ismej.2013.44 1430 46 Wright, J. P., Gurney, W. S. C., & Jones, C. G. (2004). Patch dynamics in a landscape 1431 47 48 1432 modified by ecosystem engineers. Oikos, 105(2), 336-348. 49 https://doi.org/10.1111/j.0030-1299.2004.12654.x 1433 50 Wright, J. P., & Jones, C. G. (2004). Predicting Effects of Ecosystem Engineers on Patch-1434 51 Scale Species Richness from Primary Productivity. Ecology, 85(8), 2071-2081. 1435 52 1436 https://doi.org/10.1890/02-8018 53 54 Yallop, M. L., de Winder, B., Paterson, D. M., & Stal, L. J. (1994). Comparative structure, 1437 55 primary production and biogenic stabilization of cohesive and non-cohesive marine 1438 56 sediments inhabited by microphytobenthos. Estuarine, Coastal and Shelf Science, 1439 57 39(6), 565-582. https://doi.org/10.1016/S0272-7714(06)80010-7 1440 58 59 1441 Zika, U., & Peter, A. (2002). The introduction of woody debris into a channelized stream: 60 39

2 3 4 5 6 7 8 9 10 11 12	1442 1443 1444 1445 1446 1447 1448 1449	effect on trout populations and habitat. <i>River Research and Applications</i> , <i>18</i> (4), 355–366. https://doi.org/10.1002/rra.677 Zumsteg, A., Luster, J., Göransson, H., Smittenberg, R. H., Brunner, I., Bernasconi, S. M., Frey, B. (2012). Bacterial, Archaeal and Fungal Succession in the Forefield of a Receding Glacier. <i>Microbial Ecology</i> , <i>63</i> (3), 552–564. https://doi.org/10.1007/s00248-011-9991-8
14 15 16 17 18 19	1450 1451 1452 1453	Miller, H. R., & Lane, S. N. (2019). Biogeomorphic feedbacks and the ecosystem engineering of recently deglaciated terrain. <i>Progress in Physical Geography: Earth</i> and Environment, 43(1), 24–45. https://doi.org/10.1177/0309133318816536
19 20 21 22 23 24 25 26 27 28 20 31 32 33 43 56 37 89 40 41 23 44 56 47 48 90 51 253 54 55 56	1454 1455 1456 1457	Polvi, L. E., & Sarneel, J. M. (2018). Ecosystem engineers in rivers: An introduction to how and where organisms create positive biogeomorphic feedbacks. <i>Wiley</i> <i>Interdisciplinary Reviews: Water</i> , 5(2), e1271. https://doi.org/10.1002/wat2.1271
57 58 59 60		40







The Valsorey floodplain (Valais, Switzerland) in October 2018. The floodplain shows clearly primary succession patterns, which are confined to terraces (red lines mark the edges) on both sides of the river that are not reworked by the main braidplain (sub-figures A and B). Well-developed and visible biofilm communities tend to be restricted to channels located on terraces (sub-figure C) because disturbances are not too frequent to destroy the mats.

385x581mm (300 x 300 DPI)







A) Biofilm formation in a stream on a terrace in the Val d'Otemma, Valais, Switzerland; B) A biofilm mat that has formed in a stream in the Val d'Otemma supplied with hillslope-sourced groundwater showing successional colonization of the channel margin by vegetation. The stream is on a terrace about 1 m above the morphodynamically active channel. Note to the right there are stable bar surfaces, also on the terrace, but largely void of primary production due to severe moisture limitations related to well-drained glaciogenic sediments.

320x213mm (300 x 300 DPI)





Channels on the active plain

Abandoned channels