

Quantifying the stability of planktic foraminiferal physical niches between the Holocene and Last Glacial Maximum

Waterson, Amy; Edgar, Kirsty; Schmidt, Daniela N.; Valdes, Paul J.

DOI:

[10.1002/2016PA002964](https://doi.org/10.1002/2016PA002964)

License:

Creative Commons: Attribution (CC BY)

Document Version

Publisher's PDF, also known as Version of record

Citation for published version (Harvard):

Waterson, A, Edgar, K, Schmidt, DN & Valdes, PJ 2017, 'Quantifying the stability of planktic foraminiferal physical niches between the Holocene and Last Glacial Maximum', *Paleoceanography*, vol. 32, no. 1, pp. 74-89. <https://doi.org/10.1002/2016PA002964>

[Link to publication on Research at Birmingham portal](#)

General rights

Unless a licence is specified above, all rights (including copyright and moral rights) in this document are retained by the authors and/or the copyright holders. The express permission of the copyright holder must be obtained for any use of this material other than for purposes permitted by law.

- Users may freely distribute the URL that is used to identify this publication.
- Users may download and/or print one copy of the publication from the University of Birmingham research portal for the purpose of private study or non-commercial research.
- User may use extracts from the document in line with the concept of 'fair dealing' under the Copyright, Designs and Patents Act 1988 (?)
- Users may not further distribute the material nor use it for the purposes of commercial gain.

Where a licence is displayed above, please note the terms and conditions of the licence govern your use of this document.

When citing, please reference the published version.

Take down policy

While the University of Birmingham exercises care and attention in making items available there are rare occasions when an item has been uploaded in error or has been deemed to be commercially or otherwise sensitive.

If you believe that this is the case for this document, please contact UBIRA@lists.bham.ac.uk providing details and we will remove access to the work immediately and investigate.

RESEARCH ARTICLE

10.1002/2016PA002964

Key Points:

- Ecological niche models (ENMs) used to test assumption of planktic foraminifer niche stability 21 kya to present
- High environmental niche stability of planktic foraminifera between the Last Glacial Maximum and today
- Regional calibrations predict global occurrences less well than global calibrations due to niche underrepresentation or multiple genotypes

Supporting Information:

- Supporting Information S1
- Data Set S1

Correspondence to:

A. M. Waterson,
A.Waterson@bristol.ac.uk

Citation:

Waterson, A. M., K. M. Edgar, D. N. Schmidt, and P. J. Valdes (2017), Quantifying the stability of planktic foraminifer physical niches between the Holocene and Last Glacial Maximum, *Paleoceanography*, 32, 74–89, doi:10.1002/2016PA002964.

Received 19 APR 2016

Accepted 16 DEC 2016

Accepted article online 21 DEC 2016

Published online 24 JAN 2017

©2016. The Authors.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

Quantifying the stability of planktic foraminiferal physical niches between the Holocene and Last Glacial Maximum

A. M. Waterson^{1,2} , K. M. Edgar³ , D. N. Schmidt² , and P. J. Valdes¹ 

¹School of Geographical Sciences, University of Bristol, Bristol, UK, ²School of Earth Sciences, University of Bristol, Bristol, UK, ³Now at the School of Geography, Earth and Environmental Sciences, University of Birmingham, Birmingham, UK

Abstract The application of transfer functions on fossil assemblages to reconstruct past environments is fundamentally based on the assumption of stable environmental niches in both space and time. We quantitatively test this assumption for six dominant planktic foraminiferal species (*Globigerinoides ruber* (pink), *G. ruber* (white), *Trilobatus sacculifer*, *Truncorotalia truncatulinoides*, *Globigerina bulloides*, and *Neogloboquadrina pachyderma*) by contrasting reconstructions of species realized and optimum distributions in the modern and during the Last Glacial Maximum (LGM) using an ecological niche model (ENM; MaxEnt) and ordination framework. Global ecological niche models calibrated in the modern ocean have high predictive performance when projected to the LGM for subpolar and polar species, indicating that the environmental niches of these taxa are largely stable at the global scale across this interval. In contrast, ENMs had much poorer predictive performance for the optimal niche of tropical-dwelling species, *T. sacculifer* and *G. ruber* (pink). This finding is supported by independent metrics of niche margin change, suggesting that niche stability in environmental space was greatest for (sub)polar species, with greatest expansion of the niche observed for tropical species. We find that globally calibrated ENMs showed good predictions of species occurrences globally, whereas models calibrated in either the Pacific or Atlantic Oceans only and then projected globally performed less well for *T. sacculifer*. Our results support the assumption of environmental niche stability over the last ~21,000 years for most of our focal planktic foraminiferal species and, thus, the application of transfer function techniques for palaeoenvironmental reconstruction during this interval. However, the lower observed niche stability for (sub)tropical taxa *T. sacculifer* and *G. ruber* (pink) suggests that (sub)tropical temperatures could be underestimated in the glacial ocean with the strongest effect in the equatorial Atlantic where both species are found today.

1. Introduction

The distribution and abundance of organisms can provide valuable quantitative and qualitative proxy data for reconstructing palaeoenvironments [Williams *et al.*, 2007]. In the marine realm, microfossil assemblages are commonly used to estimate a range of palaeoceanographic and palaeoclimatic variables [Imbrie and Kipp, 1971; Climate: Long-Range Investigation, Mapping, and Prediction (CLIMAP) Project Members, 1976; Kucera *et al.*, 2005a]. Planktic foraminifera are perhaps the most common group employed in this manner and have an excellent fossil record, high abundance in deep-sea sediments, and a global distribution [Hemleben *et al.*, 1989]. Their abundance and geographic ranges are strongly related to surface ocean properties, most notably temperature, but also nutrient availability, water column stratification, and turbidity [Ortiz *et al.*, 1995; Bé and Hamlin, 1967; Bé and Tolderlund, 1971; Hemleben *et al.*, 1989; Morey *et al.*, 2005]. Thus, one means of reconstructing past environments is the application of transfer functions. Here a calibration based on the relationship between organisms and their environment in the modern ocean, using the relative abundance of species contributing to an assemblage, is applied to a fossil assemblage to estimate an environmental variable in the past [Berger, 1969; Imbrie and Kipp, 1971; Ortiz and Mix, 1997]. While predominantly applied to the reconstruction of sea surface temperatures (SSTs) [Imbrie and Kipp, 1971; Kipp, 1976; Kucera *et al.*, 2005a], transfer functions also exist for palaeoproductivity [Ivanova *et al.*, 2003] and thermocline depth [Andreasson and Ravelo, 1997]. Successful applications of this approach have focused on the Last Glacial Maximum (LGM; ~21,000 years ago) [CLIMAP Project Members, 1976; Telford *et al.*, 2004; Kucera *et al.*, 2005b] but have also been extended to the Pliocene where the uncertainty regarding their validity is much larger [Andersson, 1997; Dowsett *et al.*, 2011; Dowsett *et al.*, 2012]. This is because implicit in the transfer function approach is the assumption that the ecological relationship between a faunal assemblage and the environment remains stable [Kucera and Schönfeld, 2007]. This assumption is supported for the last glacial interglacial

cycles by similar species' environmental optima identified by comparison of abundance and morphometric data [Schmidt et al., 2003] but has not been rigorously tested in an ecological framework. However, even minor changes in a species niche could generate errors in paleoclimate reconstructions and hence impact resulting model-data comparisons and our assessment of climate change impacts [Schmittner et al., 2011]. In fact, niche changes may help to explain some of the mismatches between different temperature proxies at the same location during the LGM [Kucera et al., 2005a; Multiproxy Approach for the Reconstruction of the Glacial Ocean Surface (MARGO) Project Members, 2009].

Quantification of species ecological niches and estimation of niche differences primarily rely on two approaches, ordination techniques, and ecological niche models (ENMs) [Guisan et al., 2014]. Ordination uses direct observations only and compares the difference in environmental attributes at sites where a species occurs in two different time periods to identify any overlap in environmental space [Broennimann et al., 2012]. ENMs are used to estimate species environmental requirements and can be used to describe the ecological niche by relating species occurrence records to the available environment [Guisan and Zimmermann, 2000]. ENMs have been successfully used to assess the retention of niche-related ecological traits in the past [Martinez-Meyer and Peterson, 2006; Nogués-Bravo et al., 2008; Maguire and Stigall, 2009; Stigall, 2012] and to predict present and future distributions of invasive species [Langer et al., 2013; Weinmann et al., 2013]. Here we use the ENM algorithm MaxEnt which uses presence-background data and compares the environmental conditions at locations of occurrence records with randomly selected points from a background extent to create maps of environmental suitability [Phillips et al., 2006; Merow et al., 2013]. We use the term "niche" here to refer to the multivariate space of physical environmental variables that best correspond to observed species distributions and the associated distribution of potentially abiotically suitable habitats. Other factors, such as biotic interactions and dispersal ability, which limit the occupation of potential niche space, are not incorporated [Araújo and Peterson, 2012]. We augment this analysis with ordination methods which have been shown to more accurately quantify niche overlap (an indicator of niche stability) than ENMs [Broennimann et al., 2012] while being less able to optimize the contribution of different environmental variables to a species' geographical distribution which we analyze using ENMs [Broennimann et al., 2012; Guisan et al., 2014].

In this study we aim to (1) identify the environmental drivers of foraminifer distributions and (2) quantify niche stability between the modern and the LGM using both ordination and ENM techniques. We focus our study on six species dominating the main biogeographic zones (*Trilobatus sacculifer*, *Globigerinoides ruber* (pink), *G. ruber* (white), *Truncorotalia truncatulinoides*, *Globigerina bulloides*, and *Neogloboquadrina pachyderma*). We use MaxEnt to estimate environmental niches in the modern and project these models to the LGM and use an ordination framework to quantify niche margin change metrics of unfilling, expansion, and stability between the LGM and modern. We further test for the presence of nonanalogous environmental space between the modern and the LGM that could bias our analyses, and assess the sensitivity of ENMs to basin-specific versus global calibrations.

2. Methods

2.1. Species Occurrence Data

Species occurrence data for the modern and LGM were taken from the Multiproxy Approach for the Reconstruction of the Glacial Ocean Surface (MARGO) project [Kucera et al., 2005a; Barrows and Juggins, 2005; Chen et al., 2005] as archived on the PANGAEA data repository portal (<http://pangaea.de/Projects/MARGO>). Total sampled localities for the modern are Atlantic ($n = 1166$), Pacific ($n = 1468$), and Indian Oceans ($n = 685$) and for the LGM are Atlantic ($n = 748$), Pacific ($n = 265$), and Indo-Pacific Oceans ($n = 273$) (Figure 1). Samples were not included from the Red Sea and the Mediterranean because during the LGM they were characterized by very different environmental conditions [Thunell and Williams, 1989] that lead to the development of abiotic zones that may bias the data sets [Fenton et al., 2000]. All of the data sets included in MARGO have been carefully prefiltered to exclude dissolution-sensitive data points, and thus, we expect preservation to have little impact on our analyses. Certainly, preliminary ENM analyses support little sensitivity of outputs to preservation as a function of depth-dependent dissolution (Figures S3a and S3b in the supporting information). Occurrence data were treated in two different ways; first to describe the total niche (results in the supplementary information) and second, data were filtered to represent the more informative "optimum niche" (Figure 2). This is important as a shift in a species' optimal niche will impact the reliability of

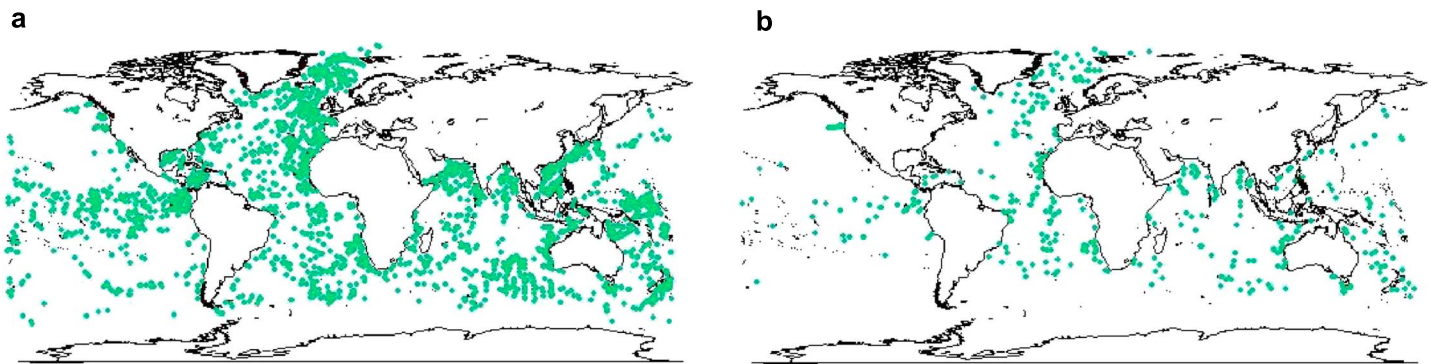


Figure 1. Global maps showing the location of core-top samples with planktic foraminiferal data available for (a) the modern and (b) the Last Glacial Maximum (~21 kyr).

transfer function application to fossil assemblages [Imbrie and Kipp, 1971; Fraile et al., 2009; Jonkers and Kučera, 2015]. Raw abundance data were converted to relative abundance and then filtered by using defined abundance thresholds for the focal species to create a subset of data representing the species optima (Figure S7). Thresholds were selected where there were strong increases in species relative abundance compared to background values. Species' optima were then compared to an independent assessment of the optimal niche based on maximum test body sizes determined by Schmidt et al. [2004, 2006]. Total species occurrences are total modern/ optimal modern: *N. pachyderma* (942/309), *G. bulloides* (3191/325), *T. truncatulinoides* (1949/262), *G. ruber* white (2817/527), *T. sacculifer* (2602/591), and *G. ruber* (pink) (743/19). Total species occurrences for the total LGM/optimal LGM are *N. pachyderma* (552/392), *G. bulloides* (975/88), *T. truncatulinoides* (496/40), *G. ruber* white (759/78), *T. sacculifer* (596/39), and *G. ruber* (pink) (238/19). Geographically filtered occurrences (one presence per environmental grid cell) were used in all MaxEnt analyses to remove the potential inclusion of duplicate records in time or space. Multiple records from one core increase the presence of the species in the analysis of the realized niche as minor changes in abundance do result in differences between absence or presence. These multiple records are unlikely to have any influence on the optimal niche as the differences are very small. Site water depths range from ~50 to 5500 m in each ocean basin [Kucera et al., 2005a; Barrows and Juggins, 2005]. *Neogloboquadrina pachyderma* historically incorporated two morphotypes based on coiling direction which are now known as separate species [Darling et al., 2006]. *Neogloboquadrina pachyderma* is the sinistral morphotype, and *N. incompta* is the dextral type; the former is abundant in cooler polar waters, whereas the latter is adapted to subpolar to temperature environments. Here we focus on *N. pachyderma* sinistral only.

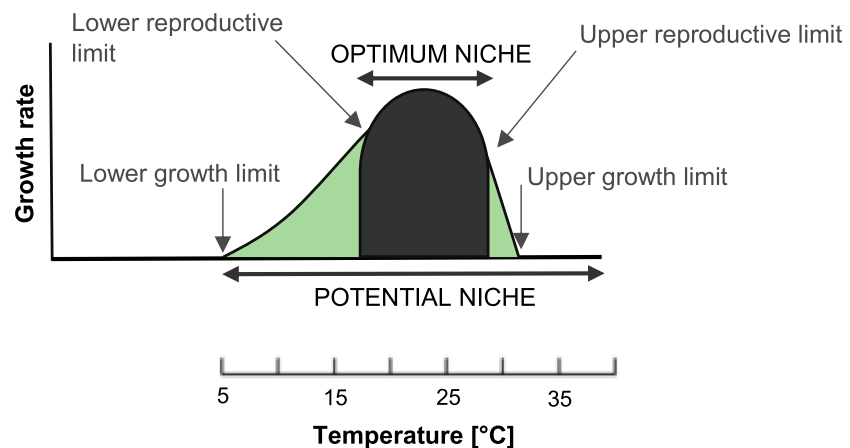


Figure 2. Idealized temperature growth rate model for a planktic foraminifera species modified after Schmidt et al. [2006]. The potential environmental niche is divided into optimum growth, reproductive range, and growth limits.

Species-specific ecological characteristics prior to ENM and ordination analyses ensure the inclusion of physiologically important environmental variables within the model framework. *Globigerinoides ruber* is a (sub) tropical dinoflagellate symbiont bearing species with a temperature optima of $\sim 26^{\circ}\text{C}$ [Bijma et al., 1990; Schmidt et al., 2003] living in the mixed layer. Genetic analyses indicate a wide genetic diversity within this morphospecies (six distinct genotypes in total), suggesting multiple ecologies [Aurahs et al., 2009, 2011]. This is clearly highlighted by distinct differences in the distribution and controls on *G. ruber* (white) and *G. ruber* (pink) (Figures 3 and 4) in the modern ocean, which are genetically different species [Aurahs et al., 2011]. *Globigerinoides ruber* (white) is more cosmopolitan in (sub)tropical waters [Kucera, 2007] with temperature optima in the mixed layer between 18 and 25°C [Hecht, 1976; Lombard et al., 2009] and can also tolerate more oligotrophic waters than *G. ruber* (pink) [Bé, 1959; Bé and Tolderlund, 1971]. *Trilobatus sacculifer* is a symbiotic species which occupies the mixed layer and dominates tropical oligotrophic assemblages [Reiss et al., 1980; Bijma and Hemleben, 1994]. It exhibits large morphological variability, e.g., individuals with a terminal sac-like chamber (*T. sacculifer*) and without sac (*T. trilobus*); however, genetic analyses indicate the presence of just one genotype in the species [André et al., 2012]. *Truncorotalia truncatulinoides* is a deep-dwelling asymbiotic species that occupies subtropical, temperate, and subpolar regions [Parker, 1962]. Its subsurface habitat in subtropical and tropical waters versus a closer to mixed layer habitat in colder waters [Mulltza et al., 1997] makes its environmental preferences difficult to ascertain [Lohmann and Schweitzer, 1990]. *Truncorotalia truncatulinoides* is composed of five genetic species adapted to specific hydrographic conditions that are mainly subdivided on the basis of temperature [Quillévéré et al., 2013]. *Globigerina bulloides* is living in the mixed layer of colder water masses [Bé and Tolderlund, 1971; Naidu and Malmgren, 1996] and highly productive coastal upwelling zones [Thiede and Jünger, 1992]. Consequently *G. bulloides* distribution patterns are sensitive to food availability [Ortiz et al., 1995; Schiebel et al., 1997] and exhibit two temperature optima at annual SST of 10.4°C and 26.6°C representing its general temperature habitat but also the dominance in tropical and subtropical upwelling zones [Schmidt et al., 2003]. *Neogloboquadrina pachyderma* sinistral is a polar species with highest abundances between 0 and 3°C [Kucera, 2007; Lombard et al., 2009] and an optimum temperature of $\sim 0.7^{\circ}\text{C}$ [Schmidt et al., 2003] often residing in the pycnocline if food availability facilitates this [Van Nieuwenhove et al., 2016].

2.2. Environmental Data

Environmental data are derived from the UK Met Office Unified Model Hadley Centre Coupled Model version 3 (HadCM3); a fully coupled Atmosphere-Ocean General Circulation Model. The oceanic model component has a $1.25^{\circ} \times 1.25^{\circ}$ horizontal resolution with 20 vertical levels. The model was run for preindustrial (“modern”) and LGM conditions; an in-depth description is in Gordon et al. [2000] and Cox et al. [2001]. The models were initialized from previous preindustrial and LGM simulations and run for a further 1100 years to spin up surface and intermediate waters. Boundary conditions for LGM simulations (greenhouse gases, orbit, ice sheet, and land-sea mask) are based on the Paleoclimate Modelling Intercomparison Project 3 protocol (<http://pmp3.lsce.ipsl.fr/>). The LGM ocean was initialized from cold ocean conditions; hence, the deep ocean only experiences minimal temperature drift during model spin up (0.4°C in 1100 years). A bilinear interpolation was applied to convert environmental variables from the GCM to 10 minute resolution for use in ENM analyses. This ensured the use of environmental variables at a resolution that adequately captured the foraminiferal physical niche.

For ENM analyses, environmental variables that best describe the ecological limitations on planktic foraminifera distributions were selected [Bé and Tolderlund, 1971; Hemleben et al., 1989; Rutherford et al., 1999; Morey et al., 2005]. Initially, a large number of variables were considered: downward surface solar flux (W m^{-2}), mixed layer depth (between 0 and 300 m water depth), potential temperature difference (between 0 and 200 m water depth), brunt-vaisala frequency (measure of temperature stratification in the first 96 m), annual mean temperature (over surface 5 m), temperature seasonality (maximum-minimum monthly temperature), mean temperatures of the wettest and driest quarters, and mean temperatures of the warmest and coldest quarters (over surface 5 m). However, intercorrelated variables are problematic for ENMs, leading to over-fit models and a reduction in predictive ability [Peterson et al., 2007]. Thus, we only retained variables with a Pearson’s pairwise correlation coefficient < 0.7 (Table S1 in the supporting information). Exploratory MaxEnt analyses found that ENMs incorporating either the mean temperature of the warmest or coldest quarter better predicted LGM foraminifer distributions than annual mean SST. This is unsurprising as absolute

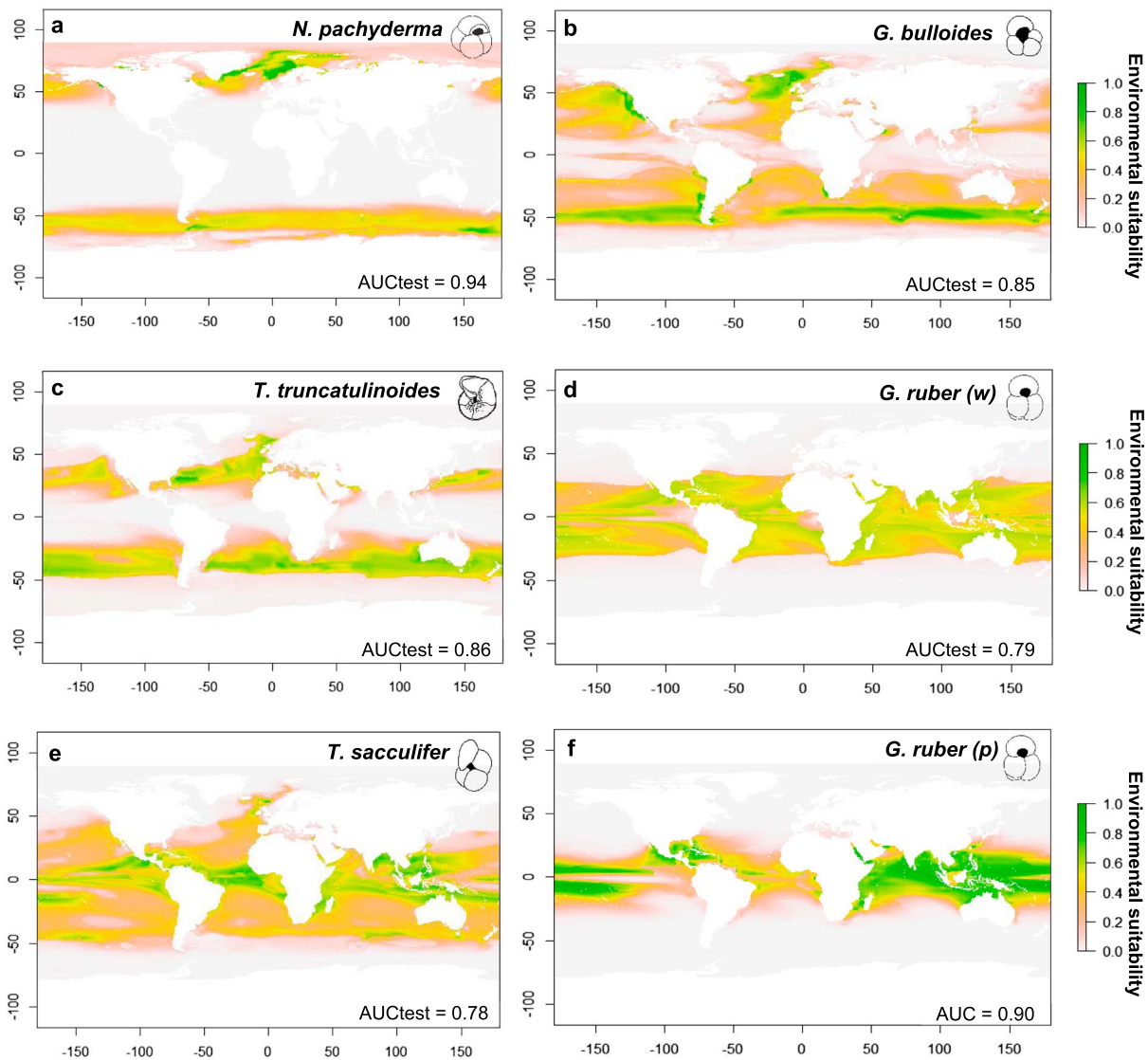


Figure 3. MaxEnt ENMs calibrated in the modern ocean and projected globally: (a) *Neogloboquadrina pachyderma*, (b) *Globigerina bulloides*, (c) *Truncorotalia truncatulinoides*, (d) *Globigerinoides ruber* (white), (e) *Trilobatus sacculifer*, and (f) *G. ruber* (pink). Models calibrated with mean temperature of the coldest quarter. Environmental suitability maps for models calibrated by using mean temperature of the warmest quarter are in Figure S8b. The dark green color (and higher values) indicates high environmental suitability for species, the light-green/yellow color indicates intermediate suitability, and the pink/white color (lower values) indicates poor suitability for species. Average AUCtest values are a measure of predictive performance and show the fit of MaxEnt models to test occurrence data not included in model calibration. AUCdiff values are reported in Table S8).

upper and lower temperature limits more directly impact species and result in lower fecundity, slower growth and smaller sizes [Hecht, 1976; Schmidt et al., 2004; Lombard et al., 2009]. Final ENM analyses therefore use either warm or cold quarter temperatures (in separate calibrations due to high correlations between these two variables) instead of the annual mean, as well as, temperature seasonality, mixed layer depth (MLD), and brunt vaisala frequency. Downward solar flux (W^{-2}) was not included because it is highly correlated with SST such that the individual impacts of the two factors cannot be disentangled.

As the carbon cycle is not sufficiently resolved in HadCM3, we were unable to obtain a measure of oceanic net primary productivity for use in the ENM despite foraminiferal abundance and distributions being influenced by food availability [Rutherford et al., 1999; Bé and Hamlin, 1967; Bé and Tolderlund, 1971; Hemleben et al., 1989; Morey et al., 2005]. We considered using the UVic Earth System Climate Model [Weaver et al., 2001; Meissner et al., 2003] rather than HadCM3 in our analyses because it generates both SST and carbonate chemistry in the oceanic component. However, the UVic vertical resolution is too coarse to resolve the surface

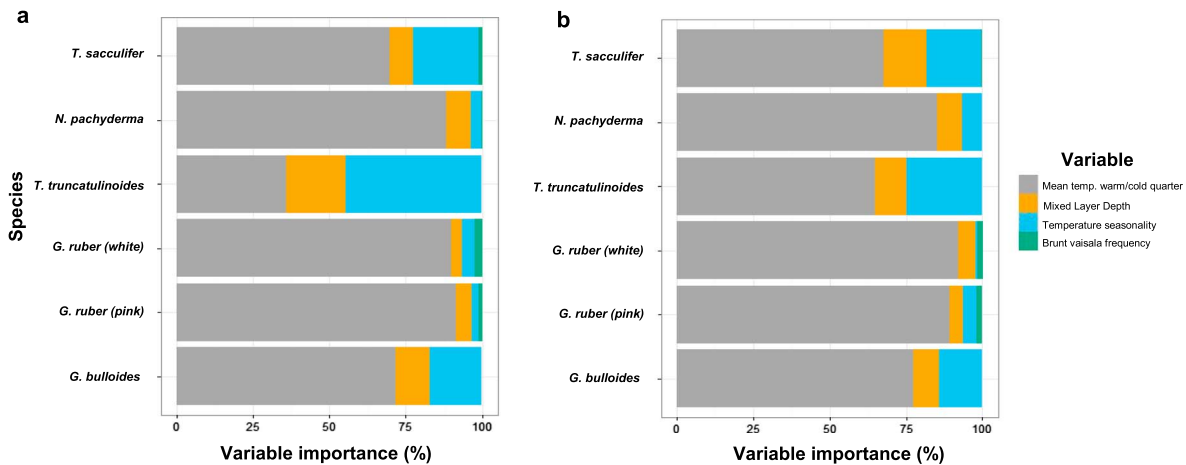


Figure 4. Relative contribution (%) of selected environmental variables on modeled habitat suitability of modern planktic foraminifera from MaxEnt models calibrated with either (a) mean temperature of the warmest quarter or (b) mean temperature of the coldest quarter.

ocean foraminiferal habitat rendering a measure of stratification, a more important variable, unobtainable [Rutherford et al., 1999; Schmidt et al., 2004].

2.3. MaxEnt

MaxEnt (3.3.3 k [Phillips et al., 2006]) ENMs were calibrated on modern global planktic foraminiferal occurrences and projected to the LGM, using the R “dismo” package [Hijmans et al., 2016]. MaxEnt has shown high predictive performance in comparison with other ENMs, displays good consistency when identifying variable importance, and is a widely used ENM allowing for comparison with other studies [Elith et al., 2006]. A fivefold cross-validation procedure [Elith et al., 2010] was used to create global models for the modern and calculate area under the curve (AUC) statistics (predictive performance measure). AUC values of 1 indicate a perfect model fit, and 0.5 represents a no better than random fit [Fielding and Bell, 1997]. Default MaxEnt model parameters were used, with the exception of the regularization parameter set to 2.0. MaxEnt assesses the importance of each environmental variable to model fit using percentage contribution, permutation importance, and jackknife tests (see Tables S6 and S7 and Figures S1a–S1f) for more details). Geographical extent used for model calibration is typically tied to specific hypotheses of the dispersal capacity of taxa being studied [Anderson and Raza, 2010; Barve et al., 2011]. We applied a global calibration extent to all focal species except *G. ruber* (pink). This species’ modern range is restricted to the Atlantic so training extent was limited to this basin only. A global calibration was chosen for the remaining species on the basis of molecular evidence for genetic mixing of Antarctic and Arctic planktic foraminiferal populations, implying a lack of barriers to dispersal [Darling et al., 2000]. This is supported by fossil studies which indicate that the majority of species ranges are wide-reaching, but large populations are sustained only in regions with suitable abiotic and biotic conditions [Norris, 2000; Sexton and Norris, 1980]. Modern ENMs were projected to global LGM environmental conditions, and a binomial test was used to assess the ability of modern ENMs to predict LGM fossil occurrences. This determines if a model prediction is better than random by assessing the statistical significance of agreement between ENM projections and LGM fossil distributions. Three thresholds were used to define binary environmentally suitable and unsuitable model predictions (0%, or the “least training presence threshold” [in the sense of, Pearson et al., 2007], 90% and 50% fossil occurrences included (see the supporting information for further details).

2.4. Testing the Impact of Global Versus Regional Calibrations

The presence of foraminifera genotypes [de Vargas et al., 1999, 2001; Kucera and Darling, 2002; Darling and Wade, 2008], which are often ecotypes with a limited distribution compared to the morphotype, may impact transfer function efficiency [Kucera and Darling, 2002]. We tested this by assessing ENM sensitivity to the range of environmental conditions present in individual ocean basins compared with the global range. ENMs were calibrated in the modern Atlantic and Pacific Oceans for *Trilobatus sacculifer* and *Truncorotalia truncatulinoides*. These species were selected as extreme cases because *T. truncatulinoides* has five genetic

types [de Vargas *et al.*, 2001, 2004; Quillévéré *et al.*, 2013] while *T. sacculifer* has only one [André *et al.*, 2012]; therefore, any noise introduced by a basin-specific calibration for *T. sacculifer* cannot be attributed to the presence of cryptic species. Total species occurrences for Pacific calibrations were *T. sacculifer* (991) and *T. truncatulinoides* (1037) and for Atlantic calibrations were *T. sacculifer* (982) and *T. truncatulinoides* (553). Restricting model calibrations to single ocean basins also impacts the range of the environmental variables incorporated in our ENM analyses. For instance, the Pacific Ocean has a lower salinity and MLD range than the global ocean. Whereas, the Atlantic Ocean is characterized by both lower seasonality and a lower temperature of the warmest quarter than in the global ocean.

2.5. Testing Niche Stability: Ordination Framework

Measures of niche margin unfilling and expansion (environmental niche space occurring only in the LGM or modern, respectively) and stability (niche space occurring in both time slices) were quantified by using an ordination framework [Broennimann *et al.*, 2012] with the R “ecospat” package [Broennimann *et al.*, 2016]. These metrics also provide information on the directionality of niche change between the LGM and modern. The framework reduces dimensions in environmental space using a principle components analysis (PCA), and niche quantification analyses are performed within the first two PCA axes. Niche change analyses were calculated relative to the combined modern and LGM species ranges (pooled-range level [Guisan *et al.*, 2014]). Species percentage occupancy per environmental grid cell is calculated by dividing the density of occurrences with the density of the environmental conditions present across the study area [Broennimann *et al.*, 2012]. The framework uses a kernel density function to generate a “smoothed” density of occurrences in environmental space; this helps to minimize unrealistic holes in the niche, which may occur due to low sampling. All ENM and ordination analyses were performed in the free R environment [R Core Team, 2015].

Nonanalogous climates or environments occur over time with climate change, and these can result in unreliable ENM predictions. We tested for the presence of nonanalogous climates using the ExDet software package; this measures the similarity of variables between two time intervals by assessing deviation from the mean and correlation between environmental variables. It also identifies which variables are most important to detected similarity [Mesgaran *et al.*, 2014].

3. Results

3.1. Understanding Foraminifers Modern Environmental Niche

AUC was used as measure of predictive performance for species occurrence data withheld from the model calibration (~20%); average AUCtest scores ranged between 0.78 and 0.90, indicating that the MaxEnt ENMs are able to discriminate presence from background locations [Peterson *et al.*, 2011] (Figure 3). AUCdiff (difference between AUCtrain and AUCtest) ranged between 0.0006 and 0.0190 (Table S8). For ease we present results for ENMs calibrated with mean temperature of the coldest quarter from herein, unless otherwise stated. Measures of ENM predictive performance (AUC and binomial test scores) and variable importance scores for warm quarter calibrations can be found in Tables S6 and S7 and Figures S1a–f). At the global scale, SST makes the greatest contribution to modern ENM fit for all species (Figure 4). For *T. truncatulinoides*, while SST was most important (64.7%), seasonality (24.7%) and MLD (10.6%) also make substantial contributions to the model fit.

Models fit at and projected at the regional scale were slightly better or equivalent than species global calibrations, and variable importance was largely similar in the Atlantic and Pacific, for investigated taxa *T. truncatulinoides* and *T. sacculifer*. Average AUCtest scores for *T. sacculifer* were 0.82 for the Atlantic and 0.81 for the Pacific, compared to 0.78 for the global calibration. For *T. truncatulinoides*, average AUCtest was 0.86 for the Atlantic, 0.89 for the Pacific, and 0.86 for the global calibration. However, regional calibrations showed poorer predictive performance at the global scale for *T. sacculifer* compared to globally calibrated models (average AUCtest scores for *T. sacculifer*: 0.52 Atlantic, 0.64 Pacific, and 0.78 global), indicating that the regional calibrations do not capture the global niche as well. In contrast, for *T. truncatulinoides* the predictive performance of regional calibrations at the global scale was comparable with or better than global calibrations (average AUCtest values were 0.87 (cold) and 0.89 (warm) for the Atlantic, and 0.80 for the Pacific and 0.86 (global) (Figure 5 and Table S9).

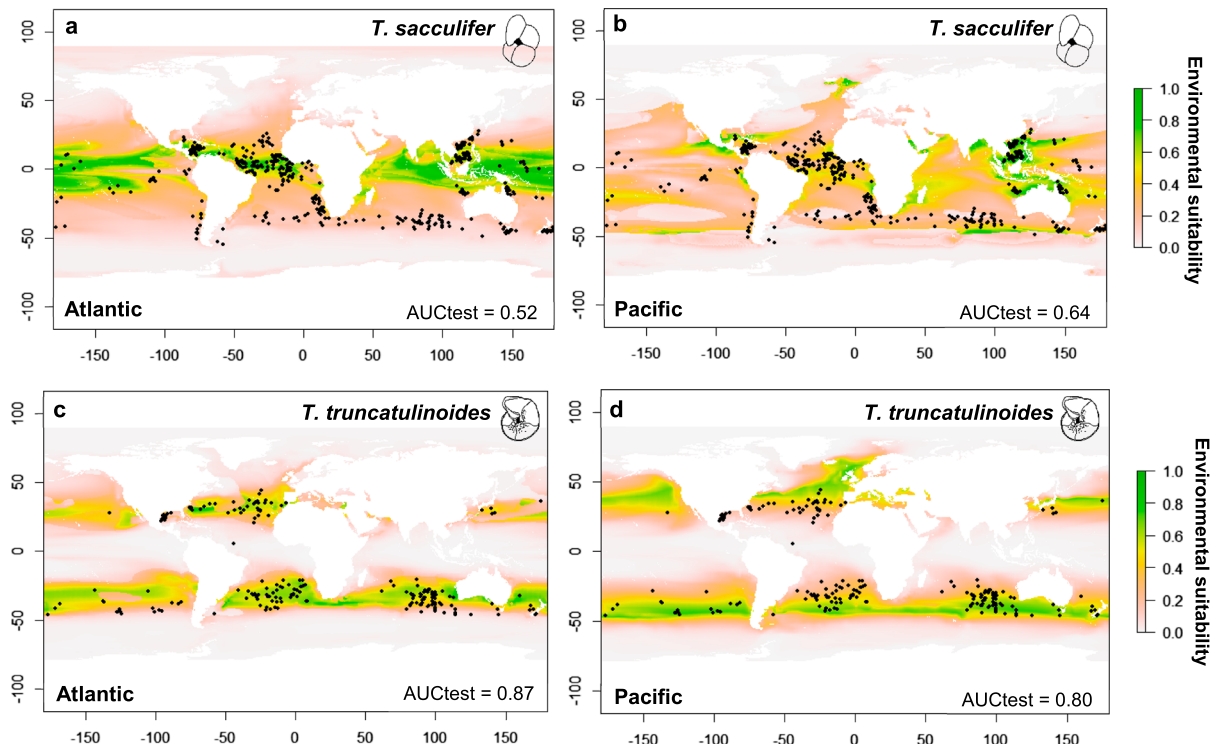


Figure 5. MaxEnt basin-specific ENMs for *T. sacculifer* and *T. truncatulinoides* calibrated in either the (a and c) Atlantic or (b and d) Pacific Oceans and projected to global modern environmental conditions. Models are calibrated with temperature of the coldest quarter. Environmental suitability maps for models calibrated by using mean temperature of the warmest quarter are in Figure S8d. Species occurrences (black circle) overlay the model projections of environmental suitability. Average AUCtest values are measures of predictive performance and show the fit of MaxEnt model projections to the remainder of global foraminifera occurrences not included in model calibration. AUCdiff values are reported in Table S9).

3.2. Stability of Foraminifers Niche Through Time

Grid cells with at least one environmental variable outside of the univariate range are confined to small regions close to Antarctica during the LGM (Figure S10). The brunt vaiala frequency is most influential in these nonanalogue areas (Figure S5b). Fortunately, very few LGM foraminifera fossil occurrences fall within these nonanalogue regions; therefore, the influence of these areas on the model is expected to be minimal. Modern ENMs were projected to LGM environmental layers to test for stability in the niche between the two time slices. Average binomial test scores were significant at all three thresholds ($p < 0.01$), for cold and warm temperature quarter calibrations (with the exception of *T. truncatulinoides* at the least training presence threshold). These results suggest that modern ENMs produce LGM predictions of environmental suitability that are consistent with the distribution of LGM fossil distributions. However, the greatest mismatches between ENM predictions and LGM fossil occurrences are observed for *T. truncatulinoides*, *T. sacculifer*, and *G. ruber* (pink), suggesting that modern models characterize the LGM optimal niche less well for these species (Figure 6).

The cold quarter calibrations show slightly higher thresholds for environmental suitability for tropical species, suggesting that this temperature variable may be more critical in driving niche stability between the LGM and modern for these planktic foraminifera than the temperature of the warmest quarter. Conversely, the warm quarter temperature calibration gives a higher threshold for environmental suitability for *N. pachyderma*, suggesting that the warmer end of the temperature range better describes the niche of this species in the LGM and modern.

Ordination results suggest that the six planktic foraminiferal species exhibit differing degrees of niche margin stability between the modern and LGM. *Neogloboquadrina pachyderma*, *G. bulloides*, *G. ruber* (white), and *G. ruber* (pink) have niches more similar than expected by chance ($p < 0.05$), whereas niche similarity tests were non-significant for *T. sacculifer* and *T. truncatulinoides* (i.e., the hypothesis that the LGM and modern niche are no more similar than by chance cannot be rejected) (Figure 7 and Table S10). Species occupying polar and

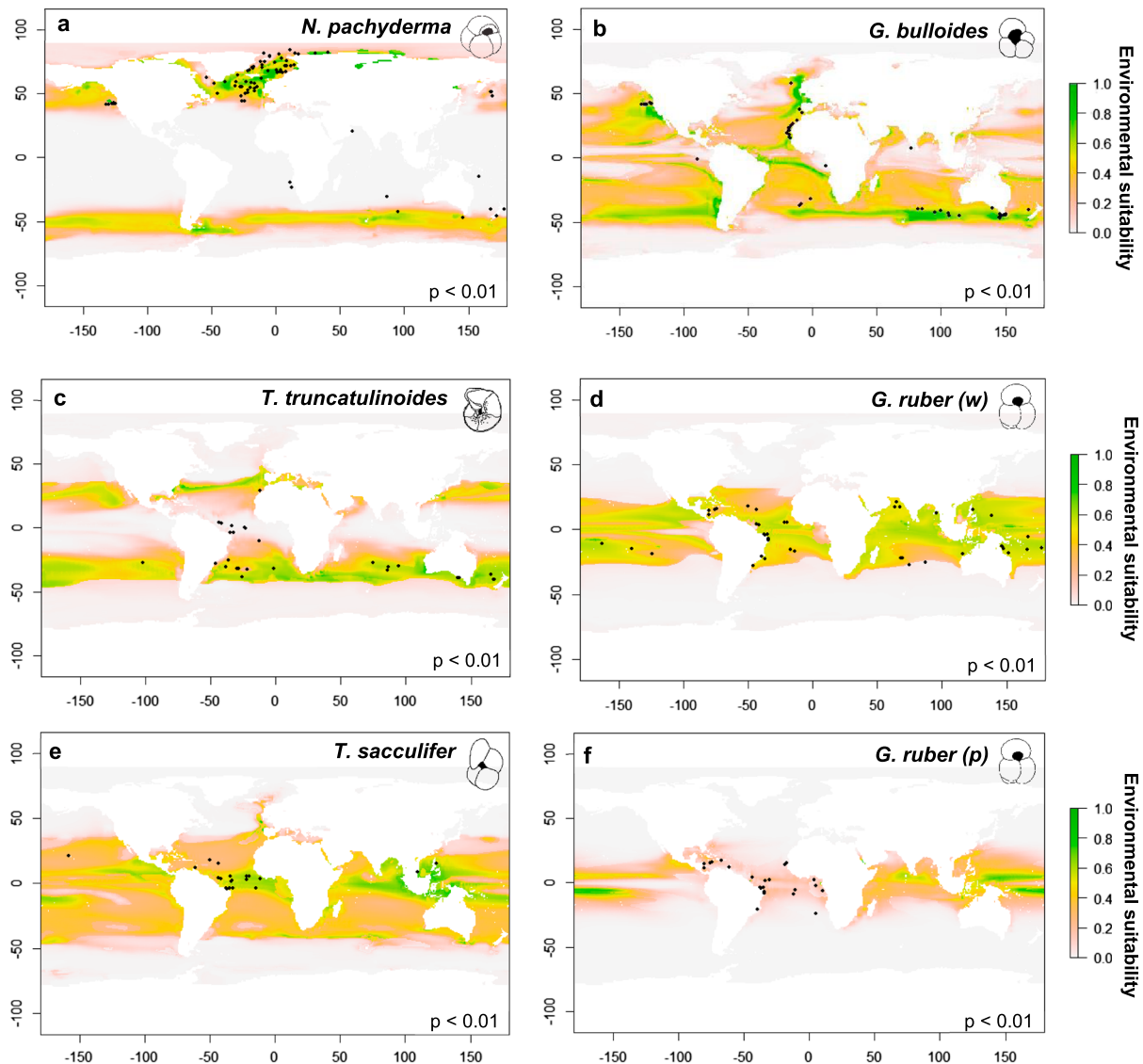


Figure 6. MaxEnt global ENMs calibrated in the modern and projected to the LGM: (a) *Neogloboquadrina pachyderma*, (b) *Globigerina bulloides*, (c) *Truncorotalia truncatulinoides*, (d) *Globigeroidea ruber* (white), (e) *Trilobatus sacculifer*, and (f) *G. ruber* (pink). LGM species occurrences (black circle) overlay the model projections of LGM environmental suitability. Models are calibrated with mean temperature of the coldest quarter. Environmental suitability maps for models calibrated by using mean temperature of the warmest quarter are in Figure S8c. The dark green color (and higher values) indicates high environmental suitability for species, the light-green/yellow color indicates intermediate suitability, and the pink/white color (lower values) indicates poor suitability for species. Average binomial test scores (P values) are a measure of predictive performance and show the fit of MaxEnt model projections to LGM foraminifera species distributions.

subpolar waters show the greatest niche stability between the modern and the LGM (*N. pachyderma*: 99%, *G. bulloides*: 95%, alongside subtropical species *G. ruber* (white): 91%) and minimal niche margin expansion between the LGM and modern (0–9%). All species have negligible or no niches occupied solely during the LGM, excluding *N. pachyderma*, in which niche unfilling is most pronounced (~10%). The largest niche expansion since the LGM is recorded in *T. sacculifer* (58%), *G. ruber* (pink) (26%), and *T. truncatulinoides* (19%). The new habitat is predominantly expressed by a change in niche center in the direction of PC1 (PC1 = 62.35% and PC2 = 11.38%), represented primarily by temperature of the warmest and coldest quarter, and other highly correlated variables including downward solar flux and potential temperature difference (Figure S6), suggesting that these three species were able to benefit from the increasing SSTs and associated changes in the water column between the LGM and modern (Figure 7 and Table S10).

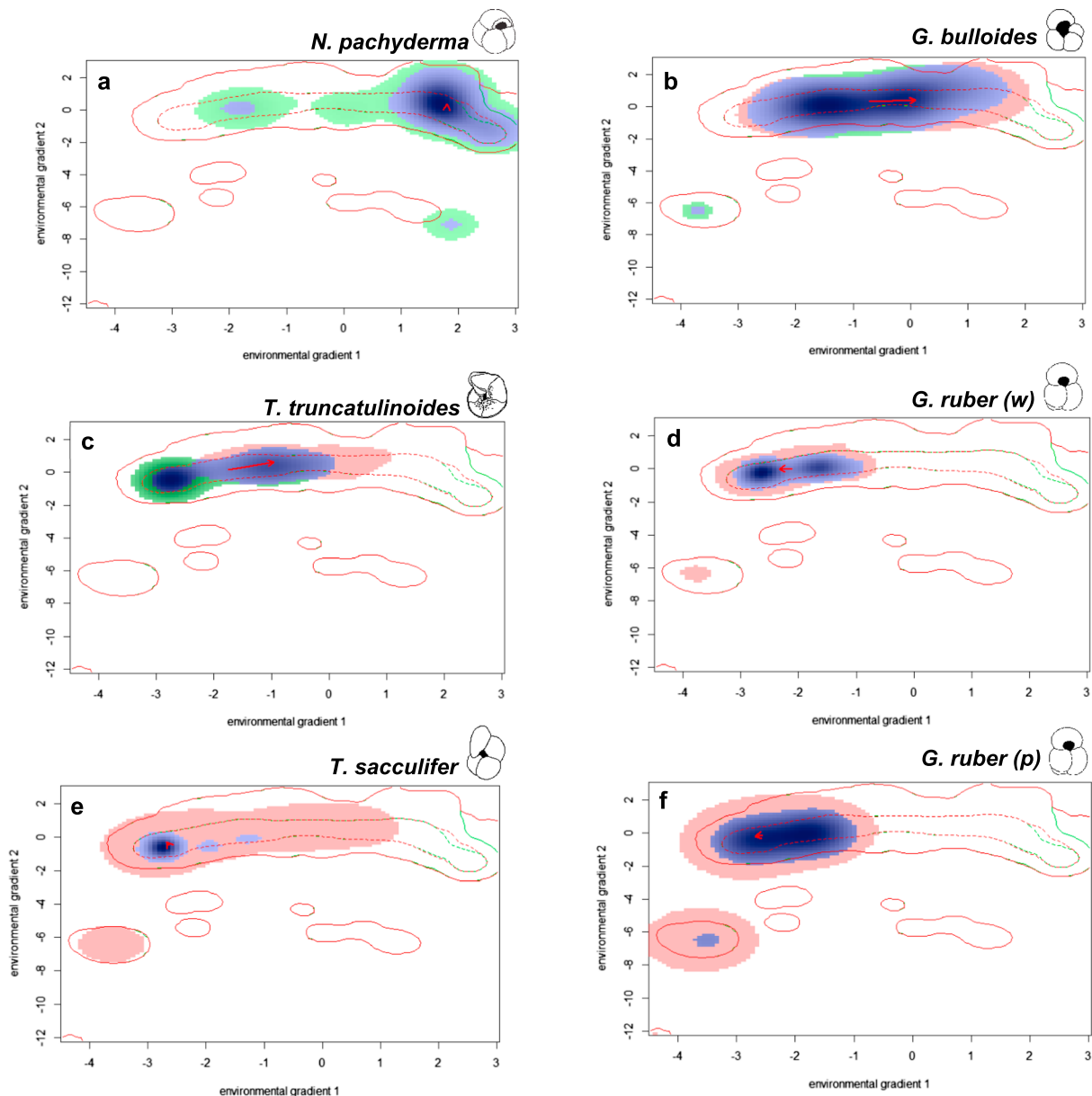


Figure 7. Planktic foraminifera niche change between the modern and LGM in environmental space determined by ordination techniques. The solid contour lines illustrate the full range (100%) of possible (background) environmental space in the two timeslices, and the dashed lines are 50%. The shading shows the density of species occurrences per grid cell. PC1 accounts for 62.48% total variation and is primarily represented by temperature variables, downward solar flux, and potential temperature difference. PC2 accounts for 11.3% total variation and is represented by the remaining variables: salinity, mixed layer depth, temperature seasonality, and brunt-vaiala frequency (see Figure S6). The blue pixels show the stable niche (common between modern and LGM), the green pixels show the unfilled niche (LGM only), and the red pixels show the expansion of the niche (modern only). The red arrow indicates the change in the direction of the center of the species niche from the LGM to the modern.

4. Discussion

4.1. Environmental Drivers of Planktic Foraminiferal Distributions in the Modern Ocean

Culture experiments and in situ observations clearly highlight temperature as the most important variable driving modern planktic foraminifer distributions [Bé et al., 1985; Bijma et al., 1990; Ortiz et al., 1995; Morey et al., 2005]. MaxEnt results identify SST as the most important variable on modern planktic foraminifer species distributions (Figure 4), reinforcing the major role this parameter plays in governing biogeographic distributions [Bé and Tolderlund, 1971]. The environmental variables included in our ENM analyses focus solely

on the physical aspects of the foraminiferal niche and omit synergistic factors such as food and light, which modify species distribution under favorable temperatures [Berger, 1969; Bé et al., 1985; Ortiz et al., 1995].

A further potentially important chemical variable that we omit here is seawater carbonate ion concentration, which impacts the geochemical composition of planktic foraminifer tests [Russell et al., 2004; Spero et al., 1997] and calcification [Barker and Elderfield, 2002]. However, its impact on species distribution and diversity is currently unclear. Comprehensive modeling projects are underway which address several of these key oceanic variables [Bopp et al., 2013]; thus, we expect future studies to benefit from those models that incorporate both physical and chemical components of the surface ocean.

4.2. Species-Specific Drivers of Biogeography

While temperature is the dominant variable for *G. ruber* (pink) and *G. ruber* (white), distinct differences in the distribution of the two genetically different “species” [Aurahs et al., 2011] in the modern ocean are clearly highlighted (Figures 3 and 4). *Globigerinoides ruber* (white) has a broader temperature optimum (18–25°C [Hecht, 1976; Lombard et al., 2009]) in (sub)tropical waters compared to *G. ruber* (pink) which has a more tropical distribution [Kucera, 2007]. Our results corroborate the importance of SST as the most dominant variable for *T. sacculifer* based on experimental results [Bijma et al., 1990]. We find a slightly better model “fit” for *T. sacculifer* at the basin-specific scale but transferring the calibration to the global scale showed lower predictive performance in comparison to global ENMs, suggesting that geographically-constrained calibration data sets do not capture the entire environmental niche of *T. sacculifer* (Figure 5). This is most likely a function of the limited range of environmental space sampled in an individual ocean in comparison to the global ocean. We find that while SST is the dominant variable for *T. truncatulinoides*, seasonality and MLD also make significant contributions, and is supported by the well-documented dependence of its habitat on water column structure [Lohmann and Schweitzer, 1990; Mulitza et al., 1997; McKenna and Prell, 2004]. Regional calibrations resulted in comparable performance when projected to a global scale for *T. truncatulinoides* (Figure 5), suggesting that individual ocean basins encompass the entire environmental range of the morphospecies, as already suggested by the presence of all genotypes in each major basin for this species [Quillévére et al., 2013]. SST is the dominant variable for *G. bulloides* model fit, however to a lesser degree than in most other species, as expected due to its dominance in high productivity upwelling areas which explains the relatively high contribution of MLD in our results. Upwelling is highly seasonal at a local scale; thus, this may also explain the contribution of temperature seasonality for *G. bulloides*. SST is the dominant variable for *N. pachyderma* model fit, supporting observations that the species’ distribution is constrained primarily by its thermal limits [Parker, 1962].

Observational data show that planktic foraminiferal distributions are sensitive to seasonal extremes and that some species respond to environmental change by altering the timing of their presence, with subsequent impacts on predator-prey relationships [Jonkers and Kučera, 2015]. Such responses are important for understanding niche dynamics over time, and we acknowledge that accounting for these is often underestimated by the fallacy of the mean. For those species that are able to alter their depth in response to seasonality changes may still appear to occupy the same temperature despite a shift in the niche.

4.3. Ecological Niche Stability Between the LGM and Modern

During the LGM, SST cooling was most pronounced at midlatitudes relative to little or no changes in the tropics and at high latitudes (Figure S9) [Kucera et al., 2005a]. LGM oceans displayed a higher salinity and greater temperature seasonality in contrast to the modern, with colder winters and an expansion in sea ice extent [de Vernal et al., 2005]. Simulations suggest that the ocean thermal structure changed between the LGM and pre-industrial [Telford et al., 2013]. The Western Pacific warm pool contracted, and there was substantive (>3°C) cooling of the eastern equatorial Pacific and eastern boundary currents [Kucera et al., 2005a]. Significant cooling of eastern boundary currents led to steepened SST gradients in the midlatitude North Atlantic, while the subtropical gyres experienced little change.

Our use of optima occurrence data has a substantial impact on ENM predictions to the modern and LGM and ordination analyses, in comparison to occurrence data used in realized niche analyses (Figures S2–S5). For instance, analysis of species’ optimum environmental niche rather than realized niche suggests significant changes in the occupation of physical environmental niche space between the LGM and modern in *T. truncatulinoides*, *T. sacculifer*, and *G. ruber* (pink). This highlights the importance of using species optima for widely

distributed taxa by removing occurrences that represent marginal niche conditions. Our ENM results suggest that niche stability is best described by cold quarter temperature for tropical species (*T. sacculifer* and *G. ruber*) and warm quarter temperature for polar species (*N. pachyderma*). This has potential consequences for modern analogue techniques as foraminiferal assemblages may represent colder (or warmer) temperatures in the LGM. While this influence is likely to be small it may explain some of the noise observed among proxy signals in some ocean regions during the LGM [MARGO Project Members, 2009].

Based on our findings the optimum environmental niche of three of the six investigated species (*N. pachyderma*, *G. bulloides*, and *G. ruber* (white)) over the past 20 kyr did not change significantly, supporting evidence from biometric [Schmidt *et al.*, 2003] and genetic analyses that the most recent splits in the foraminiferal tree predate the LGM by 100 kyr [de Vargas *et al.*, 2001]. In contrast, ordination results show significant niche expansion for *T. truncatulinoides*, *T. sacculifer*, and *G. ruber* (pink) from the LGM toward the modern primarily driven by temperature. This suggests that these species were able to benefit from increasing SSTs and associated changes in the water column structure between the LGM and present-day, which allowed them to expand their range. ENM predictions to the LGM for *T. truncatulinoides* (Figure 6) show fossil occurrences in the equatorial Atlantic in areas of low environmental suitability and might reflect changes in water column structure and thermocline depth [Schneider *et al.*, 1996]. Large areas in the North Atlantic where *T. truncatulinoides* is commonly found today were unsuitable during the LGM due to changes in the Gulf Stream [Billups *et al.*, 2016].

Modern ENMs for *T. sacculifer* represent the environmental niche well in tropical regions. Unexpectedly, while the model suggests lower suitability in the South Atlantic subtropical front, an abundance of optima occurrences are found in this area (Figure S8a). Bé *et al.* [1981] suggesting that feeding frequency on zooplankton is important for *T. sacculifer's* optimal niche. Nutrient-rich frontal zones could provide such a habitat; thus, the lack of food availability in our model could help to explain this mismatch.

Modern ENMs are unable to characterize the environmental niche of *G. ruber* (pink) well or predict its LGM distribution (Figures 3 and 6). This indicates that the variables we use here do not adequately describe the environmental niche of this species. In particular, our analyses indicate high environmental suitability for *G. ruber* (pink) in the Indian Ocean from which it is absent, indicating that factors other than the physical environment may impact its distribution and that inclusion of biotic factors may help to inform future analyses.

Comparison of GCM reconstructions to multiproxy data sets indicates that the models correctly represent the large-scale features of LGM climate; however, in some areas climate proxies show a wide spread in estimated LGM temperatures. Regions identified by MARGO Project Members [2009] as areas with the largest ranges between different proxies include the equatorial Atlantic and the eastern equatorial upwelling region, which suggest colder temperatures. We observe the greatest niche change in tropical species, *T. sacculifer* and *G. ruber* (pink), with expansion from the LGM to the modern, toward regions warmer than those occupied during the LGM. Lower observed niche stability for these taxa means that the absence of these tropical species in LGM assemblages within these regions will likely weight transfer function results toward colder temperature reconstructions.

As you go further back in time differentiation across phylogenies should increasingly display a reduction in conservatism, i.e., niche stability simply breaks down over time [Peterson, 2011]. This will vary between groups and is dependent on the time scales investigated. While our findings suggest that the optimal niche of our focal species is largely (>70%) conservative over the past 20 kyr, the plant record for example, documents nonanalogue floras in the Quaternary [Jackson and Overpeck, 2000; Veloz *et al.*, 2012]. In contrast to the terrestrial realm, there has been limited investigation of niche conservatism in the marine realm with which to compare our results, Saupe *et al.* [2014] found stability of environmental preferences of marine mollusc species between the Pliocene (~3 Ma) and modern, suggesting that their response to future climate change will be to track suitable habitats or face extinction where the rate of change is too rapid.

4.4. Next Steps in Using ENMs in Paleooceanographic Research

A breakdown of niche conservatism is expected over geological timescales longer than the 20 kyr investigated here. Key target intervals for future research are the early Pleistocene, prior to the environmental and evolutionary changes which shaped modern diversity patterns, and past warm interval marine isotope

stage 11 (424–374 kyr), which has been the focus of a large data collection and modeling effort; therefore, the necessary environmental and biotic data are readily available for developing ENMs. ENMs have primarily been used with species presence-only data because this is often the most readily available in large-scale biodiversity databases [Peterson *et al.*, 2011]. However, the microfossil community are extremely fortunate in that highly spatially and temporally resolved occurrence and abundance data sets are generated routinely and are available for many taxonomic groups (e.g., coccolithophores, radiolarians, ostracods, diatoms, and foraminifers) unlike in the terrestrial realm where ENMs are most commonly employed. The inclusion of abundance data in ENM analysis would allow the quantification of not just range stability but also how species' optimal niche changes through time (Figure 2), i.e., abundance shifts that may have occurred within the overall niche of a species and could provide vital clues for understanding adaptation and evolution of species.

5. Conclusions

Global-scale ENM and ordination analyses overall support the assumption of planktic foraminiferal environmental niche stability between the modern and LGM in agreement with previous morphological and genetic approaches. SST is consistently the most important environmental variable to model fit in almost all global ENMs over mixed layer depth, or seasonality this suggests that the thermal limits of the selected species has remained largely constant between the LGM and modern. Ordination results show that the stability of the optimal niche is high for the species that dominate the major subpolar and polar biogeographic zones in the modern ocean. Species occupying tropical regions show lower niche stability and greater niche margin expansion between the LGM and modern. This implies that caution should be taken when applying transfer functions to foraminifera assemblages over this interval that include high abundances of these taxa.

Future studies would benefit from incorporating other factors into our model framework such as light, productivity, and carbonate chemistry. The use of abundance data would also allow us to address range stability and further quantification of the optimal niche through time.

Acknowledgments

Financial support was provided in the form of an NE/J020389/1 Natural Environment Research Council grant to D.N.S. and P.V. funding AW, Leverhulme Early Career Fellowship ECF-2013-608 to K.M.E., and a Royal Society Wolfson Merit Award to D.N.S. We would also like to thank Elena Couce, Katrin Meisner, and Chris Yesson for helpful discussions. Climate variables used in ENM and ordination analyses for the modern and LGM are available at: <http://www.bridge.bris.ac.uk/resources/simulations>. Planktic foraminifera occurrence data can be found at <http://pangaea.de/Projects/MARGO>.

References

- Andersson, C. (1997), Transfer function vs. modern analog technique for estimating Pliocene sea-surface temperatures based on planktic foraminiferal data, western Equatorial Pacific Ocean, *J. Foramin. Res.*, *27*, 123–132.
- Anderson, R. P., and A. Raza (2010), The effect of the extent of the study region on GIS models of species geographic distributions and estimates of niche evolution: Preliminary tests with montane rodents (genus *Nephelomys*) in Venezuela, *J. Biogeogr.*, *37*(7), 1378–1393.
- André, A., A. Weiner, F. Quillévéré, R. Aurahs, R. Morard, C. J. Douady, T. de Garidel-Thoron, G. Escarguel, C. de Vargas, and M. Kucera (2012), The cryptic and the apparent reversed: Lack of genetic differentiation within the morphologically diverse plexus of the planktonic foraminifer *Globigerinoides sacculifer*, *Paleobiology*, *39*, 21–39.
- Andreason, D. J., and A. C. Ravelo (1997), Tropical Pacific Ocean thermocline depth reconstructions for the Last Glacial Maximum, *Paleoceanography*, *12*, 395–413, doi:10.1029/97PA00822.
- Araújo, M. B., and A. T. Peterson (2012), Uses and misuses of bioclimatic envelope modeling, *Ecology*, *93*(7), 1527–1539.
- Aurahs, R., G. W. Grimm, V. Hemleben, C. Hemleben, and M. Kucera (2009), Geographical distribution of cryptic genetic types in the planktonic foraminifer *Globigerinoides ruber*, *Mol. Ecol.*, *18*, 1692–1706.
- Aurahs, R., Y. Treis, K. Darling, and M. Kucera (2011), A revised taxonomic and phylogenetic concept for the planktonic foraminifer species *Globigerinoides ruber* based on molecular and morphometric evidence, *Mar. Micropaleontol.*, *79*, 1–14.
- Barker, S., and H. Elderfield (2002), Foraminiferal calcification response to glacial-interglacial changes in atmospheric CO₂, *Science*, *297*, 833–836.
- Barrows, T. T., and S. Juggins (2005), Sea surface temperatures around the Australian margin and Indian Ocean during the last glacial Maximum, *Quat. Sci. Rev.*, *24*(7–9), 951–998.
- Barve, N., V. Barve, A. Jiménez-Valverde, A. Lira-Noriega, S. P. Maher, A. T. Peterson, J. Soberón, and F. Villalobos (2011), The crucial role of the accessible area in ecological niche modeling and species distribution modeling, *Ecol. Modell.*, *222*(11), 1810–1819.
- Bé, A. W. H. (1959), Ecology of recent planktonic foraminifera: Part I: Areal distribution in the Western North Atlantic, *Micropalaeontology*, *5*, 77–1000.
- Bé, A. W. H., and D. S. Tolderlund (1971), Distribution and ecology of planktonic foraminifera, in *The Micropalaeontology of Oceans*, edited by B. M. Funnell and W. R. Riedel, pp. 105–150, Cambridge Univ. Press, London.
- Bé, A. W. H., and W. H. Hamlin (1967), Ecology of recent planktonic foraminifera, *Micropalaeontology*, *13*(1), 87–106.
- Bé, A. W. H., D. A. Caron, and O. R. Anderson (1981), Effects of feeding frequency on life processes of the planktonic foraminifer *Globigerinoides sacculifer* in laboratory culture, *J. Mar. Biol. Assoc. U. K.*, *61*, 257–277.
- Bé, A. W. H., J. K. B. Bishop, M. S. Sverdløve, and W. D. Gardner (1985), Standing stock, vertical distribution and flux of planktonic foraminifera in the Panama Basin, *Mar. Micropalaeontol.*, *9*(4), 307–333.
- Berger, W. H. (1969), Ecological patterns of living planktonic foraminifera, *Deep Sea Res. Oceanogr. Abstr.*, *16*(1), 1–24.
- Bijma, J., and C. Hemleben (1994), Population dynamics of the planktic foraminifer *Globigerinoides sacculifer* (Brady) from the central Red Sea, *Deep Sea Res. Part 1; Oceanogr. Res. Papers*, *41*, 485–510.
- Bijma, J., W. W. Faber, and C. Hemleben (1990), Temperature and salinity limits for growth and survival of some planktonic foraminifera in laboratory cultures, *J. Foramin. Res.*, *20*(2), 95–116.

- Billups, K., C. Hudson, H. Kunz, and I. Rew (2016), Exploring Globorotalia truncatulinoides coiling ratios as a proxy for subtropical gyre dynamics in the northwestern Atlantic Ocean during late Pleistocene Ice Ages, *Paleoceanography*, *31*, 553–563, doi:10.1002/2016PA002927.
- Bopp, L., et al. (2013), Multiple stressors of ocean ecosystems in the 21st century: Projections with CMIP5 models, *Biogeosciences*, *10*, 6225–6245.
- Broennimann, O. B., et al. (2016), Ecospat: Spatial ecology miscellaneous methods. [Available at: <https://cran.r-project.org/web/packages/ecospat/>]
- Broennimann, O., et al. (2012), Measuring ecological niche overlap from occurrence and spatial environmental data, *Global Ecol. Biogeogr.*, *21*(4), 481–497.
- Chen, M. T., C. C. Huang, U. Pflaumann, C. Waelbroeck, and M. Kucera (2005), Estimating glacial wester Pacific seas-surface temperature: Methodological overview and data compilation of surface sediment planktic foraminifer faunas, *Quat. Sci. Rev.*, *24*(7–9), 1049–1062.
- CLIMAP Project Members (1976), The surface of the ice-age Earth, *Science*, *191*, 1131–1137.
- Cox, P. M., R. A. Betts, C. Jones, S. A. Spall, and I. Totterdell (2001), Modelling vegetation and the carbon cycle as interactive elements of the climate system.
- Darling, K. F., and C. M. Wade (2008), The genetic diversity of planktic foraminifera and the global distribution of ribosomal RNA genotype, *Mar. Micropaleontol.*, *67*, 216–238.
- Darling, K. F., M. Kucera, D. Kroon, and C. M. Wade (2006), A resolution for the coiling direction paradox in *Neogloboquadrina pachyderma*, *Paleoceanography*, *21*, PA2011, doi:10.1029/2005PA001189.
- Darling, K. F., C. M. Wade, I. A. Stewart, D. Kroon, R. Dingle, and A. J. L. Brown (2000), Molecular evidence for genetic mixing of Arctic and Antarctic subpolar populations of planktonic foraminifers, *Nature*, *405*(6782), 43–47.
- de Vargas, C., R. Norris, L. Zaninetti, S. W. Gibb, and J. Pawlovski (1999), Molecular evidence of cryptic speciation in planktonic foraminifers and their relation to oceanic provinces, *Proc. Natl. Acad. Sci. U.S.A.*, *96*, 2864–2868.
- de Vargas, C., S. Renaud, H. Hübner, and J. Pawlovski (2001), Pleistocene adaptive radiation in Globorotalia truncatulinoides: Genetic, morphologic, and environmental evidence, *Paleobiology*, *27*, 104–125.
- de Vargas, C., A. G. Saez, L. K. Medlin, and H. R. Thierstein (2004), Super-species in the calcareous plankton, in *Coccolithophores From Molecular Processes to Global Impact*, edited by H. R. Thierstein and J. R. Young, pp. 271–298, Springer, Berlin.
- de Vernal, A., et al. (2005), Reconstruction of sea-surface conditions at middle to high latitudes of the Northern Hemisphere during the Last Glacial Maximum (LGM) based on dinoflagellate cyst assemblages, *Quat. Sci. Rev.*, *24*(7–9), 897–924.
- Dowsett, H. J., A. M. Haywood, P. J. Valdes, M. M. Robinson, D. J. Lunt, D. J. Hill, D. K. Stoll, and K. M. Foley (2011), Sea surface temperatures of the mid-Piacenzian Warm Period: A comparison of PRISM3 and HadCM3, *Palaeogeogr. Palaeoclimatol. Palaeoecol.*, *309*, 83–91.
- Dowsett, H. J., et al. (2012), Assessing confidence in Pliocene sea surface temperatures to evaluate predictive models, *Nat. Clim. Change*, *2*, 365–371.
- Elith, J., M. Kearney, and S. Phillips (2010), The art of modelling range-shifting species, *Meth. Ecol. Evol.*, *1*(4), 330–342.
- Elith, J., et al. (2006), Novel methods improve predictions of species' distributions from occurrence data, *Ecography*, *29*, 129–151.
- Fenton, M., S. Geiselhart, E. J. Rohling, and C. Hemleben (2000), Planktonic zones in the Red Sea, *Mar. Micropaleontol.*, *40*, 277–294.
- Fielding, A. H., and J. F. Bell (1997), A review of methods for the assessment of prediction errors in conservation presence/absence models, *Environ. Conservat.*, *24*, 38–49.
- Fraile, I., M. Schulz, S. Multiza, U. Merkel, M. Prange, and A. Paul (2009), Modeling the seasonal distribution of planktonic foraminifera during the Last Glacial Maximum, *Paleoceanography*, *24*, PA2216, doi:10.1029/2008PA001686.
- Gordon, C., C. Cooper, C. A. Senior, H. Banks, J. M. Gregory, T. C. Johns, J. F. B. Mitchell, and R. A. Wood (2000), The simulation of SST, sea ice extents and ocean heat transports in a version of the Hadley Centre coupled model without flux adjustments, *Clim. Dyn.*, *16*, 147–168.
- Guisan, A., and N. E. Zimmermann (2000), Predictive habitat distribution models in ecology, *Ecol. Modell.*, *135*(2–3), 147–186.
- Guisan, A., B. Petitpierre, O. Broennimann, C. Daehler, and C. Kueffer (2014), Unifying niche shift studies: insights from biological invasions, *Trends Ecol. Evol.*, *29*(5), 260–269.
- Hecht, A. D. (1976), Size variations in planktonic foraminifera: Implications for quantitative paleoclimatic analysis, *Science*, *192*, 1330–1332.
- Hemleben, C., M. Spindler, and O. R. Anderson (1989), *Modern Planktonic Foraminifera*, Springer, Berlin.
- Hijmans, R. J., S. Phillips, J. Leathwick, and J. Elith (2016), Dismo: Species Distribution Modelling. [Available at: <http://cran.r-project.org/web/packages/dismo/>]
- Imbrie, J., and N. Kipp (1971), A new micropaleontological method for quantitative paleoclimatology: Application to late Pleistocene Caribbean core V28-238, in *The late Cenozoic Glacial Ages*, edited by K. K. Turekian, pp. 77–181, Yale Univ. Press, New Haven, Conn.
- Ivanova, E. M., R. Schiebel, A. Deo Singh, G. Schmiedl, H. S. Niebler, and C. Hemleben (2003), Primary production in the Arabian Sea during the last 135,000 years, *Palaeogeogr. Palaeoclimatol. Palaeoecol.*, *197*(1–2), 61–82.
- Jackson, S. T., and J. T. Overpeck (2000), Response of plant populations and communities to environmental changes of the Late Quaternary, *Paleobiology*, *26*(4), 194–220.
- Jonkers, L., and M. Kucera (2015), Global analysis of seasonality in the shell flux of extant planktonic foraminifera, *Biogeosciences*, *12*(7), 2207–2226.
- Kipp, N. (1976), New transfer function for estimating past sea-surface conditions from sea-bed distribution of planktonic foraminifer assemblages in the North Atlantic, *Geol. Soc. Am. Mem.*, *145*, 3–42.
- Kucera, M. (2007), Planktonic foraminifera as tracers of past oceanic environments, in *Developments in Marine Geology*, vol. 1, edited by C. Hillaire-Marcel and A. De Vernal, pp. 213–262, Elsevier, Amsterdam.
- Kucera, M., and J. Schönfeld (2007), The origin of modern oceanic foraminifer faunas and Neogene climate change, in *Deep-Time Perspectives on Climate Change: Marrying the Signal From Computer Models and Biological Proxies*, *The Micropaleontological Society, Special Publications*, edited by M. Williams et al., The Geological Society, London.
- Kucera, M., and K. F. Darling (2002), Cryptic species of planktonic foraminifera: Their effect on palaeoceanographic reconstructions, *Philos. Trans. R. Soc. Lond. Ser. A*, *360*, 695–718.
- Kucera, M., et al. (2005a), Reconstruction of sea-surface temperatures from assemblages of planktonic foraminifera: Multi-technique approach based on geographically constrained calibration data sets and its application to glacial Atlantic and Pacific Oceans, *Quat. Sci. Rev.*, *24*(7–9), 951–998.
- Kucera, M., A. Rosell-Melé, R. Schneider, C. Waelbroeck, and M. Wienelt (2005b), Multiproxy approach for the reconstruction of the glacial ocean surface (MARGO), *Quat. Sci. Rev.*, *24*(7–9), 813–819.
- Langer, M., A. E. Weinmann, S. Lötters, J. M. Bernhard, and D. Rödder (2013), Climate-driven range extension of *Amphistegina* (Protista, Foraminiferida): Models of current and predicted future ranges, *PLoS One*, *8*(2), e54443.

- Lohmann, G. P., and P. N. Schweitzer (1990), Globorotalia truncatulinoides' Growth and chemistry as probes of the past thermocline:1. Shell size, *Paleoceanography*, 5, 55–75, doi:10.1029/PA005i001p00055.
- Lombard, F., L. Labeyrie, E. Michel, H. J. Spero, and D. W. Lea (2009), Modelling the temperature dependent growth rates of planktic foraminifera, *Mar. Micropaleontol.*, 70, 1–7.
- Maguire, K. C., and A. L. Stigall (2009), Using ecological niche modeling for quantitative biogeographic analysis: A case study of Miocene and Pliocene Equinae in the Great Plains, *Paleobiology*, 35(4), 587–611.
- MARGO Project Members (2009), Constraints on the magnitude and patterns of ocean cooling at the Last Glacial Maximum, *Nat. Geosci.*, 2, 127–132.
- Martinez-Meyer, E., and A. T. Peterson (2006), Conservatism of ecological niche characteristics in North American plant species over the Pleistocene-to-Recent transition, *J. Biogeogr.*, 33, 1779–1789.
- McKenna, V. S., and W. L. Prell (2004), Calibration of the Mg/Ca of Globorotalia truncatulinoides (R) for the reconstruction of marine temperature gradients, *Paleoceanography*, 19, PA2006, doi:10.1029/2000PA000604.
- Meissner, K. J., A. J. Weaver, H. D. Matthews, and P. M. Cox (2003), The role of land surface dynamics in glacial inception: A study with the UVic Earth System Model, *Clim. Dyn.*, 21, 515–537.
- Merow, C., M. J. Smith, and J. A. Silander Jr. (2013), A practical guide to MaxEnt for modeling species' distributions: What it does, and why inputs and settings matter, *Ecography*, 36(10), 1058–1069.
- Mesgaran, M. B., R. D. Cousens, and B. L. Webber (2014), Here be dragons: A tool for quantifying novelty due to covariate range and correlation change when projecting species distribution models, *Divers. Distrib.*, 20(10), 1147–1159.
- Morey, A. E., A. C. Mix, and N. G. Pisias (2005), Planktonic foraminiferal assemblages preserved in surface sediments correspond to multiple environmental variables, *Quat. Sci. Rev.*, 24(7–9), 925–950.
- Mulitza, S., A. Dürkoop, W. Hale, G. Wefer, and H. S. Niebler (1997), Planktonic foraminifera as recorders of past surface-water stratification, *Geology*, 25, 335–338.
- Naidu, P. D., and B. A. Malmgren (1996), A high resolution record of Late Quaternary upwelling along the Oman Margin, Arabian Sea based on planktonic foraminifera, *Paleoceanography*, 11, 129–140, doi:10.1029/95PA03198.
- Nogués-Bravo, D., J. Rodríguez, J. Hortal, P. Batra, and M. B. Araújo (2008), Climate change, humans, and the extinction of the Woolly Mammoth, *PLoS Biol.*, 6(4), 0685–0692.
- Norris, R. D. (2000), Pelagic species diversity, biogeography, and evolution, *Paleobiology*, 26, 236–258.
- Ortiz, J. D., and A. C. Mix (1997), Comparison of Imbrie-Kipp transfer function and modern analog temperature estimates using sediment trap and core top foraminiferal faunas, *Paleoceanography*, 12, 175–190, doi:10.1029/96PA02878.
- Ortiz, J. D., A. C. Mix, and R. W. Collier (1995), Environmental control of living symbiotic and asymbiotic foraminifera of the California Current, *Paleoceanography*, 10, 987–1009, doi:10.1029/95PA02088.
- Parker, F. L. (1962), Planktonic foraminiferal species in Pacific sediments, *Micropalaeontology*, 8(2), 219–254.
- Pearson, R. G., C. J. Raxworthy, M. Nakamura, and A. T. Peterson (2007), Predicting species distributions from small numbers of occurrence records: A test case using cryptic geckos in Madagascar, *J. Biogeogr.*, 34(1), 102–117.
- Peterson, A. T. (2011), Ecological niche conservatism: A time-structured review of evidence, *J. Biogeogr.*, 38, 817–827.
- Peterson, A. T., M. Papeş, and M. Eaton (2007), Transferability and model evaluation in ecological niche modeling: A comparison of GARP and Maxent, *Ecography*, 30(4), 550–560.
- Peterson, A. T., J. Soberón, R. J. Pearson, R. P. Anderson, M. Nakamura, E. Martínez-Meyer, and M. B. Araújo (2011), *Ecological Niches and Geographical Distributions, Monographs in Population Biology*, vol. 49, Princeton Univ. Press, New Jersey.
- Phillips, S. J., R. P. Anderson, and R. E. Schapire (2006), Maximum entropy modeling of species geographic distributions, *Ecol. Modell.*, 190(3–4), 231–259.
- Quillévéré, F., R. Morard, G. Escarguel, C. J. Douady, Y. Ujiie, T. de Garidel-Thoron, and C. de Vargas (2013), Global scale same-specimen morpho-genetic analysis of *Truncorotalia truncatulinoides*: A perspective on the morphological species concept in planktonic foraminifera, *Palaeogeogr. Palaeoclimatol. Palaeoecol.*, 391, 2–12.
- R Core Team (2015), R: A language and environment for statistical computing, R Foundation for Statistical Computing, Vienna. [Available at <http://www.R-project.org/> (version 3.2.1). Ecospat package (version 2.0) and Dismo package (version 1.0–15).]
- Reiss, Z., B. Luz, A. Almogi-Labin, E. Halicz, A. Winter, M. Wolf, and D. A. Ross (1980), Late Quaternary paleoceanography of the Gulf of Aqaba (Elat), Red Sea, *Quat. Res.*, 14(3), 294–308.
- Russell, A. D., B. Hönisch, H. J. Spero, and D. W. Lea (2004), Effects of seawater carbonate ion concentration and temperature on shell U, Mg, and Sr in cultured planktonic foraminifera, *Geochim. Cosmochim. Acta*, 68, 4347–4361.
- Rutherford, S., S. D. Hondt, and W. Prell (1999), Environmental controls on the geographic distribution of zooplankton diversity, *Nature*, 400, 749–753.
- Saupe, E. E., J. R. Hendricks, R. W. Prtell, H. J. Dowsett, A. Haywood, S. J. Hunter, and B. S. Lieberman (2014), Macroevolutionary consequences of profound climate change on niche evolution in marine molluscs over the past three million years, *Proc. R. Soc. B*, 281, 20141995, doi:10.1098/rspb.2014.1995.
- Schiebel, R., J. Bijma, and C. Hemleben (1997), Population dynamics of the planktic foraminifer *Globigerina bulloides* from the eastern North Atlantic, *Deep Sea Res., Part 1*, 44(9–10), 1701–1713.
- Schmidt, D. N., S. Renaud, and J. Bollmann (2003), Response of planktic foraminiferal size to late Quaternary climate change, *Paleoceanography*, 18(2), 1039, doi:10.1029/2002PA000831.
- Schmidt, D. N., S. Renaud, J. Bollmann, R. Schiebel, and H. R. Thierstein (2004), Size distribution of Holocene planktic foraminiferal assemblages: Biogeography, ecology and adaptation, *Mar. Micropalaeontol.*, 50, 319–338.
- Schmidt, D. N., D. Lazarus, J. R. Young, and M. Kucera (2006), Biogeography and evolution of body size in marine plankton, *Earth Sci. Rev.*, 78(3), 239–266.
- Schmittner, A., N. M. Urban, J. D. Shakun, N. M. Mahowald, P. U. Clark, P. J. Bartlein, A. C. Mix, and A. Rosell-Melé (2011), Climate sensitivity estimated from temperature reconstructions of the Last Glacial Maximum, *Science*, 334, 1385–1388.
- Schneider, R. R., P. J. Müller, G. Ruhland, G. Meineke, H. Schmidt, and G. Wefer (1996), Late Quaternary surface temperatures and productivity in the East-Equatorial South Atlantic: Response to changes in trade/monsoon wind forcing and surface water advection, in *The South Atlantic: Present and Past Circulation*, edited by G. Wefer et al., pp. 527–551, Springer, Berlin.
- Sexton, P. F., and R. D. Norris (2008), Dispersal and biogeography of marine plankton: Long-distance dispersal of the foraminiferal *Truncorotalia truncatulinoides*, *Geology*, 36(11), 899–902.
- Spero, H. J., J. Bijma, D. W. Lea, and B. E. Bemis (1997), Effect of seawater carbonate concentration on foraminiferal carbon and oxygen isotopes, *Nature*, 390, 497–500.

- Stigall, A. L. (2012), Using ecological niche modelling to evaluate niche stability in deep time, *J. Biogeogr.*, *39*(4), 772–781.
- Telford, R. J., C. Andersson, H. J. B. Birks, S. Juggins (2004), Biases in the estimate of transfer function prediction errors, *Paleoceanography*, *19*, PA4014, doi:10.1029/2004PA001072.
- Telford, R. J., C. Li, and M. Kucera (2013), Mismatch between the depth habitat of planktonic foraminifera and the calibration depth of SST transfer functions may bias reconstructions, *Clim. Past*, *9*, 859–870.
- Thiede, J., and B. Jünger (1992), Faunal and floral indicators of ocean coastal upwelling (NW African and Peruvian Continental Margins) from Summerhayes, C. P., Prell, W. L. & Emeis, K. C. (eds.), 1992, *Upwelling Systems: Evolution Since the Early Miocene*. *Geol. Soc. Spec. Publ.*, *64*, 47–76.
- Thunell, R. C., and D. F. Williams (1989), Glacial-Holocene salinity changes in the Mediterranean Sea: Hydrographic and depositional effects, *Nature*, *338*, 493–496.
- Van Nieuwenhove, N., C. Hillaire-Marcel, H. A. Bauch, and A. de Vernal (2016), Sea surface density gradients in the Nordic Seas during the Holocene as revealed by paired microfossil and isotope proxies, *Paleoceanography*, *31*, 380–398, doi:10.1002/2015PA002815.
- Veloz, S. D., J. W. Williams, J. L. Blois, F. He, B. Otto-Bleisner, and Z. Liu (2012), No-analog climates and shifting realized niches during the late quaternary: Implications for 21st-century predictions by species distribution models, *Global Change Biol.*, *18*, 1698–1713.
- Weaver, A. J., et al. (2001), The UVic earth system climate model: Model description, climatology, and applications to past, present and future climates, *Atmos. Ocean*, *39*(4), 361–428.
- Weinmann, A. E., D. Rödder, S. Lötters, and M. Langer (2013), Traveling through time: The past, present and future biogeographic range of the invasive foraminifera *Ammonia* spp. in the Mediterranean Sea, *Mar. Micropalaeontol.*, *105*, 30–39.
- Williams, M., A. M. Haywood, F. J. Gregory, and D. N. Schmidt (Eds.) (2007), *Deep-Time Perspectives on Climate Change: Marrying the Signal from Computer Models and Biological Proxies*, *The Micropalaeontological Society, Special Publications*, The Geol. Soc., London.
- Zeebe, R. E., and D. Wolf-Gladrow (2001), *CO₂ in seawater: equilibrium, kinetics and isotopes*, *Elsevier Oceanography Series*, vol. 65, Elsevier, New York.