

Fungal Genetics Reports

Volume 14

Article 3

Catabolite effects on enzymes

M. Kapoor

D. Bray

Follow this and additional works at: <https://newprairiepress.org/fgr>



This work is licensed under a [Creative Commons Attribution-Share Alike 4.0 License](https://creativecommons.org/licenses/by-sa/4.0/).

Recommended Citation

Kapoor, M., and D. Bray (1969) "Catabolite effects on enzymes," *Fungal Genetics Reports*: Vol. 14, Article 3. <https://doi.org/10.4148/1941-4765.2028>

This Research Note is brought to you for free and open access by New Prairie Press. It has been accepted for inclusion in Fungal Genetics Reports by an authorized administrator of New Prairie Press. For more information, please contact cads@k-state.edu.

Catabolite effects on enzymes

Abstract

Catabolite effects on enzymes

K apoor, M. and D. Bray. Catabolite effects on some enzymes of *Neurospora*

During an experiment designed to study the effect of growth conditions on the activity and synthesis of glutamine synthetase, several interesting observations were made. *Neurospora crassa* pe (Y 8743m) (FGSC #37) was used as a source of enzymes in this study. All cultures were prepared in Vogel's minimal medium with sucrose

or glucose as a carbon source, and mycelial powders were obtained as described in the communication on phosphofructokinase in this issue of the *Neurospora* Newsletter. The cultures were grown for 30 hours at 28°C. There was no autolysis in cultures with low concentration of sucrose. At the end of 30 hours not all of the sugar in the medium was exhausted, a very small amount remaining. Crude extracts were prepared by extracting at 3°C lyophilized mycelium powder in 0.05 M phosphate buffer (5 x 10⁻⁴ M in EDTA and 10⁻⁴ M in β-mercaptoethanol) pH 7.5, for 30 minutes, straining the mixture through four layers of cheesecloth and centrifuging the supernatant at 27,000 x g for 15 minutes. The residue was discarded and the supernatant was used without further purification.

Table 1. Effect of sucrose on some enzymes of *Neurospora*.

Sucrose concentration	Specific activity (OD/mg protein)			
	GDH-D	GDH -T	GluN-S'ase	PK
0.1%	0.75	0.05	0.005	0.12
	0.75	0.05	0.000	0.12
0.5%	0.56	0.37	0.18	1.30
	0.53	0.38	0.17	1.40
1.0%	0.45	0.70	0.32	2.00
	0.43	0.70	0.32	2.20
1.5%	0.23	0.81	0.41	2.1
	0.24	0.79	0.41	2.1
2.0%	0.15	0.86	0.42	1.75
	0.15	0.86	0.42	2.00
2.5%	0.12	0.98	0.45	2.59
	0.19	0.98	0.44	2.30

Activities of glutamine synthetase (GluN-S'ase), NAD-specific glutamate dehydrogenase (GDH-D), NADP-specific glutamate dehydrogenase (GDH -T) and pyruvate kinase (PK) were determined in extracts of mycelia obtained from cultures grown in different concentrations of sucrose. Arrays of the activities of the two glutamate dehydrogenases were performed with a Gilford model 2000 recording spectrophotometer by following the initial decrease in OD at 340 mμ accompanying the reductive amination of α-ketoglutarate in the presence of ammonia and reduced NAD or reduced NADP (K apoor and Smith 1968 *Can. J. Microbiol.* 14:609). GluN-S'ase was assayed by measuring the formation of γ-glutamyl hydroxamate from L-glutamate and hydroxylamine in the presence of ATP (K apoor and Bray 1968 *Biochemistry* 7:3583). PK was measured by following the decrease in OD at 340 mμ in the following reaction mixture at 25°C: Tris-HCl, pH 8.0, 100 μmoles; MgCl₂ 10 μmoles; ADP 1 μmole; reduced NAD 0.14 μmole; PEP 0.6 μmole; LDH (Sigma) 100 μg and enzyme preparation in a total volume of 3 ml.

Table 1 shows the specific activities of these enzymes in crude extracts of *Neurospora* mycelium grown at concentrations varying from 0.1% to 2.5%. Glutamine synthetase is not repressed by sucrose and

neither is pyruvate kinase; both these enzymes show an increase in specific activity in the presence of sucrose up to 1.5% but no further increase was noted at 2% and 2.5% sucrose. A study of the response of the two GDH's towards sucrose in the growth medium revealed a dramatic feature of regulation of GDH-T and GDH-D. Whereas GDH-D is subject to catabolite repression by sucrose and glucose, GDH-T is induced under the same conditions, thus demonstrating a reciprocal relationship between these two enzymes. It is already known that in the presence of glutamate or ammonia in the medium GDH-D is induced with a simultaneous repression

of GDH-T (Sanwal and Lata 1962 Arch. Biochem. Biophys. 98:420). It has been suggested that GDH-D is primarily a catabolic enzyme and that GDH-T serves an anabolic function in the cell. Our studies are in agreement with this suggestion in so far as it is GDH-D alone that is subject to catabolite repression and that GDH-T is induced under the same conditions. ■ ■ ■ Department of Biology, University of Calgary, Calgary, Alberta, Canada.