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- 1 Characterizing neonatal vitamin D deficiency in the modern era: a maternal-
- 2 neonatal birth cohort from Southern Europe

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Abstract

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Absence of adequate maternal vitamin D supplementation and decreased maternal ultraviolet exposure during pregnancy are key determinants for the manifestation of neonatal hypovitaminosis D at birth. These parameters may vary, according to countryspecific dietary patterns, health policies and sunshine exposure. We aimed to investigate differences in calcium metabolism and anthropometric profiles according to neonatal vitamin D status at birth, in a sunny region of Northern Greece. A secondary aim was to identify maternal parameters as risk factors for developing neonatal vitamin D deficiency at birth. A total of 129 mother-neonate pairs were included in the study and classified into three groups, according to neonatal 25-hydroxy-D [25(OH)D)] concentrations at birth [deficiency (<30 nmol/l), insufficiency (30-50 nmol/l) and sufficiency (>50 nmol/l)]. Neonatal biochemical and anthropometric profiles and maternal demographic, social, dietary and biochemical profiles were comparatively evaluated between the three groups. Univariate and multivariate logistic regression was performed to identify independent associations of maternal factors with neonatal vitamin D status. Vitamin D deficient-neonates manifested higher parathyroid hormone $(7.20\pm2.60 \text{ vs } 5.50\pm1.50 \text{ pg/ml}, p=0.01)$ and lower corrected calcium $(10.70\pm0.70 \text{ vs})$ 11.30±1.30 mg/dl, p=0.02) concentrations compared with vitamin D-insufficient neonates. Mothers of vitamin D deficient and insufficient neonates had a lower total of 25(OH)D (31.7±19.2 and 36.5±22.3 vs 53.3±39.0 nmol/l, p<0.01) and 25(OH)D₃ $(27.4\pm17.5 \text{ and } 33.3\pm19.9 \text{ vs } 47.3\pm36.7 \text{ nmol/l}, p<0.01 \text{ and } p=0.04, \text{ respectively})$ concentrations respectively, compared with those of vitamin D-sufficient neonates. Maternal use of alcohol during pregnancy was associated with a 5.57-fold higher risk for neonatal vitamin D deficiency at birth (OR 5.57, 95% CI1.17-26.56, p=0.03). Newborns with vitamin D deficiency presented a 6.89-fold higher risk of having been

- 57 given birth by vitamin D deficient mothers (OR 6.89, 95% CI 3.09-15.38, p<0.01). In
- 58 conclusion, neonatal vitamin D deficiency is associated with maternal 25(OH)D
- 59 concentrations at birth and maternal alcohol use. Further studies are required to replicate
- these findings in other regions and populations.

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62 **Keywords:** Vitamin D; pregnancy; neonatal health; calcium; rickets.

1. Introduction

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65	Despite the availability of supplementation and numerous published guidelines for its
66	prevention [1,2], the resurgence of nutritional rickets (NR) secondary to vitamin D
67	deficiency and/or dietary calcium (Ca) deficiency is becoming increasingly prevalent
68	worldwide, highlighting the potential risks of not gaining sufficient vitamin D through
69	diet, supplementation or exposure to ultraviolet (UVB) radiation. NR remains an
70	important global, public health problem, with an established adverse impact on the
71	skeletal and mental development of neonates and infants [3-5]. Recent reports detail the
72	importance of maternal serum vitamin D concentrations in determining neonatal
73	vitamin D status [6-10].
74	Although guidelines resulted in an improvement of the management of maternal
75	hypovitaminosis D in the daily clinical setting [11], many aspects of such an approach
76	are largely affected by country-specific dietary patterns, public health policies and
77	variation of UVB exposure, due to cultural and life-style reasons [8,12]. In this context,
78	results from regional studies are essential for establishing appropriate preventive
79	strategies during pregnancy, to optimize vitamin D status in the mother and the
80	neonate, in a country-specific approach. The aim of this maternal-neonatal birth cohort
81	study was to investigate differences in Ca metabolism and anthropometric profiles
82	according to neonatal vitamin D status at birth, in a sunny region of Northern Greece.
83	A secondary aim was to identify maternal parameters as risk factors for developing
84	neonatal vitamin D deficiency at birth.

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2. Methods

87 2.1 Study criteria

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The inclusion criterion was full-term pregnancy (gestational week 37-42). Maternal exclusion criteria were primary hyperparathyroidism, secondary osteoporosis, heavy alcohol use (≥7 alcohol units per week or ≥6 units at any time during pregnancy), hyperthyroidism, nephritic syndrome, inflammatory bowel disease, rheumatoid arthritis, osteomalacia, obesity [body mass index (BMI)>30 kg/m²], gestational diabetes and use of medications affecting Ca or vitamin D status (e.g. corticosteroids), except for Ca and vitamin D supplements. Neonatal exclusion criteria were being small-for-gestational age (SGA) and presence of severe congenital anomalies. Informed consent was obtained from all mothers. The protocol received approval from the Bioethics Committee of the Aristotle University of Thessaloniki, Greece (approval number 1/19-12-2011).

Pregnant women on regular follow-up, were recruited from the Maternity Unit of

2.2 Demographics and dietary assessment

At enrolment, maternal demographic and social characteristics, as well as dietary habits, were recorded. Ca and vitamin D dietary intake during the last month of pregnancy were assessed through a validated, semi-quantitative, food frequency questionnaire that includes 150 foods and beverages [13]. For each dietary item, participants were asked to report their frequency of dairy products consumption and portion size. From these data, calculations were made for estimations of consumed quantities (in gr per day) based on a food composition database, modified to accommodate the particularities of the Greek diet [14] for estimating daily dietary Ca and vitamin D intake. Maternal education was classified as elementary (primary),

standard (secondary) and higher (tertiary and holding of academic degrees). Maternal alcohol use during pregnancy was treated as a dichotomous variable, defined either as none (subdivided in never drinking alcohol or drinking alcohol but not during pregnancy) or light (1-2 units per week or at any one time during pregnancy) / moderate (3-6 units per week or at any one time during pregnancy) [15].

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2.3 Biochemical and hormonal assays

Blood samples were obtained from mothers by antecubital venipuncture 30-60 min before delivery. Umbilical cord blood was collected immediately after clamping, from the umbilical vein. Serum and umbilical cord specimens were stored at -20°C prior to analysis for the following parameters: Ca, phosphorus (P), parathyroid hormone (PTH), 25-hydroxyvitamin D₂ [25(OH)D₂] and D₃ [25(OH)D₃]. Serum Ca and P determinations were performed using the Cobas INTEGRA clinical chemistry system (D-68298; Roche Diagnostics, Mannheim, Germany). The inter- and intra-assay coefficients of variation were 0.99% and 3.5% for Ca, and 1.3% and 2.5% for P, respectively. PTH determinations were performed using the electrochemiluminescence immunoassay ECLIA (Roche Diagnostics GmbA, Mannheim, Germany). Reference range for PTH was 15-65 pg/ml, functional sensitivity 6.0 pg/ml, within-run precision 0.6-2.8% and total precision 1.6-3.4%. Concentrations of 25(OH)D₂ and 25(OH)D₃ were determined using novel assay, liquid chromatography-tandem mass spectrometry (LC-MS/MS), with lower limits of quantification (LLOQ): 25(OH)D₂ (0.5 ng/ml), 25(OH)D₃ (0.5 ng/ml). Briefly, the assay involves analyte purification using liquid-liquid extraction followed by chromatographical separation using a chiral column in tandem with a rapid resolution microbore column. Full method validation parameters have been previously reported [16,17].

2.4 Neonatal and maternal vitamin D status

Neonates were classified into three groups according to 25(OH)D status: Group A (deficiency, 25(OH)D < 30 nmol/l), Group B (insufficiency, $30 \le 25(OH)D \le 50 \text{ nmol/l}$) and Group C (sufficiency 25(OH)D > 50 nmol/l) [2]. Respectively, mothers were classified according to neonatal 25(OH)D concentrations at birth, into the following groups: those who gave birth to deficient (group A), insufficient (group B) or sufficient neonates (group C). Maternal vitamin D status was also included in subsequent uniand multivariate analysis as follows: vitamin D sufficiency $\ge 50 \text{ nmol/l}$ and Vitamin D insufficiency $\le 50 \text{ nmol/l}$ [18].

2.5 Maternal and neonatal anthropometry

At enrolment, maternal and neonatal anthropometry was recorded. Maternal prepregnancy BMI was either normal (18-25 kg/m²) or overweight (25-30 kg/m²). All neonatal anthropometric measurements were performed by the same trained nurse, between 12 and 72 hours of age. The following measurements were recorded: birth weight, height, neck-rump, upper arm, femur and knee heel lengths; head, chest, abdominal, upper arm and middle thigh circumferences; and abdominal skin fold thickness. The birth weight of the neonates was measured on regularly calibrated scales. The knee-heel length was measured with a hand-held BK5 infant knemometer (Force Technology, Brondby, Denmark). Instrument software calculated the mean of 10 sequential readings and generated a printed report of all readings and the calculated mean. Neonatal height was measured to the nearest mm using an Ellard newborn length board (Ellard Instrumentation Ltd., Seattle, WA). Abdominal, upper arm and middle thigh head, mid-upper arm, and maximal head circumferences were measured using a plastic encircling tape (Child Growth Foundation, London, UK). Abdominal skinfold was measured using Holtain calipers (Holtain, Crymych, UK).

2.6 UVB measurements

UVB radiation includes wavelengths from 280 to 320 nm. UVB data for the broad geographical region of Thessaloniki, Greece were collected at the Laboratory of Atmospheric Physics, School of Physics, Aristotle University of Thessaloniki.

The daily integral of vitamin D effective UVB radiation (09:00 to 16:00 local time) was used as the most representative parameter for UVB exposure. These hours were selected as indicatives, since they are related to the beginning and the end of the working period for the majority of the population. Individual sunlight exposure was recorded for each participant during that period. Finally, mean UVB exposure during the previous 45 days (daily integral) before blood sample collection (estimated mean half-life of vitamin D) was calculated for each participant.

2.7 Statistical analysis

Kolmogorov-Smirnov and Shapiro-Wilk tests were used to check the normality of distribution of the continuous variables in the whole study sample and in the neonates' vitamin D status groups, respectively. Normally distributed continuous data were presented as mean \pm standard deviation (SD) and non-normally distributed as median (interquartile range) (IQR). One-way analysis of variance (ANOVA) was used to compare means among groups. Chi-square test was used for between-group comparisons, in case of categorical variables. Categorical data were presented as

absolute numbers and frequencies (percentages). Univariate and multivariate logistic regression was performed to identify independent associations of maternal factors with neonatal vitamin D status. Maternal factors significantly associated to neonatal vitamin D status in the univariate analysis were selected for the multivariate logistic regression, indicating each factor's effect on neonates' vitamin D deficiency after adjusting for the other factors. Statistical analysis was performed using IBM Statistics software SPSS 23.0 for Windows. A p-value <0.05 was considered as significant.

3. Results

A total of consecutive 129 mother-neonate pairs were included in the analysis. Recruitment period was from February to November. Mean gestational age was 38.6 ± 1.5 weeks, and 53.6% of births occurred between September and March. Mean birth weight of neonates was 3269 ± 408 gr and mean neonatal 25(OH)D concentrations were 37.5 ± 27.4 nmol/l. Neonates at the range of severe vitamin D deficiency manifested mean 25(OH)D concentrations of 15.9 ± 7.5 nmol/L compared with 38.4 ± 5.0 nmol/l and 71.0 ± 30.5 nmol/l of insufficient and sufficient ones, respectively. Vitamin D deficient neonates manifested higher PTH $(7.20\pm2.60 \text{ vs } 5.50\pm1.50 \text{ pg/ml}, \text{ p=0.01})$ and lower corrected Ca $(10.70\pm0.70 \text{ vs } 11.30\pm1.30 \text{ mg/dl}, \text{ p=0.02})$ concentrations compared with vitamin D insufficient neonates, being not different from neonates with sufficiency [(PTH: $7.2\pm2.6 \text{ vs } 6.3\pm2.0 \text{ pg/ml}, \text{ p=0.22})$ and (corrected Ca: $10.7\pm0.7 \text{ vs } 10.8\pm0.7, \text{ p=1.00}$)]. Epimers did not differ among various groups (**Table 1**). Vitamin D insufficient neonates had higher lower leg length compared with sufficient ones $(15.6\pm1.5 \text{ vs } 13.0\pm1.9 \text{ cm}, \text{ p<0.01})$, whereas manifested lower knee-heel length compared with vitamin D deficient ones $(8.0\pm1.3 \text{ vs } 9.0\pm0.8 \text{ cm}, \text{ p<0.01})$ (**Table 2**).

Mothers of vitamin D deficient and insufficient neonates had lower total 25(OH)D
$[31.7\pm19.2 \text{ and } 36.5\pm22.3 \text{ vs } 53.3\pm39.0 \text{ nmol/l}, p<0.01] \text{ and } 25(OH)D_3 \ [27.4\pm17.5 \text{ and } 25(OH)D_3 \ [27$
33.3±19.9 vs 47.3±36.7 nmol/l, p<0.01 and p=0.04, respectively] concentrations,
compared with sufficient ones (Table 3). The 37.5% of mothers of sufficient neonates
at birth had higher education, compared with 10.6% of mothers of insufficient ones
(p<0.01) (Table 4).
In a univariate analysis, various maternal factors (age, smoking, alcohol consumption,
vitamin D status, Ca supplementation, dietary daily Ca and vitamin D intake during
the third trimester, pre-pregnancy BMI, delivery BMI, education level, UVB exposure)
were included, to assess their impact on neonatal vitamin D status. Maternal use of
alcohol during pregnancy (≥1 unit per week) was associated with a 5.57-fold higher
probability for neonatal vitamin D deficiency at birth (OR 5.57, 95% CI 1.17-26.56,
p=0.03). These results remained significant in a multivariate analysis [OR 6.18
(adjusted for maternal vitamin D status and UVB exposure)], 95% CI 1.18-32.51,
p=0.03] (Table 5). Risk of neonatal vitamin D deficiency was 6.89-fold increased for
infants born by vitamin D deficient (<50 nmol/l), when compared with vitamin D
sufficient mothers (>50 nmol/l) (OR 6.89, 95% CI 3.09-15.38, p<0.01). The results
remained significant in the multivariate analysis [OR (adjusted for alcohol consumption
and UVB exposure)] 7.62, 95% CI 3.27-17.78] (Table 5). UVB exposure during
summer months, was associated with 3.51-fold higher probability for neonatal vitamin
D deficiency at birth (OR 3.51, 95% CI1.87-11.42, p=0.03); this association did not
retain its significance in the multivariate analysis.

4. Discussion

The present study captures the current status of neonatal vitamin D deficiency at birth in a sunny Mediterranean region, characterized by both PTH and corrected Ca concentrations within the normal range, however, significantly decreased compared with insufficient neonates. Neonatal birth anthropometry was not affected by neonatal vitamin D deficiency, with the exclusion of a significant increase in knee-heel length. It was also demonstrated that maternal alcohol use and maternal vitamin D insufficiency (<50 nmol/l) during pregnancy are risk factors for the development of neonatal vitamin D deficiency at birth. Neonatal vitamin D deficiency is a major risk factor for the development of acute and chronic metabolic complications including NR, hypocalcemia and impairment of optimal skeletal development of the offspring [1-3]. Recent reports underline the importance of maternal circulating vitamin D concentrations in determining neonatal vitamin D status [6-8]. Preventive strategies are primarily focused on the sufficiency of maternal vitamin D status during pregnancy, either through appropriate vitamin D supplementation or sunshine exposure. In accordance with our previous findings [19] regarding a high prevalence of maternal hypovitaminosis D during pregnancy in the Mediterranean countries, results from this cohort describe a high prevalence of neonatal vitamin D sufficiency at birth, in this region. The main reasons behind this phenomenon seem to be racial, social and cultural particularities, as well as the absence of preventive strategies, including food fortification policies in our region, that mitigate the benefits of sun exposure. In detail, dark skin, dressing habits, and sunshine avoidance, especially during the hot summer months, have been associated with an increased prevalence of maternal hypovitaminosis D during pregnancy [12,19]. PTH becomes a major regulator of mineral and bone homeostasis within the first hours after birth [20]. Parathyroid glands increase the synthesis and secretion of PTH, which

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acts to raise serum Ca, lower P, stimulate calcitriol synthesis, inhibit calcitriol catabolism, reabsorb Ca in the kidney tubules, and up regulate bone formation [21]. The significant increase of PTH concentrations - albeit within the normal range - in vitamin D deficient neonates observed in this study, might result from an in utero adaptive up-regulation of fetal PTH, as a result of maternal-fetal hypovitaminosis D, to maintain adequate Ca supply for the developing infant [9]. On the other hand, absence of correlation between 25(OH)D and intact PTH in cord blood has been previously reported [22-24], indicating that the well documented rise of PTH in adults with secondary hyperparathyroidism, is not evident in neonates after birth, as a result of a temporary PTH suppression. Further studies are necessary in order to evaluate the magnitude of neonatal PTH variation in the context of neonatal vitamin D status at birth. Neonatal anthropometry at birth was not remarkably affected by neonatal vitamin D deficiency in our cohort. Although lower mean neonatal knee-heel length at birth has been associated with lower maternal 25(OH)D levels between 28 and 32 weeks of gestation [25], this observation was not replicated in vitamin D deficient neonates at birth. Our results are in agreement with the findings reported by Czech-Kowalska et al [26], who failed to establish an association between neonatal vitamin D status and neonatal anthropometry and adiposity in a similar to our study population, including appropriate for gestational age neonates. Contrariwise, Sauder et al. [27] proved a relationship between higher neonatal 25(OH)D concentrations and lower birthweight and neonatal adiposity, after adjustment for gestational age at birth. Therefore, the fact that SGA infants were not included in our analysis, might have had an impact on the pattern of our findings.

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Neonatal 25(OH)D concentrations at birth roughly follow the maternal pattern in the deficient and insufficient mother groups, while resembling uniform distribution in the group of mothers with sufficient vitamin D status [6]. The results of this study suggest that both mothers of deficient and insufficient neonates manifested significant differences in their vitamin D status compared to those of sufficient ones, indicating that both profiles fall within the same pathophysiological high-risk zone, for development of neonatal vitamin D deficiency at birth. On the other hand, mothers who gave birth to vitamin D sufficient neonates demonstrated mean total 25(OH)D concentrations >50 nmol/l, identifying a potential, maternal, low-threshold to be targeted during pregnancy. Of course, these values are country-specific and might not be applied to other regions and countries. The present study revealed an association between maternal alcohol use, defined as ≥ 1 unit per week, and hypovitaminosis D during pregnancy and the development of neonatal vitamin D deficiency. There is a paucity of studies concerning the effects of alcohol on vitamin D status during pregnancy. In a cohort of Ukrainian pregnant women [28], alcohol-exposed women had lower 25(OH)D concentrations than low/unexposed women during spring and winter. Of major interest, vitamin D concentrations were lower in patients with alcohol use disorders, whose last alcohol intake was within the last 30 days compared with those who had abstained >30 days from alcohol [29]. In non-pregnant rats, chronic alcohol consumption can lead to depletion of vitamin D stores [30]. Multiple factors have been proposed as potential explanations for the inverse association between alcohol use and vitamin D status, including poor diet, malabsorption and restricted exposure to natural sunlight, which are all commonly seen in heavy drinkers [28]. A direct effect of alcohol on vitamin D biodynamics has been also postulated, possibly related to alcoholic liver disease that disrupts protein

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synthesis, resulting in low levels of vitamin D binding protein and – subsequently - of the active form of vitamin D [31]. Shankar et al. [32] have demonstrated that alcohol reduces circulating 1,25 (OH)2 D₃ levels by inducing CYP24A1, resulting from renal oxidative stress due to local ethanol metabolism. On the other hand, a systematic review on the relationship between alcohol use and vitamin D [33] concluded that studies reporting positive associations between alcohol units per day and vitamin D serum concentrations had better study designs and larger sample sizes compared with those that established the opposite findings. Greater exposure to sunlight due to the homeless status of many alcoholics was suggested by the authors as a plausible explanation for this observation. It is evident that the relationship between alcohol and vitamin D is mediated by complex factors, equally related to the social and health consequences of alcohol abuse, thus warranting further evaluation by future studies. Two characteristics of this cohort deserve special attention. First, the study did not observe differences between maternal groups in dietary Ca and vitamin D intake during pregnancy, nor an impact of these parameters on the risk of neonatal hypovitaminosis D. A systematic review and meta-analysis on micronutrient intake during pregnancy in developed countries [34], reported that dietary vitamin D intake among pregnant women is insufficient; still, it has a limitted impact on neonatal vitamin D status. In addition, recent findings from a birth-cohort of 567 women from northern Sweden [35], indicated that more than half of the women participated, had intake levels of vitamin D lower than those recommended (median level of 0.85 µg/MJ/day vs. recommended level of 0.98 µg/MJ/day). These results are in accordance with our findings, derived from a sunny region of Southern Europe. Moreover, the vast majority of the women included in this analysis regularly followed the typical Mediterranean diet during

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pregnancy. Given that the analysis lacks a detailed evaluation of dietary protein and fat intake as potential food sources of vitamin D, future studies are needed to elucidate the hypothesis about a potential neutral effect of this dietary pattern on maternal vitamin D status. A second finding was the inverse association (although not confirmed in the multivariate analysis) between UVB exposure and vitamin D equilibrium. Whole-body sun exposure at the right time of the year is required, in order for circulating vitamin D levels to be affected [28,29]. In contrast, partial exposure (5-10%) of total body surface to intense sunlight produces only a limited amount of vitamin D. It could be hypothesized that, under high-temperature climatic conditions that are present during spring and summer months in Greece, most pregnant women limit their outdoor activities during the morning and afternoon hours. In this context, high UVB radiation, especially during hot summer months, might comprise a risk factor for reduced maternal sunshine exposure which could in turn adversely affect maternal vitamin D status during pregnancy, leading to a high prevalence of neonatal vitamin D deficiency [36,37]. Additional studies are required in this direction. This study has several limitations, including the relatively small sample size, which may explain why additional parameters known to affect maternal vitamin D status during pregnancy and result in neonatal vitamin D deficiency, were not found to be regulators of 25(OH)D concentrations. In this regard, skin color has been reported to be a determinant of vitamin D status [38]. However, in this study we chose not to include a rough estimate of skin pigmentation as, for example, the Fitzpatrick's scale, given that most women of Greek origin belong to a relatively homogenous group of mild dark or white skin phototypes [36]. Finally, the exclusion of obese pregnant

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358 women from our cohort could be considered as an additional limitation, since maternal 359 obesity is a well-known risk factor for maternal vitamin D deficiency [39]. In conclusion, this study reported results from a maternal-neonatal Greek cohort, 360 361 primarily focusing on neonatal vitamin D deficiency. It demonstrated a high prevalence 362 of vitamin D deficient neonates at birth. This phenomenon was primarily mediated by 363 maternal 25(OH)D concentrations at birth and maternal alcohol use. Adequate and appropriate sun exposure may not be indispensable for avoiding maternal 364 365 hypovitaminosis D, even in the sunny Mediterranean region. These data could provide 366 a targeted approach based on specific-population characteristics for future vitamin D 367 supplementation studies and help to recognize parameters necessary for developing 368 health policies to prevent neonatal vitamin D deficiency in this region.

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378 References

- 1. Callaghan AL, Moy RJ, Booth IW, Debelle G, Shaw NJ. Incidence of symptomatic
- vitamin D deficiency. ArchDisChild. 2006;91:606-7.
- **2.** Munns CF, Shaw N, Kiely M, Specker BL, Thacher TD, Ozono K, Michigami T,
- Tiosano D, Mughal MZ, Mäkitie O, Ramos-Abad L, Ward L, DiMeglio LA,
- Atapattu N, Cassinelli H, Braegger C, Pettifor JM, Seth A, Idris HW, Bhatia V, Fu
- J, Goldberg G, Sävendahl L, Khadgawat R, Pludowski P, Maddock J, Hyppönen E,
- Oduwole A, Frew E, Aguiar M, Tulchinsky T, Butler G, Högler W. Global
- Consensus Recommendations on Prevention and Management of Nutritional
- 387 Rickets. J Clin Endocrinol Metab. 2016;101:394-415.
- 388 3. Pettifor JM. Vitamin D deficiency and nutritional rickets in children. In: Feldman
- D, Pike JW, Adams J, editors, Vitamin D. 3rd ed. London: Elsevier Inc; 2011.
- 390 p.1107-28.
- 391 4. Maiya S, Sullivan I, Allgrove J, Yates R, Malone M, Brain C, Archer N, Mok Q,
- Daubeney P, Tulloh R, Burch M.Hypocalcaemia and vitamin D deficiency: an
- important, but preventable, cause of life-threatening infant heart failure. Heart.
- 394 2008;94:581-4.
- 395 5. Pettifor JM. Nutritional rickets. In: Glorieux FH, Pettifor JM, Juppner H, editors,
- Pediatric Bone: Biology and Diseases. 2nd ed. Amsterdam, The Netherlands:
- 397 Elsevier; 2012: p.625-54.
- 398 **6.** Karras SN, Shah I, Petroczi A, Goulis DG, Bili H, Papadopoulou F, Harizopoulou
- V, Tarlatzis BC, Naughton DP. An observational study reveals that neonatal
- vitamin D is primarily determined by maternal contributions: implications of a new
- assay on the roles of vitamin D forms. Nutr J. 2013;12:77.

- 7. Pilz S, Zittermann A, Obeid R, Hahn A, Pludowski P, Trummer C, Lerchbaum E,
- 403 Pérez-López FR, Karras SN, März W. The Role of Vitamin D in Fertility and during
- 404 Pregnancy and Lactation: A Review of Clinical Data. Int J Environ Res Public
- 405 Health. 2018;15:E2241.
- **8.** Karras SN, Wagner CL, Castracane VD. Understanding vitamin D metabolism in
- 407 pregnancy: From physiology to pathophysiology and clinical
- 408 outcomes. Metabolism. 2018;86:112-23.
- 409 9. Wagner CL, Hollis BW, Kotsa K, Fakhoury H, Karras SN. Vitamin D
- administration during pregnancy as prevention for pregnancy, neonatal and
- postnatal complications. Rev Endocr Metab Disord. 2017;18:307-22.
- 412 **10.** Anastasiou A, Karras SN, Bais A, Grant WB, Kotsa K, Goulis DG. Ultraviolet
- radiation and effects on humans: the paradigm of maternal vitamin D production
- during pregnancy. Eur J ClinNutr. 2017;71:1268-72.
- 415 **11.** Holick MF, Binkley NC, Bischoff-Ferrari HA, Gordon CM, Hanley DA, Heaney
- 416 RP, Murad MH, Weaver CM; Endocrine Society. Evaluation, treatment, and
- prevention of vitamin D deficiency: an Endocrine Society clinical practice
- 418 guideline. J Clin Endocrinol Metab. 2011;96:1911-30.
- 419 **12.** Karras SN, Anagnostis P, Paschou SA, Kandaraki E, Goulis DG. Vitamin D status
- during pregnancy: time for a more unified approach beyond borders? Eur J
- 421 ClinNutr. 2015;69:874-7.
- 422 **13.** Gnardellis C, Trichopoulou A, Katsouyanni K, Polychronopoulos E, Rimm EB,
- Trichopoulos D. Reproducibility and validity of an extensive semi-quantitative food
- frequency questionnaire among Greek school teachers. Epidemiology. 1995;6:74-
- 425 7.

- 426 **14.** Trichopoulou A, GeorgaK. Composition tables of foods and Greek dishes. Athens
- 427 Greece: Parisianos, 2004.
- 428 **15.** Connelly R, Platt L. Cohort profile: UK Millennium Cohort Study (MCS). Int J
- 429 Epidemiol. 2014;43:1719-25.
- 430 **16.** Shah I, James R, BarkerJ, Petroczi A, Naughton DP. Misleading measures
- inVitamin D analysis: a novel LC-MS/MS assay to account for epimers and isobars.
- 432 Nutr J. 201114;10:46.
- 433 17. Shah I, Petroczi A, Naughton DP. Method for simultaneous analysis of eight
- analogues of vitamin D using liquid chromatography tandem mass spectrometry.
- 435 Chem Central J.2012;6:112.
- **18.** Ross AC. The 2011 report on dietary reference intakes for calcium and vitamin D.
- 437 Public Health Nutr. 2011;14:938-9.
- 438 19. Karras S, Paschou SA, Kandaraki E, Anagnostis P, Annweiler C, Tarlatzis BC,
- Hollis BW, Grant WB, Goulis DG. Hypovitaminosis D in pregnancy in the
- Mediterranean region: a systematic review. Eur J Clin Nutr. 2016;70:979-86.
- **20.** Kovacs CS. Bone development and mineral homeostasis in the fetus and neonate:
- roles of the calciotropic and phosphotropic hormones. Physiol Rev.2014;94:1143-
- 443 218.
- 21. Kovacs CS. The role of vitamin D in pregnancy and lactation: insights from animal
- models and clinical studies. Annu Rev Nutr. 2012;32:97-123.
- 22. Bowyer L, Catling-Paull C, Diamond T, Homer C, Davis G, Craig ME, Vitamin D,
- PTH and calcium levels in pregnant women and their neonates. Clin Endocrinol
- 448 (Oxf). 2009;70:372-7.
- 23. Czech-Kowalska J, Pludowski P, Dobrzanska A, Kryskiewicz E, Karczmarewicz
- E, Gruszfeld D, Pleskaczynska A, Golkowska M. Impact of vitamin D

- supplementation on markers of bone mineral metabolism in term infants. Bone.
- 452 **2012;51:781-6**.
- 453 **24.** Young BE, McNanley TJ, Cooper EM, McIntyre AW, Witter F, Harris ZL, O'Brien
- KO. Young BE, McNanley TJ, Cooper EM, McIntyre AW, Witter F, Harris ZL,
- O'Brien KO. Maternal vitamin D status and calcium intake interact to affect fetal
- skeletal growth in utero in pregnant adolescents. Am J Clin Nutr. 2012;95:1103-12.
- 457 **25.** Morley R, Carlin JB, Pasco JA, Wark JD. Maternal 25-hydroxyvitamin D and
- parathyroid hormone concentrations and offspring birth size. J Clin Endocrinol
- 459 Metab. 2006;91:906-12.
- 26. Czech-Kowalska J, Gruszfeld D, Jaworski M, Bulsiewicz D, Latka-Grot J,
- Pleskaczynska A, Lygas J, Wygledowska G, Pawlus B, Zochowska A, Borszewska-
- Kornacka MK, Dobrzanska A. Determinants of Postpartum Vitamin D Status in the
- Caucasian Mother-Offspring Pairs at a Latitude of 52 degrees N: A Cross-Sectional
- 464 Study. Ann Nutr Metab. 2015;67:33-41.
- 27. Sauder KA, Koeppen HJ, Shapiro ALB, Kalata KE, Stamatoiu AV, Ringham BM,
- Glueck DH, Norris JM, Dabelea D. Prenatal Vitamin D Intake, Cord Blood 25-
- Hydroxyvitamin D, and Offspring Body Composition: The Healthy Start Study.
- 468 Nutrients. 2017;9:E790.
- **28.** Carlson CR Jr, Uriu-Adams JY, Chambers CD, Yevtushok L, Zymak-Zakutnya N,
- Chan PH, Schafer JJ, Wertelecki W, Keen CL. Vitamin D Deficiency in Pregnant
- 471 Ukrainian Women: Effects of Alcohol Consumption on Vitamin D Status. J Am
- 472 CollNutr. 2017;36:44-56.
- 29. Neupane SP, Lien L, Hilberg T, Bramness JG. Vitamin D deficiency in alcohol-use
- disorders and its relationship to comorbid major depression: a cross-sectional study
- of inpatients in Nepal. Drug Alcohol Depend. 2013;133:480-5.

- 476 **30.** Gascon-Barre M. Interrelationships between vitamin D3 and 25-hydroxyvitamin
- D3 during chronic ethanol administration in the rat. Metabolism. 1982;31:67-72.
- 478 **31.** Bikle DD, Halloran BP, Gee E, Ryzen E, Haddad JG. Free 25-hydroxyvitamin D
- levels are normal in subjects with liver disease and reduced total 25-hydroxyvitamin
- 480 D levels. J Clin Invest. 1986;78:748-52.
- 481 **32.** Shankar K, Liu X, Singhal R, Chen JR, Nagarajan S, Badger TM, Ronis MJ.
- 482 Chronic ethanol consumption leads to disruption of vitamin D3 homeostasis assoc
- iated with induction of renal 1,25 dihydroxyvitamin D3-24-
- 484 hydroxylase (CYP24A1). Endocrinology. 2008;149:1748-56.
- 485 **33.** Tardelli VS, Lago MPPD, Silveira DXD, Fidalgo TM. Vitamin D and alcohol: A
- review of the current literature. Psychiatry Res. 2017;248:83-6.
- 487 **34.** Blumfield ML, Hure AJ, Macdonald-Wicks L, Smith R, Collins CE. A systematic
- 488 review and meta-analysis of micronutrient intakes during pregnancy in developed
- 489 countries. Nutr Rev. 2013;71:118-32.
- 490 **35.** Stråvik M, Jonsson K, Hartvigsson O, Sandin A, Wold AE, Sandberg AS, Barman
- M. Food and Nutrient Intake during Pregnancy in Relation to Maternal
- Characteristics: Results from the NICE Birth Cohort in Northern Sweden.
- 493 Nutrients. 2019;11:E1680.
- 494 **36.** Karras SN, Anagnostis P, Annweiler C, Naughton DP, Petroczi A, Bili E,
- Harizopoulou V, Tarlatzis BC, Persinaki A, Papadopoulou F, Goulis DG. Maternal
- vitamin D status during pregnancy: the Mediterranean reality. Eur J Clin Nutr.
- 497 2014;68:864-9.
- 498 37. Grant CC, Stewart AW, Scragg R, Milne T, Rowden J, Ekeroma A, Wall C,
- 499 Mitchell EA, Crengle S, Trenholme A, Crane J, Camargo CA Jr. Vitamin D during

500	pregnancy and infancy and infant serum 25-hydroxyvitamin D concentration.
501	Pediatrics 2014;133:e143-53.
502	38. Zaidi Z. Skin of colour: Characteristics and disease. J Pak Med Assoc. 2017;67:292-
503	9.
504	39. Sotunde OF, Laliberte A, Weiler HA. Maternal risk factors and newborn infant
505	vitamin D status: a scoping literature review. Nutr Res. 2019;63:1-2.
506	

Table 1. Biochemical parameters of neonates born with Vitamin D deficiency, insufficiency or sufficiency.

Parameter	Group A (n=50)	Group B (n=47)	Group C (n=32)	Comparisons
	Deficiency	Insufficiency	Sufficiency	
Total 25(OH)D (nmol/l)	15.9 ± 7.5	38.4 ± 5.0	71.0 ± 30.5	A vs B p<0.01
				A vs C p<0.01
				B vs C p<0.01
25(OH)D ₂ (nmol/l)	2.6 ± 3.2	3.4 ± 4.4	6.9 ± 7.8	A vs C p=0.01
				B vs C p=0.02
25(OH)D ₃ (nmol/l)	13.4 ± 7.6	35.8 ± 6.1	64.1 ± 29.8	A vs B p<0.01
				A vs C p<0.01
				B vs C p<0.01
3-epi-25(OH)D ₂ (nmol/l)	3.6 ± 5.4	4.8 ± 4.0	3.4 ± 3.9	p=0.91
3-epi-25(OH)D ₃ (nmol/l)	4.3 ± 7.6	3.3 ± 3.9	7.2 ± 8.5	B vs C p=0.03
PTH (pg/ml)	7.2 ± 2.6	5.5 ± 1.5	6.3 ± 2.0	A vs B p=0.01
Corrected calcium (mg/dl)	10.7 ± 0.7	11.3 ± 1.3	10.8 ± 0.7	A vs B p=0.02
Phosphorus (mg/dl)	5.7 ± 0.5	5.5 ± 0.5	5.8 ± 0.6	p=0.65
Total bilirubin (mg/dl)	1.6 ± 0.4	1.9 ± 0.5	1.7 ± 0.4	p=0.11

 $Vitamin\ D\ deficiency\ <30\ nmol/l,\ insufficiency\ 30\text{-}50\ nmol/l,\ sufficiency\ >50\ nmol/l.\ Values\ are\ presented\ as\ mean\ \pm\ standard\ deviations.$

Significant differences are presented in bold. Abbreviations: PTH: parathyroid hormone; 25(OH)D: 25-hydroxy-Vitamin D

Table 2. Demographic and anthropometric parameters of neonates born with Vitamin D deficiency, insufficiency or sufficiency.

Parameter	Group A (n=50)	Group B (n=47)	Group C (n=32)	Comparisons
	Deficiency	Insufficiency	Sufficiency	
Male (%)	23 (46)	29 (61.7)	13 (40.6)	p=0.12
Season of birth	42 / 8	36 / 11	27 / 5	p=0.35
(Sep-Mar / Apr-Aug)				
Height (cm)	50.3 ± 1.9	50.8 ± 1.7	50.2 ± 2.3	p=0.73
Birth weight (kg)	3290.9 ± 331.2	3334.4 ± 355.8	3145.7 ± 428.2	p=1.00
Apgar score at 1 min	7.8 ± 0.5	8.0 ± 0.2	8.0 ± 0.0	p=0.20
Apgar score at 5 min	8.7 ± 0.8	9.0 ± 0.2	8.9 ± 0.5	p=0.10
Head circumference (cm)	34.7 ± 4.0	34.8 ± 0.8	33.6 ± 1.9	p=1.00
Neck-rump length (cm)	17.6 ± 2.0	17.9 ± 1.2	17.9 ± 2.5	p=1.00
Chest circumference (cm)	30.8 ± 2.0	31.0 ± 1.5	31.3 ± 2.1	p=1.00
Abdominal circumference (cm)	28.2 ± 2.0	28.3 ± 1.6	28.8 ± 2.5	p=1.00
Abdominal circumference	25.8 ± 1.4	26.0 ± 1.3	26.3 ± 2.0	p=1.00
(iliac)				
Upper arm length (cm)	13.2 ± 0.7	13.7 ± 0.6	13.9 ± 1.5	p=0.17
Upper arm circumference (cm)	9.8 ± 0.7	9.8 ± 0.7	9.8 ± 0.7	p=1.00

Lower arm-radial circumference	9.0 ± 0.6	9.0 ± 0.5	9.0 ± 0.7	p=1.00
(cm)				
Thigh circumference (high)	15.0 ± 1.0	15.4 ± 1.4	15.5 ± 1.7	p=0.79
(cm)				
Thigh circumference (middle)	13.3 ± 1.0	13.3 ± 1.0	13.3 ± 1.4	p=1.00
(cm)				
Lower leg-calf circumference	10.4 ± 0.7	10.3 ± 0.7	10.3 ± 0.9	p=1.00
(maximum) (cm)				
Lower leg length (cm)	13.9 ± 0.9	15.6 ± 1.5	13.0 ± 1.9	B vs C p<0.01
Knee-heel length (cm)	9.0 ± 0.8	8.0 ± 1.3	8.8 ± 0.9	A vs B p<0.01
Femur length (cm)	9.6 ± 0.9	10.0 ± 0.5	9.4 ± 1.1	p=0.19
Skin fold-subscapular (cm)	2.8 ± 0.4	2.6 ± 0.4	2.9 ± 0.7	p=0.14
Skin fold-abdominal (cm)	2.9 ± 0.7	2.8 ± 0.3	2.9 ± 0.5	p=0.82
Skin fold-anterior thigh (cm)	3.6 ± 0.6	3.7 ± 0.4	3.7 ± 0.5	p=1.00

Vitamin D deficiency <30 nmol/l, insufficiency 30-50 nmol/l, sufficiency >50 nmol/lValues are presented as mean ± standard deviation or as absolute values (percentage). Significant differences are presented in bold. **Abbreviations:** min: minute(s); cm: centimeter(s); kg: kilogram(s).

Table 3. Biochemical parameters of mothers who gave birth to neonates with Vitamin D deficiency, insufficiency or sufficiency.

Group A (n=50)	Group B (n=47)	Group C (n=32)	Comparisons
Deficiency	Insufficiency	Sufficiency	
31.7 ± 19.2	36.5 ± 22.3	53.3 ± 39.0	A vs C p<0.01
			B vs C p=0.01
4.2 ± 5.4	3.9 ± 6.0	6.0 ± 8.0	p=1.00
27.4 ± 17.5	33.3 ± 19.9	47.3 ± 36.7	A vs C p<0.01
			B vs C p=0.04
4.0 ± 5.1	3.1 ± 3.9	7.5 ± 13.9	p=1.00
4.5 ± 4.9	3.4 ± 3.6	7.7 ± 14.1	p=1.00
32.2 ± 9.1	30.8 ± 15.5	31.9 ± 13.1	p=1.00
8.1 ± 1.2	8.7 ± 0.9	8.4 ± 1.0	p=0.076
9.5 ± 0.5	9.9 ± 0.3	9.9 ± 0.4	p=0.06
3.6 ± 0.6	3.6 ± 0.6	3.5 ± 0.7	p=1.00
	Deficiency 31.7 ± 19.2 4.2 ± 5.4 27.4 ± 17.5 4.0 ± 5.1 4.5 ± 4.9 32.2 ± 9.1 8.1 ± 1.2 9.5 ± 0.5	Deficiency Insufficiency 31.7 ± 19.2 36.5 ± 22.3 4.2 ± 5.4 3.9 ± 6.0 27.4 ± 17.5 33.3 ± 19.9 4.0 ± 5.1 3.1 ± 3.9 4.5 ± 4.9 3.4 ± 3.6 32.2 ± 9.1 30.8 ± 15.5 8.1 ± 1.2 8.7 ± 0.9 9.5 ± 0.5 9.9 ± 0.3	Deficiency Insufficiency Sufficiency 31.7 ± 19.2 36.5 ± 22.3 53.3 ± 39.0 4.2 ± 5.4 3.9 ± 6.0 6.0 ± 8.0 27.4 ± 17.5 33.3 ± 19.9 47.3 ± 36.7 4.0 ± 5.1 3.1 ± 3.9 7.5 ± 13.9 4.5 ± 4.9 3.4 ± 3.6 7.7 ± 14.1 32.2 ± 9.1 30.8 ± 15.5 31.9 ± 13.1 8.1 ± 1.2 8.7 ± 0.9 8.4 ± 1.0 9.5 ± 0.5 9.9 ± 0.3 9.9 ± 0.4

Vitamin D deficiency < 30 nmol/l, insufficiency > 50 nmol/l. Values are presented as mean \pm standard deviation.

Significant differences are presented in bold. Abbreviations: PTH: parathyroid hormone; 25(OH)D: 25-hydroxy-Vitamin D

Table 4. Demographic and anthropometric parameters of mothers who gave birth to neonates with Vitamin D deficiency, insufficiency or sufficiency.

Parameter	Group A (n=50)	Group B (n=47)	Group C (n=32)	Comparisons
	Deficiency	Insufficiency	Sufficiency	
Age (years)	33.0 ± 4.5	32.5 ± 5.1	32.5 ± 5.0	p=1.00
Height (cm)	169.4 ± 8.9	168.2 ± 7.2	166.8 ± 6.0	p=1.00
Father's height (cm)	183.3 ± 10.7	181.2 ± 10.3	181.6 ± 8.0	p=1.00
Weight before	64.3 ± 9.1	66.8 ± 10.5	62.6 ± 8.7	p=0.63
pregnancy (kg)				
Weight at term (kg)	78.5 ± 10.0	81.6 ± 12.1	75.7 ± 9.2	p=0.49
BMI before	22.6 ± 3.7	23.9 ± 3.8	22.6 ± 3.2	p=0.36
pregnancy (kg/m²)				
BMI at term (kg/m²)	27.4 ± 3.3	28.2 ± 6.1	27.2 ± 3.1	p=1.00
Gestation weeks	39.0 ± 1.7	38.9 ± 1.6	38.9 ± 1.0	p=1.00
Previous live births	0.9 ± 0.7	0.8 ± 0.8	1.2 ± 1.0	p=1.00
Daily calcium	416.6 ± 304.8	474.3 ± 292.7	403.4 ± 9280.6	p=1.00
supplementation (mg)				
Daily dietary calcium	786.2 ± 401.8	796.5 ± 334.0	776.2 ± 338.7	p=1.00
intake during 3 rd trimester				

(mg)				
Daily dietary vitamin D	2.7 ± 1.4	2.5 ± 1.5	3.0 ± 1.4	p=1.00
intake during 3 rd trimester				
(mcg)				
UVB exposure (Wh/m²)	0.2 ± 0.1	0.2 ± 0.1	0.2 ± 0.1	p=0.55
Smoking (%)	5 (10)	4 (8.5)	0 (0)	p=0.80
Higher education (%)	11 (22)	5 (10.6)	12 (37.5)	B vs C p<0.01
Alcohol consumption (%)	None 32 (64)	None 40 (85.1)	None 29 (90.6)	p=0.09
	Light 11 (22)	Light 5 (10.6)	Light 2 (6.3)	
	Moderate 7 (14)	Moderate 2 (4.3)	Moderate 1 (3.1)	

Vitamin D deficiency <30 nmol/l, insufficiency 30-50 nmol/l, sufficiency >50 nmol/l. Values are presented as mean ± standard deviation. Frequencies are presented as absolute values (percentage). Significant differences are presented in bold. **Abbreviations:** BMI: Body Mass Index; UVB: Ultraviolet B radiation.

Table 5. Maternal factors associated to neonatal vitamin D status.

Maternal factors	Univariate analysis		Multivariate analysis			
	OR	95% CI	p-value	OR (adjusted)	95% CI	p-value
Alcohol consumption	5.57	1.17-26.56	0.03	6.18¶	1.18-32.51	0.03
Vitamin D status	6.89	3.09-15.38	<0.01	7.62§	3.27-17.78	<0.01

Vitamin D deficient: 25(OH)D<30 nmol/l, Vitamin D non-deficient: 25(OH)D≥30 nmol/l in univariate and multivariate analysis. Significant differences are presented in bold. The maternal factors that were significantly associated to neonates' vitamin D status in the univariate analysis (alcohol consumption, vitamin D status and UVB exposure) were selected for the multivariate analysis. ¶adjusted for vitamin D status and UVB exposure, §adjusted for alcohol consumption and UVB exposure. **Abbreviations:** OR: odds ratio; CI: confidence interval.