

## Supplemental material

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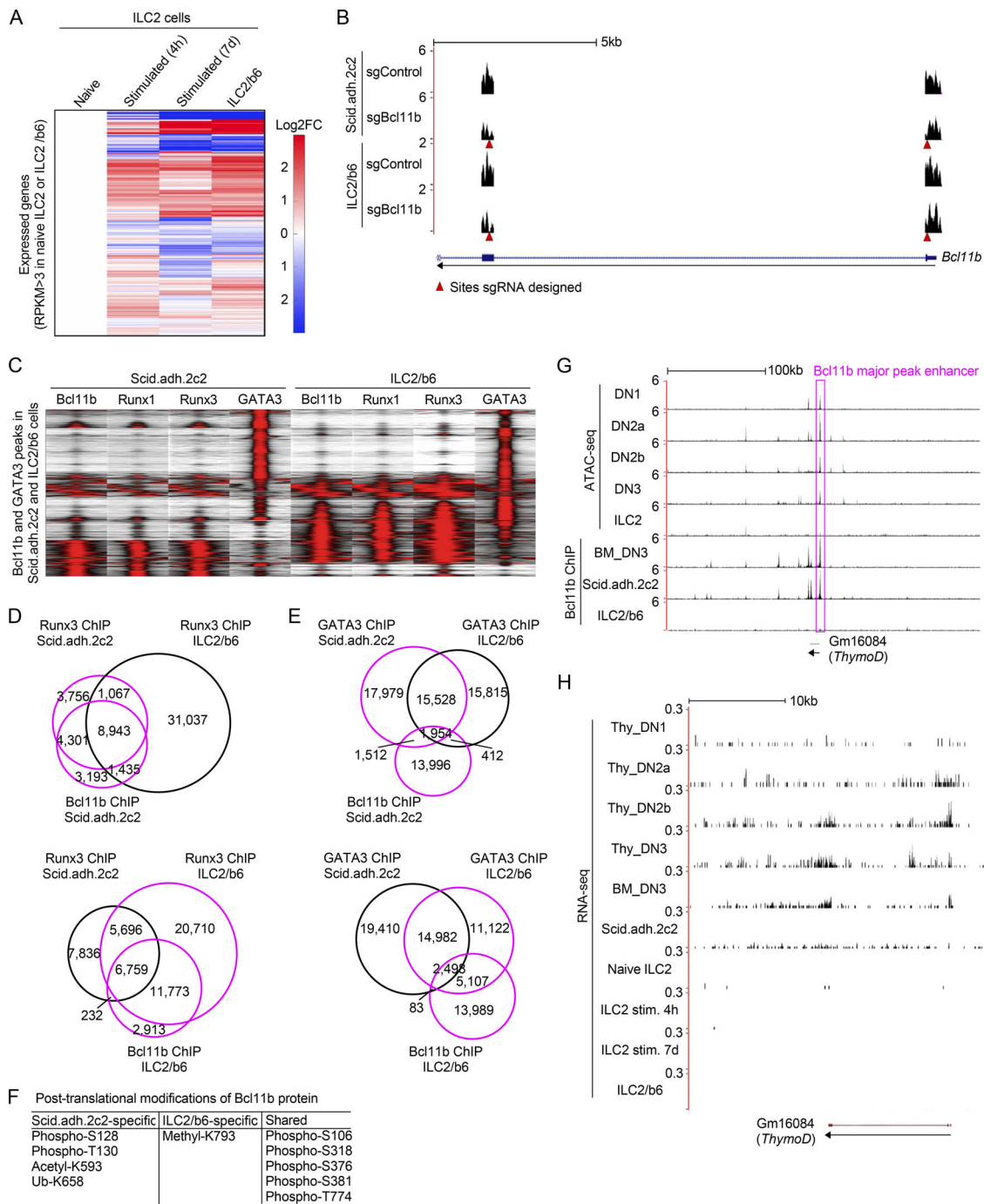


Figure S1. **Characterization of ILC2 and pro-T cell transcriptomes, Runx binding patterns, Bcl11b modifications, and activities in the Bcl11b enhancer region.** (A) Heat maps show hierarchical clustering analyses of the expression of all expressed genes, which have RPKM > 3 in naive ILC2 cells or an ILC2 cell line, ILC2/b6 cells, in naive ILC2, stimulated ILC2 for 4 h or 7 d (Shih et al., 2016; Yagi et al., 2014), and ILC2/b6 cells. (B) Representative RNA-seq tracks are shown for *Bcl11b*-deficient Scid.adh.2c2 or ILC2/b6 cells at the *Bcl11b* locus (around exon1 and 2). Red arrowheads show sites against which sgRNA was designed. (C) Tag count distributions for Bcl11b, Runx1, Runx3, and GATA3 in Scid.adh.2c2 and ILC2/b6 cells are shown. All Bcl11b and GATA3 binding sites identified in the DN3 and ILC2/b6 cells were included in the analysis. (D) Venn diagrams show the number of Runx3 ChIP peaks in Scid.adh.2c2 and ILC2/b6 cells with Bcl11b ChIP peaks in Scid.adh.2c2 cells (top) or ILC2/b6 cells (bottom). (E), Venn diagrams show the number of GATA3 ChIP peaks in Scid.adh.2c2 and ILC2/b6 cells with Bcl11b ChIP peaks in Scid.adh.2c2 cells (top) or ILC2/b6 cells (bottom). (F) Post-translational modifications of Bcl11b protein detected by mass spectrometry analysis are shown. (G) Representative ATAC-seq tracks for thymic DN subsets and mature small intestine ILC2 cells (Yoshida et al., 2019; downloaded from GEO accession no. GSE100738) and binding profiles of Bcl11b in DN3, Scid.adh.2c2, and ILC2/b6 cells are shown around the Bcl11b major peak enhancer (magenta rectangle) and *ThymoD* locus. The low activity state shown here in mature ILC2 cells contrasts with the state inferred to exist in ILC2 precursors, based on evidence in Fig. 5. (H) Representative RNA-seq tracks (mapped with STAR v2.4.0) at the *ThymoD* locus are shown for thymic DN subsets (Yoshida et al., 2019; GSE100738), in vitro cultured bone marrow precursor-derived DN3, Scid.adh.2c2, mature naive ILC2, mature ILC2 stimulated (stim.) for 4 h or 7 d (Shih et al., 2016; Yagi et al., 2014), and ILC2/b6 cells. Data from this study are based on two replicate RNA-seq results (A, B, and H) and are based on reproducible ChIP-seq peaks in two replicate samples (C–E and G).

Tables S1 and S2 are provided online as separate Excel files. Table S1 shows DEGs after Cas9-mediated disruption of *Bcl11b* gene in *Scid.adh.2c2* and ILC2/b6 cells. Table S2 shows *Bcl11b*-interacting molecules in *Scid.adh.2c2* and ILC2/b6 cells identified by LC-MS/MS.

## References

- Hosokawa, H., M. Romero-Wolf, M.A. Yui, J. Ungerback, M.L.G. Quiloan, M. Matsumoto, K.I. Nakayama, T. Tanaka, and E.V. Rothenberg. 2018a. *Bcl11b* sets pro-T cell fate by site-specific cofactor recruitment and by repressing *Id2* and *Zbtb16*. *Nat. Immunol.* 19:1427–1440. <https://doi.org/10.1038/s41590-018-0238-4>
- Shih, H.Y., G. Sciumè, Y. Mikami, L. Guo, H.W. Sun, S.R. Brooks, J.F. Urban Jr., F.P. Davis, Y. Kanno, and J.J. O’Shea. 2016. Developmental acquisition of regulomes underlies innate lymphoid cell functionality. *Cell.* 165:1120–1133. <https://doi.org/10.1016/j.cell.2016.04.029>
- Yagi, R., C. Zhong, D.L. Northrup, F. Yu, N. Bouladoux, S. Spencer, G. Hu, L. Barron, S. Sharma, T. Nakayama, et al. 2014. The transcription factor *GATA3* is critical for the development of all *IL-7R $\alpha$* -expressing innate lymphoid cells. *Immunity.* 40:378–388. <https://doi.org/10.1016/j.immuni.2014.01.012>
- Yoshida, H., C.A. Lareau, R.N. Ramirez, S.A. Rose, B. Maier, A. Wroblewska, F. Desland, A. Chudnovskiy, A. Mortha, C. Dominguez, et al. Immunological Genome Project. 2019. The cis-regulatory atlas of the mouse immune system. *Cell.* 176:897–912.e20. <https://doi.org/10.1016/j.cell.2018.12.036>