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# **INTERACTIONS OF RIBONUCLEOTIDES WITH ALUMINUM AND IRON OXIDES UNDER VARYING ENVIRONMENTAL CONDITIONS**



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### Introduction

Mineral surfaces might have concentrated building blocks of biomolecules in the context of the origin of life, facilitating their polymerization. Mineral surfaces have also been shown to protect biomolecules from degradation, facilitating their survival in soils. Previous work has highlighted the predominant role of the edges of mineral particles in the adsorption of nucleotides (Feuillie et al, 2013), implying oxide-like adsorption sites. The adsorption mechanism is not fully understood yet.

Here we present experimental and surface complexation modeling results on the adsorption of nucleotides with alumina and hematite, 2 oxide minerals presenting the same structure, as a function of pH, ionic strength and ligand-to-solid ratio.



Above : Crystal maker structure of  $\alpha$ alumina  $Al_2O_3$  (red = O, blue = Al) Below : TEM image of rhombohedral hematite nanoparticles used in this study.



# **Methods**

- Experimental approach : **batch adsorption experiments** 
  - Adsorption at room temperature until equilibrium
  - Wide range of pH (3 11), ionic strength (from 0 to 0.1 M of NaCI), and surface loading
  - Centrifugation and filtration of the supensions
  - **Ion Chromatography** analysis of the supernatant (ICS-5000 – Dionex; AminoPac PA10 column)



Characteristics of alumina and hematite used in this study. a: Halter & Pfeifer, 2001; b: Fukushi & Sverjensky, 2007; c: Cornell & Schwertmann, 2003; d: Sverjensky, 2005

|                   | Corundum - Al <sub>2</sub> O <sub>3</sub> | Hematite - Fe <sub>2</sub> O <sub>3</sub>   |
|-------------------|---|---|
| Provenance        | Well studied commercial alumina a, b      | Forced hydrolysis of acid Fe <sup>III</sup> salt solutions at 98°C following Schwertmann & Cornell (2000) |
| рН <sub>zPC</sub> | <b>9.1</b> <sup>a, b</sup>                | 8.5 - 9.5 <sup>c, d</sup>   |
| Particle size     | ~ 5 microns                               | ~ 40 nm   |
| Surface area      | 3.52 m²/g                                 | ~ 52 m²/g   |

Theoretical approach : surface complexation calculations using the **Extended Triple Layer Model (ETLM)** 

- 1 site model
- Takes into account the electrical work linked to the desorption of chemisorbed water
- Establish the speciation of surface species, the stoichiometry and the thermodynamic equilibrium constants for the adsorption of nucleotides on oxide minerals







High adsorption at low pH; maximum adsorption at pH 5; > 2  $\mu$ mol / m<sup>2</sup> No ionic strength effect above pH 5

Below pH 5, when ionic strength increases the adsorption decreases  $\implies$  similar to phosphate adsorption (He et al, 1997)









GMP

### Adsorption of GMP on Fe<sub>2</sub>O<sub>3</sub>



## Other nucleotides on Al<sub>2</sub>O<sub>3</sub> (Feuillie et al, 2015)

5 g/L ; equilibrium reached after 24 h of adsorption

log K = 18.9



inner-sphere is figured in red, the bidentate outer-sphere complex in blue.

pH profiles of the adsorption of GMP on hematite nanoparticles obtained at an ionic strength of (a) NaCl 0 M, (b) NaCl 0.01 M, and (c) NaCl 0.1 M. Experimental error is ~ 5%. Symbols are experimental data; dashed lines are guides for the eyes and do not represent modeling data.

High adsorption, similar to alumina : above 2  $\mu$ mol / m<sup>2</sup> High adsorption at low pH; maximum adsorption at pH 5.5 – 6 Similar ionic strength effect at low pH Contrary to alumina, hematite shows adsorption even at high pH

Surface complexation calculations coming soon...

#### Summary

- High adsorption of GMP on both Alumina and Hematite, **above 2**  $\mu$ **mol/m**<sup>2</sup> Same behaviour at low pH for both minerals, but higher adsorption at high pH for hematite
- **One mechanism** for all nucleotides adsorption **on alumina** : 1 monodentate inner-sphere at low pH, 1 bidentate outer-sphere complex at high pH, involving phosphate groups
- More strongly bound at low pH : Availability of nucleotides for polymerization in the context of the origin of life and in modern soils ?



*References*: Feuillie et al, 2013, GCA 120 97–108; Halter & Pfeifer, 2001, C. Appl. Geochem., 16 (7-8), 793–802; Fukushi & Sverjensky, 2007, GCA 71 (15), 3717–3745; Cornell & Schwertmann, 2003, The iron oxides: structures, properties, reactions, occurence and uses, p 85-86; Sverjensky, 2005, GCA 69 (2) 225–257; Feuillie et al, 2015, Langmuir 31 (1) 240–248; He et al, 1997, Soil Sci. Soc. Am. J., 61 (3), 784–793.