



Biodegradation of 3,5-dinitrosalicylic acid by *Phanerochaete chrysosporium*

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ABSTRACT

Despite intensive efforts put on prevention of environment pollution by nitroaromatic compounds, these xenobiotics have not been eliminated from the biosphere. The physicochemical properties make nitroaromatics extremely recalcitrant to biodegradation. Therefore, microbial degraders of these pollutants are sought after. This paper reports preliminary results of the study on degradation of 3,5-dinitrosalicylic acid (DNS) by a basidiomycetous fungus *Phanerochaete chrysosporium* under stationary conditions in a culture medium containing 0.05–0.5% v/v of DNS. The results obtained suggest that the fungus degrades DNS through the reductive pathway.

KEY WORDS: nitroaromatic compounds, white-rot fungi, fungal biodegradation

Introduction

Aromatic nitrocompounds are chemicals containing one or more nitro (-NO₂) groups attached to the aromatic ring. These substances are known to be highly toxic and probably carcinogenic for humans as well as highly recalcitrant to

natural biodegradation, thus they pose very serious threat to the environment (Williams *et al.* 2015, Lipczynska-Kochany 1995, Price 1997, Gong *et al.* 2003). Their content in natural environment increases every year due to

military operations, combustion of fossil fuels or civil application of explosives based on nitrated aromatic compounds (Sekhar & Wignes 2016, Rezaei 2010, Anasonye *et al.* 2015). There is a number of commercial applications of aromatic nitrocompounds. Vast majority of nitroaromatic products are employed in military, for instance chemicals like hexahydro-1,3,5-trinitro-1,3,5-triazine (RDX) or 2,4,6-trinitrotoluene (TNT) are examples of the most commonly applied explosives, whereas substances like 2,4-dinitrophenol (DNP) were widely used as defoliants, e.g. during Vietnam war (Haberman 2014, Grundlingh 2011). Nevertheless, aromatic nitrocompounds are also used in other areas, as e.g. pharmaceuticals (an antibiotic Chloramphenicol) or general purpose chemical reagents such as nitro-based aromatics like nitrobenzene or 3,5-dinitrosalicylic acid (DNS), which is used for reducing sugars assays (Kumar 2016).

The low affinity to water combined with high electron-withdrawing character are main factors responsible for the highly hazardous character of aromatic nitrocompounds (Shen *et al.* 2009, Mathieu and Alaime 2015). Most of the nitrocompounds are viscous liquids or solid compounds characterized by poor water dissolvability. Moreover, not only nitro groups impart the xenobiotic character, but also explosophoric properties (Infante-Castillo & Hernandez-Rivera 2012), what may explain their abundance in biosphere.

3,5-dinitrosalicylic acid is an organic, aromatic compound with physicochemical properties resembling other nitroaromatic compounds. The biggest difference lies in its solubility in water (Apelblat & Manzurrola 1999), making DNS potentially more susceptible to biodegradation than chemicals like TNT or DNP.

There is a small number of microorganisms that degrade nitroaromatic compounds. One could distinguish two types of them: bacteria, that degrade nitrocompounds via reductive pathway through nitroso- and hydroxyamino- derivatives (Singh *et al.* 2015, Claus 2013, Lenke & Knackmuss 2016), often yielding products that are sometimes claimed to be more toxic than the substrate (for example amino- and hydroxyamino- derivatives of TNT) (Kulkarni & Chaudari 2007), and fungi, capable of even complete mineralizing of toxic nitroaromatics.

One of the fungal species, being under intense research is the basidiomycete fungus *Phanerochaete chrysosporium*, producing enzymes evincing very low substrate-specificity. Due to that, this white-rot family fungus is capable of degrading a wide range of aromatic substances, including lignin alcohols (Nousiainen *et al.* 2014, Asina *et al.* 2016), polychlorinated biphenyls (Cvancarova *et al.* 2012) and nitroaromatics (Spain 1995).

Therefore, this organism was investigated in terms of 3,5-dinitrosalicylic acid biodegradation. The results of preliminary investigations on the biodegradation of DNS by *P. chrysosporium* as well as the putative biodegradation pathway are presented in this paper.

Materials and methods

Experimental setup

The *P. chrysosporium* strain was maintained on agar slants with a modified Czapek medium, containing glucose instead of sucrose (glucose 30 g/l, sodium nitrate 2 g/l, dipotassium phosphate 1 g/l, magnesium sulphate 0.5 g/l, potassium chloride 0.5 g/l, ferrous sulphate 0.01 g/l) and enriched with oat flakes (5 mg per 5 ml) as a source of thiamine (Maza *et al.* 2015). Liquid cultures under stationary

conditions were carried out in 200 ml Erlenmeyer's flasks, in the modified Czapek's medium (20 ml) enriched with 50 mg of sugar beet pulp instead of oat flakes. The sterile medium (after autoclaving at 121 °C for 15 min) was inoculated with 1 ml of fungal spores washed from the agar slants using sterile demineralized water (5 ml per slant). The fungus was cultivated at 30 °C for 7 days and then 20 ml of a tested (0.05%, 0.25% and 0.5%) DNS solution in water (its pH was adjusted to 5.0 with 0.1 M NaOH) was added to each flask and the flasks were weighted because the metabolic activity of *P. chrysosporium* in the DNS-containing culture medium was monitored, among others, based on the decrease in the weight, caused by carbon dioxide emission from the culture.

The controls without DNS contained 20 ml sterile demineralized water instead of DNS solution. To eliminate the error caused by water evaporation during the culture, which also causes a decrease in weight, three 40 ml portions of the sterile culture medium were incubated along with the inoculated samples. The decrease in the weight of the culture medium controls was compared with that of the inoculated samples and controls. Another set of three controls, containing 20 ml of the sterile culture medium (not inoculated with the fungus) and supplemented after the first 7 days with 20 ml of suitable DNS solution was prepared and used to monitor the spontaneous DNS degradation under experimental conditions.

Extraction of 3,5-DNS

Every fourteen days the content of three weighted flasks was filtered through filter papers and 6 ml of each filtrate was analysed for the content of residual DNS and its degradation products. DNS was extracted from each filtrate three times with 12 ml of ethyl acetate (analytical grade) and then the extracts were pooled,

and 6 ml aliquots of the pooled extracts were evaporated. The efficiency of extraction was above 98%. The solids (residues of DNS and its degradation products) were dissolved in 1 ml of methanol (analytical grade), and the solutions were filtered using syringe filters (4.5 µm), transferred into eppendorf tubes and subjected to HPLC-UV analysis.

HPLC-UV analysis

The extracted compounds were analysed using an HPLC Knauer system equipped with a C18 RP column (Supelco 4.8 µm x 4.8 mm x 10 cm), a Knauer HPLC pump and an UV-Vis detector. The mobile phase contained 70% methanol and 30% acetonitrile (both HPLC-grade). The temperature of column was set at 25 °C and the injection volume was 20 µl.

The measurements of absorbance were conducted at the wavelengths of 210, 254, 278 and 340 nm.

Presentation of results

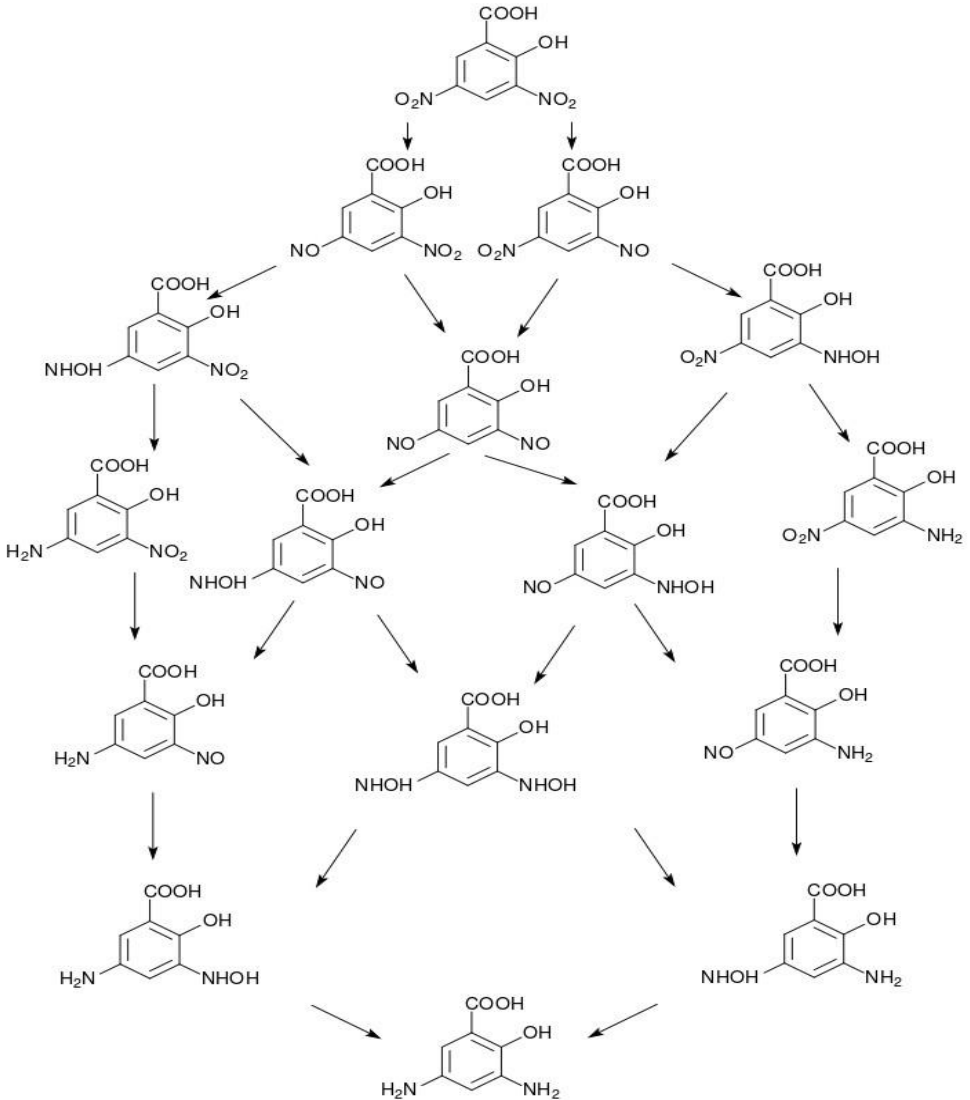
The results presented in figures 2, 3 and 5 are means + SD of triplicate samples. All structures presented in this work as well as a theoretical pathway of DNS biodegradation by *P. chrysosporium* were prepared using a BKchem GNU software (ver. 0.13.0). The figures were created using a RStudio Open Source Edition ver. 3.4.0.0 and MS Excel 2007.

Results

Putative biodegradation pathway

The investigation of TNT biodegradation (Spain 1995, Bayman *et al.* 1997) showed that not only oxidoreductases (LiP, MnP, laccase) take part in detoxication and biodegradation, but also a range of nitroreductases (Maza *et al.* 2015). Based on that, the putative pathway concerning initial steps of 3,5-DNS biodegradation was proposed (Fig. 1).

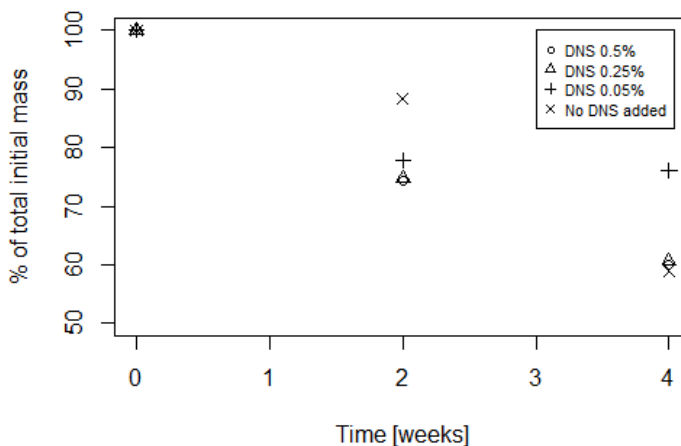
Figure 1. Putative denitrification pathway of 3,5-DNS by *P. chrysosporium* involving nitroreductases and leading to 3,5-diaminosalicylic acid through nitroso- and hydroxyamino- derivatives.



It was assumed that the presence of 3,5-dinitrosalicylic acid might affect the general fungal metabolism and influence the rate of carbon dioxide emission by the growing *P. chrysosporium*. Production of carbon dioxide increases with the intensity of oxidative metabolism, and when the latter is growing, the decrease in

weight of the culture medium is faster. Therefore, each flask with the fungus was weighted before filtration and its weight was compared with the initial one. The dynamics of weight decrease with time, either in the absence or presence of 0.05, 0.25 and 0.5% DNS, is shown in Fig. 2.

Figure 2. The average weight decrease with time.

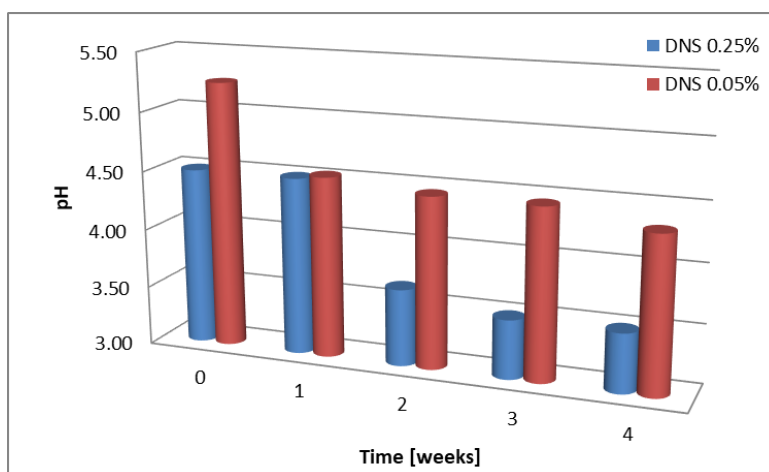


The data shown in Fig. 2 demonstrate that the presence of 0.05–0.5% DNS in the culture medium only slightly affected the dynamics of the medium’s weight decrease during the growth of *P. chrysosporium*. It was also visible that at these DNS concentrations, the fungus could grow like in the DNS-free medium. The decrease in weight of the sterile culture medium controls was negligible and therefore it was not presented in Fig. 2.

As *P. chrysosporium* is known to produce organic acids (oxalic) and acidify

the environment (Bonnamme & Jeffries 1990), changes in pH of the culture medium with time were monitored. The results of these measurements are presented for the DNS concentrations of 0.05% and 0.25% in Fig. 3. It was observed that in the medium containing 0.05% DNS, the values of pH were slightly higher on the same days than in the other samples that suggested the occurrence of basic intermediates of DNS conversion. At the higher DNS concentrations, the pH values were nearly the same as in the DNS-free samples.

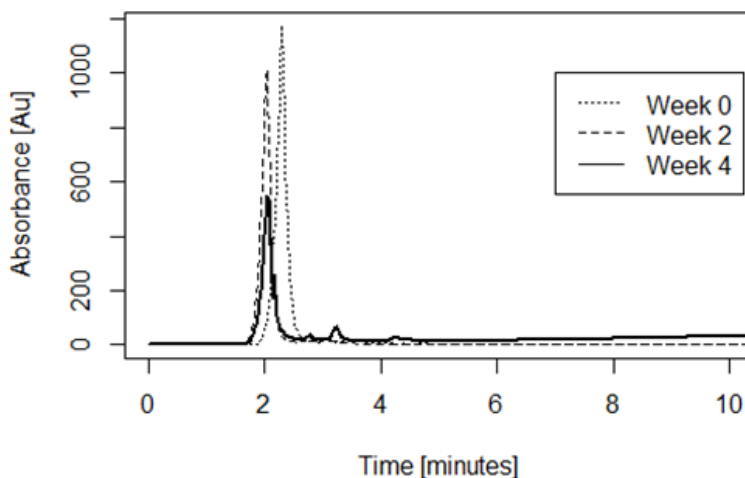
Figure 3. The change of pH with time.



The analysis of extracts from culture medium samples using HPLC-UV showed that DNS concentration decreased with time of *P. chrysosporium* growth (Fig. 4). Because the analysis of the DNS-containing controls, which were not

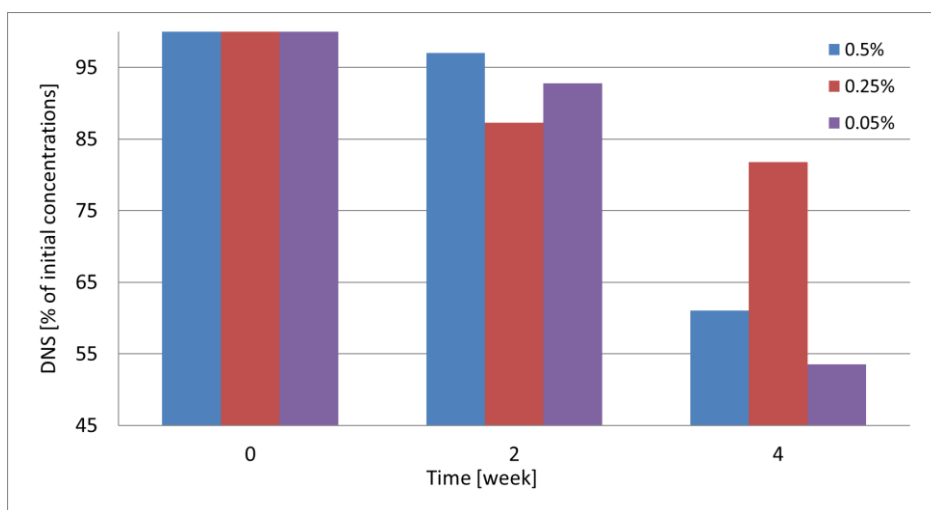
inoculated with the fungus, showed that this compound was not spontaneously degraded under the experimental conditions, so the reduction of DNS content in the inoculated samples may be ascribed to its degradation by the fungus.

Figure 4. An exemplary chromatogram of three samples. The first peak at 2 min corresponds to DNS whereas the visible small peaks may represent the unknown intermediates of DNS degradation.



The decrease in DNS concentration in the culture medium and occurrence of peaks that may represent DNS conversion products suggest that *P. chrysosporium* metabolizes 3,5-dinitrosalicylic acid. The collective results of HPLC-UV analyses are shown in Fig. 5.

Figure 5. A decrease in 3,5-DNS content with time.



The relatively large reduction of DNS content in the culture medium supplemented with 0.05% and 0.5% DNS after 4 week cultivation may be ascribed to its partial precipitation from the culture medium, which was not observed at 0.25% concentration. The results presented in Fig. 5 demonstrate that the rate of DNS removal for the highest and lowest concentrations is significant, up to 50% of initial substrate concentration. The lesser extent of DNS content reduction at its initial concentration of 0.25%, compared to that at the lowest initial concentration (of 0.05%), may suggest that in the latter case DNS was metabolized by the fungus like 'a co-substrate' and therefore its concentration was decreased below 55% of the initial one within 4 weeks. When the initial concentration of DNS was increased 10-fold (to 0.5%) the fungus was unable to efficiently metabolize the substrate and had to precipitate its excess to survive. Also at the initial concentration of 0.25%, the metabolism of DNS was disturbed, however, the fungus was able to synthesize enzymes that are responsible for this process and therefore precipitation of DNS was less intensive. The observed significant drop in pH in the latter case might be favorable for the solubility of degradation products, which contain amino groups (Fig. 1).

Discussion

The assumption of the reductive character of 3,5-dinitrosalicylic acid biodegradation by *P. chrysosporium* is based on the gradual decrease in DNS concentration in the culture medium filtrates. Furthermore, these filtrates contained putative intermediate products that absorbed light at wavelengths

characteristic of nitroaromatic compounds. Furthermore, the gradual browning of culture medium during the 4 week cultivation may be also caused by reduction of the nitro groups attached to the aromatic ring of DNS. The gradual decrease in pH of the culture medium was ascribed to the synthesis of organic acids by *P. chrysosporium*. It may be also considered one of the mechanisms enabling the removal of DNS from the environment by precipitation.

The results obtained in this study are consistent with findings concerning the removal of other pollutants like 2,4,6-trinitrotoluene or 2,4-dinitrophenol by *P. chrysosporium* and other wood-decaying fungi. These organisms are capable of either partial or complete degradation of TNT (Spain *et al.* 2000, Tashes *et al.* 1990). However, the reported initial concentrations of the substrates were in most cases much lower than in our work that was beneficial for the biodegradation process. Although further studies are necessary to describe in detail the DNS degradation pathway, the observed browning of culture medium and decrease in the DNS concentration provide evidence that *P. chrysosporium* may be used for the removal of this nitroaromatic compound from aqueous systems.

Summary

The reported results suggest that *P. chrysosporium* metabolizes 3,5-dinitrosalicylic acid. Further experiments will focus on determination of the structure of intermediates that appear in the culture medium, and identification of the genes encoding nitroreductases in the genome of the fungus. Also the conditions of DNS degradation will be optimized to increase the rate of this process.

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Streszczenie

Związki nitrowe to szeroka grupa ksenobiotyków, które ze względu na swoją silną toksyczność, wyjątkową odporność na rozkład biologiczny oraz skłonność do bioakumulacji, stanowią bardzo poważny problem dla biosfery. Prowadzi się obecnie wiele badań nad mikroorganizmami, które zdołały wykształcić szlaki metaboliczne pozwalające na rozkład takich związków jak 2,4,6-trinitrotoluen, kwas pikrynowy czy kwas 3,5-dinitrosalicylowy. Jednym z takich mikroorganizmów jest podstawczak *Phanerochaete chrysosporium*, należący do grupy grzybów białej zgnilizny drewna.

Artykuł ten poświęcony jest badaniom nad rozkładem kwasu 3,5-dinitrosalicylowego przez *P. chrysosporium* w warunkach hodowli stacjonarnej w pożywce zawierającej 0,05–0,5% masowego kwasu 3,5-dinitrosalicylowego. Uzyskane wyniki wskazują na zdolność wybranego mikroorganizmu do rozkładu substratu na drodze redukcji grup nitrowych.