

Impact of Small-Quantity Lipid-Based Nutrient Supplement on Hemoglobin, Iron Status and Biomarkers of Inflammation in Pregnant Ghanaian Women¹⁻⁷

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⁵Abbreviations: AGP, Alpha-1 glycoprotein; CRP, C-reactive protein; EFA, Essential Fatty Acid; GA, Gestational age; GW, Gestational weeks; IDA, Iron Deficiency Anemia; IFA, Iron and folic acid; ; LNS, Lipid-based Nutrient Supplement; MMN, Multiple Micronutrients; SQ-LNS, Small-quantity lipid-based nutrient supplement; TfR, Transferrin receptor; ZPP, Zinc Protoporphyrin.

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Contributor statement

SA-A, AL, PA, MZ, SV, and KGD designed the research; MZ was responsible for the development and production of the LNS used in the study based on the specifications agreed

upon by the iLiNS Project; SA-A, AL, and HO conducted the research; LMB and BO performed laboratory analysis; SA-A performed the statistical analysis; AL, PA, and KGD advised on the analysis; SA-A and KGD wrote the manuscript; and AL, HO, PA, MZ, LMB, BO, and SV reviewed the draft manuscript. All authors read and approved the final manuscript.

1 Abstract

2 We examined hemoglobin (Hb, g/L), iron status (zinc protoporphyrin, ZPP, $\mu\text{mol/mol}$ heme, and
3 transferrin receptor, TfR, mg/L), and inflammation (C-reactive protein, CRP and alpha-1
4 glycoprotein, AGP) in pregnant Ghanaian women who participated in a randomized controlled
5 trial. Women ($n=1320$) received either 60 mg Fe + 400 μg folic acid (IFA); 18 micronutrients
6 including 20 mg Fe (MMN); or small-quantity lipid-based nutrient supplements (SQ-LNS, 118
7 kcal/d) with the same micronutrient levels as in MMN, plus 4 additional minerals (LNS) daily
8 during pregnancy. Intention-to-treat analysis included 349, 354, and 354 women in the IFA,
9 MMN and LNS groups, respectively, with overall baseline mean Hb and anemia (Hb <100)
10 prevalence of 112 and 13.3%, respectively. At 36 gestational weeks, overall Hb was 117 and
11 anemia prevalence was 5.3%. Compared with the IFA group, the LNS and MMN groups had
12 lower mean Hb (120 ± 11 vs 115 ± 12 and 117 ± 12 , respectively; $p<0.001$), higher mean ZPP
13 (42 ± 30 vs 50 ± 29 and 49 ± 30 ; $p=0.010$) and TfR (4.0 ± 1.3 vs 4.9 ± 1.8 and 4.6 ± 1.7 ; $p<0.001$), and
14 greater prevalence of anemia (2.2% vs 7.9% and 5.8%; $p=0.019$), elevated ZPP (>60) [9.4% vs
15 18.6% and 19.2%; $p=0.003$] and elevated TfR (>6.0) [9.0% vs 19.2% and 15.1%; $p=0.004$]. CRP
16 and AGP concentrations did not differ among groups. We conclude that among pregnant women
17 in a semi-urban setting in Ghana, supplementation with SQ-LNS or MMN containing 20 mg
18 iron resulted in lower Hb and iron status but had no impact on inflammation, when compared
19 with iron (60 mg) plus folic acid (400 μg). The amount of iron in such supplements that is most
20 effective for improving both maternal Hb/iron status and birth outcomes requires further
21 evaluation. This trial was registered at ClinicalTrials.gov as: NCT00970866.

22

- 23 **Keywords** lipid-based nutrient supplements, LNS, prenatal supplementation, multiple
24 micronutrients, hemoglobin, iron status, inflammation

25 **Introduction**

26 Poor nutrient intake during pregnancy has been associated with several adverse
27 consequences. It is estimated that up to 50% of the anemia among pregnant women in many
28 developing country settings is due to iron deficiency (van den Broek et al., 1998) usually as a
29 result of low dietary iron intake (World Health Organization, 1992) and poor iron bioavailability
30 due to over-reliance on plant-based diets high in inhibitors of iron absorption such as phytate
31 (Tatala et al., 1998). Consequences of anemia include reduced work capacity and increased risk
32 of mortality for the mother, and premature delivery, low birth weight and poor mental
33 development for the infant (Ren et al., 2007, International Anemia Consultative Group (INACG),
34 2002). While normal pregnancy is found to be associated with an increased inflammatory
35 response (Picklesimer et al., 2008, Sacks et al., 1998), this response may be modified by macro-
36 or micro-nutrients (Roberts et al., 2003). Higher intakes of folic acid (Bertran et al., 2005) and
37 vitamin B₆ (Friso et al., 2001) have been associated with lower concentration of C-reactive
38 protein (CRP), a common biomarker of inflammation. Several other dietary factors including
39 essential fatty acids, EFAs (Rallidis et al., 2003) and antioxidants (Brighenti et al., 2005, Devaraj
40 and Jialal, 2000) are also associated with the reduction of CRP concentration. Elevated CRP
41 concentration in pregnancy is related to the development of pre-eclampsia and preterm delivery
42 (Elovitz, 2006).

43 A major recommendation for increasing nutrient intake among pregnant women in
44 developing countries is the one developed by WHO (WHO), which is the consumption of
45 iron/folic acid (IFA) supplements containing 30-60 mg iron and 400 µg folic acid. In meta-
46 analyses, this strategy, compared with no iron or placebo, reduced the risk of maternal anemia by
47 69-70 % (Imdad and Bhutta, 2012, Pena-Rosas et al., 2012) and iron deficiency by 57% (Pena-

48 Rosas et al., 2012), but appeared to increase the risk of reported side effects (relative risk (RR) =
49 2.36; 95% CI: 0.96 -5.82) particularly at iron doses of 60 mg or higher (Pena-Rosas et al., 2012).
50 The WHO/UNICEF/UNU UNIMMAP (United Nations International Multiple Micronutrient
51 Preparation) formulation containing 15 vitamins and minerals was more recently developed
52 (UNICEF/WHO/UNU) to combat other possible deficiencies, e.g. for vitamins A, C and B₁₂,
53 which may also contribute to anemia (World Health Organization, 1992). The dose of iron in the
54 UNIMMAP was set at 30 mg (below 60 mg) for the following reasons: (a) the presence of
55 vitamins A, B₂ and C in the UNIMMAP would enhance the absorption and utilization of iron,
56 and therefore the lower amount of iron should be sufficient, (b) a lower iron dose would be
57 associated with less negative side effects and therefore better adherence, (c) including 60 mg of
58 iron would mean including at least 30 mg of zinc (to avoid possible negative influence of iron on
59 zinc absorption), bringing the total amount of metals to 90 mg, which is likely to increase
60 negative side effects, and (d) UNIMMAP may be used in conjunction with additional iron /folic
61 acid tablets in individual cases of more severe anemia (assuming it is caused by iron deficiency).
62 Meta-analyses suggested that supplementation with UNIMMAP and similar products (mostly
63 containing 11 or more micronutrients including 30 mg iron) had the same effect on maternal
64 hemoglobin and iron status as iron (usually at 60 mg dose) with or without folic acid (Allen and
65 Peerson, 2009, Haider et al., 2011), while also reducing the risk of low birth weight (Fall et al.,
66 2009, Haider and Bhutta, 2012, Ramakrishnan et al., 2012).

67 Our group developed small quantity (20 g/d) lipid-based nutrient supplements (SQ-LNS)
68 for pregnant and lactating women (Arimond et al., 2013) to provide micronutrients together with
69 EFAs, using a minimum food base that supplies a small amount of energy (118 kcal/d) and high
70 quality protein (2.6 g/d). In many populations, total energy intake among pregnant and lactating

71 women may be adequate, but the EFA content of the usual diet may be low (Michaelsen et al.,
72 2011). The micronutrient composition of the SQ-LNS was generally based on the UNIMMAP
73 formulation and a similar product used in Guinea Bissau (Kaestel et al., 2005), but we further
74 reduced the daily iron dose to 20 mg, based on evidence that 20 mg day⁻¹ may be an adequate
75 dose to prevent iron deficiency anemia during pregnancy (even for women who are iron deficient
76 at entry to prenatal care) and causes fewer gastrointestinal side effects, compared to higher doses
77 of iron (Zhou et al., 2009). We estimated (Arimond et al., 2013) that in addition to iron coming
78 from the usual diet, the 20 mg of iron from a daily supplement would meet the recommended
79 dietary allowance (RDA) of 27 mg iron during pregnancy (and be close to the 30 mg/d dose in
80 the UNIMMAP formulation) while not greatly exceeding the RDA (9 mg/d) for iron during
81 lactation (IOM, 2001, Arimond et al., 2013).

82 Currently, there is a growing interest in the potential use of Small-Quantity Lipid-based
83 Nutrient Supplements (SQ-LNS) among pregnant women in developing-country settings
84 (Hambidge et al., 2014, Research Engagement on Food Innovation for Nutritional Effectiveness
85 (REFINE), 2013), because of evidence suggesting a positive impact of the product on certain
86 pregnancy outcomes (Adu-Afarwuah et al., 2015). However, little is known about the impact of
87 SQ-LNS on maternal outcomes such as anemia, iron status and inflammation. We previously
88 reported (Adu-Afarwuah et al., 2015), that compared to IFA and a multiple micronutrient
89 (MMN) capsule with most of the same micronutrients as the SQ-LNS, the SQ-LNS promoted
90 fetal growth in vulnerable women, particularly primiparas, whilst the occurrence of serious
91 adverse events did not differ between the 3 groups. In the current analysis, we compare the effect
92 of the 3 supplementation regimens (IFA, MMN and SQ-LNS), on maternal hemoglobin (Hb),

93 iron status, and two biomarkers of inflammation (CRP and alpha-1 glycoprotein, AGP) during
94 pregnancy.

95 **Methods**

96 **Study setting, design, participants and blinding**

97 The iLiNS DYAD study in Ghana was conducted in several adjoining semi-urban
98 communities in the Yilo Krobo and the Lower Manya Krobo Districts about 70 km north of
99 Accra, Ghana. Details of the study setting, participants, design, randomization and masking
100 schemes, and other key procedures have been reported elsewhere (Adu-Afarwuah et al., 2015).
101 In brief, the study was designed as a partially double-blind, parallel, individually randomized,
102 controlled trial with three equal-size groups. Pregnant women attending usual ante-natal clinics
103 in four main health facilities in the area between December 2009 and December 2011 completed
104 a screening questionnaire if they were ≥ 18 years old, ≤ 20 weeks gestation (as determined by the
105 antenatal clinics mostly by fundal height), and had an antenatal card complete with history and
106 examination. Informed consent for the screening was obtained by trained study workers at the
107 antenatal clinics. Following screening, women were excluded if the antenatal card indicated HIV
108 infection, asthma, epilepsy, tuberculosis or any malignancy. Additional exclusion criteria were
109 known milk or peanut allergy, not residing in the area, intention to move within the next two
110 years, unwillingness to receive field workers or take study supplement, participation in another
111 trial, or gestational age (GA) > 20 weeks before completion of the enrolment process.

112 Women who passed the screening were visited in their homes, where details of the study
113 were provided, and those willing to participate were recruited, after signing or thumb-printing
114 informed consent. Recruited women remaining eligible underwent a baseline laboratory
115 assessment after consent, and were immediately randomized to receive one of three treatments
116 daily: (a) 60 mg iron plus 400 μ g folic acid (hereafter, IFA supplement or group); (b) multiple
117 micronutrient capsule containing 18 vitamins and minerals (including 20 mg iron) (hereafter,

118 MMN supplement or group); and (c) SQ-LNS with similar micronutrients as the MMN
119 supplement, plus other minerals and macronutrients (hereafter, LNS supplement or group).
120 Group allocations were developed by the Study Statistician at UC Davis using a computer-
121 generated (SAS version 9.3) randomization scheme (in blocks of nine), and were placed in
122 sealed, opaque envelopes. At each enrolment, a Study Nurse offered nine envelopes at a time,
123 and the woman picked one to reveal the allocation. Allocation information was kept securely by
124 the Field Supervisor and the Study Statistician only.

125 The compositions of the 3 supplements were reported previously (Adu-Afarwuah et al.,
126 2015), as well as the considerations underlying the concentrations of the nutrients in the MMN
127 and SQ-LNS (Arimond et al., 2013). Apart from iron which was kept at 20 mg/day in the MMN
128 and SQ-LNS, the vitamin and mineral contents were either 1x or 2x the RDA for pregnancy, or
129 in a few cases, the maximum amount that could be included in the supplement given technical
130 and organoleptic constraints. The IFA and MMN supplements were provided as capsules in
131 blister packs, and were intended to be consumed with water after a meal, one capsule per day
132 throughout pregnancy. The LNS supplement was in 20-g sachets, and was intended to be mixed
133 with any prepared food, one sachet per day throughout pregnancy. To maintain blinding, two
134 individuals independent of the study placed color-coded stickers behind the blister packs (three
135 different colors for IFA and three for MMN supplements) so that the capsules were known to the
136 study team and participants only by the colors of the stickers. Laboratory staff and data analysts
137 had no knowledge of group assignment until all preliminary analyses had been completed and
138 the allocation codes were broken. The study was registered on ClinicalTrials.gov (Identifier:
139 NCT00970866) and was approved by ethics committees of the University of California, Davis,

140 the Ghana Health Service, and the University of Ghana Noguchi Memorial Institute for Medical
141 Research.

142

143 **Procedures**

144 We collected socio-demographic information at baseline, and determined GA mostly by
145 ultrasound biometry (Aloka SSD 500, Tokyo, Japan). During follow-up, field workers visited
146 women in their homes every two weeks, whereupon they delivered a fresh supply of supplement
147 and monitored supplement intakes. At each of laboratory assessments at baseline and at 36 GW,
148 women's weight (Seca 874) and height (Seca 217) were measured, and peripheral malaria
149 parasitemia (Clearview Malarial Combo, Vision Biotech, South Africa), hemoglobin, Hb
150 (HemoCue AG, Wetzikon, Switzerland) and zinc protoporphyrin, ZPP (hematofluorometer, Aviv
151 Biomedical Co. NJ, USA), were determined using venous blood (Adu-Afarwuah et al., 2015).

152 We used the original Aviv cover-slides and 3-level control material for the ZPP measurements,
153 after red blood cells were washed three times with normal saline. Plasma samples obtained after
154 blood was centrifuged at 1,252 x g for 15 min were stored in Ghana at -20 °C, before being air-
155 freighted on dry ice to UC Davis, where soluble transferrin receptor (TfR, mg/L), C-reactive
156 protein (CRP, mg/L), and alpha-1 glycoprotein (AGP, g/L) concentrations were determined
157 using a Cobas Integra 400 plus Automatic Analyzer (Roche Diagnostic Corp., Indianapolis, IN).

158 At 36 GW, the continuous outcomes measures were Hb (g/L), ZPP ($\mu\text{mol/mol}$ heme),
159 and plasma TfR (mg/L), CRP (mg/L), and AGP (g/L) concentrations, whilst the binary outcome
160 measures were the percentages of women with low Hb, high Hb, and elevated ZPP, TfR, CRP
161 and AGP.

162

163 Sample size and data analysis

164 For the Ghana iLiNS-DYAD Study, an effect size (Cohen's d: difference between group
165 means divided by the pooled standard deviation) of 0.3 (considered a small-to-moderate effect
166 size) (Cohen) was the basis for sample size calculation. Thus, our sample size was based on
167 detecting an effect size of 0.3 between any two groups for any continuous variable at 36 GW,
168 with a two-sided 5% test and 80% power. As described previously (Adu-Afarwuah et al., 2015),
169 we enrolled 1320 pregnant women into the study, but after excluding 177 who received both IFA
170 and MMN supplements during pregnancy because of a temporary mislabeling of supplements, as
171 well as 86 in the LNS group who were pregnant during the same time period, 1,057 women were
172 included in the current analysis. Based on a sample size of 827 women (~275 per group) for
173 whom data were available at 36 GW, we had 94% power to detect an effect size of 0.3 between
174 any two groups for hemoglobin, ZPP, or TfR. This would allow a difference of 3.4 g/L in Hb, 8.9
175 $\mu\text{mol/mol}$ heme in ZPP, and 0.5 mg/L in TfR (given SD of 11.0, 30.0, and 2.0, respectively) to
176 be detected between any 2 groups.

177 We posted the statistical analysis plan (www.ilins.org) before analysis. Statistical
178 analysis, by intention-to-treat, was performed using SAS for Windows Release 9.3 (Cary, NC,
179 USA). Background socio-demographic characteristics were summarized as mean \pm SD for
180 continuous variables, or number of participants and percentages for categorical variables. As
181 done previously (Adu-Afarwuah et al., 2015), we used 2 indices, namely assets index and
182 housing index as proxy indicators for socioeconomic status, and calculated household food
183 insecurity access (HFIA) score (Coates et al., 2007) as a measure of degree of household food
184 insecurity. Higher values of the assets and housing indices represented higher socioeconomic
185 status, and higher values of the food insecurity index represented higher food insecurity.

186 We calculated adherence to treatment as percentage of days from enrolment to the home
187 visit closest to the laboratory assessment at 36 GW, when women reported consuming the
188 supplement. We used Hb <100 g/L as our primary definition for low Hb (representing anemia).
189 This was based on previous WHO (WHO, 2007, WHO/UNICEF/UNU, 2001) and International
190 Nutritional Anemia Consultative Group, INACG (Nestel and INACG Steering Committee, 2002)
191 documents that suggest lowering the standard 110 g/L cut-off by 10 g/L for pregnant women of
192 African extraction to achieve adequate sensitivity and specificity for screening purposes
193 (WHO/UNICEF/UNU, 2001). In addition, we defined low Hb using the standard cut-off of Hb
194 <110 g/L, based on a recent WHO recommendation (WHO, 2011) to maintain that cut-off (110
195 g/L) without any adjustment, because of scarce evidence to support the adjustment. A meta-
196 analysis (Haider et al., 2013) revealed that Hb cut-offs ranging from <100 g/L to 115 g/L have
197 been used in studies to define anemia in pregnant women. We defined high Hb as >130 g/L
198 (Pena-Rosas et al., 2012), elevated ZPP (proxy for iron deficiency) as >60 $\mu\text{mol/mol}$ heme
199 (Walsh et al., 2011) and elevated TfR (proxy for tissue iron deficiency) as > 6.0 mg/L (Pfeiffer et
200 al., 2007, Vandevijvere et al., 2013). Because there is no generally accepted cut-off value for
201 TfR, we derived the 6.0 mg/L cut-off based on the evidence that TfR values obtained using the
202 Automatic Analyzer assay (as used in this study) were on average 30% lower than values
203 obtained with the ELISA assay (Pfeiffer et al., 2007). Therefore, we reduced by 30% the 8.5
204 mg/L cut-off value used when TfR was determined using ELISA (Vandevijvere et al., 2013) to
205 obtain the cut-off of approximately 6.0 mg/L for our analysis. Because we used 2 cut-offs to
206 define anemia, we also defined iron deficiency anemia (IDA) in two ways: first as Hb <100 g/L
207 and at least one marker of iron deficiency (ZPP > 60 ($\mu\text{mol/mol}$ heme) or TfR >6.0 mg/L
208 (Pfeiffer et al., 2007, Vandevijvere et al., 2013)), and second, as Hb <110 g/L and at least one

209 marker of iron deficiency. We defined elevated inflammatory markers using the cut-off values of
210 >5.0 mg/L for CRP and >1.0 g/L for AGP (Thurnham and McCabe), and categorized women
211 with inflammation as either elevated CRP only (indicative of incubation phase of infection),
212 elevated CRP and AGP (indicative of early convalescence) or elevated AGP only (indicative of
213 late convalescence) (Thurnham and McCabe).

214 At 36 GW, we calculated overall mean (\pm SD) values and percentages for Hb and markers
215 of iron status and inflammation. We compared groups by using general linear models
216 (continuous outcomes) and logistic regression models (binary), with Tukey-Kramer adjustment
217 for multiple comparisons. Along with the group comparisons, we calculated pairwise mean
218 differences (continuous outcomes, ANOVA) and relative risks (binary outcomes, Logistic
219 regression) with their 95% CI and p-values. Relative risks were calculated using Poisson
220 regression (Spiegelman and Hertzmark, 2005). In addition, we analyzed changes in the
221 prevalence of anemia, high Hb, and elevated ZPP, TfR, CRP and AGP from enrolment using
222 mixed model logistic regression (SAS PROC GLIMMIX). Where the mixed model logistic
223 regression failed to converge because of sparse data, we used generalized estimating equations
224 model (SAS PROC GENMOD). We analyzed each outcome twice, first without any covariate
225 adjustments, and then with adjustment for covariates significantly associated ($p < 0.10$) with the
226 outcome in a bivariate analysis. Because ZPP, TfR, AGP and CRP are not normally distributed,
227 we calculated the group means (\pm SD or SE), group percentages, and pair-wise mean differences
228 and relative risks with their 95% CI based on untransformed data, but generated the p-values for
229 group or pair-wise comparisons using logarithmically transformed data.

230 To investigate the possible effect of group differences in adherence to treatment, we
231 performed a per-protocol analysis, which was restricted to women with adherence $\geq 70\%$. We

232 evaluated potential interaction of treatment group with pre-specified baseline variables for
233 maternal characteristics, anemia and iron status. These variables were: age, years of schooling,
234 BMI, gestational age at enrolment, household assets index, housing index, food insecurity access
235 score, season at enrolment (dry or wet), primiparous, anemia, and elevated ZPP, TfR, and AGP
236 or CRP. Where an interaction was significant ($\alpha < 0.10$), we performed subgroup analysis by
237 including an interaction term between treatment and the effect modifier in the ANCOVA or
238 logistic regression model. For continuous effect modifiers, we used data from all participants to
239 create a linear regression model to predict the values of the outcome at the 10th and 90th
240 percentile of the effect modifier distribution. Each effect modifier was considered separately in
241 the models to avoid collinearity.

242 In a sensitivity analysis aimed at correcting for the effect of inflammation (CRP and
243 AGP) on the Hb and iron status outcomes, we repeated the above analyses using values of Hb
244 and iron status markers corrected for inflammation (WHO, 2007). These corrected values were
245 calculated by grouping women into 3 inflammation categories, estimating the correction factor
246 (CF) for each inflammation category, and multiplying the Hb and iron status values of each
247 woman by the inflammation category-specific CF (Grant et al., 2012). The 3 inflammation
248 categories were: reference (normal CRP and AGP), incubation (raised CRP and normal AGP),
249 and early (raised CRP and AGP) or late (normal CRP and raised AGP) convalescence [these two
250 phases of convalescence were combined because of small sample sizes and little indication of
251 differences]. For ZPP at 36 GW, women were grouped into 2 inflammation categories (normal
252 versus any inflammation), since the 3-category grouping did not yield consistent results.

253

254 **Results**

255 We collected data from December 2009 to August 2012. The study flow diagram, as well
256 as the main reasons women were not eligible, or were eligible but not enrolled, were reported
257 elsewhere (Adu-Afarwuah et al., 2015). Among eligible women, those enrolled (n=1320) and
258 those not enrolled (n=606) did not differ in most background characteristics (results not shown).
259 The background characteristics of the 1057 women included in the current analysis are presented
260 in **Table 1**. These characteristics were well-balanced across the three groups. On average,
261 women were about 26 years of age, had about 7 years of formal education, and BMI of about 25
262 kg/m³. Nearly all of the women said they were married or living with a partner, slightly more
263 than a third were primiparous, and nearly 10% tested positive for malaria. The average GA at
264 enrolment was 16 weeks. At baseline, we obtained Hb values for all 1057 women, ZPP values
265 for 1055 women, and TfR, CRP and AGP values for 1032 women.

266 At 36 GW, dropout (4.4%) was low, and did not differ among groups ($p = 0.65$). Women
267 who dropped out – mainly because of miscarriage (2.8%) and movement from the study site
268 (1.2%) –did not differ in the baseline characteristics from those who were present at 36 GW,
269 except for GA (weeks) at enrolment, which was significantly lower ($p = 0.001$) for the former
270 (14.5) than for the latter (16.3). Mean (\pm SD) adherence (% of days from enrolment to the home
271 visit closest to the laboratory assessment at 36 GW when supplement was reportedly consumed)
272 was lower ($p=0.001$) in the LNS group (68 ± 24) compared to the IFA (74 ± 21) and the MMN
273 (72 ± 23) groups. Women usually reported mixing the LNS supplement with porridge, but
274 sometimes mixed it with other foods including soups and stews, or consumed it alone. We
275 obtained Hb values for 827 women, and ZPP, TfR, CRP and AGP values for 822 women; the
276 number (%) of women without Hb values did not differ between groups ($p = 0.10$). The women

277 with Hb values did not differ from those without Hb values in most of the baseline
278 characteristics, except that the latter had lower mean housing index (-0.17 ± 1.09 vs. 0.04 ± 0.99 ;
279 $p = 0.011$), BMI (24 ± 4.0 vs. 25 ± 4.7 kg/m²; $p = 0.022$) and GA at enrolment (16 ± 3.1 vs. $16 \pm$
280 3.3 weeks; $p = 0.042$).

281 In the intention-to-treat analysis, the overall mean (\pm SD) values at baseline and 36 GW
282 were 112 ± 12 and 117 ± 12 , respectively, for Hb (g/L), 45 ± 32 and 47 ± 30 for ZPP (μ mol/mol
283 heme) and 4.1 ± 2.5 and 4.5 ± 1.7 for TfR (mg/L). From baseline to 36 GW, the prevalence of
284 anemia decreased significantly from 13% to 5.3% ($p < 0.001$), whilst the reverse was true for the
285 prevalence of high Hb (4.9% vs 12%, $p < 0.001$) and elevated TfR (9.2% to 15%; $p = 0.001$),
286 with a moderate change in the prevalence of elevated ZPP (13% vs. 16%; $p = 0.06$). The mean
287 (\pm SD) CRP (mg/L) and AGP (g/L) at baseline (6.9 ± 11 and 0.6 ± 0.2 , respectively) were slightly
288 greater than at 36 GW (5.7 ± 17 and 0.5 ± 0.2 , respectively), which was also reflected in the
289 percentages of women with elevated CRP and AGP at baseline vs. 36 GW (38.2% vs 24.1%; $p <$
290 0.001 and 6.3% vs 2.8%; $p < 0.001$, respectively).

291

292 **Main group comparisons at 36 gestational weeks**

293 **Table 2** shows the unadjusted mean (\pm SD) Hb (g/L), ZPP (μ mol/mol heme), TfR (mg/L),
294 CRP (mg/L) and AGP (g/L) concentrations, by intervention group, at baseline and 36 GW in the
295 intention-to-treat analysis. Baseline values did not differ significantly among groups. At 36 GW,
296 mean Hb was significantly ($p < 0.001$) greater in the IFA group (120 ± 11) than in the LNS
297 (115 ± 12) or MMN (117 ± 12) group; ZPP was significantly ($p < 0.001$) lower in the IFA group (43
298 ± 30) than in the LNS (50 ± 29) or MMN (49 ± 30) group; and TfR was significantly ($p < 0.001$)
299 lower in the IFA group (4.0 ± 1.3) than in the LNS (4.9 ± 1.8) or MMN (4.6 ± 1.7) group. Further

300 (Table 3), the percentage of women with anemia defined either as Hb <100 g/L or Hb <110 g/L
301 was significantly lower in the IFA group compared with the LNS and MMN groups, and when
302 using the latter definition, this percentage was also significantly greater in the LNS compared
303 with the MMN group. Compared with the IFA group, the LNS and MMN groups had greater
304 percentages of women with elevated ZPP (9.4% vs 19% and 19%, respectively; $p = 0.003$) and
305 elevated TfR (9.0% vs 19% and 15 %, respectively; $p = 0.004$). Differences among the 3 groups
306 in the prevalence of IDA were marginally significant ($p = 0.07$) when the Hb cut-off of 100 g/L
307 was used in the definition of IDA, but were significant ($p < 0.001$) when the Hb cut-off of 110
308 g/L was used. In the latter situation, the risk of IDA was significantly greater in the LNS group
309 compared with the IFA group ($p < 0.001$), and marginally greater in the MMN compared to the
310 IFA group, and in the LNS compared with the MMN group. The prevalence of high Hb did not
311 differ among groups ($p = 0.15$).

312 From baseline to 36 GW, the decrease in the prevalence of anemia (based on our primary
313 definition of Hb <100 g/L) and increase in the prevalence of high Hb were significant for all
314 groups; the change in prevalence over time differed between groups for anemia (P -interaction =
315 0.099) but not for high Hb (P -interaction = 0.95). For elevated ZPP, the increase in prevalence
316 from baseline was marginally significant in the LNS group ($p = 0.06$) and non-significant in the
317 other two groups, and the change in prevalence over time did not differ between groups (P -
318 interaction = 0.14). For elevated TfR, the increase in prevalence was significant for the LNS ($p =$
319 0.003) and MMN ($p = 0.009$) groups only, and the change in prevalence over time did not differ
320 between groups (P -interaction = 0.17). Apart from the prevalence of anemia defined using the
321 Hb cut-off of 110 g/L, the LNS and MMN groups did not differ significantly in any of the
322 continuous (Table 2) or binary (Table 3) Hb and iron status outcomes.

323 There were no significant differences among groups in 36 GW mean (\pm SD) concentrations
324 of CRP ($p = 0.98$) and AGP ($p = 0.35$) (Table 2), or the percentages of women in the incubation
325 phase of infection (elevated CRP only; $p = 0.26$), early convalescence (both CRP and AGP
326 elevated; $p = 0.67$) or late convalescence (elevated AGP only; $p = 1.00$). From baseline to 36
327 GW, the decrease in the percentage of women in the incubation phase of infection was
328 significant for all groups. For percentage of women in early convalescence, only the decrease in
329 the IFA group ($p = 0.010$) was significant, and for percentage of women in late convalescence
330 (where the mixed model logistic regression did not converge because of sparse data, and hence
331 generalized estimating equations model was used), only the decrease in the MMN group ($p =$
332 0.024) was significant.

333 Adjustments by covariates significantly associated with Hb and the iron status (ZPP and
334 TfR) and inflammatory (CRP and AGP) outcomes (including the baseline value for each
335 outcome) did not alter the unadjusted results (data not shown). In addition, correcting the Hb and
336 iron status values for inflammation in the sensitivity analysis (results not shown) did not change
337 the above findings.

338 The per-protocol analysis (results not shown) revealed that among women with adherence
339 to treatment $\geq 70\%$ (samples sizes at 36 gestational weeks: 213 in IFA, 222 in MMN and 166 in
340 the LNS group, for Hb), the above findings remained unchanged, except for the fact that the
341 prevalence of anemia (defined as Hb < 100 g/L or < 110 g/L) did not differ between the LNS and
342 MMN groups, and the prevalence of IDA (if definition included Hb < 110 g/L) was greater only
343 in the LNS compared with the IFA group.

344

345 **Effect modification**

346 Interactions of treatment group with BMI, season of enrolment and elevated CRP
347 concentration at baseline were not significant for any of the outcomes. As shown in **Table 4**, the
348 effect of intervention on ZPP concentration at 36 GW was modified by baseline anemia (Hb <
349 100 g/L; *P*-interaction = 0.099), elevated ZPP (*P*-interaction = 0.013) and TfR (*P*-interaction =
350 0.090), GA at enrolment (*P*-interaction = 0.041), and household assets score (*P*-interaction =
351 0.061). Specifically, the difference in mean (\pm SD) ZPP concentration at 36 GW among
352 intervention groups (MMN and LNS groups compared with the IFA group) was greater among
353 women with anemia at baseline, elevated baseline ZPP, greater GA at enrolment, or lower
354 household assets scores . Similarly, the difference in risk of elevated ZPP among intervention
355 groups was greater in women with greater GA at enrolment (*P*-interaction = 0.057), and elevated
356 TfR at baseline (*P*-interaction = 0.049)

357 The difference in mean TfR concentration at 36 GW among intervention groups was
358 greater among women who were anemic at baseline (*P*-interaction = 0.079), did not have
359 elevated AGP at baseline (*P*-interaction = 0.051), or had greater GA at enrolment (*P*-interaction
360 = 0.016), whereas the difference in risk of elevated TfR among intervention groups was greater
361 in women with less food insecurity.

362 For elevated CRP and AGP at 36 GW, there were significant interactions between
363 intervention group and elevated ZPP at baseline (*P*-interaction = 0.008) and household assets
364 score (*P*-interaction = 0.091), but the stratified analyses (Table 4) did not show consistent results.

365

366 **Discussion**

367 In the iLiNS-DYAD-Ghana study, pregnant women who were provided with standard
368 iron (60 mg) and folic acid supplements from ≤ 20 GW had significantly greater mean Hb, lower
369 mean ZPP and TfR and lower prevalence of anemia and iron deficiency (elevated ZPP and TfR)
370 at 36 GW than pregnant women who were provided with either the MMN or LNS supplements,
371 both of which contained 20 mg iron. Overall, however, the prevalence of anemia (Hb < 100 g/L)
372 at 36 GW was relatively low (2.2-7.9%), and iron deficiency was evident in <20% of women in
373 all groups. At 36 GW, the three groups did not differ in the percentage of women with high Hb
374 (12.3% overall) or inflammation (CRP and AGP). These findings remained unchanged when
375 analyses were restricted to women who were more adherent to treatment.

376 A few weaknesses of our study were described previously (Adu-Afarwuah et al., 2015),
377 namely: (a) a fully double-blind study design was not possible because of the physical
378 differences between the SQ-LNS in the form of sachets, and the other two supplements (MMN
379 and IFA) in the form of capsules, and (b) adherence to treatment was assessed by self-report and
380 not direct observation. However, none of the individuals involved in sample collection,
381 laboratory measurements or data analysis had any knowledge of group assignment. The good
382 collaboration with the antenatal clinics (which gave women the confidence to cooperate),
383 relatively low rate of attrition, intense follow-up of participants, and detailed attention we paid to
384 ensuring data quality were notable strengths of the study.

385 Several possible explanations may be relevant for the observed lower Hb and higher ZPP
386 and TfR values in the MMN and LNS groups compared to the IFA group. First, the 20 mg iron
387 dose used in the MMN and LNS supplements may have been too low for this Ghanaian
388 population of pregnant women. In most (but not all) similar studies, the iron dose of the iron +

389 folic acid supplement was 60 mg/day and that of the multiple micronutrient supplement was 30
390 mg/day (Allen and Peerson, 2009, Roberfroid et al., 2011, Mei et al., 2014), although in
391 Indonesia (Suprpto et al., 2002), Mexico (Ramakrishnan et al., 2004), Nepal (Christian et al.,
392 2003) and Tanzania (Makola et al., 2003), both the iron + folic acid and multiple micronutrient
393 supplement groups received iron doses of at least 50 mg/ day. In these studies (even for those
394 that used multiple micronutrients containing 30 mg Fe), pregnant women consuming the multiple
395 micronutrient supplements generally did not differ in Hb or iron status indicators compared to
396 those consuming iron + folic acid (Allen and Peerson, 2009, Mei et al., 2014, Roberfroid et al.,
397 2011). A study of Australian women (Zhou et al., 2009) suggested that 20 mg iron per day may
398 be an adequate dose to prevent iron deficiency anemia during pregnancy compared with higher
399 doses of iron. However, the diet of the women in our sample, as is typical of Ghana, is mainly
400 plant-based and high in phytate (Gibson, 1994), which reduces iron absorption (Baech et al.,
401 2003), so dietary iron needs during pregnancy may be higher in Ghana than in Australia.

402 Another possibility is that the relatively high dose of zinc (30 mg) in the MMN and LNS
403 supplements may have interfered with iron absorption, as suggested by the results of a
404 supplementation trial in Nepal (Christian et al., 2003). The lack of a good biomarker of zinc
405 status at the individual level makes it difficult to explore this potential mechanism for the
406 differences in iron status between the IFA and other two groups.

407 It is noteworthy that there were no significant differences in mean Hb or iron status
408 between the LNS and MMN groups, despite the fact that SQ-LNS is a food-based supplement
409 rather than a capsule and some differences in composition could have affected these outcomes.
410 For instance, it is possible that the calcium or phytate in SQ-LNS could have limited the
411 absorption of iron in the LNS group. The lack of differences in Hb and iron status between these

412 two groups suggests that the iron or other micronutrient content (which was identical in these
413 two supplements, except for the macro-minerals) was the most critical factor.

414 In Ghana, anemia (Ghana Statistical Service (GSS) et al., 2009) and infections including
415 malaria (Yatich et al., 2009) are common among pregnant women even in relatively high income
416 communities, and evidence (Mockenhaupt et al., 2000) suggests that malaria is a major risk
417 factor for anemia. Thus, it is noteworthy that the prevalence of both anemia and elevated CRP
418 declined significantly in all three groups between baseline and 36 weeks gestation.

419 It is important to consider the implications of the observed lower mean Hb of the SQ-LNS
420 and MMN groups compared to the IFA group at 36 weeks gestation. Low Hb concentration or
421 anemia in the first or second trimester of pregnancy is linked with poor pregnancy outcomes
422 including low birth weight (Murphy et al., 1986), but no such association has been established
423 for low Hb concentration or anemia in the third trimester (Allen, 2000). Further, Hb
424 concentrations substantially above 110–119 g/L during pregnancy may be independent of iron
425 status and have been linked with poorer health outcomes for the mother and fetus (Yip, 2000,
426 Zhou et al., 1998). Therefore, the higher mean Hb concentrations observed for women in the IFA
427 group compared to those in the LNS and MMN groups in the third trimester (36 GW) may not
428 necessarily be beneficial with respect to birth outcomes. In fact, we previously demonstrated that
429 in this same study (Adu-Afarwuah et al., 2015) the prenatal consumption of LNS (compared to
430 IFA) was associated with greater birth weight, weight-for-age z-score and BMI-for-age z-score,
431 and that, in first-time mothers, prenatal LNS supplementation also increased birth length and
432 head circumference and reduced the proportion of infants with low birth weight, low birth length,
433 and small-for-gestational age. In this cohort, there was no relationship between Hb at 36 wk
434 gestation and infant birth size, and there was actually a significant *negative* relationship between

435 maternal iron status at 36 wk and birth size (Oaks et al., 2015). Thus, the difference between the
436 LNS and IFA groups in mean maternal Hb, ZPP, and TfR concentrations needs to be weighed
437 against the difference (in the opposite direction) in birth outcomes. In two sets of analyses (Garn
438 et al., 1981, Steer et al., 1995) each involving a large number of pregnant women, the lowest risk
439 of adverse birth outcomes including low birth weight was seen in women with Hb ~95-105 g/L
440 (Steer et al., 1995) or Hb ~100 -110 g/L (Garn et al., 1981). Appropriate cut-offs for ZPP and
441 TfR in pregnancy are not well documented, particularly with respect to functional outcomes.
442 Therefore, it is difficult to judge whether our results should be interpreted as “improvements” in
443 maternal iron status in the IFA group compared to the MMN or LNS group.

444 We conclude that among pregnant women in a semi-urban setting in Ghana,
445 supplementation with SQ-LNS or MMN containing 20 mg iron resulted in lower Hb and iron
446 status but had no impact on inflammation, when compared with iron (60 mg) plus folic acid (400
447 µg) treatment. The amount of iron in such supplements that is most effective for improving both
448 maternal Hb/iron status and birth outcomes requires further evaluation.

449

450 **Key messages**

- 451 1. In this semi-urban Ghanaian population, the prevalence of anemia (Hb <100 g/L) among
452 pregnant women who received IFA, MMN or SQ-LNS was reduced from 13% at <20
453 gestational weeks (GW) to 5% at 36 GW.
- 454 2. Provision of IFA (with 60 mg Fe) was associated with a greater mean concentration of
455 Hb and lower prevalence of anemia at 36 GW than provision of MMN or SQ-LNS (both
456 with 20 mg Fe).

- 457 3. The prevalence of high Hb (>130 g/L) or elevated inflammatory biomarkers (CRP and
458 AGP) at 36 GW was not affected by the type of prenatal supplement provided.
- 459 4. More research is needed to determine the concentration of iron in MMN and SQ-LNS
460 supplements that is most effective for improving both maternal Hb/iron status and birth
461 outcomes.

462 **References**

- 463 Adu-Afarwuah S., Lartey A., Okronipa H., Ashorn P., Zeilani M., Peerson J.M., et al. (2015)
464 Lipid-based nutrient supplement increases the birth size of infants of primiparous women
465 in Ghana. *The American Journal of Clinical Nutrition* **101**, 835-846.
- 466 Allen L.H. (2000) Anemia and iron deficiency: effects on pregnancy outcome. *The American*
467 *Journal of Clinical Nutrition* **71**, 1280S-1284S.
- 468 Allen L.H. & Peerson J.M. (2009) Impact of multiple micronutrient versus iron-folic acid
469 supplements on maternal anemia and micronutrient status in pregnancy. *Food and*
470 *Nutrition Bulletin* **30**, S527-532.
- 471 Arimond M., Zeilani M., Jungjohann S., Brown K.H., Ashorn P., Allen L.H., et al. (2013)
472 Considerations in developing lipid-based nutrient supplements for prevention of
473 undernutrition: experience from the International Lipid-Based Nutrient Supplements
474 (iLiNS) Project. *Maternal & Child Nutrition*.
- 475 Baech S.B., Hansen M., Bukhave K., Jensen M., Sorensen S.S., Kristensen L., et al. (2003)
476 Nonheme-iron absorption from a phytate-rich meal is increased by the addition of small
477 amounts of pork meat. *The American Journal of Clinical Nutrition* **77**, 173-179.
- 478 Bertran N., Camps J., Fernandez-Ballart J., Arija V., Ferre N., Tous M., et al. (2005) Diet and
479 lifestyle are associated with serum C-reactive protein concentrations in a population-
480 based study. *Journal of Laboratory and Clinical Medicine* **145**, 41-46.
- 481 Brighenti F., Valtuena S., Pellegrini N., Ardigo D., Del Rio D., Salvatore S., et al. (2005) Total
482 antioxidant capacity of the diet is inversely and independently related to plasma
483 concentration of high-sensitivity C-reactive protein in adult Italian subjects. *The British*
484 *Journal of Nutrition* **93**, 619-625.

- 485 Christian P., Shrestha J., LeClerq S.C., Khattry S.K., Jiang T., Wagner T., et al. (2003)
486 Supplementation with micronutrients in addition to iron and folic acid does not further
487 improve the hematologic status of pregnant women in rural Nepal. *The Journal of*
488 *Nutrition* **133**, 3492-3498.
- 489 Coates J., Swindale A. & Bilinsky P. (2007) Household Food Insecurity Access Scale (HFIAS)
490 for Measurement of Food Access: Indicator Guide (V.3) [Internet]. Food and Nutrition
491 Technical Assistance Project, Academy for Educational Development, Washington, D.C
492 [cited 2013 Aug 12]. Available from: [http://www.fao.org/fileadmin/user_upload/eufao-](http://www.fao.org/fileadmin/user_upload/eufao-fsi4dm/doc-training/hfias.pdf)
493 [fsi4dm/doc-training/hfias.pdf](http://www.fao.org/fileadmin/user_upload/eufao-fsi4dm/doc-training/hfias.pdf).
- 494 Cohen J. *Statistical Power Analysis in the Behavioral Sciences* (2nd edition). Hillsdale (NJ):
495 Lawrence Erlbaum Associates, Inc., 1988.
- 496 Devaraj S. & Jialal I. (2000) Alpha tocopherol supplementation decreases serum C-reactive
497 protein and monocyte interleukin-6 levels in normal volunteers and type 2 diabetic
498 patients. *Journal of Free Radicals in Biology & Medicine* **29**, 790-792.
- 499 Elovitz M.A. (2006) Anti-inflammatory interventions in pregnancy: Now and the future.
500 *Seminars in Fetal & Neonatal Medicine* **11**, 327-332.
- 501 Fall C.H., Fisher D.J., Osmond C. & Margetts B.M. (2009) Multiple micronutrient
502 supplementation during pregnancy in low-income countries: a meta-analysis of effects on
503 birth size and length of gestation. *Food and Nutrition Bulletin* **30**, S533-546.
- 504 Friso S., Jacques P.F., Wilson P.W.F., Rosenberg I.H. & Selhub J. (2001) Low Circulating
505 Vitamin B6 Is Associated With Elevation of the Inflammation Marker C-Reactive Protein
506 Independently of Plasma Homocysteine Levels. *Circulation* **103**, 2788-2791.

- 507 Garn S.M., Keating M.T. & Falkner F. (1981) Hematological status and pregnancy outcomes.
508 *The American Journal of Clinical Nutrition* **34**, 115-117.
- 509 Ghana Statistical Service (GSS), Ghana Health Service (GHS) & ICF Macro (2009) Ghana
510 Demographic and Health Survey 2008. Accra, Ghana: GSS, GHS, and ICF Macro.
- 511 Gibson R.S. (1994) Zinc nutrition in developing countries. *Nutrition Research Reviews* **7**, 151-
512 173.
- 513 Grant F.K., Suchdev P.S., Flores-Ayala R., Cole C.R., Ramakrishnan U., Ruth L.J., et al. (2012)
514 Correcting for inflammation changes estimates of iron deficiency among rural Kenyan
515 preschool children. *The Journal of Nutrition* **142**, 105-111.
- 516 Haider B.A. & Bhutta Z.A. (2012) Multiple-micronutrient supplementation for women during
517 pregnancy. *The Cochrane Database of Systematic Reviews* **11**, CD004905.
- 518 Haider B.A., Olofin I., Wang M., Spiegelman D., Ezzati M., Fawzi W.W., et al. (2013) Anaemia,
519 prenatal iron use, and risk of adverse pregnancy outcomes: systematic review and meta-
520 analysis. *BMJ*, **346**, f3443.
- 521 Haider B.A., Yakoob M.Y. & Bhutta Z.A. (2011) Effect of multiple micronutrient
522 supplementation during pregnancy on maternal and birth outcomes. *BMC Public Health*
523 **11 Suppl 3**, S19.
- 524 Hambidge K.M., Krebs N.F., Westcott J.E., Garces A., Goudar S.S., Kodkany B.S., et al. (2014)
525 Preconception maternal nutrition: a multi-site randomized controlled trial. *BMC*
526 *Pregnancy & Childbirth* **14**, 111.
- 527 Imdad A. & Bhutta Z.A. (2012) Routine iron/folate supplementation during pregnancy: effect on
528 maternal anaemia and birth outcomes. *Paediatric and Perinatal Epidemiology* **26 Suppl**
529 **1**, 168-177.

- 530 International Anemia Consultative Group (INACG) (2002) Report of the 2001 International
531 Anemia Consultative Group Symposium. Why is iron important and what to do about it:
532 a new perspective. Washington, DC, INACG Secretariat.
- 533 IOM (2001) *Dietary Reference Intakes for Vitamin A, Vitamin K, Arsenic, Boron, Chromium,*
534 *Copper, Iodine, Iron, Manganese, Molybdenum, Nickel, Silicon, Vanadium and Zinc.*
535 National Academy Press, Washington, DC.
- 536 Kaestel P., Michaelsen K.F., Aaby P. & Friis H. (2005) Effects of prenatal multimicronutrient
537 supplements on birth weight and perinatal mortality: a randomised, controlled trial in
538 Guinea-Bissau. *The European Journal of Clinical Nutrition* **59**, 1081-1089.
- 539 Makola D., Ash D.M., Tatala S.R., Latham M.C., Ndossi G. & Mehansho H. (2003) A
540 micronutrient-fortified beverage prevents iron deficiency, reduces anemia and improves
541 the hemoglobin concentration of pregnant Tanzanian women. *The Journal of Nutrition*
542 **133**, 1339-1346.
- 543 Mei Z., Serdula M.K., Liu J.M., Flores-Ayala R.C., Wang L., Ye R., et al. (2014) Iron-
544 containing micronutrient supplementation of Chinese women with no or mild anemia
545 during pregnancy improved iron status but did not affect perinatal anemia. *The Journal of*
546 *Nutrition* **144**, 943-948.
- 547 Michaelsen K.F., Dewey K.G., Perez-Exposito A.B., Nurhasan M., Lauritzen L. & Roos N.
548 (2011) Food sources and intake of n-6 and n-3 fatty acids in low-income countries with
549 emphasis on infants, young children (6-24 months), and pregnant and lactating women.
550 *Maternal & Child Nutrition* **7 Suppl 2**, 124-140.
- 551 Mockenhaupt F.P., Rong B., Günther M., Beck S., Till H., Kohne E., et al. (2000) Anaemia in
552 pregnant Ghanaian women: importance of malaria, iron deficiency, and

- 553 haemoglobinopathies. *Transaction of the Royal Society of Tropical Medicine & Hygiene*
554 **94**, 477-483.
- 555 Murphy J.F., O'Riordan J., Newcombe R.G., Coles E.C. & Pearson J.F. (1986) Relation of
556 haemoglobin levels in first and second trimesters to outcome of pregnancy. *Lancet* **1**,
557 992-995.
- 558 Nestel P. & INACG Steering Committee (2002) Adjusting Hemoglobin Values in Program
559 Surveys [Internet]. INACG, Washington, DC [cited 2014-Jan 10]. Available from:
560 http://pdf.usaid.gov/pdf_docs/PNACQ927.pdf.
- 561 Oaks B., Stewart C., Laugero K., Adu-Afarwuah S., Lartey A., Baldiviez L., et al. (2015)
562 Associations of maternal cortisol, inflammation, hemoglobin, iron status, and BMI with
563 birth outcomes in pregnant women in Ghana. *The Federation of American Societies for*
564 *Experimental Biology Journal* **29**.
- 565 Pena-Rosas J.P., De-Regil L.M., Dowswell T. & Viteri F.E. (2012) Daily oral iron
566 supplementation during pregnancy. *The Cochrane Database of Systematic Reviews* **12**,
567 CD004736.
- 568 Pfeiffer C.M., Cook J.D., Mei Z., Cogswell M.E., Looker A.C. & Lacher D.A. (2007) Evaluation
569 of an automated soluble transferrin receptor (sTfR) assay on the Roche Hitachi analyzer
570 and its comparison to two ELISA assays. *Clinica Chimica Acta; International Journal of*
571 *Clinical Chemistry* **382**, 112-116.
- 572 Picklesimer A.H., Jared H.L., Moss K., Offenbacher S., Beck J.D. & Boggess K.A. (2008)
573 Racial differences in C-reactive protein levels during normal pregnancy. *American*
574 *Journal of Obstetrics and Gynecology* **199**, 523.e521-523.e526.

- 575 Rallidis L.S., Paschos G., Liakos G.K., Velissaridou A.H., Anastasiadis G. & Zampelas A.
576 (2003) Dietary α -linolenic acid decreases C-reactive protein, serum amyloid A and
577 interleukin-6 in dyslipidaemic patients. *Atherosclerosis* **167**, 237-242.
- 578 Ramakrishnan U., Grant F.K., Goldenberg T., Bui V., Imdad A. & Bhutta Z.A. (2012) Effect of
579 multiple micronutrient supplementation on pregnancy and infant outcomes: a systematic
580 review. *Paediatric and Perinatal Epidemiology* **26 Suppl 1**, 153-167.
- 581 Ramakrishnan U., Neufeld L.M., Gonzalez-Cossio T., Villalpando S., Garcia-Guerra A., Rivera
582 J., et al. (2004) Multiple micronutrient supplements during pregnancy do not reduce
583 anemia or improve iron status compared to iron-only supplements in Semirural Mexico.
584 *The Journal of Nutrition* **134**, 898-903.
- 585 Ren A., Wang J., Ye R.W., Li S., Liu J.M. & Li Z. (2007) Low first-trimester hemoglobin and
586 low birth weight, preterm birth and small for gestational age newborns. *International*
587 *Journal of Gynaecology and Obstetrics: The Official Organ of the International*
588 *Federation of Gynaecology and Obstetrics* **98**, 124-128.
- 589 Research Engagement on Food Innovation for Nutritional Effectiveness (REFINE) (2013)
590 Ongoing studies using Lipid Based Nutrient Supplement (LNS) [Internet]. REFINE.
591 Boston, MA. [cited 2015 Jun 23]. Available from:
592 <http://refinenutrition.org/research/REFINE-product-LNS.pdf>.
- 593 Roberfroid D., Huybregts L., Habicht J.P., Lanou H., Henry M.C., Meda N., et al. (2011)
594 Randomized controlled trial of 2 prenatal iron supplements: is there a dose-response
595 relation with maternal hemoglobin? *The American Journal of Clinical Nutrition* **93**,
596 1012-1018.

- 597 Roberts J.M., Balk J.L., Bodnar L.M., Belizán J.M., Bergel E. & Martinez A. (2003) Nutrient
598 Involvement in Preeclampsia. *The Journal of Nutrition* **133**, 1684S-1692S.
- 599 Sacks G.P., Studena K., Sargent I.L. & Redman C.W.G. (1998) Normal pregnancy and
600 preeclampsia both produce inflammatory changes in peripheral blood leukocytes akin to
601 those of sepsis. *American Journal of Obstetrics and Gynecology* **179**, 80-86.
- 602 Spiegelman D. & Hertzmark E. (2005) Easy SAS calculations for risk or prevalence ratios and
603 differences. *American Journal of Epidemiology* **162**, 199-200.
- 604 Steer P., Alam M.A., Wadsworth J. & Welch A. (1995) Relation between maternal haemoglobin
605 concentration and birth weight in different ethnic groups. *BMJ*, **310**, 489-491.
- 606 Suprpto B., Widardo & Suhanantyo (2002) Effect of low-dosage vitamin A and riboflavin on
607 iron-folate supplementation in anaemic pregnant women. *Asia Pacific Journal Clinical*
608 *Nutrition* **11**, 263-267.
- 609 Tatala S., Svanberg U. & Mduma B. (1998) Low dietary iron availability is a major cause of
610 anemia: a nutrition survey in the Lindi District of Tanzania. *The American Journal of*
611 *Clinical Nutrition* **68**, 171-178.
- 612 Thurnham D.I. & McCabe G.P. Influence of infection and inflammation on biomarkers of
613 nutritional status with an emphasis on vitamin A and iron. In: World Health
614 Organization. Report: Priorities in the assessment of vitamin A and iron status in
615 populations, Panama City, Panama, 15–17 September 2010 [Internet]. World Health
616 Organization, Geneva; 2012 [cited 2014 Nov 14]. Available from:
617 http://www.who.int/nutrition/publications/micronutrients/background_paper4_report_assessment_vitAandIron_status.pdf.
618

- 619 UNICEF/WHO/UNU Composition of a multi-micronutrient supplement to be used in pilot
620 programmes among pregnant women in developing countries [Internet]. New York:
621 UNICEF. 1999 [cited 2015 Jan 18]. Available from:
622 [http://apps.who.int/iris/bitstream/10665/75358/1/UNICEF-WHO-multi-](http://apps.who.int/iris/bitstream/10665/75358/1/UNICEF-WHO-multi-micronutrients.pdf?ua=1)
623 [micronutrients.pdf?ua=1](http://apps.who.int/iris/bitstream/10665/75358/1/UNICEF-WHO-multi-micronutrients.pdf?ua=1).
- 624 van den Broek N.R., Letsky E.A., White S.A. & Shenkin A. (1998) Iron status in pregnant
625 women: which measurements are valid? *The British Journal of Haematology* **103**, 817-
626 824.
- 627 Vandevijvere S., Amsalkhir S., Van Oyen H., Egli I. & Moreno-Reyes R. (2013) Iron status and
628 its determinants in a nationally representative sample of pregnant women. *Journal of the*
629 *Academy of Nutrition and Dietetics* **113**, 659-666.
- 630 Walsh T., O'Broin S.D., Cooley S., Donnelly J., Kennedy J., Harrison R.F., et al. (2011)
631 Laboratory assessment of iron status in pregnancy. *Clinical Chemistry and Laboratory*
632 *Medicine : CCLM / FESCC* **49**, 1225-1230.
- 633 WHO Guideline: Daily iron and folic acid supplementation in pregnant women [Internet]. World
634 Health Organization, Geneva; 2012 [cited 2014 Jul 13]. Available from:
635 http://apps.who.int/iris/bitstream/10665/77770/1/9789241501996_eng.pdf?ua=1.
- 636 WHO (2007) Assessing the iron status of populations: Report of a joint World Health
637 Organization/ Centers for Disease Control and Prevention technical consultation on the
638 assessment of iron status at the population level, 2nd Ed. [Internet]. World Health
639 Organization, Geneva [cited 2015 Oct 22]. Available from:
640 http://www.who.int/nutrition/publications/micronutrients/anaemia_iron_deficiency/97892
641 [41596107.pdf](http://www.who.int/nutrition/publications/micronutrients/anaemia_iron_deficiency/9789241596107.pdf)

- 642 WHO (2011) Hemoglobin concentrations for the diagnosis of anemia and assessment of severity
643 [Internet]. Vitamin and Mineral Nutrition Information System. Geneva, World Health
644 Organization (WHO/NMH/NHD/MNM/11.1) [cited 2015 Oct 21]. Available from:
645 <http://www.who.int/vmnis/indicators/haemoglobin.pdf>.
- 646 WHO/UNICEF/UNU (2001) Iron deficiency anaemia assessment, prevention, and control: a
647 guide for programme managers. WHO/NHD/01.3. [Internet]. World Health Organization,
648 Geneva [cited 2014 Dec 01]. Available from:
649 http://www.who.int/nutrition/publications/en/ida_assessment_prevention_control.pdf.
- 650 World Health Organization (1992) The Prevalence of Anaemia in Women. WHO/MCH
651 /MSM/92.2.
- 652 Yatich N.J., Yi J., Agbenyega T., Turpin A., Rayner J.C., Stiles J.K., et al. (2009) Malaria and
653 Intestinal Helminth Co-infection Among Pregnant Women in Ghana: Prevalence and
654 Risk Factors. *The American Journal of Tropical Medicine and Hygiene* **80**, 896-901.
- 655 Yip R. (2000) Significance of an abnormally low or high hemoglobin concentration during
656 pregnancy: special consideration of iron nutrition. *The American Journal of Clinical*
657 *Nutrition* **72**, 272S-279S.
- 658 Zhou L.-M., Yang W.-W., Hua J.-Z., Deng C.-Q., Tao X. & Stoltzfus R.J. (1998) Relation of
659 hemoglobin measured at different times in pregnancy to preterm birth and low birth
660 weight in Shanghai, China. *American Journal of Epidemiology* **148**, 998-1006.
- 661 Zhou S.J., Gibson R.A., Crowther C.A. & Makrides M. (2009) Should we lower the dose of iron
662 when treating anaemia in pregnancy? A randomized dose-response trial. *European*
663 *Journal of Clinical Nutrition* **63**, 183-190
- 664

TABLE 1

Background characteristics of pregnant Ghanaian women whose hemoglobin, and iron status and inflammatory markers were analyzed at 36 gestational weeks, by intervention group¹

Background characteristics	IFA (N = 349)	MMN (N = 354)	LNS (N = 354)
Age, y	27 ± 5.3 [349]	27 ± 5.7 [354]	27 ± 5.4 [354]
Formal education, y	7.6 ± 3.5 [349]	7.5 ± 3.6 [354]	7.7 ± 3.7 [354]
Body Mass Index, kg/m ²	25 ± 4.3 [342]	25 ± 5.0 [348]	25 ± 4.4 [349]
Low BMI, n/N (%)	11/342 (3.2)	8/348 (2.3)	6/349 (1.7)
Gestational age at enrolment, weeks	16.3 ± 3.3 [346]	16.2 ± 3.2 [353]	16.2 ± 3.3 [349]
Assets index ²	0.09 ± 0.98 [342]	0.1 ± 0.9 [349]	-0.01 ± 0.91 [348]
Housing index ²	0.04 ± 0.99 [342]	-0.03 ± 1.03 [349]	0.00 ± 1.00 [348]
HFIA Score ³	2.9 ± 4.6 [345]	2.5 ± 3.9 [346]	2.5 ± 3.9 [348]
Married or co-habiting, n/N (%)	320/349 (91.7)	332/354 (93.8)	328/354 (92.7)
Primiparous women, n/N (%)	131/349 (37.5)	110/354 (31.1)	128/354 (36.2)
Tested positive for malaria ⁴ , n/N (%)	31/349 (8.9)	30/354 (8.5)	40/354 (11.3)

¹ IFA= Iron-Folic Acid group; MMN=Multiple Micronutrients group. LNS= Small Quantity Lipid-based Nutrient Supplement group. HFIA is Household Food Insecurity Access Score. N=total number of participants in the group in question; n = number of participants positive on the variable in question; % = percent of participants positive on the variable in question. Values are Mean ± SD [N] or n/N (%).

² Proxy indicators for household socioeconomic status; higher values represent higher socioeconomic status.

³ HFIA (Household food insecurity access) is a proxy indicator for household food insecurity (Coates et al., 2007); higher values represent higher food insecurity

⁴ Rapid Diagnostic Test (Clearview Malarial Combo, Vision Biotech, South Africa), which detected *P. falciparum* and non-*P. falciparum* histidine-rich protein-2

TABLE 2

Unadjusted hemoglobin, and iron status and inflammatory markers of pregnant Ghanaian women at baseline and 36 gestational weeks, by intervention group, and pair-wise comparison of groups¹

Variable	IFA ² [N=349]	MMN ² [N=354]	LNS ² [N=354]	P ³	Comparison of IFA and MMN		Comparison of IFA and LNS		Comparison of MMN and LNS	
					Mean difference (95 % CI)	p	Mean difference (95 % CI)	p	Mean difference (95 % CI)	p
Hb, g/L										
Baseline	112 ± 13 [349]	111 ± 12 [354]	112 ± 12 [354]		-1 (-4, 1)		0 (-2, 2)		1 (-1, 3)	
36 GW	120 ± 11 [270]	117 ± 12 [291]	115 ± 12 [266]	<0.001	-3 (-6, -1)	0.002	-5 (-7, 3)	<0.001	-2 (-4, 1)	0.18
ZPP, µmol/mol heme										
Baseline	43 ± 28 [347]	46 ± 36 [354]	45 ± 33 [354]		3 (-3, 8)		2 (-4, 8)		-1 (-6, 5)	
36 GW	42 ± 30 [267]	49 ± 30 [291]	50 ± 29 [264]	<0.001	6 (0, 12)	<0.001	7 (1, 13)	<0.001	1 (-5, 7)	0.91
TfR, mg/L										
Baseline	4.0 ± 1.9 [338]	4.0 ± 1.7 [348]	4.3 ± 3.5 [346]		0.0 (-0.4, 0.5)		0.3 (-0.1, 0.8)		0.3 (-0.1, 0.8)	
36 GW	4.0 ± 1.3 [266]	4.6 ± 1.7 [291]	4.9 ± 1.8 [265]	<0.001	0.5 (0.2, 0.9)	<0.001	0.8 (0.5, 1.2)	<0.001	0.3 (-0.0, 0.6)	0.07
CRP, mg/L										
Baseline	7.9 ± 14 [338]	5.8 ± 8.3 [348]	6.9 ± 11 [346]		-2.1 (-4.1, -0.1)		-1.1 (-3.1, 1.0)		1.0 (-1.0, 3.0)	
36 GW	5.6 ± 18 [266]	5.7 ± 14 [291]	5.9 ± 18 [265]	0.85	0.1 (-3.3, 3.5)	0.95	0.3 (-3.2, 3.8)	0.97	0.2 (-3.2, 3.6)	0.84
AGP, g/L										
Baseline	0.7 ± 0.2 [338]	0.6 ± 0.2 [348]	0.6 ± 0.2 [346]		0.0 (-0.1, 0.0)		0.0 (-0.1, 0.0)		0.0 (0.0, 0.0)	

36 GW	0.5 ± 0.2 [266]	0.5 ± 0.2 [291]	0.5 ± 0.2 [265]	0.30	0.0 (0.0, 0.1)	0.65	0.0 (0.0, 0.0)	0.80	0.0 (-0.1, 0.0)	0.27
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¹IFA= Iron-Folic Acid group; MMN=Multiple Micronutrients group; LNS= Small Quantity Lipid-based Nutrient Supplement group. AGP, CRP, GW, Hb, TfR, and ZPP are alpha-1 acid glycoprotein, C-reactive protein, gestational weeks, hemoglobin, transferrin receptor, and zinc protoporphyrin, respectively. Analyses are based on ANOVA (SAS, PROC GLM). Group means (±SD) and pair-wise mean difference (95% CI) were calculated using untransformed data. Except for Hb, all p-values for group or pair-wise comparisons were generated from log-transformed data; untransformed data were used for comparisons of mean Hb values. N=total number of participants in the group in question.

² Values are Mean ± SD [N].

³P-values, with Tukey-Kramer adjustments, compare all three groups.

TABLE 3

Number (percentage) of pregnant Ghanaian women with abnormal hemoglobin, and iron status and inflammatory markers at baseline and 36 weeks of gestation, by intervention group and pairwise relative risks (RR) between groups¹

	IFA ² [N=349]	MMN ² [N=354]	LNS ² [N=354]	P ³	Comparison of IFA and MMN ⁴		Comparison of IFA and LNS ⁴		Comparison of MMN and LNS ⁴	
					RR (95 % CI)	p	RR (95 % CI)	p	RR (95 % CI)	p
Low Hb (< 100 g/L)⁵										
Baseline	39/349 (11.2)	51/354 (14.4)	47/354 (13.3)		1.29 (0.81, 2.05)		1.19 (0.74, 1.91)		0.92 (0.59, 1.43)	
36 GW	6/270 (2.2)	17/291 (5.8)	21/266 (7.9)	0.019	2.63 (0.88, 7.86)	0.09	3.55 (1.22, 10.3)	0.013	1.35 (0.65, 2.83)	0.60
Low Hb (< 110 g/L)⁶										
Baseline	139/349 (39.8)	157/354 (44.4)	134/354 (37.9)		1.11 (0.90, 1.37)		0.95 (0.76, 1.19)		0.85 (0.69, 1.06)	
36 GW	38/270 (14.1)	69/291 (23.7)	88/266 (33.1)	<0.001	1.68 (1.10, 2.59)	0.011	2.35 (1.56, 3.53)	<0.001	1.40 (1.01, 1.92)	0.038
High Hb⁷										
Baseline	21/349 (6.0)	15/354 (4.2)	16/354 (4.5)		0.70 (0.33, 1.52)		0.75 (0.35, 1.60)		1.07 (0.47, 2.43)	
36 GW	42/270 (15.6)	32/291 (11.0)	28/266 (10.5)	0.15	0.71 (0.42, 1.18)	0.25	0.68 (0.40, 1.16)	0.20	0.96 (0.54, 1.70)	0.98
Elevated ZPP⁸										
Baseline	40/347 (11.5)	54/354 (15.3)	46/354 (13.0)		1.32 (0.84, 2.09)		1.13 (0.70, 1.81)		0.85 (0.55, 1.32)	
36 GW	25/267 (9.4)	56/291 (19.2)	49/264 (18.6)	0.003	2.06 (1.21, 3.48)	0.003	1.98 (1.16, 3.40)	0.007	0.96 (0.64, 1.46)	0.98
Elevated TfR⁹										
Baseline	29/338 (8.6)	29/348 (8.3)	37/346 (10.7)		0.97 (0.54, 1.75)		1.25 (0.72, 2.17)		1.28 (0.74, 2.23)	
36 GW	24/266 (9.0)	44/291 (15.1)	51/265 (19.2)	0.004	1.68 (0.96, 2.94)	0.08	2.13 (1.24, 3.67)	0.003	1.27 (0.82, 1.97)	0.40
IDA¹⁰										
Baseline	20/338 (5.9)	22/348 (6.3)	22/346 (6.4)		1.07 (0.53, 2.15)		1.07 (0.53, 2.17)		1.01 (0.51, 1.99)	
36 GW	2/264 (0.8)	11/291 (3.8)	11/263 (4.2)	0.07	4.99 (0.83, 29.9)	0.09	5.52 (0.92, 33.1)	0.06	1.11 (0.42, 2.95)	0.97
IDA¹¹										
Baseline	36/338 (10.7)	37/348 (10.6)	37/346 (10.7)		1.00 (0.59, 1.68)		1.00 (0.60, 1.69)		1.01 (0.60, 1.68)	
36 GW	12/264 (4.5)	27/291 (9.3)	41/263 (15.6)	<0.001	2.04 (0.93, 4.49)	0.08	3.43 (1.63, 7.20)	<0.001	1.68 (0.97, 2.90)	0.07
Elevated CRP only¹²										

Baseline	113/338 (33.4)	108/348 (31.0)	121/346 (35.0)		0.93 (0.72, 1.20)		1.05 (0.82, 1.34)		1.13 (0.87, 1.45)	
36 GW	49/266 (18.4)	70/291 (24.1)	59/265 (22.3)	0.26	1.31 (0.89, 1.93)	0.24	1.21 (0.81, 1.81)	0.52	0.93 (0.64, 1.33)	0.87
Elevated CRP+AGP ¹³										
Baseline	26/338 (7.7)	15/348 (4.3)	11/346 (3.2)		0.56 (0.27, 1.17)		0.41 (0.18, 0.94)		0.74 (0.30, 1.84)	
36 GW	7/266 (2.6)	9/291 (3.1)	5/265 (1.9)	0.67	1.18 (0.37, 3.77)	0.94	0.72 (0.18, 2.79)	0.83	0.61 (0.17, 2.22)	0.64
Elevated AGP only ¹⁴										
Baseline	3/338 (0.9)	5/348 (1.4)	5/346 (1.4)		1.62 (0.30, 8.88)		1.63 (0.30, 8.93)		1.01 (0.23, 4.38)	
36 GW	1/266 (0.4)	0/291 (0.0)	1/265 (0.4)	1.00						

¹IFA= Iron-Folic Acid group; MMN=Multiple Micronutrients group; LNS=Small Quantity Lipid-based Nutrient Supplement group. AGP, CRP, GW, Hb, IDA, TfR, and ZPP are alpha-1 acid glycoprotein, C-reactive protein, gestational weeks, hemoglobin, iron deficiency anemia, transferrin receptor, and zinc protoporphyrin, respectively. N=total number of participants in the group in question.

²Values are number of participants positive on the variable in question/N (% of participants positive on the variable in question).

³P-values compare all three groups, with Tukey-Kramer adjustment, using logistic regression (SAS PROC LOGISTIC).

⁴Relative Risks, RR(95% CI) and their p-values are based on Poisson regression (Spiegelman and Hertzmark, 2005).

⁵Nestel and INACG Steering Committee, 2002, WHO, 2007, WHO/UNICEF/UNU, 2001.

⁶WHO, 2011.

⁷Hb >130 g/L (Pena-Rosas et al., 2012).

⁸ZPP > 60.0 (µmol/mol heme).

⁹TfR >6.0 mg/L (Pfeiffer et al., 2007, Vandevijvere et al., 2013).

¹⁰Hb <100 g/L (Nestel and INACG Steering Committee, 2002, WHO, 2007, WHO/UNICEF/UNU, 2001) and at least one marker of iron deficiency (ZPP > 60.0 (µmol/mol heme) or TfR >6.0 mg/L (Pfeiffer et al., 2007, Vandevijvere et al., 2013)).

¹¹Hb <110 g/L (WHO, 2011) and at least one marker of iron deficiency (ZPP > 60.0 (μmol/mol heme) or TfR >6.0 mg/L (Pfeiffer et al., 2007, Vandevijvere et al., 2013)).

¹²CRP >5.0 mg/L and AGP not >1.0 g/L (Thurnham and McCabe).

¹³ CRP >5.0 mg/L and AGP >1.0 g/L (Thurnham and McCabe).

¹⁴ CRP not > 5.0 mg/L and AGP >1.0 g/L (Thurnham and McCabe).

TABLE 4

Effect of intervention on iron status and inflammatory outcomes of pregnant Ghanaian women, stratified by baseline characteristics¹

Outcomes	IFA ²	MMN ²	LNS ²	P ³	P ⁴	Comparison of MMN and IFA (n = 349)		Comparison of LNS and IFA (n = 354)		Comparison of LNS and MMN (n = 354)	
						Difference or RR	P	Difference or RR	P	Difference or RR	P
ZPP, µmol/mol heme⁵											
Baseline anemia				0.099							
No [†]	44 ± 2 [233]	47 ± 2 [245]	48 ± 2 [226]		0.09	3 (-2, 9)	0.34	5 (-0, 11)	0.08	2 (-4, 8)	0.71
Yes	36 ± 6 [26]	54 ± 5 [41]	53 ± 5 [33]		0.013	18 (3, 34)	0.017	18 (1, 34)	0.032	-1 (-16, 14)	0.99
Elevated baseline ZPP				0.013							
No	42 ± 2 [230]	46 ± 2 [242]	46 ± 2 [226]		0.19	3 (-2, 9)	0.34	4 (-2, 10)	0.20	1 (-5, 7)	0.94
Yes	47 ± 5 [29]	64 ± 5 [44]	72 ± 5 [33]		0.001	18 (3, 32)	0.015	25 (9, 41)	0.001	8 (-7, 22)	0.43
Elevated baseline TfR				0.090							
No	43 ± 2 [239]	47 ± 2 [261]	49 ± 2 [233]		0.06	4 (-2, 9)	0.28	6 (0, 12)	0.048	2 (-4, 8)	0.65
Yes	41 ± 6.5 [20]	63 ± 6 [25]	59 ± 6 [26]		0.017	22 (3, 41)	0.017	18 (-1, 36)	0.07	-4 (-22, 14)	0.84
GA at enrolment				0.041							
At 10 th percentile [‡]	46 ± 3	51 ± 3	44 ± 3		0.28	5 (-5, 15)	0.44	-1 (-11, 9)	0.97	-6 (-16, 4)	0.29
At 90 th percentile	40 ± 3	48 ± 3	54 ± 3		0.002	7 (-1, 16)	0.12	13 (5, 22)	0.001	6 (-3, 15)	0.25
Assets score				0.061							
At 10 th percentile	40 ± 3	46 ± 3	54 ± 3		0.003	5 (-4, 15)	0.38	14 (4, 24)	0.002	8 (-1, 18)	0.11
At 90 th percentile	46 ± 3	50 ± 3	46 ± 3		0.37	5 (-4, 13)	0.44	-0 (-9, 9)	1.00	-5 (-14, 4)	0.46
Elevated ZPP⁶											
GA at enrolment				0.057							
At 10 th percentile [§]	11.1 (5.7, 20.5)	23.2 (15.3, 33.6)	11.6 (6.4, 20.0)		0.06	3.0 (0.9, 4.6)	0.12	1.0 (0.4, 2.5)	1.00	0.5 (0.2, 1.1)	0.11
At 90 th percentile	5.8 (2.9, 11.4)	14.9 (9.6, 22.3)	20.7 (14.1, 29.2)		0.004	2.4 (1.0, 5.7)	0.049	3.4 (1.5, 7.7)	0.003	1.4 (0.8, 2.5)	0.46

Outcomes	IFA ²	MMN ²	LNS ²	P ³	P ⁴	Comparison of MMN and IFA (n = 349)		Comparison of LNS and IFA (n = 354)		Comparison of LNS and MMN (n = 354)	
						Difference or RR	P	Difference or RR	P	Difference or RR	P
Elevated baseline TfR					0.049						
No [¶]	17/234 (7.4)	44/256 (17.4)	32/229 (14.0)		0.008	2.2 (1.2, 4.0)	0.006	1.8 (1.0, 3.4)	0.08	0.8 (0.5, 1.3)	0.61
Yes	1/20 (3.0)	3/25 (10.0)	8/25 (31.3)		0.014	1.3 (0.4, 4.6)	0.41	2.7 (0.8, 9.4)	0.016	2.1 (0.7, 6.1)	0.14
TfR, mg/L ⁷											
Baseline anemia					0.079						
No	4.1 ± 0.1 [232]	4.5 ± 0.1 [245]	4.8 ± 0.1 [226]		<0.001	0.4 (0.1, 0.8)	0.004	0.7 (0.4, 1.0)	<0.001	0.3 (-0.0, 0.6)	0.11
Yes	3.5 ± 0.3 [26]	4.8 ± 0.3 [41]	5.0 ± 0.3 [33]		<0.001	1.3 (0.4, 2.1)	0.002	1.5 (0.6, 2.4)	<0.001	0.2 (-0.6, 1.1)	0.76
Elevated baseline AGP					0.051						
No	4.1 ± 0.1 [238]	4.6 ± 0.1 [272]	4.9 ± 0.1 [250]		<0.001	0.6 (0.3, 0.9)	<0.001	0.8 (0.5, 1.2)	<0.001	0.3 (-0.02, 0.6)	0.07
Yes	4.0 ± 0.4 [20]	4.0 ± 0.4 [14]	3.3 ± 0.5 [9]		0.503	-0.0 (-1.2, 1.2)	1.00	-0.7 (-2.1, 0.8)	0.51	-0.66 (-2.2, 0.8)	0.56
GA at enrolment					0.016						
At 10 th percentile	4.4 ± 0.2	5.0 ± 0.2	4.8 ± 0.2		0.023	0.9 (0.1, 1.1)	0.019	0.4 (-0.1, 0.9)	0.16	-0.2 (-0.7, 0.3)	0.64
At 90 th percentile	3.8 ± 0.1	4.3 ± 0.1	4.9 ± 0.1		<0.001	0.5 (0.0, 0.9)	0.041	1.1 (0.7, 1.6)	<0.001	0.7 (0.2, 1.1)	0.002
Elevated TfR ⁸											
HFIA score					0.047						
At 10 th percentile	5.3 (2.9, 9.7)	15.7 (10.8, 22.2)	15.3 (10.5, 21.7)		0.004	2.3 (1.1, 4.7)	0.006	2.5 (1.2, 5.0)	0.007	1.1 (0.6, 1.8)	1.00
At 90 th percentile	9.7 (5.1, 17.5)	4.9 (1.7, 13.5)	18.7 (10.3, 31.5)		0.06	0.7 (0.2, 2.0)	0.50	1.9 (0.8, 4.6)	0.27	2.8 (0.9, 8.7)	0.06
Elevated CRP ⁹											
Elevated baseline ZPP					0.008						
No	41/225 (18.3)	72/238 (30.1)	47/221 (21.4)		0.013	1.6 (1.1, 2.4)	0.014	1.2 (0.8, 1.8)	0.71	0.7 (0.5, 1.1)	0.11
Yes	9/29 (30.1)	6/43 (13.5)	11/33 (32.0)		0.09	0.5 (0.2, 1.1)	0.17	1.1 (0.5, 2.1)	0.99	2.1 (0.9, 4.9)	0.10

Outcomes	IFA ²	MMN ²	LNS ²	P ³	P ⁴	Comparison of MMN and IFA (n = 349)		Comparison of LNS and IFA (n = 354)		Comparison of LNS and MMN (n = 354)	
						Difference or RR	P	Difference or RR	P	Difference or RR	P
Elevated AGP ¹⁰											
Assets score				0.091							
At 10 th percentile	1.0 (0.2, 4.8)	1.0 (0.2, 4.5)	2.8 (0.9, 8.4)		0.39	1.0 (0.1, 9.8)	1.00	2.8 (0.6, 14.1)	0.51	2.8 (0.3, 28.0)	0.48
At 90 th percentile	4.3 (1.5, 11.3)	4.4 (1.7, 10.8)	0.7 (0.1, 3.9)		0.14	1.0 (0.2, 4.5)	1.00	0.16 (0.0, 1.3)	0.16	0.2 (0.0, 1.4)	0.14

¹IFA= Iron-Folic Acid group; MMN=Multiple Micronutrients group; LNS=Small Quantity Lipid-based Nutrient Supplement group. GA, Hb, HFIA, TfR, and ZPP are gestation age, hemoglobin, household food insecurity access, transferrin receptor, and Zinc protoporphyrin, respectively. Baseline anemia is Hb <100 g/L (Nestel and INACG Steering Committee, 2002, WHO, 2007, WHO/UNICEF/UNU, 2001). Analyses are based on ANCOVA (SAS PROC MIXED, with SLICE option) for continuous outcomes or logistic regression (SAS PROC GLIMMIX, with SLICE option) for binary outcomes. Linear regression modeling was used to predict the outcome at the 10th and 90th percentile of continuous baseline effect modifiers.

²Values are mean ± SE [total number of participants], or mean ± SE, or percent of participants positive on the variable in question (95% CI) or number of participants positive on the variable in question/total number of participants (percent of participants positive on the variable in question). Values are adjusted for variables significantly associated with the outcome variable in bivariate analysis.

³P-values are for interaction with the outcome in question

⁴P-values compare all three groups in each stratum.

⁵ZPP values are adjusted for age, parity, season at enrolment, and baseline Hb, ZPP, and TfR concentrations.

⁶Elevated ZPP (ZPP >60 µmol/mol heme) adjusted for age, education, and baseline Hb, ZPP and TfR concentrations.

⁷TfR values are adjusted for age, season at enrolment, and baseline Hb, ZPP, TfR and AGP concentrations.

⁸Elevated TfR (TfR > 6.0 mg/L) adjusted for age, gestational age enrolment, season at enrolment, and baseline Hb, ZPP, and TfR concentrations.

⁹Elevated CRP (CRP > 5.0 mg/L) adjusted for BMI, household food insecurity access score, and baseline ZPP, AGP and CRP concentrations.

¹⁰Elevated AGP (AGP > 1.0 g/L) adjusted for age, parity, and season as enrolment.

[†]Values are group mean \pm SE [number of participants], and difference in means (95% CI) and p-values. All such values.

[‡]Values are group mean \pm SE, and difference in means (95% CI) and p-values. All such values.

[§]Values are group % (95% CI), and relative risk (95% CI) and p-values. All such values.

[¶]Values are number of participant (%), and relative risk (95% CI) and p-values. All such values.