



International Journal of Sciences: Basic and Applied Research (IJSBAR)

ISSN 2307-4531
(Print & Online)

<http://gssrr.org/index.php?journal=JournalOfBasicAndApplied>



The Relationship of Inflammatory Regulation and Pain Intensity in SIRS Patients

Dwi Pantja Wibowo^{a*}, Suryani As'ad^b, Fransiscus Suhadi^c, I Wayan Suranadi^d,
Ilhamjaya Patellongi^e, Irawan Yusuf^f, Mohammad Ramli Ahmad^g, Syafrie
Kamsul Arif^h, Andi Husni Tanraⁱ

^{a,c}Anesthesiology and Intensive Care Department, Bintaro Premier Hospital, Banten, Indonesia

^bClinical Nutrition Department, Hasanuddin University, Makassar, Indonesia

^dAnesthesiology and Intensive Care Department, Udayana University, Denpasar, Indonesia

^{e,f}Physiology Department, Hasanuddin University, Makassar, Indonesia

^{g,h,i}Anesthesiology and Intensive Care Management Department, Hasanuddin University, Makassar, Indonesia

^aEmail: dwipantja@yahoo.com

Abstract

Aims of this study were to investigate the changes in inflammatory regulation as shown by proinflammatory cytokines (IL-6) and anti-inflammatory cytokines (IL-10) from patients with systemic inflammatory responses syndrome (SIRS) that affect pain intensity changes with the marked increase of critical-care pain observation tools (CPOT) and decreased of the pain pressure threshold (PPT). A cross-sectional analysis to compare the values of IL-6, IL-10, PPT, and CPOT of SIRS patients and patients without SIRS. Of the 46 patients who were the subjects of the study, there were 21 SIRS patients and 25 patients non-SIRS. Patients with SIRS had higher CPOT values than patients without SIRS; CPOT values (3.3 vs. 1.2), significant with $p = 0.001$ ($p < 0.05$). The PPT scores of patients with SIRS were lower than those without SIRS (4.24 vs. 7.37), significant with $p = 0.001$ ($p < 0.05$). We conclude that in SIRS patients there is an increase in both proinflammatory (IL-6) and anti-inflammatory (IL-10) cytokines, but none of those cytokines had a relationship with pain intensity.

Keyword: SIRS; CPOT; IL-6; IL-10.

* Corresponding author.

1. Introduction

After an inflammatory stimulation by sepsis, there will be an increase of proinflammatory mediators followed by an increase of anti-inflammatory mediators [1]. Interleukin-6 is a 22- to 27-kDa glycoprotein secreted by many types of cell, such as macrophages, monocytes, eosinophils, hepatocytes, and glial cells. This interleukin is one of the earliest and important mediators of induction and control of acute phase protein synthesis and releases during pain stimuli, such as trauma, infection, surgery, and burns. After an injury, plasma concentrations of IL-6 are detectable within 60 minutes, with a peak between 4 and 6 hours, and it can persist for up to 10 days. It is considered the most appropriate marker of the degree of tissue damage during a surgical procedure in which excessive and prolonged increase is associated with greater postoperative morbidity. Patients with severe sepsis for <48hrs have shown a tight correlation between the elevation of IL-6, the severity of the SIRS and subsequent mortality. Interleukin-10 is an 18-kDa nonglycosylated peptide synthesized in immune cells and neuroendocrine and neural tissues. It inhibits proinflammatory cytokines, especially TNF, IL-1, and IL-6, produced by activated macrophages and monocytes, stimulating endogenous production of anti-inflammatory cytokines. Levels of TNF α and IL-10 were higher in patients with SIRS and MODS, as compared to the healthy volunteers [2,3,4].

There were reports that a number of patients who survived sepsis developed long-term complications such as persistent pain [5], this study attempted to find the cause of chronic pain which usually begins with the occurrence of inflammatory pain with inadequate therapy by finding a link between increased proinflammatory mediators (IL-6) and increased anti-inflammatory mediators (IL-10) with pain intensity (CPOT) and pain excitatory threshold (PPT).

2. Materials and Method

2.1. Collection of Samples

This is an observational study with longitudinal research model. Subjects in this study were patients who admitted to the adult intensive care unit of Bintaro Premier Hospital from April 2015 to December 2015. We included all subjects who fulfilled inclusion criteria: 1) age ≥ 18 years old, 2) indicated to admit ICU, 3) no hepatic failure, 4) no renal impairment and excluded subjects with incomplete data and whose families refuse to be the subjects of our research. Subjects were categorized into two clinical diagnose: SIRS and nonSIRS as we use the SIRS criteria: 1) body temperature $>38^{\circ}\text{C}$ or $<36^{\circ}\text{C}$, 2) heart rate $>90/\text{minute}$, 3) respiratory rate $>20/\text{minute}$ or PaCO₂ <32 mmHg, 4) white blood cells count >12000 cu/mm or <4000 cu/mm or immature neutrophils $>10\%$ as to categorize the subjects.

All samples who fulfilled inclusion and exclusion criteria and willing to participate in the study and to sign informed consent recruited as study samples.

2.2. Measurement of IL-6, and IL-10 using ELISA

Examination the plasma levels of IL-6, and IL-10 were using ELISA direct methods.

2.3. Classification of pain scale by using CPOT criteria

We measure the pain level by using CPOT criteria [6] 1) Facial expression (0-2), 2) Body movement (0-2), 3) Muscle tension (0-2), 4) or Ventilation compliance or vocalization (0-2). We also measure the pain threshold with pressure algometer (PPT) [7] with measurable pressure (kg/m²) at the tendon of extensor carpi radialis [8].

2.4. Data Analysis

Data analysis using the SPSS statistics (IBM Corp. Released 2011, version 20 Armonk, NY, US). The measures expressed as a mean and standard deviation. We evaluate the association between two qualitative variables with Chi Square Test and the association between a qualitative variable and quantitative variable using Mann-Whitney U-test. We performed correlation test with calculating the determinant (R). A probability value less than 5% was considered statistically significant.

2.5. Ethical Clearance

Ethical approval for this study obtained from Mochtar Riady Institute for Nanotechnology as well as Bintaro Premier Hospital’s ethical board, NO: 02.1403014. We obtained written informed consent from all patient’s family.

3. Results

Of the 205 adult patients admitted during the study period, there were 25 SIRS patients who met the inclusion criteria; four patients excluded for rejecting and lacking the data. Of the group of patients, non-SIRS 25 patients were willing to be the subjects of the study, thus during the collection period, we obtained 46 patients as research subjects with 21 patients SIRS (45.7%) and 25 patients, nonSIRS (54.3%). We should consider the population of this study because there are age differences in the two sample groups.

Table 1: Characteristics of Subjects

Characteristics	SIRS		p
	Yes (n=21)	No (n=25)	
	Mean±SD	Mean±SD	
Age (years)	57.9±11.1	51.0±4.8	0.014
BMI (kg/m ²)	25.06±4.90	26.10±5.59	0.511
MAP	87.56±18.10	93.53±8.69	0.177
HR (x/minute)	104.2±21.8	74.5±11.5	<0.001
RR (x/minute)	23.8±8.0	16.5±2.3	0.001
pCO2 (mmHg)	36.1±8.2	36.3±4.9	0.943
WBC (/mm ³)	16743.33±9145.02	5956.00±1668.85	<0.001

Measurements of the pain scale were using the CPOT [1], while measurements of the pain threshold were using algometer in assessing PPT [9]. Both of these measurements were done directly by the researcher to avoid the measurement bias. The CPOT value in SIRS patients was higher than in the non-SIRS group (3.3 versus 1.2: $p = 0.001$). The mean value of PPT in the SIRS group was significantly lower than in the non-SIRS group (4.24 versus 7.37: $p = 0.001$). From these two measures, it concluded that SIRS patients have higher pain with a lower pain threshold. Furthermore, from these findings, it is necessary to analyze the factors that cause it.

Table 2: Pain Score (CPOT), Pain Threshold (PPT), and PGE2

Parameters	SIRS		p
	Yes (n=21)	No (n=25)	
	Mean±SD	Mean±SD	
PPT (kg)	4.24±1.26	7.37±1.31	0.001
CPOT	3.3±1.8	12±0.6	0.001

SIRS is a spectacular inflammatory reaction that is also known as a cytokine storm, the release of various cytokines into the blood circulation [10]. The first phase characterized by an increase in TNF- α , IL-1 β , and IL-6. While the second phase is hypo-inflammation and characterized by increased concentrations of IL-10 and some other anti-inflammatory cytokines. While the next step is a balance between proinflammation and anti-inflammatory [11,12]. To maintain homeostasis, levels of anti-inflammatory cytokines such as interleukin-10 also increased when there was an increase in proinflammatory cytokines [13,14, 15]. Although both proinflammatory and anti-inflammatory levels were significantly increased (IL-6: 129.18 versus 9.70, $p < 0.001$ and IL-10: 114.40 versus 12.03, $p < 0.001$), it turned out that when traced in SIRS patients, the proinflammatory cytokines are more dominant than anti-inflammatory ones. This shown by the difference in the ratio of IL-6 / IL-10 levels in the SIRS group significantly higher than in the no-SIRS group (5.04 versus 0.64, $p < 0.001$). The explanation of this is that the cause of SIRS with all its manifestations is due to the dominance of the higher inflammatory mediators than the anti-inflammatory mediators [16]. Or it can also be said SIRS occurs due to the failure of suppression from the anti-inflammatory mediator. (Table 3).

Table 3: Comparison of inflammatory status between the two groups

Inflammatory status	SIRS		p
	Yes (n=21)	No (n=25)	
	Mean±SD	Mean±SD	
IL-6 (pg/mL)	129.18±110.51	9.70±29.17	<0.001
IL-10 (pg/mL)	114.40±181.32	12.03±27.05	<0.001
IL-6/IL-10 ratio	5.04±7.49	0.64±1.51	<0.001

Pain that occurs in patients with SIRS in the ICU is an inflammatory pain caused by an increase in inflammatory mediators. This pain is a pain that is adaptive and protective with a decrease in the threshold of excitatory pain [17]. The reduced limit of excitatory pain caused by the influence of mediators on nociceptors resulting in increased excitability of outer nociceptor membrane [18]. The arachidonic acid present in the cell wall phospholipid produced when the injury occurs, and the cascade process becomes prostaglandin, prostacyclin, and thromboxane and causes inflammation and pain. This process is the enzyme cyclooxygenase (COX-1) which is the essential protein that causes changes in arachidonic acid into protective compounds and COX-2 enzymes that produce prostaglandins with inflammatory effects, including pain [19]. Although both proinflammatory (IL-6) and anti-inflammatory (IL-10) inflammatory mediators increased in the SIRS group, none were correlated with decreased pain threshold value (PPT) or with increased pain intensity (CPOT) in this study (Table 6 and 7).

Table 6: Correlation between PPT, IL-6, IL-10, and IL-6/IL-10 ratio

	Bivariate		Partial	
	R	<i>p</i>	R	<i>p</i>
PPT vs IL-6	-0.569	<0.001	-0.244	0.069
PPT vs IL-10	-0.515	<0.001	-0.161	0.148
PPT vs IL-6/IL-10	-0.480	<0.001	-0.046	0.381

Table 7: Correlation between CPOT, IL-6, IL-10, and IL-6/IL-10 ratio

	Bivariate		Partial	
	R	<i>p</i>	R	<i>p</i>
CPOT vs IL-6	0.727	<0.001	0.120	0.217
CPOT vs IL-10	0.607	<0.001	0.157	0.152
CPOT vs IL-6/IL-10	0.590	<0.001	0.024	0.438

4. Discussion

From this study, we suspect that SIRS which is the embryo of sepsis as one of the causes of post sepsis chronic pain as we found the hypersensitivity of pain characterized by a decrease in PPT value and an increase of CPOT.

This study showed that in SIRS patients increased proinflammatory cytokines (IL-6) and anti-inflammatory cytokines (IL-10) differed in levels. Critically ill patients in the intensive care unit (ICU) almost always feel pain during treatment. In a study of 158 patients who had been treated in ICU with mechanical ventilation, 47% reported feeling anxious and afraid of their actions, and 36% still remembered the pain they experienced [20]. In

another study conducted by interview, 64% of ICU cardiac surgery patients reported moderate to severe pain [21]. A significant problem is a pain that persists after the patient leaves the ICU. Of these variables, statistically significant causes of the post-treatment pain in ICU were age and sepsis [22].

Pain treatment for ICU patients can be done by decreasing the effects of inflammatory mediators from pain by using anti-inflammatory drugs as an adjunctive therapy in other analgesics drugs [23]. With precaution and to avoid the contraindications such as renal insufficiency, active peptic ulcer, coagulation disorders [24]. To prevent the occurrence of pain hypersensitivity due to both peripheral and central sensitization, ICU physicians may use the multimodal analgesia [25]. Due to the presence of inflammatory dysregulation in SIRS patients, it is important to consider inflammation with anti-inflammatory drugs so that the inflammatory process becomes controlled. Although indirect correlated with the occurrence of pain hypersensitivity, increases in both proinflammatory mediators and anti-inflammatory mediators should be suspected to play a role in the occurrence of hypersensitivity of pain through other mechanisms or other compounds. Therefore it is necessary to do further analysis to find it.

5. Conclusion

Systemic inflammation characterized by SIRS results in changes in the levels of proinflammatory cytokines (IL-6). Accompanied by shifts in the levels of anti-inflammatory cytokines (IL-10) that will decrease excitatory pain threshold (PPT) due to peripheral and central sensitization resulting in increased pain (CPOT) on SIRS patients in ICU. Therefore, to reduce pain hypersensitivity in patients with SIRS, inflammatory controls, such as with anti-inflammatory administration (especially COX-2 inhibitors) or administration of antihyperalgesic drugs are warranted. For the management of sepsis which is the most complicated form of SIRS in the ICU, in addition to efforts to eradicate germs with adequate antibiotics, hemodynamic control or other vital body support, the equally important effort according to the results of this study is to control Inflammatory and surely overcome the resulting hyperalgesia. Options for the administration of antiinflammation that may control the inflammatory dysregulation.

Acknowledgements

We give our gratitude to all contributors who collected samples as well as patients and their families, whose participations and help make this work possible.

6. Footnote

Conflicts of Interest: The authors do not have any direct financial relationships with any trademarks mentioned in the paper that might lead to a conflict of interest for any of the author. The authors declare no potential conflict of interest.

References

- [1] Oberholzer, A, C Oberholzer, and L L Moldawer. "Sepsis syndrome: understanding the role of innate

- and acquire immunity." *Shock* 16 (2001): 83-96.
- [2] De Oliveira CM, Sakata RK, Machado I, Gerola LR, Salomao R. "Cytokines, and Pain." *Rev Bras Anesthesiol* 61, no.2 (2011): 255-265
- [3] Jaffer U, Wade RG, Gourlay T. "Cytokines in the systemic inflammatory response syndrome." *Intensive Care and Cardiovascular Anesthesia* 2 (2010): 161-175.
- [4] Iskander KN, Osuchowski MF, Stearns-Kurosawa DJ, Kurosawa S, Stepien D, Valentine C, Remick DG. "Sepsis: Multiple Abnormalities, Heterogeneous Responses, and Evolving Understanding." *Physiol Rev* 93 (2013): 1247-1288
- [5] Gelinas, C, L Fillon, K A Puntilo, C Viens, and M Fortier. "Validation of the critical-care pain observation tool in adult patients." *American Journal of Critical Care* 15, no. 4 (2006): 420-442.
- [6] Nurnberg, D, F Sabonchi, PV Sackey, and G Bjorling. "A preliminary validation of Swedish version of the critical-care pain observation tool in adults." *Acta Anaesthesiologica* 55, no. 4 (2011): 379-386.
- [7] Ji, R R, T Kohno, K A Moore, and C J Wolf. "Central sensitization and LTP: Do pain and memory share similar mechanisms?" *Trends Neurosci* 23 (2003): 696-705.
- [8] Herridge, M S. "One-year outcomes in survivors of the acute respiratory distress syndrome." *N Engl J Med* 348 (2003): 683-693.
- [9] Kawasaki, Y, L Zhang, Jen-Kun Cheng, and Ru-Rong Ji. "Cytokine mechanisms of central sensitization: distinct and overlapping role of interleukin-1 β , interleukin-6, and tumor necrosis factor- α in regulating synaptic and neuronal activity in the superficial spinal cord." *The Journal of Neuroscience* 28, no. 20 (2008): 5189-5194.
- [10] Bone, R C. "Sir Isaac Newton, sepsis, SIRS, and CARS." *Crit Care Med* 24 (1996): 1125-1128.
- [11] Oberholzer, A, C Oberholzer, and L L Moldawer. "Sepsis syndrome: understanding the role of innate and acquire immunity." *Shock* 16 (2001): 83-89.
- [12] Carson, William F, and L Kunkel Steven. "Immune cell dysfunction as a consequence of severe sepsis." In *Killer, Severe Sepsis and Septic Shock - Understanding a Serious*, by Richardo Fernandez, 125-154. Michigan: Intech, 2012.
- [13] Woolf, CJ, and Q Ma. "Nociceptors-noxious stimulus detectors." *Neuron* 55, no. 3 (2007): 353-364.
- [14] Shankar, Ravi, Kurt A Melstrom, and Richard L Gamelli. "Inflammation and Sepsis: past, present, and the future." *Journal of Burn Care & Research* 28, no. 4 (July 2007): 1-6.
- [15] Shankar-Hari, M, G S Phillips, and M L Levy. "Developing a new definition and assessing new clinical criteria for septic shock: for the Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3)." *JAMA* 315, no. 8 (2016): 775-785.
- [16] Battle, Ceri E, Simon Lovett, and Hayley Hutchings. "Chronic pain in survivors of critical illness: a retrospective analysis of incidence and risk factors." *Critical Care* 17, no. R101 (2013): 1-8.
- [17] Bergbom-Engberg, I, and H Haljamae. "Assessment of patients' experience of discomforts during respiratory therapy." *Crit Care Med* 17 (1989): 1068-1072.
- [18] Puntillo, K A. "Pain experiences of intensive care unit patients." *Heart Lung* 19, no. 5 Pt 1 (1990): 526-533.
- [19] Lee, M, S Silverman, H Hansen, V Patel, and L Manchikanti. "A comprehensive review of opioid-induced hyperalgesia." *Pain Physician* 14 (2011): 145-161.

- [20] Mattia, C, G Savoia, F Paoletti, O Piazza, D Albanese, and B Amantea. "SIAARTI recommendations for analgosedation in intensive care unit." *Minerva Anesthesiol* 72 (2006): 769-805.
- [21] Ching, Tang. "The effect of epidural clonidine on perioperative cytokine response, postoperative pain and bowel function in a patient undergoing colorectal surgery." *Anesth Analg* 99 (2004): 502-507.
- [22] McMahon, S B, W B Cafferty, and F Marchand. "immune and glial cell factors as pain mediators and modulators." *Exp Neurol* 192 (2005): 444-462.
- [23] Very, W A, T M Cunha, C A Parade, S Poole, F Q Cunha, and S H Ferreira. "Hyper nociceptive role of cytokines and chemokines: targets for analgesic drug development?" *Pharmacol Ther* 112, no. 1 (Oct 2006): 116-138.
- [24] Rapanos, T, P Murphy, J P Szalai, L Burlacoff, J Lam-McCulloch, and J Kay. "Rectal indomethacin reduces postoperative pain and morphine use after cardiac surgery." *Can J Anaesth* 46 (1999): 725-730.
- [25] Hynninen, M S, D C Cheng, I Hossain, J Carrol, S S Aumbhagavan, and R Yue. "Non-steroidal anti-inflammatory drugs in the treatment of postoperative pain after cardiac surgery." *CanJ Anaesth* 47 (2000): 1182-1187.