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Same species, different prerequisites: investigating body condition and foraging success in young reef sharks between an atoll and an island system

Ornella C. Weideli^{1,2}, Ian A. Bouyoucos^{1,3}, Yannis P. Papastamatiou⁴, Gauthier Mescam⁵, Jodie L. Rummer³ & Serge Planes^{1,6}

Acquiring and storing energy is vital to sharks of all age-classes. Viviparous shark embryos receive endogenous maternal energy reserves to sustain the first weeks after birth. Then, in order to maintain body condition, sharks must start foraging. Our goal was to understand whether maternal energy investments vary between blacktip reef sharks (*Carcharhinus melanopterus*) from two populations and to what extent body condition and the initiation of foraging might be affected by presumably variable maternal investments. A total of 546 young sharks were captured at St. Joseph atoll (Seychelles) and Moorea (French Polynesia) between 2014 and 2018, and indices of body condition and percentage of stomachs containing prey were measured. Maternal investment was found to be site-specific, with significantly larger, heavier, and better conditioned individuals in Moorea. Despite these advantages, as time progressed, Moorea sharks exhibited significant decreases in body condition and were slower to initiate foraging. We suggest that the young sharks' foraging success is independent of the quality of maternal energy resources, and that other factors, such as prey availability, prey quality, and/or anthropogenic stressors are likely responsible for the observed differences across sites. Insights into intraspecific variations in early life-stages may further support site-specific management strategies for young sharks from nearshore habitats.

Acquiring and storing energy reserves to maintain body functions and survival is vital to animals of all age-classes¹. To estimate energy reserves during various life-stages, body condition, as a proxy of animal health, is commonly used², with animals in good body condition presumably associated with relatively larger energy reserves^{2,3}. At birth, an animal's body condition is determined by the parents, notably by the mother⁴. Depending on maternal size and age at parturition, the diet, as well as the environmental conditions to which the mother was exposed during gestation, the offspring's size, body mass, and body condition can vary among and within species. Indeed, coral reef fishes from high quality habitats pass on larger yolk reserves to their offspring than parents living in low quality habitats⁵. In the first weeks after birth, young animals with no parental care are required to gradually incorporate autonomous foraging activities to their daily routine to sustain their energy reserves. Hence, young animals depend on prey resources and habitat quality, in addition to remaining maternal energy resources. While strong positive relationships between parental energy reserves and factors such as offspring condition and time to exogenous feeding have been noted for teleost fishes and marine reptiles⁵⁻⁸, little work has been done on

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maternal energy investment in elasmobranchs. As maternal investment may vary with life-history traits (e.g., size, body condition) and habitat, it is also important to understand if and to what extent the level of maternal energy investment affects the offspring's condition and foraging development during the first weeks of life.

Elasmobranchs occur across a range of heterogeneous habitats and experience variable environmental conditions and levels of anthropogenic threats that differentially affect life-history traits⁹. While intraspecific differences in life-history traits may be less distinctive in sharks with broad movement patterns, genetically and geographically isolated sharks with restricted movements and site-fidelity are known to exhibit pronounced intraspecific differences in size at birth, growth rates, and litter sizes^{9,10}. Adult reef-sharks from the family Carcharhinidae have been the focus of a number of studies investigating such differences¹⁰, but fewer studies have characterized intra-specific variability among populations of young animals. Barker *et al.*¹¹, for example, reported that larger sized female lemon sharks (*Negaprion brevirostris*) from Florida's Marquesas Keys (USA), give birth to larger offspring than the smaller sized females from a nearby nursery¹¹. Likewise, Hussey *et al.*¹² revealed an increase in maternal reproductive output (larger neonatal mass) with increasing maternal size in two carcharhinid sharks. While such size and body mass measurements can help assess characteristics of young shark populations¹³, their relationship (e.g., body mass per unit size) provides invaluable information on energy reserves, overall body condition and fitness^{3,3,14,15}. Despite the importance of energy reserves, it is unclear as to whether maternal energy investment varies across shark populations adopting different life-history traits.

At birth, viviparous sharks receive endogenous energy reserves, primarily stored as lipids in livers, from their mothers^{12,16–18}. Although this maternal energy allocation can lead to significantly enlarged livers in neonatal sharks (up to 20% of total body mass¹²), this endogenous energy transfer is finite. As opposed to marine mammals, where exogenous maternal energy provisioning (e.g., lactation) can last months to years, depending on species¹⁹, sharks receive no maternal aftercare. This results in energy resources being utilized within the first weeks, as demonstrated by decreasing condition indices^{12,17}. Similarly, Duncan and Holland¹⁶ reported mass loss in young sharks following parturition, most likely a sign of depleted energy reserves. To counteract such declines, young sharks are required to incorporate autonomous foraging to their daily routine. As it is difficult to directly observe young sharks foraging in the wild, biomarkers that indirectly estimate when young sharks shift from relying on maternal energy resources to feeding autonomously have recently been established^{20,21}. Although biomarkers, such as bulk stable isotopes, alone^{20,22} or in combination with fatty acids²¹, provide insights into autonomous foraging developments and broad estimates of body condition^{23,24}, isotopic turnover rates impede timely and precise estimates of body condition as well as foraging development. To date, Hussey *et al.*¹² executed the only study that simultaneously and precisely assessed changes in body condition and estimated foraging development in early life-stages of sharks¹² via a combination of lethal and non-lethal measurements to calculate such developments across different umbilical scar healing stages. Given that the study only focused on a single location that precludes investigations on variabilities among populations, further work is needed to understand if and how body condition and the development of autonomous foraging may vary across species inhabiting different habitats.

To examine intraspecific variability in body condition and foraging development during the first weeks of life, we collected life-history data from neonatal and juvenile blacktip reef sharks (*Carcharhinus melanopterus*), a species with high levels of genetic population structure^{25,26}, from two remote habitats in the Indo-Pacific Ocean. While Moorea (French Polynesia) is a remote island with human-impacted shorelines in the South Pacific, St. Joseph atoll (Seychelles), located in the western Indian Ocean, consists of a small and uninhabited ring of islands with adjacent shallow reef flats. Our objectives were to use non-lethal methods to determine (1) whether maternal energy investment varies between *C. melanopterus* populations potentially adopting different life-history strategies, and (2) if and to what extent body condition and foraging development might be affected by presumably variable maternal investments. We hypothesized that better conditioned neonates (e.g., neonates with higher energy stores) would show a slower decrease in body condition and a faster acquisition of foraging skills during the first weeks of life. Considering the steady increase in human activities in nearshore areas that is resulting in declining prey abundance and habitats^{16,27,28} as well as predicted higher water temperatures due to climate change^{29,30}, a better understanding of maternal energy investments, body condition, and autonomous foraging development during early life-stages of sharks has important implications for conservation³¹. Insights into potential intraspecific differences in such characteristics may further support site-specific management strategies for sharks from remote and potentially prey-limited habitats^{32,33}.

Results

Intraspecific variation in maternal energy investments. In Moorea, during the parturition seasons in 2016/2017 and 2017/2018, a total of 313 neonatal and juvenile *C. melanopterus* were captured and measured. Of those, 163 individuals (52%) were categorized as neonates (based on the presence of open or semi-healed umbilical scars) ranging from 368 to 466 mm L_{PC} (418.42 ± 18.90 mm, Fig. 1a) and weighting 670 to 1500 g (1025.22 ± 148.75 g, Fig. 1b). At St. Joseph, during the parturition seasons in 2014/2015, 2015/2016 and 2016/2017, a total of 233 neonatal and juvenile *C. melanopterus* were collected. Of those, 173 individuals (74%) were categorized as neonates ranging from 287 to 459 mm L_{PC} (372.22 ± 27.66 mm, Fig. 1a) and weighting 300 to 1375 g (694.99 ± 182.71 g, Fig. 1b). Neonatal *C. melanopterus* from Moorea were significantly larger (two sample *t*-test: $t = 17.769$, $df = 334$, $p < 0.0001$) and heavier at birth (two sample *t*-test: $t = 17.917$, $df = 325$, $p < 0.0001$) than individuals from St. Joseph. Mean water temperatures during the pupping seasons were significantly lower in Moorea (29.5 °C ± 0.003) compared to St. Joseph (30.0 °C ± 0.003; two sample *t*-test: $t = -101.87$, $df = 1040400$, $p < 0.0001$; see Supplementary Information S1).

Body condition, as calculated via three methods, differed significantly across locations. Neonatal sharks from Moorea were heavier for any given size than those at St. Joseph ($F_{1,234} = 20.89$, $p < 0.001$), and length-body mass results suggest positive allometric growth (Fig. 2). Independent indices of body condition were significantly higher in Moorea sharks compared to St. Joseph sharks, as calculated by Fulton's K (two sample *t*-test:

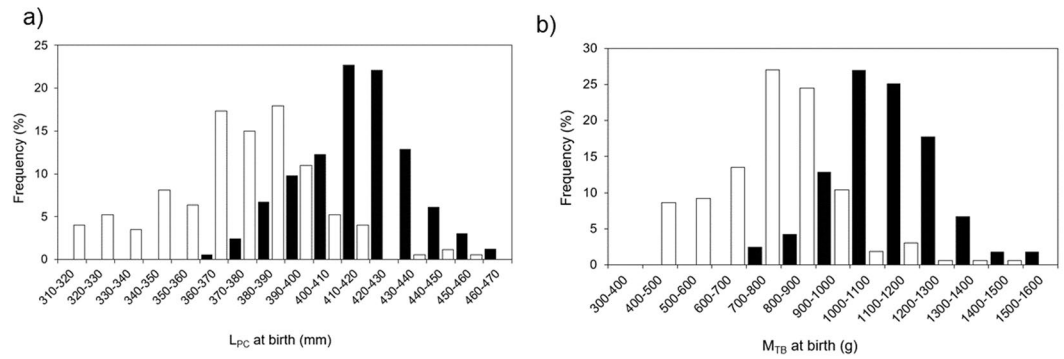


Figure 1. Percentage frequency histogram of (a) precaudal length (L_{PC}) and (b) total body mass (M_{TB}) in neonatal *Carcharhinus melanopterus* (USS1 and USS2) from Moorea (black, $n = 163$) and St. Joseph (white, $n = 173, 164$ respectively).

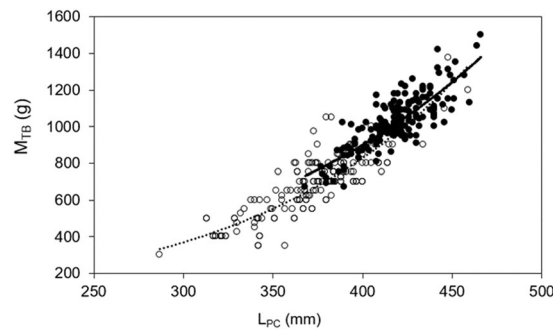


Figure 2. Relationship between total body mass (M_{TB}) and precaudal length (L_{PC}) of neonatal *Carcharhinus melanopterus* (USS1 and USS2) from Moorea (black, $y = 0.042472x^{2.70}$, $r^2 = 0.69$, $n = 163$) and St. Joseph ($y = 0.013947x^{2.98}$, $r^2 = 0.73$, $n = 164$).

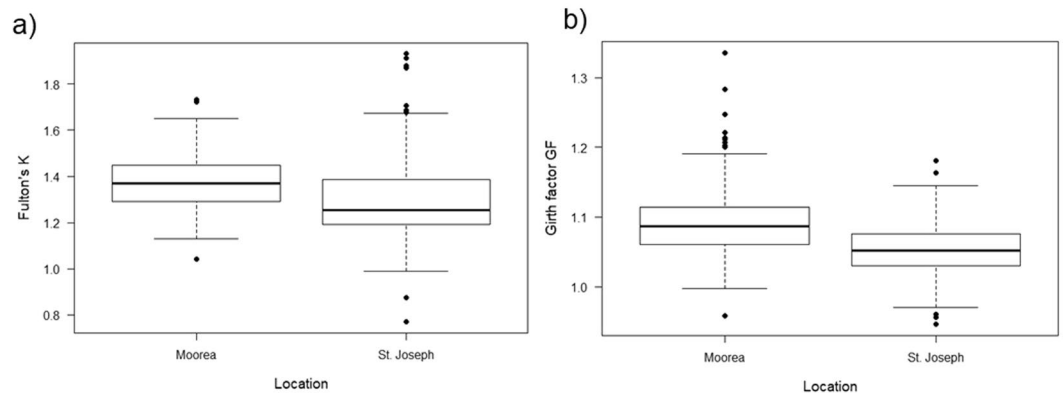


Figure 3. Comparison of body condition indices across locations. (a) Fulton's K for neonatal and juvenile *Carcharhinus melanopterus* from Moorea ($n = 313$) and St. Joseph ($n = 224$). (b) Girth factor GF for neonatal and juvenile *Carcharhinus melanopterus* from Moorea ($n = 313$) and St. Joseph ($n = 91$). Boxes indicate the interquartile range with the median shown by horizontal lines, minimum and maximum values shown by whiskers, and points representing outliers.

$t = 6.083$, $df = 535$, $p < 0.0001$; Fig. 3a) and GF (two sample t -test: $t = 7.036$, $df = 402$, $p < 0.0001$; Fig. 3b). Linear regressions revealed no relationships between L_{PC} and Fulton's K (St. Joseph: $F_{1,222} = 2.247$, $p = 0.135$), and L_{PC} and GF (Moorea: $F_{1,311} = 1.035$, $p = 0.310$; St. Joseph: $F_{1,89} = 3.397$, $p = 0.069$; see Supplementary Information S2). However, linear regressions revealed significant negative relationships between L_{PC} and Fulton's K in Moorea sharks ($F_{1,311} = 11.280$, $r^2 = 0.04$, $p = 0.0009$; see Supplementary Information S2).

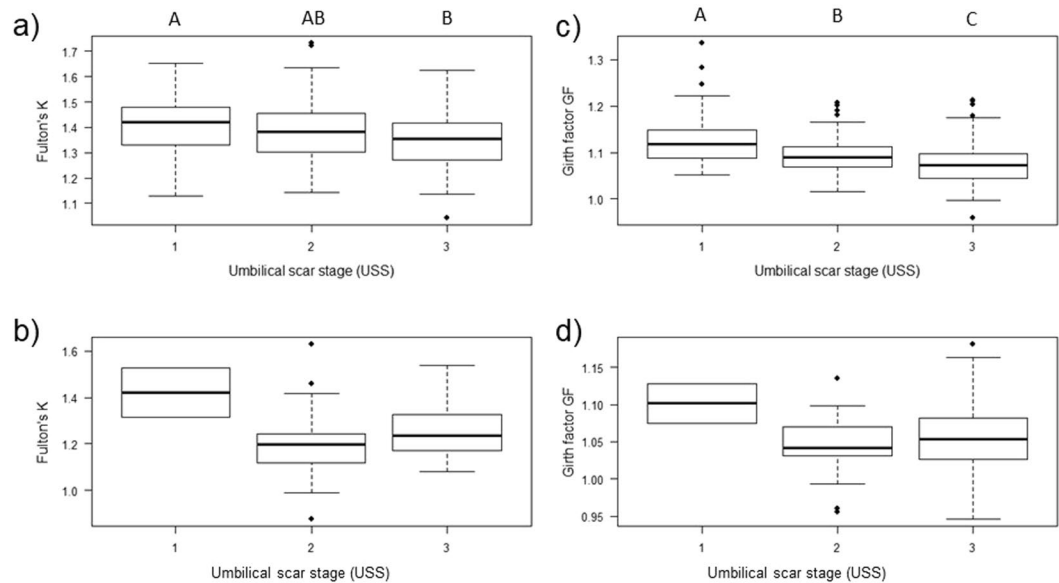


Figure 4. Transition of body condition indices with increasing umbilical scar stages (USS) in *Carcharhinus melanopterus*. Fulton's K at (a) Moorea and (b) St. Joseph, and girth factor GF at (c) Moorea, and (d) St. Joseph. Boxes indicate the interquartile range with the median shown by horizontal lines, minimum and maximum values shown by whiskers, and black dots represent outliers. Letters above plots in (a) and (c) indicate statistically significant differences between groups. Sample size in Moorea: USS1 $n = 59$, USS2 $n = 104$, USS3 $n = 150$. Sample size at St. Joseph: USS1 $n = 2$, USS2 $n = 29$, USS3 $n = 60$.

Intraspecific variation in change of body condition. In cases where body condition indices did not conform to a normal distribution (Shapiro-Wilks test, $p < 0.05$), non-parametric one-way and multiple comparison tests were applied. Body condition, as estimated by Fulton's K in young sharks from Moorea, differed significantly among increasing umbilical scars stages (ANOVA, $F_{2,310} = 6.907$, $p = 0.001$). Specifically, pair-wise comparisons showed statistical differences between USS1 and USS3 (Tukey's HSD, $t = -0.06$, $p = 0.001$) and decreasing, albeit non-significant, body condition between USS1 and USS2 (Tukey's HSD, $t = -0.03$, $p = 0.179$) and between USS2 and USS3 (Tukey's HSD, $t = -0.03$, $p = 0.097$, Fig. 4a). Body condition, as estimated by girth factor GF in young sharks from Moorea, decreased significantly as umbilical scar stages increased (Kruskal-Wallis test, $\chi^2 = 48.513$, $df = 2$, $p < 0.001$). Pair-wise comparisons reported significant differences between all three umbilical scar stage classes (Dunn test, USS1/USS2: $p = 0.0006$; USS1/USS3: $p < 0.0001$; USS2/USS3: $p = 0.0003$, Fig. 4c). On the contrary, no significant differences were found between Fulton's K and GF with increasing umbilical scar healing stages in young sharks from St. Joseph (Kruskal-Wallis test, $\chi^2 = 8.6627$, $df = 2$, $p = 0.056$ and $\chi^2 = 2.8051$, $df = 2$, $p = 0.246$, respectively; Fig. 4b,d).

The 45 individuals that were recaptured in Moorea during one parturition season were at liberty from 4 to 72 days (see Supplementary Information S3), and linear regressions showed significant negative relationships between changes in Fulton's K and time at liberty ($F_{1,43} = 5.41$, $p = 0.025$, $r^2 = 0.11$, Fig. 5a). Linear regression further revealed decreasing, albeit non-significant, relationships between GF and time at liberty ($F_{1,45} = 2.75$, $p = 0.104$; Fig. 5b). Similarly, linear regression indicated significant negative relationships between change in body condition with body condition at initial capture (Fulton's K; $F_{1,43} = 28.46$, $r^2 = 0.40$, $p < 0.0001$; Fig. 6a; GF: $F_{1,43} = 31.71$, $r^2 = 0.42$, $p < 0.0001$, Fig. 6b). When differences in body condition indices were regressed against one another, data showed that changes in Fulton's K could be predicted by changes in GF ($F_{1,43} = 16.83$, $r^2 = 0.28$, $p = 0.0002$; see Supplementary Information S4), suggesting that estimates of either condition index were consistent within individuals.

Intraspecific variation in foraging success. In Moorea, over the scope of one parturition season (year 2016/2017), 165 gastric lavages in *C. melanopterus* resulted in 78 full (47%), and 87 empty (53%) stomachs. At St. Joseph, over the scope of two parturition seasons (years 2015/2016 and 2016/2017), 109 gastric lavages in young *C. melanopterus* provided 93 full (85%) and 16 empty (15%) stomachs, leading to a significant bias of stomach fullness with locations ($X_2 = 36.60$, $p < 0.0001$). When separated by USS, the frequency of stomachs containing prey items (increased foraging success) increased from 30% (USS1; $n = 17$) and 47% (USS2; $n = 51$) to 51% by USS3 in Moorea ($n = 97$; Fig. 7). At St. Joseph at USS2, 100% of sampled stomachs had prey items in them ($n = 8$), and 84% of 101 individuals at USS3 had stomachs containing prey (Fig. 7). The smallest acrylic tubes (2.5 cm outer diameter) were still too large to be used with the smallest individuals from St. Joseph, resulting in a lack of sampled USS1 individuals.

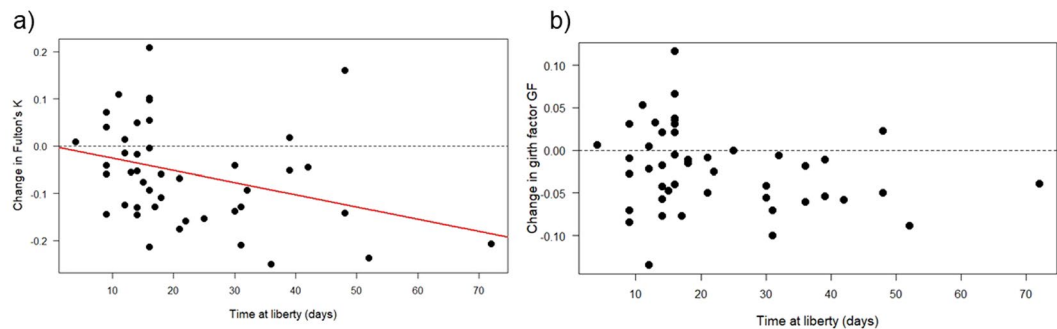


Figure 5. Changes in body condition indices with time at liberty in *Carcharhinus melanopterus* from Moorea. (a) Change in Fulton's K over time at liberty, and (b) change in girth factor GF over time at liberty. Data were obtained from neonatal sharks that were measured twice within the same parturition season (min. 4 days, max. 72 days, $n = 45$). The regression line for predicting changes in Fulton's K from time at liberty is shown in red ($y = 0.001 - 0.003x$, $r^2 = 0.11$). Note that each dot represents the change of body condition in one individual and that negative values (below the dashed line) depict a decrease of body condition in an individual shark.

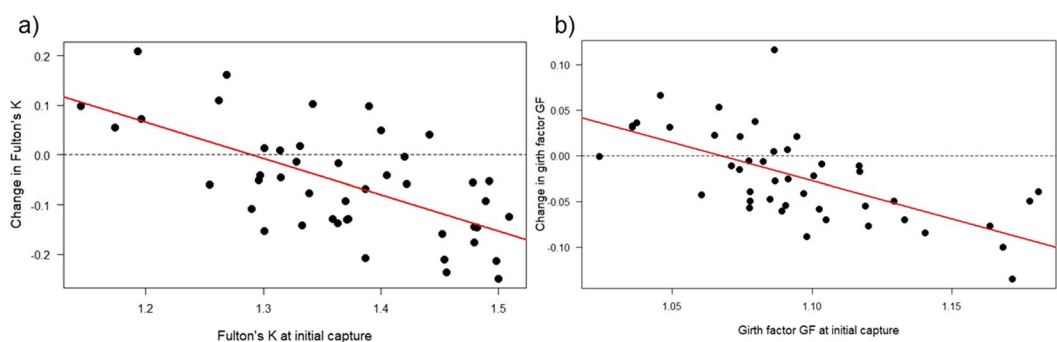


Figure 6. Changes in body condition indices versus body condition indices at initial capture in neonatal *Carcharhinus melanopterus* from Moorea. (a) Change in Fulton's K versus Fulton's K at initial capture, and (b) change in girth factor GF versus girth factor GF at initial capture. Data were obtained from sharks that were measured twice within the same parturition season (min. 4 days, max. 72 days, $n = 45$). The regression lines are shown in red (K: $y = 0.94 - 0.73x$, $r^2 = 0.40$ and GF: $y = 0.90 - 0.84x$, $r^2 = 0.42$, respectively). Note that each dot represents the change of body condition in one individual and that negative values (below the dashed line) depict a decrease of body condition in an individual shark.

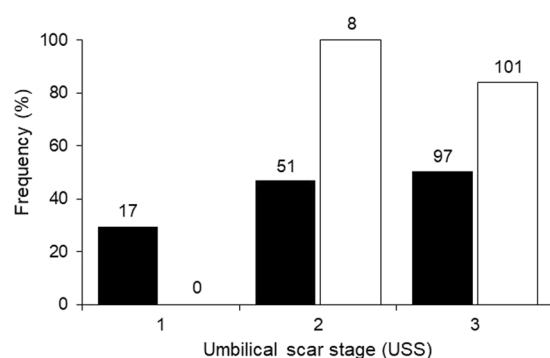


Figure 7. Frequency histogram of percentage stomachs containing prey with increasing umbilical scar stage (USS) in *Carcharhinus melanopterus* from Moorea (black, $n = 165$) and St. Joseph (white, $n = 109$). Numbers above each column represent the total sample size of sharks for a given umbilical scar stage (USS) on which gastric lavages were performed.

Discussion

This study represents the first non-lethal investigation of body condition and concurrent estimates of autonomous foraging development in young *C. melanopterus* from two isolated shark populations. Our data provide compelling evidence that maternal investment is site-specific, with significantly larger sizes, greater body masses,

and larger body condition measurements in Moorea sharks when compared to St. Joseph sharks. Furthermore, our data suggest that, despite this better head start, young sharks in Moorea exhibited significant decreases in body condition and developed foraging habits slower than sharks from St. Joseph (e.g., fewer than half of the stomachs lavaged from sharks in Moorea had contents during later development stages). These differences in foraging success likely explain the significant decrease in body condition in Moorea sharks, while sharks at St. Joseph maintained their body condition. Likewise, data from recaptured individuals from Moorea confirm the significant decrease in body condition with increasing time at liberty (up to 72 days). Recaptured individuals that initially had higher body condition indices were most likely to exhibit declines in body condition during the first weeks/months of life.

The fact that we observed larger and heavier neonates with greater mass per unit length and higher condition indices in Moorea versus St. Joseph suggests that neonates in Moorea are being well provisioned by larger, better conditioned mothers with potentially lower fecundity. Indeed, adult *C. melanopterus* from Moorea tend to be larger³⁴ than adult *C. melanopterus* from St. Joseph³⁵ and are therefore likely to produce larger, heavier and better conditioned young¹². Body size is strongly heritable, and it's also common for geographically separated shark populations of the same species to be genetically and morphologically different^{9,10,34}. Body size, or at least body condition, can further be influenced by a species' diet. Data on natural prey abundance were not collected in either of the two study locations, but provisioning sites in Moorea are numerous, and adult *C. melanopterus* frequent such sites³⁶. While direct impacts of provisioning on body condition is either sparsely documented^{37,38} or show minimal impacts on the sharks' diet³⁹, provisioned female *C. melanopterus* may benefit from high-trophic level food, which, in turn, is likely to augment maternal investments (e.g., more endogenous energy resources) for their offspring. Further, the exclusive economic zone (EEZ) of French Polynesia banned fishing for *C. melanopterus* in 2006⁴⁰. This fishing ban may have helped protect larger and better conditioned females, which in turn give birth to larger, heavier, and better conditioned offspring¹². Lastly, it could be argued that differences in fecundity influence variable pup sizes in Moorea and St. Joseph. While the estimated average litter size of sharks from Moorea and Aldabra (~900 km southwest of St. Joseph) is three pups, Moorea's sharks demonstrate annual reproductive cycles as opposed to biennial cycles in Seychelles^{34,41,42}. This suggests that, considering the very limited data, fecundity does not explain our findings, because more frequent cycles in Moorea would likely infer smaller pups. While any of these mechanisms alone or in combination could explain the intraspecific variation in the level of maternal investment of female *C. melanopterus*, identifying the specific factors that result in female sharks in Moorea being larger and giving birth to larger, heavier, and better conditioned offspring was beyond the scope of this study.

The rate at which body condition and autonomous foraging success changed as umbilical scars began disappearing varied between Moorea and St. Joseph sharks, suggesting early development may be site-specific for young *C. melanopterus*. Although the rapid decrease of body condition in *C. melanopterus* from Moorea is not surprising, considering documented declines in body conditions in other young sharks^{12,16,17,22}, the relatively high maternal investment in Moorea was expected to lead to slower declines in body condition (e.g., due to more energy reserves at birth) and faster foraging development and success compared to sharks from St. Joseph. Our study, however, demonstrates significant declines in body condition and slower foraging development in sharks from Moorea, therefore suggesting that the quality of the maternal energy investment is not correlated with the foraging success of the young. Other factors, such as environmental conditions, prey resources, variable foraging strategies, and/or anthropogenic stressors are all likely, in some part, to be responsible for the observed differences across sites.

Environmental conditions, such as seawater temperatures, were measured in Moorea and St. Joseph. Despite significantly lower mean temperatures during pupping seasons in Moorea ($29.5\text{ }^{\circ}\text{C} \pm 0.003$) compared to St. Joseph ($30.0\text{ }^{\circ}\text{C} \pm 0.003$), temperature ranges were highly comparable (see Supplementary Information S4). These small differences in mean temperatures lead to standard metabolic rates (SMR; the cost of maintenance metabolism) of 160.5 and $162.7\text{ mg O}_2\text{ kg}^{-1}\text{ h}^{-1}$, respectively (Bouyoucos, IA, unpublished data). A difference in SMR of 1.4% is, however, negligible in maintenance costs and is therefore likely not responsible for the observed site-specific differences in changes of body condition. However, if ocean temperatures continue to increase, a decrease in body condition during early life-stages may be more pronounced, because higher water temperatures can have decelerating effects on growth⁴³.

Variable rates of decreasing body condition and foraging development in young sharks may have also been shaped by different levels of inter- and intraspecific competition in young sharks for limited prey resources. Recent studies categorize nearshore areas as resource-limited, a condition that may especially be distinctive in remote areas, where multiple juvenile shark species co-occur and compete for similar prey^{44–46}. Both Moorea and St. Joseph are inhabited by multiple populations of young sharks^{13,20}, therefore, competition is likely to occur at both locations¹⁸. Indeed, co-occurrence and potential competition in Moorea lead to isotopic niche partitioning between juvenile *C. melanopterus* and sicklefin lemon sharks (*Negaprion acutidens*); yet, body condition as well as growth rates were not affected by the coexisting species⁴⁵. Even if prey abundances were not quantitatively assessed in any of the two study sites, small reef-associated teleosts (e.g., the predominant prey of young sharks^{41,47}) are often observed in St. Joseph at site of collection (Weideli, OC, personal observation) and in 85% of stomachs investigated. These observations suggest that prey availability at St. Joseph is sufficient, resulting in potentially weak competitive interactions between young sharks. At Moorea, during gillnet deployments ($n = 175$) for this study, potential prey species were rarely observed; although this does not prove their absence. Future studies assessing competitive patterns among coexisting shark species and prey availability are, however, needed to draw further conclusions as to why body condition and foraging development during the first weeks of life change at different rates in Moorea and St. Joseph.

In addition to prey availability, the caloric value of ingested prey as well as foraging strategies may differ between sites. Juvenile scalloped hammerheads (*Sphyrna lewini*) have been reported consuming energetically poor prey^{16,48}, which may explain the observed decreases in body mass after parturition¹⁶. The liver lipids that

young sharks in Moorea receive as a maternal headstart are potentially higher in energy compared to their ingested prey. This caloric difference may help explain the body condition decrease as their umbilical scars begin to disappear, similar to the loss of maternal isotopic signals observed in young bull sharks (*Carcharhinus leucas*) and Atlantic sharpnose sharks (*Rhizoprionodon terraenovae*)²². On the contrary, small or negligible differences in energetic value of maternal energy resources compared to young sharks' prey may explain the maintained body condition observed in St. Joseph sharks. Low caloric prey may also help to explain how an increase in foraging success from 30% and 47% to 51% of stomachs containing prey (Fig. 7) can result in decreasing body condition in Moorea sharks. Similar findings have been reported by Hussey *et al.*¹², where body condition of neonatal dusky sharks (*Carcharhinus obscurus*) decreased despite increasing stomach content mass (increasing feeding activities). Nonetheless, this is highly speculative, and more stomach items, especially those from extremely young sharks (e.g., USS1 fresh umbilical scars), as well as the actual caloric value of the stomach contents are needed to better understand the relationship between decreasing body condition despite increasing foraging success.

Prey resources and their caloric value may deteriorate in nearshore areas with substantial anthropogenic impacts^{28,49}. Indeed, the abundance of small reef-associated teleosts is declining through large-scale habitat degradation^{27,50}, and artisanal fishing⁵¹. Likewise, anthropogenic habitat degradation underpins the declines in the abundance of energetically high-value prey species (e.g., small scarids) with a concurrent increase of low caloric gobies and shrimps in the shallow areas of Kāne' ohe Bay, Hawai'i (USA)⁵². This transition to lower caloric-value prey is thought to be partially responsible for the declining body mass in *S. lewini* during their first weeks of life¹⁶. Anthropogenic stressors, however, can also have direct impacts on young sharks. Increasing temperatures and salinity, for example, allowed young *C. leucas* to expand into formerly uninhabited bays⁵³ with potentially different prey resources and also into areas where artisanal nearshore fisheries frequently capture young sharks^{54,55}. Even if young sharks are not the target species in artisanal fisheries and are subsequently released, accidental capture events cause stress⁵⁶. Young *C. melanopterus*, for example, require at least 8 h recovery after a single accidental gillnet capture event; during this time, about 15% of the energy used for daily swimming is lost⁵⁶. Despite enforcement of partially protected areas (no-take zones) around Moorea⁵⁷, artisanal fishing is far more likely to occur within the coastal areas of Moorea when compared to near-pristine and uninhabited St. Joseph, with its uninterrupted reserve boundary^{35,40}. Similarly, human activities at Moorea (e.g., boat traffic, boat channel dredging, and shoreline activities) may constrain young shark habitats, with sharks potentially avoiding deeper channels or areas with boat traffic.

The observed relationship between decreasing body condition with increasing USS in Moorea sharks is further supported by data from individual sharks that were captured on multiple occasions. This is, to our best knowledge, the first evidence of a significant decrease of body condition with time at liberty in individual wild sharks (Fig. 5). Results from such recaptures also depict that individuals with higher body condition indices (K as well as GF) at initial capture had more pronounced decreases in body condition during the first weeks of life (Fig. 6). This is analogous to the findings across habitats, in which sharks from Moorea with higher maternal investments were subject to significant decreases in body condition (Fig. 4a,c) compared to sharks from St. Joseph, where such a decline was absent (Fig. 4b,d). Since all recaptured individuals at Moorea were exposed to similar environmental conditions (e.g., prey availability, prey quality, and anthropogenic stressors), other factors must have contributed to the within-population differences around Moorea. One plausible answer could be that sharks with higher initial body condition are less driven to start foraging because they can rely on ample endogenous energy resources for an extended period of time. On the contrary, individuals with lower initial body condition are forced to develop foraging skills at an earlier age, hence demonstrating a positive change in body condition between capture events. This is speculative, because unexperienced young sharks are generally considered as asynchronous opportunistic foragers⁵⁸, and dietary information were not collected from recaptured sharks. Also, body condition is only a proxy that may mask other behavioural or physiological traits that may have influenced our findings. Future work should therefore aim to collect dietary information (e.g., stomach contents or isotopic information) from recaptured sharks to validate changes in body condition between multiple capture events. Finally, prospective studies are recommended to include long-term recaptures to elucidate whether the body condition changes that are observed during early-life stages influence later development stages or if these early body condition changes are negligible for older age-classes.

In conclusion, our findings suggest and support that decreases in body condition within the first weeks of life are common for young viviparous sharks and not only result from natural depletions of maternal energy resources, but will also in some part be affected by prey availability, prey quality, foraging strategies, and/or anthropogenic stressors^{12,16}. Our approach, using two populations of *C. melanopterus*, further enabled us to discriminate between different maternal investments in which young sharks from Moorea with higher maternal energy resources were found to demonstrate significant decreases in body condition and slower foraging development compared to sharks from St. Joseph. A comparable observation was provided within the Moorea population in which better-conditioned individuals were subject to a higher loss of body condition. It is therefore expected that young sharks with relatively lower body condition are forced to develop foraging skills at an earlier life-stage, resulting in higher proportions of stomachs containing prey and a positive change in body condition between recaptures. This finding suggests that the habitat quality (e.g., prey abundance and quality) might be especially important for sharks with limited maternal energy resources, and generally for sharks that occur in isolated, nearshore habitats, where deeper surrounding waters or anthropogenically-induced channels impede or prevent dispersal to nearby, potentially prey-rich habitats.

The continued global expansion of human activities (e.g., overfishing, climate change, coastal development, and pollution) poses the greatest risk to reef-associated, shallow water shark species⁵⁹. Therefore generating site-specific information on early development of reef sharks is critical⁶⁰. During these early life-stages, young sharks not only depend on the maternal energy resources, but also rely on these nearshore areas for shelter and/or to access adequate prey resources. Therefore, to achieve sound conservation measures for *C. melanopterus* and

other viviparous reef sharks, management strategies need to come together to effectively protect breeding populations as well as young sharks and their shallow nearshore habitats.

Methods

Study location and sampling. Some of the sharks for this study were captured as part of long-term fisheries-independent surveys in Moorea, French Polynesia (17°30'S, 149°51'W). Moorea is surrounded by fringing reefs and lagoons that are adjacent to shallow nearshore areas serving as putative nursery grounds for young *C. melanopterus*^{34,45}. Juvenile *C. melanopterus* were captured using gillnets (50.0 m × 1.5 m, 5.0 cm mesh) during the parturition months (September – February) in 2016/2017 and 2017/2018. Captured individuals were immediately removed from the net, and handling time was kept to a minimum (<7 min.) to avoid excessive capture-related stress⁵⁶. Sharks captured in 2016/2017 were externally tagged using coloured T-bar anchor tags (Hallprint[®], Hindmarsh Valley, SA, Australia) and internally with passive integrated transponder (PIT) tags (Biolog-ID) in 2017/2018 to allow recaptured animals to be identified. During these sampling events, pre-caudal length (L_{PC} , the length from the tip of the snout to the precaudal notch) and three girth measurements were measured to the nearest 0.1 cm with a tape measure for each shark: 1) pectoral girth (G_{PEC}), the circumference of the shark measured at the base of the pectoral fin insertion, anterior to the dorsal fin, 2) dorsal girth (G_{DOR}), the circumference measured at the base of the first dorsal fin insertion, and 3) caudal girth (G_{CAU}), measured anterior to the caudal fin in the precaudal notch (see Supplementary Information S5). Umbilical scar stage (USS), a reliable indicator of neonatal life-stages^{12,16,61}, was quantified into three categories. USS1 was applied if scar was fully open, USS2, if scar was semi-healed, and USS3 for fully healed scars (see Supplementary Information S6). Individuals with USS1 and USS2 were considered as neonate sharks with an estimated maximal age of four weeks^{12,16,61}. Sharks with closed scars (USS3) were identified as young-of-the-year (>four weeks old) and no differentiation was made between visible and well-healed scars. We were unable to differentiate between young-of-the-year and older sharks, due to systematic size overlap between different age-classes¹¹. The USS of each shark was photographed alongside a ruler, and total body mass (M_{TB}) was measured with a hand-held scale to the nearest 10 g. After completing basic measurements, a subset of *C. melanopterus* individuals also had their stomachs flushed, similar to Bangle *et al.*⁶². Different diameters of transparent acrylic tubes (2.5, 3.2, and 3.8 cm outer tube diameter) were used according to the shark sizes (<60 cm, between 60–70 cm and >70 cm L_P , respectively). The beveled and lubricated tubes were inserted through the mouth, esophagus, and into the stomach while the sharks were kept in the water. As soon as the stomach and the tube were filled with water, the shark was turned upside down to flush the stomach. The stomach items were captured in a sieve, and the percentage of stomachs containing prey was recorded. This procedure was solely conducted on sharks in good condition (e.g., no open wounds) and was kept to a maximum of three consecutive procedures per individual. Environmental temperatures were recorded every ten minutes during parturition season with stationary Hobo[®] temperature loggers (UA-002-64, Onset Computer Corporation, Bourne, MA, USA) deployed in capture locations.

Fieldwork was further conducted in the western Indian Ocean at St. Joseph atoll (05°26'S, 53°20'E) in the Republic of Seychelles. St. Joseph is a near-pristine and non-inhabited atoll that offers shallow, protected areas for at least two species of young sharks¹³. Juvenile *C. melanopterus* were captured with gillnets (20.0 m × 1.5 m, 5.0 cm mesh) during the parturition months (October – December and March – April) in 2014/2015, 2015/2016 and 2016/2017. Captured *C. melanopterus* were immediately removed from the net, and handling time was kept to a minimum (<7 min.) to avoid excessive capture-related stress⁵⁶. Sharks were internally tagged using PIT tags (Biomark[®]) to allow recaptured sharks to be identified. The L_{PC} , girth, USS, and M_{TB} were measured for each shark, and gastric lavage was subsequently conducted using a sub-sample of sharks following Moorea's protocol. All sharks were released at site within minutes of capture. Temperatures were recorded every fifteen minutes during parturition season with stationary Hobo[®] temperature loggers (U22-001, Onset Computer Corporation, Bourne, MA, USA) distributed across the area surveyed.

Data analyses. Where applicable, data were checked for normality using Shapiro-Wilk tests prior to analyses in R version 3.5.3⁶³ within the RStudio interface ver. 1.0.153⁶⁴. For all tests, the level of statistical significance α was set at 0.05, and results are reported as means \pm SD. To investigate potential intraspecific life-history variabilities in neonates and temperature differences across habitats, mean L_{PC} , M_{TB} and water temperatures were compared with two sample *t*-tests, and frequency histograms were subsequently constructed. Total body mass for a given L_{PC} was used to estimate body condition, assuming that individuals in a good condition would be heavier than those in poorer condition of the same length. Thus, we determined allometric length–mass relationships by using the formula $\log y = \log a + b \log x$. These coefficients were used in $M_{TB} = a L_{PC}^b$, where M_{TB} is total body mass (g) and L_{PC} is length (cm).

Two independent indices of individual body condition were also calculated. The Fulton's body condition index, also known as Fulton's K ⁶⁵, calculates a morphometric index of a fish's body condition with the following equation:

$$K = 10^5 M_{TB} (L_{PC})^{-3} \quad (1)$$

We also constructed a non-lethal and morphometric condition index, based on the assumption that individuals with larger livers for a given body length are in better condition¹². Similar to Irschick & Hammerschlag⁶⁶, three measurements along the shark's body were chosen to incorporate the size and anatomical location of the liver, as well as the shark's shape, which is wider along the anterior part of the body⁶⁷. While massive body sizes prevent measuring the circumference in previous studies^{66,67}, we were able to take three circumference measurements to calculate the girth factor (GF) as a proxy for body condition using the following equation:

$$GF = [G_{PEC} + G_{DOR} + G_{CAU}] L_{PC}^{-1} \quad (2)$$

Resulting condition indices (K and GF) were compared across locations using a two sample *t*-tests. To demonstrate and validate the absence of inadvertent co-linearity between the two body condition indices and $L_{PC}^{2,3}$, linear least-squares regressions were performed for L_{PC} with K and GF, respectively.

In order to follow the transition of body condition with the closure of the umbilicus (increasing USS), analysis of variance (ANOVA) and Tukey's honest significant difference (HSD) tests were used for post-hoc multiple comparisons for Fulton's K at Moorea. As the other comparisons did not conform to a normal distribution, Kruskal-Wallis tests were applied. Post-hoc multiple comparisons were subsequently evaluated using Dunn tests, while p-values were adjusted using the Holm method to reduce type I error. In addition, individual sharks from Moorea that were captured on multiple occasions were used to further validate body condition changes during early life stages. Recaptured individuals allowed us to calculate the change in body condition between two capture events by subtracting body condition indices (K and GF) of the initial capture from the recapture event. Similarly, recaptured individuals allowed us to estimate if changes in body condition depend on the body condition at initial capture. For both calculations, values were plotted for each individual in a linear least-square regression. Furthermore, Fulton's K was linearly regressed against girth factor GF of recaptured *C. melanopterus* to demonstrate that changes in K could be predicted by changes in GF. Finally, in order to estimate level of autonomous foraging success during increasing USS, a sub-sample of *C. melanopterus* from Moorea and St. Joseph had gastric lavages performed. The obtained stomach status (% of stomachs containing prey) were compared with χ^2 test.

Ethical approval. Sharks for this study were captured as part of long-term fisheries-independent surveys in Moorea, French Polynesia and on St. Joseph, Republic of Seychelles. Ethical approval for Moorea was given by James Cook University Animal Ethics Committee protocol A2089 and permission to work with sharks in French Polynesia was obtained from the Ministère de l'Environnement (Arrete N° 9524). Research on sharks at St. Joseph was approved by, and conducted with the knowledge of Ministry of Environment, Energy, and Climate Change, Seychelles. Animal handling and tagging methods were conducted in accordance with the approved guidelines of S. Planes by the Autorisation de pratiquer des expériences sur les animaux n° 006725 (1995) from the ministry of Agriculture.

References

1. Stearns, S. C. *The Evolution of Life Histories*. (Oxford University Press, 1992).
2. Jakob, E. M., Marshall, S. D. & Uetz, G. W. Estimating Fitness: A Comparison of Body Condition Indices. *Oikos* **77**, 61–67 (1996).
3. Green, A. J. Mass/length residuals: Measures of body condition or generators of spurious results? *Ecology* **82**, 1473–1483 (2001).
4. Bernardo, J. Maternal Effects in Animal Ecology. *Am. Zool.* **36**, 83–105 (1996).
5. Donelson, J. M., McCormick, M. I. & Munday, P. L. Parental condition affects early life-history of a coral reef fish. *J. Exp. Mar. Bio. Ecol.* **360**, 109–116 (2008).
6. Gagliano, M. & McCormick, M. I. Maternal condition influences phenotypic selection on offspring. *J. Anim. Ecol.* **76**, 174–182 (2007).
7. Wallace, B. P. *et al.* Maternal investment in reproduction and its consequences in leatherback turtles. *Oecologia* **152** (2007).
8. Berkeley, S. A., Chapman, C., Sogard, S. M., Ecology, S. & May, N. Maternal age as a determinant of larval growth and survival in a marine fish, *Sebastes melanops*. *Ecology* **85**, 1258–1264 (2004).
9. Lombardi-Carlson, L. A., Cortés, E., Parsons, G. R. & Manire, C. A. Latitudinal variation in life-history traits of bonnethead sharks, *Sphyrna tiburo*, (Carcharhiniformes: Sphyrnidae) from the eastern Gulf of Mexico. *Mar. Freshw. Res.* **54**, 875–883 (2003).
10. Bradley, D. *et al.* Growth and life history variability of the grey reef shark (*Carcharhinus amblyrhynchos*) across its range. *PLoS One* **12** (2017).
11. Barker, M. J., Gruber, S. H., Newman, S. P. & Schluessel, V. Spatial and ontogenetic variation in growth of nursery-bound juvenile lemon sharks, *Negaprion brevirostris*: A comparison of two age-assigning techniques. *Environ. Biol. Fishes* **72**, 343–355 (2005).
12. Hussey, N. E. *et al.* Maternal investment and size-specific reproductive output in carcharhinid sharks. *J. Anim. Ecol.* **79**, 184–93 (2010).
13. Weideli, O. C., Papastamatiou, Y. P. & Planes, S. Size frequency, dispersal distances and variable growth rates of young sharks in a multi-species aggregation. *J. Fish Biol.* 1–9, <https://doi.org/10.1111/jfb.13968> (2019).
14. DiBattista, J. D., Feldheim, K. A., Gruber, S. H. & Hendry, A. P. When bigger is not better: Selection against large size, high condition and fast growth in juvenile lemon sharks. *J. Evol. Biol.* **20**, 201–212 (2007).
15. Hussey, N. E. *et al.* Risky business for a juvenile marine predator? Testing the influence of foraging strategies on size and growth rate under natural conditions. *Proc. R. Soc. B Biol. Sci.* **284**, 20170166 (2017).
16. Duncan, K. M. & Holland, K. N. Habitat use, growth rates and dispersal patterns of juvenile scalloped hammerhead sharks *Sphyrna lewini* in a nursery habitat. *Mar. Ecol. Prog. Ser.* **312**, 211–221 (2006).
17. Corsso, J. T., Gadig, O. B. F., Barreto, R. R. P. & Motta, F. S. Condition analysis of the Brazilian sharpnose shark *Rhizoprionodon lalandii*: evidence of maternal investment for initial post-natal life. *J. Fish Biol.* **93**, 1038–1045 (2018).
18. Hoffmayer, E. R., Parsons, G. R. & Horton, J. Seasonal and interannual variation in the energetic condition of adult male Atlantic sharpnose shark *Rhizoprionodon terraenovae* in the northern Gulf of Mexico. *J. Fish Biol.* **68**, 645–653 (2006).
19. Oftedal, O. T. Lactation in whales and dolphins: Evidence of divergence between baleen- and toothed-species. *J. Mammary Gland Biol. Neoplasia* **2**, 205–230 (1997).
20. Matich, P., Kiszka, J. J., Heithaus, M. R., Mourier, J. & Planes, S. Short-term shifts of stable isotope ($\delta^{13}C$, $\delta^{15}N$) values in juvenile sharks within nursery areas suggest rapid shifts in energy pathways. *J. Exp. Mar. Bio. Ecol.* **465**, 83–91 (2015).
21. Belicka, L. L., Matich, P., Jaffé, R. & Heithaus, M. R. Fatty acids and stable isotopes as indicators of early-life feeding and potential maternal resource dependency in the bull shark *Carcharhinus leucas*. *Mar. Ecol. Prog. Ser.* **455**, 245–256 (2012).
22. Olin, J. A. *et al.* Maternal meddling in neonatal sharks: implications for interpreting stable isotopes in young animals. *Rapid Commun. Mass Spectrom.* **25**, 1008–16 (2011).
23. Papastamatiou, Y. P., Friedlander, A. M., Caselle, J. E. & Lowe, C. G. Long-term movement patterns and trophic ecology of blacktip reef sharks (*Carcharhinus melanopterus*) at Palmyra Atoll. *J. Exp. Mar. Bio. Ecol.* **386**, 94–102 (2010).
24. Matich, P., Kiszka, J. J., Heithaus, M. R. & Bourq, B. Le & Mourier, J. Inter-individual differences in ontogenetic trophic shifts among three marine predators. *Oecologia* **1**, 621–636 (2019).
25. Vignaud, T. M. *et al.* Blacktip reef sharks, *Carcharhinus melanopterus*, have high genetic structure and varying demographic histories in their Indo-Pacific range. *Mol. Ecol.* **23**, 5193–5207 (2014).

26. Delsler, P. M. *et al.* Population genomics of *C. melanopterus* using target gene capture data: demographic inferences and conservation. *Sci. Rep.* **6**, 33753 (2016).
27. Jennings, D. E., Gruber, S. H., Franks, B. R., Kessel, S. T. & Robertson, A. L. Effects of large-scale anthropogenic development on juvenile lemon shark (*Negaprion brevirostris*) populations of Bimini, Bahamas. *Environ. Biol. Fishes* **83**, 369–377 (2008).
28. Thiault, L., Collin, A., Chlous, F., Gelcich, S. & Claudet, J. Combining participatory and socioeconomic approaches to map fishing effort in small-scale fisheries. *PLoS One* **12**(5), e0176862 (2017).
29. Cheng, L., Abraham, J., Hausfather, Z. & Trenberth, K. E. How fast are the oceans warming? *Science* (80-). 128–129 (2019).
30. Levitus, S., Antonov, J. I., Boyer, T. P. & Stephens, C. Warming of the world ocean. *Science* (80-). **287**, 2225–2229 (2000).
31. Rosa, R., Rummer, J. L. & Munday, P. L. Biological responses of sharks to ocean acidification. *Biol. Lett.* **13**, 20160796 (2017).
32. Knip, D. M., Heupel, M. R. & Simpfendorfer, C. A. Evaluating marine protected areas for the conservation of tropical coastal sharks. *Biol. Conserv.* **148**, 200–209 (2012).
33. Kinney, M. J. & Simpfendorfer, C. A. Reassessing the value of nursery areas to shark conservation and management. *Conserv. Lett.* **2**, 53–60 (2009).
34. Mourier, J., Mills, S. C. & Planes, S. Population structure, spatial distribution and life-history traits of blacktip reef sharks *Carcharhinus melanopterus*. *J. Fish Biol.* **82**, 979–93 (2013).
35. Lea, J. S. E., Humphries, N. E., von Brandis, R. G., Clarke, C. R. & Sims, D. W. Acoustic telemetry and network analysis reveal the space use of multiple reef predators and enhance marine protected area design. *Proc. R. Soc. B Biol. Sci.* **283**, 20160717 (2016).
36. Kiszka, J., Mourier, J., Gastrich, K. & Heithaus, M. Using unmanned aerial vehicles (UAVs) to investigate shark and ray densities in a shallow coral lagoon. *Mar. Ecol. Prog. Ser.* **560**, 237–242 (2016).
37. Brena, P. F., Mourier, J., Planes, S. & Clua, E. Shark and ray provisioning: functional insights into behavioral, ecological and physiological responses across multiple scales. *Mar. Ecol. Prog. Ser.* 273–283, <https://doi.org/10.3354/meps11492> (2015).
38. Maljković, A. & Côté, I. M. Effects of tourism-related provisioning on the trophic signatures and movement patterns of an apex predator, the Caribbean reef shark. *Biol. Conserv.* **144**, 859–865 (2011).
39. Abrantes, K. G., Brunnschweiler, J. M. & Barnett, A. You are what you eat: Examining the effects of provisioning tourism on shark diets. *Biol. Conserv.* **224**, 300–308 (2018).
40. Ward-Paige, C. A. & Worm, B. Global evaluation of shark sanctuaries. *Glob. Environ. Chang.* **47**, 174–189 (2017).
41. Stevens, J. D. Life-history and Ecology of Sharks at Aldabra Atoll, Indian Ocean. *Proc. R. Soc. London.* **222**, 79–106 (1984).
42. Porcher, I. F. On the gestation period of the blackfin reef shark, *Carcharhinus melanopterus*, in waters off Moorea, French Polynesia. *Mar. Biol.* **146**, 1207–1211 (2005).
43. Rosa, R. *et al.* Early-life exposure to climate change impairs tropical shark survival. *Proc. R. Soc. B Biol. Sci.* **281** (2014).
44. Kinney, M., Hussey, N., Fisk, A., Tobin, A. & Simpfendorfer, C. Communal or competitive? Stable isotope analysis provides evidence of resource partitioning within a communal shark nursery. *Mar. Ecol. Prog. Ser.* **439**, 263–276 (2011).
45. Match, P., Kiszka, J. J., Mourier, J., Planes, S. & Heithaus, M. R. Species co-occurrence affects the trophic interactions of two juvenile reef shark species in tropical lagoon nurseries in Moorea (French Polynesia). *Mar. Environ. Res.* **127**, 84–91 (2017).
46. Bethea, D., Buckel, J. & Carlson, J. Foraging ecology of the early life stages of four sympatric shark species. *Mar. Ecol. Prog. Ser.* **268**, 245–264 (2004).
47. Newman, S., Handy, R. & Gruber, S. Diet and prey preference of juvenile lemon sharks *Negaprion brevirostris*. *Mar. Ecol. Prog. Ser.* **398**, 221–234 (2010).
48. Lowe, C. G. Bioenergetics of free-ranging juvenile scalloped hammerhead sharks (*Sphyrna lewini*) in Kaneohe Bay, O'ahu, HI. *J. Exp. Mar. Biol. Ecol.* **278**, 141–156 (2002).
49. Thiault, L. *et al.* Ecological evaluation of a marine protected area network: a progressive-change BACIPS approach. *Ecosphere* **10**, e02576 (2019).
50. Lotze, H. K. *et al.* Depletion, Degradation, and Recovery Potential of Estuaries and Coastal Seas. *Science* (80-). **312**, 1806–1809 (2006).
51. Blaber, S. J. M. *et al.* Effects of fishing on the structure and functioning of estuarine and nearshore ecosystems. *ICES J. Mar. Sci.* **57**, 590–602 (2000).
52. Bush, A. Diet and diel feeding periodicity of juvenile scalloped hammerhead sharks, *Sphyrna lewini*, in Kaneohe Bay, O'ahu, Hawai'i. *Environ. Biol. Fishes* 1–11 (2003).
53. Bangley, C. W., Paramore, L., Shiffman, D. S. & Rulifson, R. A. Increased Abundance and Nursery Habitat Use of the Bull Shark (*Carcharhinus leucas*) in Response to a Changing Environment in a Warm-Temperate Estuary. *Sci. Rep.* **8**, 6018 (2018).
54. Glaus, K. B. J. *et al.* Fishing for profit or food? Socio-economic drivers and fishers' attitudes towards sharks in Fiji. *Mar. Policy* 1–9, <https://doi.org/10.1016/j.marpol.2018.11.037> (2018).
55. Glaus, K. B. J., Adrian-Kalchauer, I., Burkhardt-Holm, P., White, W. T. & Brunnschweiler, J. M. Characteristics of the shark fisheries of Fiji. *Sci. Rep.* **5**, 17556 (2015).
56. Bouyoucos, I. A., Weideli, O. C., Planes, S., Simpfendorfer, C. A. & Rummer, J. L. Dead tired: evaluating the physiological status and survival of neonatal reef sharks under stress. *Conserv. Physiol.* **6**, 1–13 (2018).
57. Loma, T. L. D. *et al.* A Framework for Assessing Impacts of Marine Protected Areas in Moorea (French Polynesia). *Pacific Sci.* **62**, 427–438 (2008).
58. Newman, S. P., Handy, R. D. & Gruber, S. H. Ontogenetic diet shifts and prey selection in nursery bound lemon sharks, *Negaprion brevirostris*, indicate a flexible foraging tactic. *Environ. Biol. Fishes* **95**, 115–126 (2011).
59. Dulvy, N. K. *et al.* Extinction risk and conservation of the world's sharks and rays. *Elife* **3**, e00590 (2014).
60. Heupel, M. R. *et al.* Reef Shark Science – Key Questions and Future Directions. *Front. Mar. Sci.* **21**, 1–14 (2019).
61. Chin, A., Mourier, J. & Rummer, J. L. Blacktip reef sharks (*Carcharhinus melanopterus*) show high capacity for wound healing and recovery following injury. *Conserv. Physiol.* **3**, 1–9 (2015).
62. Bangley, C. W., Rulifson, R. A. & Overton, A. S. Evaluating the Efficiency of Flushed Stomach-tube Lavage for Collecting Stomach Contents from Dogfish Sharks. **12**, 523–533 (2013).
63. R Core Team. R: A language and environment for statistical computing. R Foundation for Statistical Computing. Vienna, Austria. Available at: <http://www.r-project.org/> (2016).
64. RStudio Team 2016. RStudio: Integrated development for R. Boston, MA: RStudio, Inc. Available at: www.rstudio.com.
65. Fulton, T. W. The rate of growth of fishes. *22nd Annu. Rep. Fish. Board Scotl* 141–241 (1904).
66. Irschick, D. J. & Hammerschlag, N. A new metric for measuring condition in large predatory sharks. *J. Fish Biol.* **85**, 917–926 (2014).
67. Gallagher, A. J., Wagner, D. N., Irschick, D. J. & Hammerschlag, N. Body condition predicts energy stores in apex predatory sharks. *Conserv. Physiol.* **2**, 1–8 (2014).

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Author Contributions

O.C.W. designed and coordinated the study; O.C.W., I.A.B. and J.L.R. collected field data; O.C.W. analysed the data and interpreted the data with Y.P.P. and I.A.B. O.C.W. wrote the manuscript with support and inputs from I.A.B, Y.P.P., G.M., J.L.R. and S.P.; all authors gave final approval for publication. O.C.W., I.A.B., S.P. and J.L.R. secured the funding to support this study.

Additional Information

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