



Article

Effect of the antioxidant idebenone on maternal diabetes-induced embryo alterations during early organogenesis

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KEY MESSAGE

Rats with mild diabetes were used to investigate the usefulness of maternal treatments with idebenone to prevent post-implantation embryo impairments. A mitochondrial antioxidant maternal treatment prevented maternal diabetes-induced embryo alterations in prooxidant, proinflammatory and apoptotic markers, and in the expression of genes involved in mitochondrial biogenesis and function.

ABSTRACT

Research question: Can maternal treatments with idebenone, a structural analogue of coenzyme Q10, prevent alterations on markers of proinflammatory-prooxidant processes, on the expression of genes involved in mitochondrial biogenesis and function, and on the apoptotic rate in embryos from mild diabetic rats?

Design: A mild diabetic rat model was induced by neonatal-streptozotocin administration (90 mg/kg subcutaneously). Female diabetic rats and controls were mated with healthy males. From day 1 of pregnancy, control and diabetic rats were orally treated with idebenone (100 mg/kg daily). On day 10.5 of gestation, the embryos were explanted and prepared for immunohistochemical studies, for the evaluation of gene expression by reverse transcription polymerase chain reaction and for TdT (terminal deoxynucleotidyl transferase)-mediated dUDP nick-end-labelling assay analysis.

Results: Embryos from mild diabetic rats showed increased levels of nitrated proteins, 4-hydroxynonenal and matrix metalloproteinase 9, which were prevented by idebenone administration. We also found a decreased embryonic expression of cytochrome c oxidase and reduced mRNA levels of peroxisome proliferator activated receptor- γ coactivator-1- α and nuclear respiratory factor-1, both of which were prevented by idebenone administration to the diabetic pregnant rats. Embryos from mild diabetic rats also showed an increased apoptotic rate, which was diminished by idebenone treatment.

Conclusion: Maternal idebenone treatment ameliorates altered parameters related to the prooxidant-proinflammatory environment found in embryos from mild diabetic rats, suggesting a putative treatment to prevent diabetes-induced embryo alterations.

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Introduction

In diabetic pregnancies, many molecular and cellular pathways are altered during embryo development. These alterations are related to hyperglycaemia and the increase in other intrauterine metabolic substrates, which lead to the generation of a proinflammatory and prooxidant environment (Eriksson and Wentzel, 2016). During early organogenesis, the developing embryo is especially susceptible to the diabetes-induced prooxidant environment caused by the immaturity of its antioxidant system (Zaken et al., 2000). Increased embryonic oxidative stress is involved in diabetes-induced teratogenicity, and further contributes to fetal alterations and the programming of diseases in offspring (Higa and Jawerbaum, 2013; Jawerbaum and White, 2017; Ornoy, 2007; Zabihi and Loeken, 2010; Zhao and Reece, 2013).

Reactive oxygen species mainly originate in the mitochondrion, an organelle whose function is altered in the development of adult diabetic complications (Blake and Trounce, 2014). Cytochrome c oxidase (COX), the terminal oxidase of the mitochondrial electron transport chain, catalyzes the final step of electron transfer from reduced cytochrome c to oxygen to make water. It is also one of the three proton pumps that generate the proton gradient across the inner mitochondrial membrane, which powers adenosine triphosphate synthesis. The transcription of nuclear genes that encode COX subunits is regulated by nuclear respiratory factors 1 and 2 (NRF1 and NRF2) (Dhar et al., 2008). NRF1 and NRF2 also regulate most nuclear genes involved in mitochondrial biogenesis [mitochondrial transcription factors A, B1 and B2 (*Tfam*, *Tfb1m* and *Tfb2m*)]. In this context, the role of peroxisome proliferator activated receptor- γ coactivator 1- α (PGC-1 α), which coactivates different transcription factors in response to energy requirements, is also important to promote the activation of the transcriptional machinery that regulates mitochondrial biogenesis. PGC-1 α coactivates NRF1 and NRF2 as well as peroxisome proliferator activated receptor δ (PPAR δ) (Puigserver and Spiegelman, 2003; Scarpulla, 2008; Wang et al., 2003). This is a nuclear receptor that acts as a nutritional sensor and is able to bind a great variety of lipid molecules, triggering the transcription of genes involved in anti-inflammatory, anti-oxidant, metabolic and developmental processes (Coll et al., 2010; Rotman et al., 2006). It is the only PPAR isotype expressed in high levels in the embryo during early organogenesis (Braissant and Wahli, 1998). In embryos from pregestational streptozotocin-induced diabetic rats (with maternal glycaemia values over 350 mg/dl, rat model of severe diabetes), PPAR δ expression is reduced and its ligands ameliorate several altered parameters in diabetic embryopathy (Higa et al., 2007, 2012).

Oxidative and nitrate stress have been shown in embryos from mild and severe experimental models of diabetes (Gareskog et al., 2006; Jawerbaum and Gonzalez, 2006; Sakamaki et al., 1999; Singh et al., 2011; Viana et al., 2003). Mitochondrial damage has been found in embryos from diabetic rats during early organogenesis, although this is a period characterized by active glycolytic pathways and immature mitochondrial metabolism (Akazawa, 2005; Yang et al., 1995). Antioxidants have been found to have positive effects on the prevention of experimental diabetic embryopathy (Sakamaki et al., 1999; Viana et al., 2003). The putative effect of antioxidants with mitochondrial action, such as idebenone [2-[10-Hydroxydecyl]-5,6-dimethoxy-3-methyl-1,4-benzoquinone], a structural analogue of coenzyme Q10, and a potent antioxidant (Carbone et al., 2012; Geromel et al., 2002; Lin et al., 2015), has never been previously studied. Therefore, in this study, we used a rat model of mild diabetes [glycaemia values below

250 mg/dl, induced by neonatal streptozotocin administration] to address the effect of idebenone on markers of inflammatory-oxidative processes, on the expression of genes involved in mitochondrial function biogenesis, and on apoptosis in the embryo during early organogenesis.

Materials and methods

Animals

Albino Wistar rats were bred in the laboratory with free access to commercial rat chow (Asociación Cooperativa Argentina, Buenos Aires, Argentina) and water, in a lighting cycle of 12 h light–12 h dark. A mild diabetic rat model was developed by injecting 2-day-old neonates with streptozotocin (90 mg/kg, subcutaneously, Sigma-Aldrich, St Louis, MO, USA, diluted in citrate buffer 0.05M pH 4.5, Sigma-Aldrich), as described previously (Jawerbaum and White, 2010; Martinez et al., 2012). Because streptozotocin has a short half-life at neutral pH and selectively enters and accumulates in beta-cells, its cytotoxicity is largely limited to these cells at the doses we used (Eleazu et al., 2013; Jawerbaum and White, 2010).

Control rats were injected with citrate buffer only. Glycaemia was measured with Accu-Chek reagent strips and a glucometer (Bayer Diagnostics, Buenos Aires, Argentina) and confirmed before mating. Pregnant rats were fasted for 6 h before glycaemia measurements were taken on day 10.5 of pregnancy. Rats were considered diabetic when they presented fasting glucose values higher than 130 mg/dl. The reproductive characteristics of the diabetic model have been previously described (Jawerbaum and White, 2010). The animal protocol was approved on 24 July 2013 by the Institutional Committee for the Care and Use of Experimental Animals (CICUAL, Resolution CD N° 1497/2013), School of Medicine, University of Buenos Aires, and conducted according to the Guide for the Care and Use of Laboratory Animals, US National Institutes of Health (NIH Publication, 8th Edition, 2011, <http://www.ncbi.nlm.nih.gov/books/NBK54050/?report=reader>).

Animal treatment and tissue collection

Control and diabetic female adult rats were mated with untreated adult males. Mating was confirmed by the presence of sperm in vaginal smears. The presence of sperm in vaginal smears was used to confirm mating and to designate day 0.5 of gestation. Animals received treatment with either idebenone (Sigma-Aldrich, 100 mg/kg) or vehicle, orally from day 0.5–10.5 of gestation.

Animals were killed in a CO₂ chamber on day 10.5 of pregnancy, and the uteri were transferred to Petri dishes with Krebs Ringer Bicarbonate solution: 5 mM glucose, 145 mM Na⁺, 5.9 mM K⁺, 2.2 mM Ca⁺², 1.2 mM Mg⁺², 127 mM Cl⁻, 25 mM HCO₃⁻, 1.2 mM SO₄⁻² and 1.2 mM PO₄⁻³. The balls of decidual tissue were explanted from each uterus and gently opened to free the conceptuses, using a stereomicroscope and microsurgical dissecting instruments. The embryos were dissected out of the yolk sacs and evaluated morphologically under a stereomicroscope. On day 10.5 of pregnancy, the embryos of the evaluated rats are in the process of neural tube closure, as closure occurs on day 12 of pregnancy.

Viability was established by the presence of a beating heart. Malformed embryos (3% in the diabetic group) received no further analysis. Non-malformed viable embryos were further processed according to protocols described below.

Table 1 – Primer sequences and optimum cycle number for each primer pair used for gene expression analysis by semiquantitative polymerase chain reaction in rat embryo of 10.5 days of gestation.

Gene	Forward primer (5' to 3')	Reverse primer (5' to 3')	Cycles
<i>Coxlvi1</i>	TACGAAACACGACCCCACTG	AGCTAGCGACTACGCTTTCC	28
<i>Nrf1</i>	CAACAGGGAAGAAACGGAAA	GTGGCTCTGAGTTTCCGAAG	27
<i>Tfam</i>	CTGATGGCCATTACATGTGG	AAAGCCCGGAAGGTTAG	26
<i>Pparδ</i>	GAGGGGTGCAAGGGCTTCTT	CACTTGTTCGGTCTTCTTCTG	30
<i>Pgc-1α</i>	GTGCAGCCAAGACTCTGTATGG	GTCCAGGTCATTACATCAAGTTC	34
<i>L30</i>	CCATCTTGGCGTCTGATCTT	GGCGAGGATAACCAATTC	23

Analysis of mRNA levels of *cox*, *nrf1*, *tfam*, *pparδ* and *pgc-1α*

The mRNA levels of *Cox*, *Nrf1*, *Tfam*, *Pparδ* and *Pgc-1α* were evaluated by semi-quantitative reverse transcription polymerase chain reaction (RT-PCR), as previously described (Higa et al., 2014). Four pooled embryos from each rat in each experimental group ($n = 9$) were kept in RNAlater® (Ambion, Texas, USA) at -20°C and RNA was extracted with RNAzol (Molecular Research Center, Cincinnati, USA) in accordance with the manufacturer's instructions. cDNA was synthesized by incubating 2 μg of extracted RNA in a first-strand buffer containing Moloney murine leukemia virus (MMLV) enzyme from Promega (Buenos Aires, Argentina), random primer hexamers (Invitrogen, San Diego, CA, USA), in accordance with the MMLV manufacturer's instructions (final volume 25 μl). The reaction mixture was incubated at 37°C for 60 min and then at 70°C for 15 min. cDNA (2 μl , selected to work within the linear range) was amplified by polymerase chain reaction in a buffer containing deoxynucleotide triphosphates, magnesium chloride solution, Taq polymerase (GoTaq Polymerase, Promega, Wisconsin, USA) and each specific primer, in accordance with the Taq polymerase manufacturer's instructions. The primers used (Table 1), were purchased from Invitrogen. The primers for the ribosomal protein L30 were used as an internal control.

The initial step in the reaction was 95°C for 5 min. The following numbers of cycles are detailed in Table 1. Each cycle consisted of denaturation at 95°C for 15 s primer annealing at 58°C for 30 s and extension at 72°C for 15 s. The resulting products were separated on a 2% agarose gel and stained with SYBR® Safe (Invitrogen). The images were taken with the ImageQuant image acquiring system and software (GE Healthcare, Buckinghamshire, UK) and quantified with the Image J software, National Institutes of Health, Bethesda, MD, USA.

Immunohistochemical and TUNEL analysis

Immunohistochemistry was carried out to determine levels and localize 4-hydroxynonenal, nitrotyrosine, matrix metalloproteinase 9 (MMP9) and subunit IV of COX (COXIV) in embryos, as described previously (Higa et al., 2011). Two embryos from each rat ($n = 5$) in each experimental group were explanted, immediately fixed in 4% phosphate buffered saline (PBS)-paraformaldehyde and then paraffinized. The same number of serial, 5- μm sections were captured from each embryo and on each slide, providing reasonable confidence that protein localization and quantification were performed on comparable anatomical planes from each embryo. All sections were processed simultaneously under identical conditions. Sections were deparaffinized

and hydrated in xylene and a series of graded ethanol solutions, followed by three washes in PBS and PBS with 0.05% Tween-20 (PBS-T). Incubation in 0.3% hydrogen peroxide in PBS to block endogenous peroxidase was carried out for 20 min. Slides were incubated in a humidified chamber overnight with a rabbit anti-4-hydroxynonenal antibody (1:400 Abcam, Cambridge, UK), rabbit anti-nitrotyrosine antibody (1:4000, Millipore, Temecula, USA), a mouse anti-MMP9 antibody (1:150, Calbiochem, Billerica, USA) or a rabbit anti-COXIV antibody (1:300, Abcam). All primary antibodies were checked for antibody specificity and diluted in PBS-T with 1% bovine serum albumin. Negative controls were carried out in the absence of primary antibody and by replacing the primary antibody with a pooled serum of the same species that contains a spectrum of the Immunoglobulin G subclasses (Vector Laboratories Burlingame, CA, USA). Biotinylated horse anti-mouse antibody, rat adsorbed (1:200 in PBS-T/1.5% normal horse serum) or biotinylated goat anti-rabbit (1:200 in PBS-T/1.5% normal goat serum) (Vector Laboratories) was applied, followed by incubation with an avidin-biotin complex for 60 min (Vector Laboratories). Staining was visualized by adding 40% 3,3'-diaminobenzidine tetrahydrochloride chromogen-buffer plus 0.02% (v/v) hydrogen peroxide in 0.05 M Tris (pH 7.6) for 10 min. A positive staining appeared as a dark brown colour. The tissues were then counterstained with haematoxylin and dehydrated before mounting. Five sections of two embryos from each rat of each experimental group were examined by two skilled blinded observers under light microscopy (Nikon Eclipse E200) and photographed (Nikon DS-Fi1). Immunoreactivity intensity was quantified using the ImageProPlus software, Media Cybernetics, Bethesda, MD, USA. Data are shown as relative to a value of 1 assigned to the mean value for control. Similar results were obtained using a semiquantitative score (data not shown).

To study apoptosis in embryos, a TdT (terminal deoxynucleotidyl transferase)-mediated dUTP nick-end-labelling (TUNEL) assay was carried out using an in-situ cell apoptosis detection kit (DeadEnd Fluorometric TUNEL System, Promega) according to the manufacturer's instructions. The sections were examined under fluorescent microscopy (Nikon Eclipse E200) and photographed (Nikon DS-Fi1). The number of positive cells was quantified using the Image J software. Data are shown as relative to a value of 1 assigned to the mean value for controls.

Statistical analysis

Data are presented as the mean \pm standard error. Groups were compared by two-way analysis of variance in conjunction with Bonferroni's test. In all cases, differences were considered statistically significant at $P < 0.05$.

Results

4-hydroxynonenal, nitrotyrosine and MMP9 levels in embryos from control and diabetic rats treated or not with idebenone

We first evaluated control and mild diabetic animals treated or not with idebenone during gestation and found that glycaemia was increased in the diabetic group (control: 101 ± 7 mg/dl; diabetic: 213

± 29 mg/dl, $P < 0.001$) compared with the control group and that idebenone treatment was unable to reduce the increased maternal glucose values (control treated with idebenone: 98 ± 9 mg/dl; diabetic treated with idebenone: 207 ± 33 mg/dl). Oxidative stress during early organogenesis can lead to adverse developmental outcomes and disease risk later in life. We, therefore, studied 4-hydroxynonenal levels as a stable product of oxidative-stress induced lipid peroxidation, and found that its levels were increased in embryos from mild diabetic rats compared with embryos from control rats ($P < 0.001$) (Figure 1),

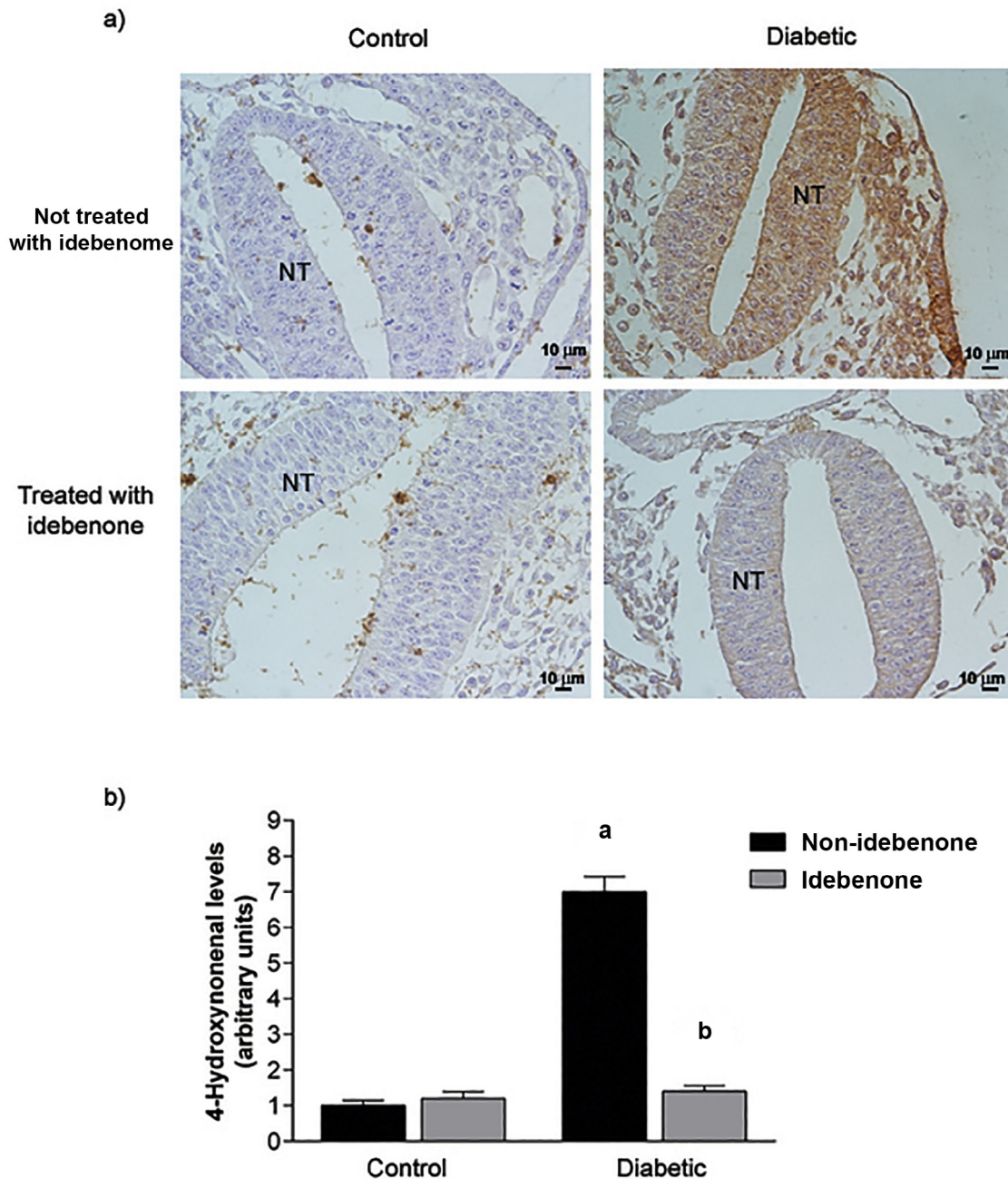


Figure 1 – Effect of maternal idebenone treatment on the levels of 4-hydroxynonenal in embryos from control and diabetic rats on day 10.5 of gestation. (a) A representative photomicrograph of a transverse section from the cephalic zone of the embryo (400X) stained with a specific anti-4-hydroxynonenal antibody; (b) graphs quantifying the immunohistochemical staining represented in (a). Values are means \pm SEM ($n = 5$ rats). Two-way analysis of variance in conjunction with Bonferroni's test was carried out. ^a $P < 0.001$ versus control rats not treated with idebenone; ^b $P < 0.001$ versus diabetic rats not treated with idebenone. NT, neural tube.

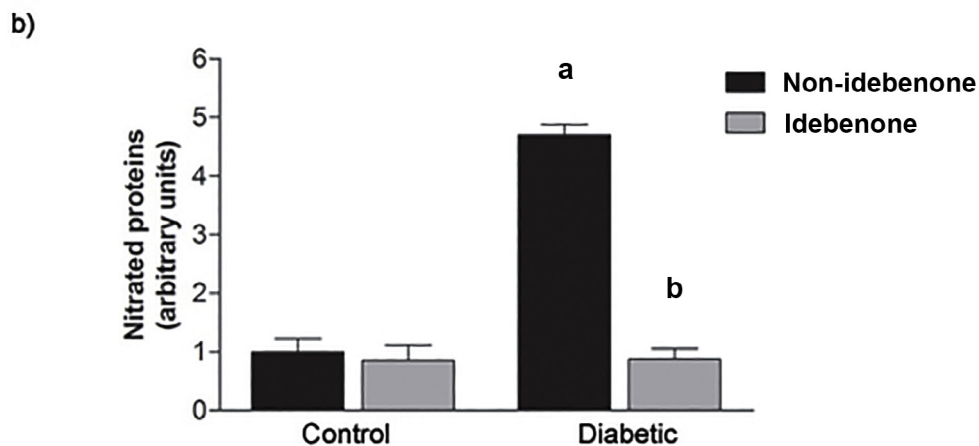
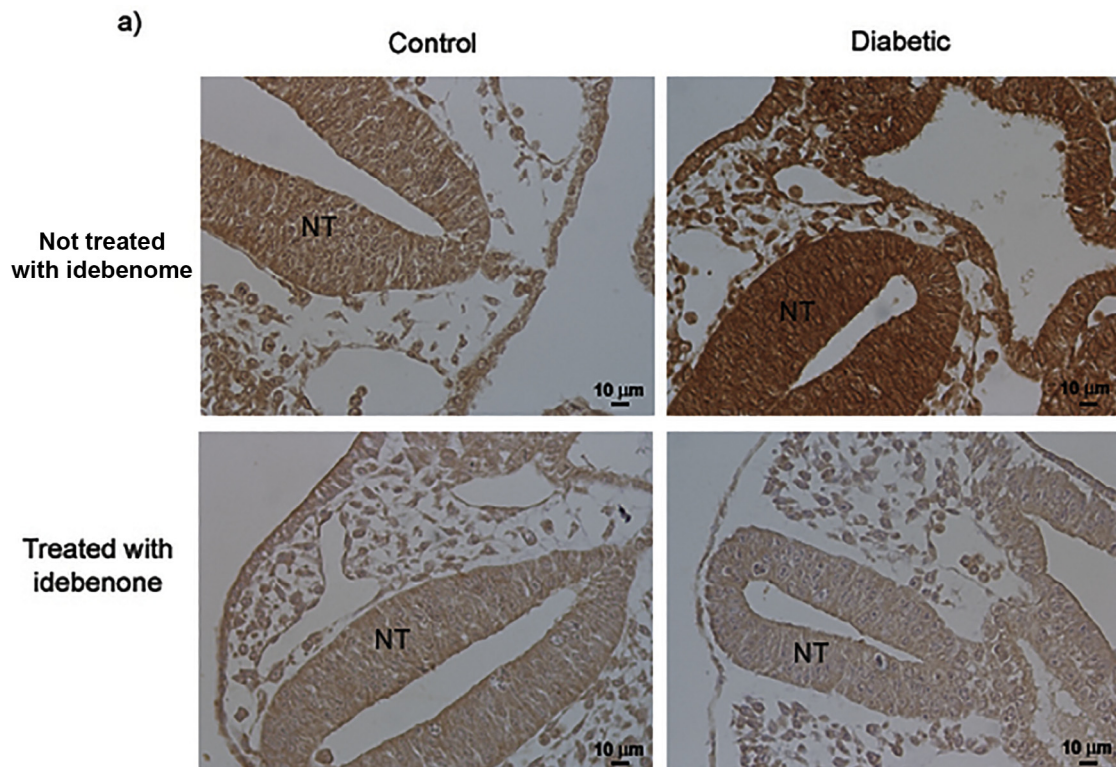


Figure 2 – Effect of maternal idebenone treatment on the levels of nitrated proteins in embryos from control and diabetic rats on day 10.5 of gestation. (a) A representative photomicrograph of a transverse section from the cephalic zone of the embryo (400X) stained with a specific anti-nitrotyrosine antibody; (b) graphs quantifying the immunohistochemical staining represented in (a). Values are means \pm SEM ($n = 5$ rats). Two-way analysis of variance in conjunction with Bonferroni's test was performed. ^a $P < 0.001$ versus control rats not treated with idebenone; ^b $P < 0.001$ versus diabetic rats not treated with idebenone NT, neural tube.

and that they were decreased in embryos from diabetic rats treated with idebenone compared with embryos from non-treated diabetic rats ($P < 0.001$) (Figure 1). We next evaluated nitrotyrosine immunostaining, a marker of the damage induced by peroxynitrites, and found that the levels of nitrated proteins in embryos from mild diabetic rats were increased compared with embryos from control rats ($P < 0.001$) (Figure 2), and they were decreased in embryos from diabetic rats treated with idebenone compared with embryos from non-treated diabetic rats ($P < 0.001$) (Figure 2). As peroxynitrites in-

crease the levels and activity of matrix metalloproteinases (MMP) [Capobianco et al., 2012], we were next interested in studying MMP9 levels, a marker of an inflammatory state when they are in excess. MMP9 immunostaining was increased in embryos from mild diabetic rats compared with embryos from control rats ($P < 0.001$) (Figure 3), and were decreased in embryos from both diabetic ($P < 0.001$) and control ($P < 0.05$) rats treated with idebenone compared with embryos from non-treated diabetic and control rats (Figure 3).

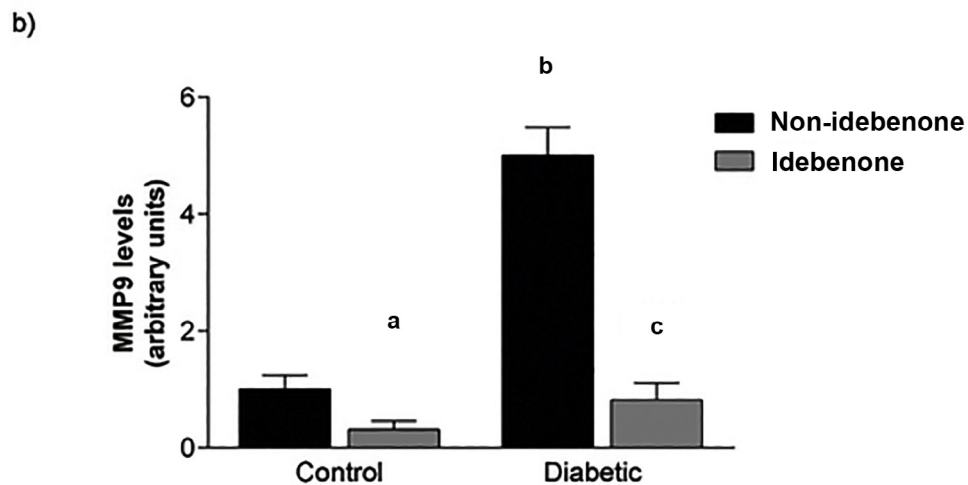
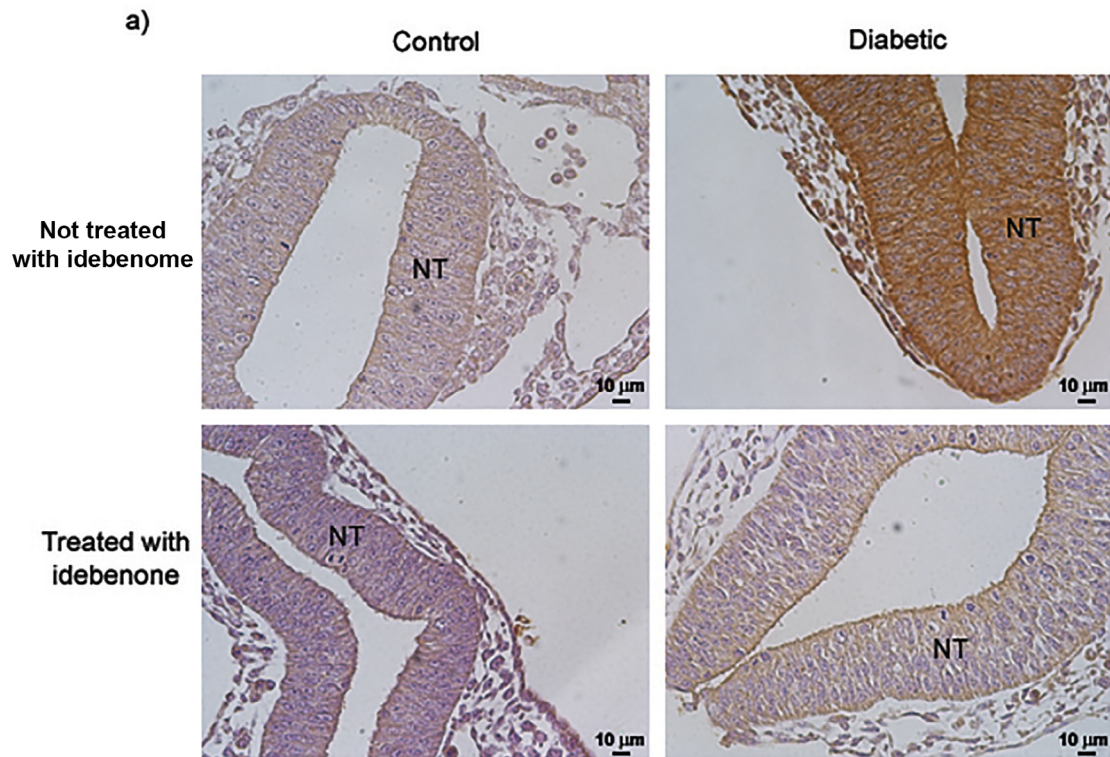


Figure 3 – Effect of maternal idebenone treatment on MMP9 levels in embryos from control and diabetic rats on day 10.5 of gestation. (a) A representative photomicrograph of a transverse section from the cephalic zone of the embryo (400X) stained with a specific anti-MMP9 antibody; (b) graphs quantifying the immunohistochemical staining represented in (a). Values are means \pm SEM ($n = 5$ rats). Two-way analysis of variance in conjunction with Bonferroni's test was performed. ^a $P < 0.05$; ^b $P < 0.001$ versus control rats not treated with idebenone; ^c $P < 0.001$ versus diabetic rats not treated with idebenone. NT, neural tube.

Markers of mitochondrial function and biogenesis in embryos from control and diabetic rats treated or not with idebenone

With COX, the last component of the mitochondrial respiratory chain, we found reduced levels in embryos from diabetic rats compared with embryos from control rats ($P < 0.05$) (Figure 4), and increased levels in embryos from both diabetic ($P < 0.001$) and control ($P < 0.05$) rats treated with idebenone (Figure 4) compared

with embryos from non-treated diabetic and control rats. The subunit IVi1 of COX [COX IVi1] is involved in the regulation of COX activity. mRNA levels of *Cox IVi1* in embryos from diabetic rats were decreased compared with embryos from control rats ($P < 0.05$) (Figure 5a), and increased in embryos from both diabetic ($P < 0.05$) and control ($P < 0.001$) rats treated with idebenone when respectively compared with embryos from non-treated diabetic and control rats (Figure 5a).

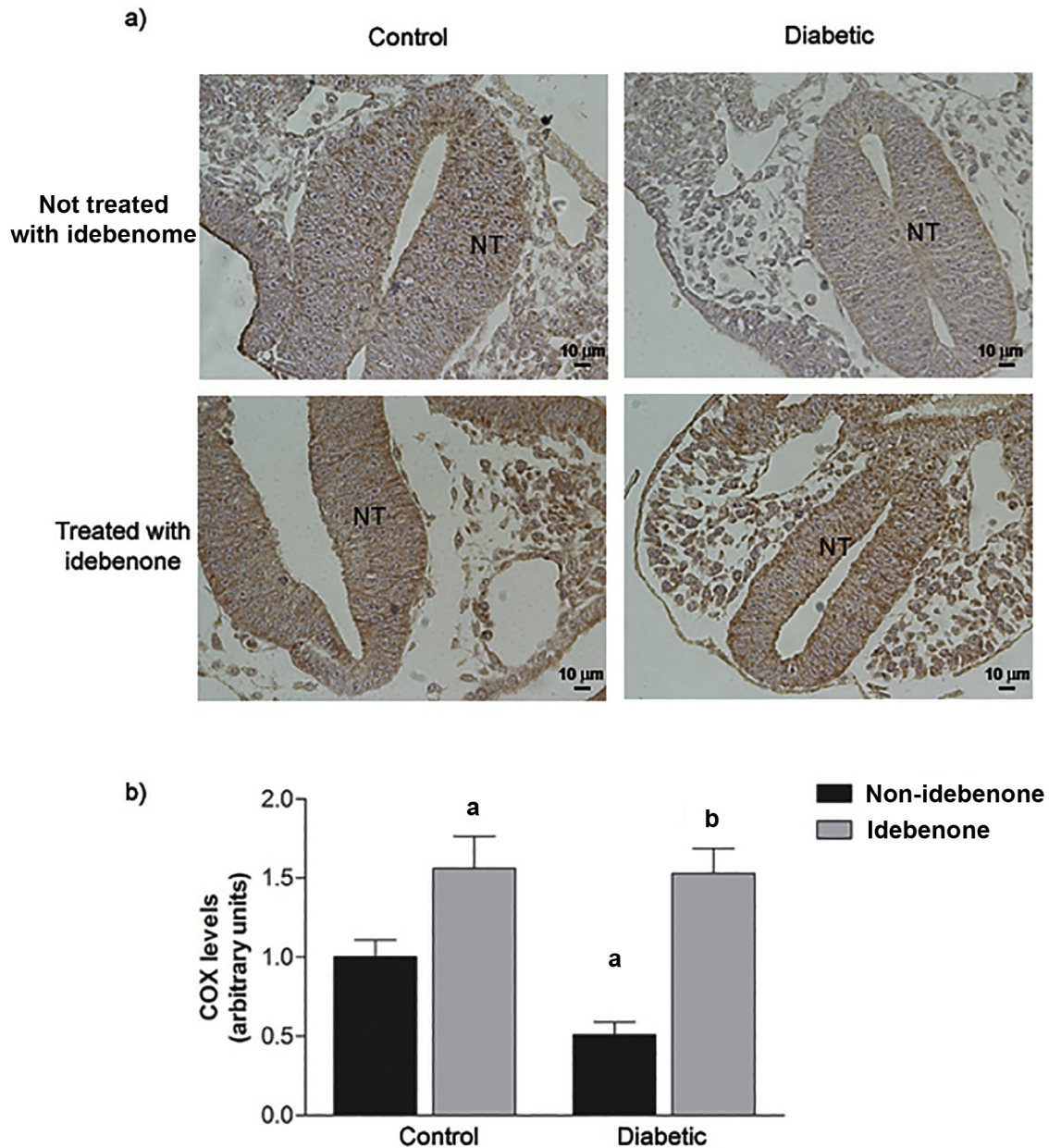


Figure 4 – Effect of maternal idebenone treatment on cytochrome c oxidase (COX) IV levels in embryos from control and diabetic rats on day 10.5 of gestation. (a) A representative photomicrograph of a transverse section from the cephalic zone of the embryo (400X) stained with a specific anti-COX IV antibody; (b) graphs quantifying the immunohistochemical staining represented in (a). Values are means \pm SEM ($n = 5$ rats). Two-way analysis of variance in conjunction with Bonferroni's test was performed. ^a $P < 0.05$ versus control rats not treated with idebenone; ^b $P < 0.001$ versus diabetic rats not treated with idebenone. NT, neural tube.

We also studied TFAM and NRF1, regulatory proteins involved in respiratory chain function. *Tfam* mRNA levels were not different in embryos from diabetic rats treated or not with idebenone when respectively compared with control rats (Figure 5b). On the other hand, *Nrf1* mRNA levels were reduced in embryos from mild diabetic rats compared with embryos from control rats ($P < 0.05$) (Figure 5c), and increased in embryos from diabetic rats treated with idebenone compared with embryos from non-treated diabetic rats ($P < 0.05$) (Figure 5c).

As NRF1 transcription is regulated by PGC-1 α , we also studied this coactivator. *Pgc-1 α* mRNA levels were reduced in embryos from

diabetic rats compared with embryos from control rats ($P < 0.05$) (Figure 6a) and increased in embryos from diabetic rats treated with idebenone compared with embryos from non-treated diabetic rats ($P < 0.05$) (Figure 6a).

The nuclear receptor PPAR δ is involved in antioxidant and developmental processes, and is coactivated by PGC-1 α . mRNA levels of *Ppar δ* were also reduced in embryos from mild diabetic rats compared with embryos from control rats ($P < 0.05$) (Figure 6b) and increased in embryos from diabetic rats treated with idebenone compared with embryos from non-treated diabetic rats ($P < 0.05$) (Figure 6b).

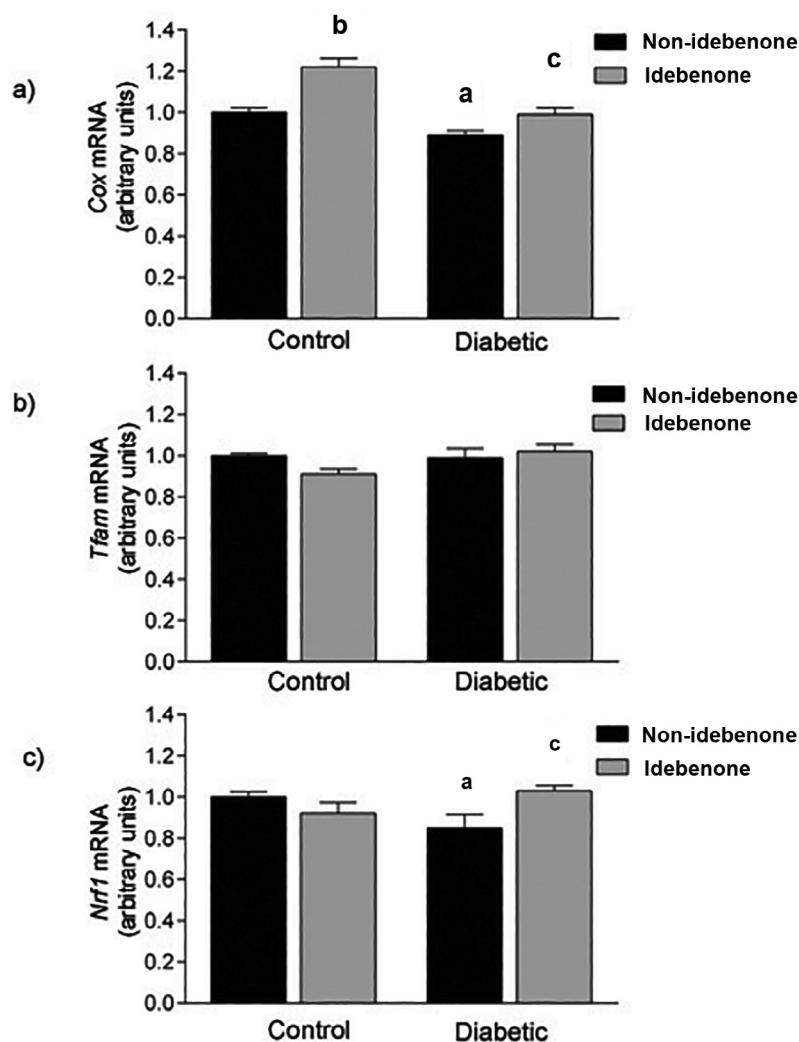


Figure 5 – Effect of maternal idebenone treatment on markers of mitochondrial function and biogenesis in embryos from control and diabetic rats on day 10.5 of gestation. (a) *Cox 1*α** mRNA levels relative to those of L30 in embryos from control and diabetic rats; (b) *Tfam* mRNA levels relative to those of L30 in embryos from control and diabetic rats; (c) *Nrf1* mRNA levels relative to those of L30 in embryos from control and diabetic rats. Values are means \pm SEM ($n = 9$ rats). Two-way analysis of variance in conjunction with Bonferroni's test was carried out. ^a $P < 0.05$; ^b $P < 0.001$ versus control rats not treated with idebenone; ^c $P < 0.05$ versus diabetic rats not treated with idebenone.

Apoptosis in embryos from control and diabetic rats treated with idebenone

To study the effect of idebenone on apoptosis, we conducted a TUNEL assay and found that the number of apoptotic cells increased in embryos from diabetic rats compared with embryos from control rats ($P < 0.001$) (Figure 7) and reduced in embryos from diabetic rats treated with idebenone compared with embryos from non-treated diabetic rats ($P < 0.05$) (Figure 7).

Discussion

The main finding of this work was that the antioxidant idebenone can prevent the increases in markers of prooxidant and proinflammatory processes in embryos from mild diabetic rats during organogenesis. The maternal treatment with idebenone was also able to prevent the alterations in markers of mitochondrial biogenesis and

function, and cell apoptosis observed in embryos of mild diabetic rats highlighting the relevance of aberrant mitochondrial pathways in diabetic embryopathy and pointing to idebenone as a putative treatment to ameliorate the development of viable embryos in a prooxidant/proinflammatory environment.

Importantly, the increased levels of prooxidant and proinflammatory markers we observed in the embryo from mild diabetic rats during early organogenesis have also been observed in fetuses (Pustovrh et al., 2005), placenta (Martinez et al., 2012) and in the postnatal heart (Capobianco et al., 2015), suggesting that embryonic alterations may affect later development.

Moreover, we recently found that maternal idebenone administration during diabetic pregnancies prevents the programming of diabetes-induced cardiac alterations in the offspring (Higa et al., 2017).

Idebenone is a synthetic antioxidant that can be orally administered. It is found both in the cellular cytosol and inside mitochondria, and can act as a free-radical scavenger, undergoing reversible reduction – oxidation reactions to mediate mitochondrial electron flow

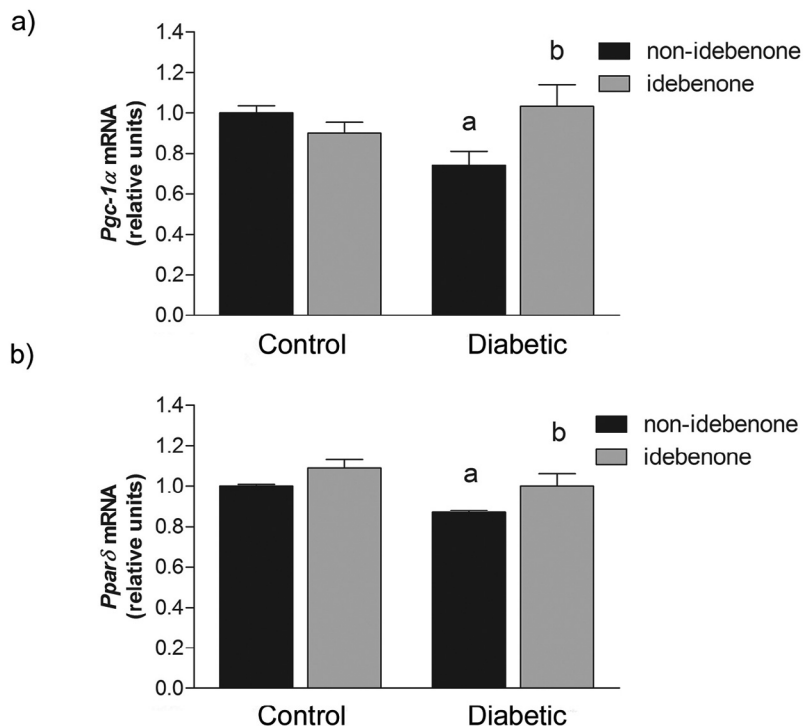


Figure 6 – Effect of maternal idebenone treatment on key regulators of cell energy homeostasis in embryos from control and diabetic rats on day 10.5 of gestation. (a) *Pgc-1α* mRNA levels relative to those of L30 in embryos from control and diabetic rats; (b) *Pparδ* mRNA levels relative to those of L30 in embryos from control and diabetic rats. Values are means \pm SEM ($n = 9$ rats). Two-way analysis of variance in conjunction with Bonferroni's test was performed. ^a $P < 0.05$ versus control rats not treated with idebenone; ^b $P < 0.05$ versus diabetic rats not treated with idebenone.

and thus increase the production of adenosine triphosphate [Carbone et al., 2012]. In this work, idebenone treatment during early organogenesis reduced the increased lipid peroxidation associated with mild diabetes, suggesting that it reduces oxidative stress, and also the increased nitrated proteins, suggesting that it reduces peroxynitrite-induced damage in embryos from diabetic rats. We have previously found increased peroxynitrites-induced damage in embryos from severe diabetic rats (glucose concentrations over 350 mg/dl) [Jawerbaum et al., 2005], but this is the first time that this damage is reported in embryos from rats with mild hyperglycaemia (glucose concentrations below 250 mg/dl). Peroxynitrites are produced by the reaction between nitric oxide and superoxide and are strong oxidizing and nitrating species that promote oxidative damage by many mechanisms [Szabo, 2003]. These compounds can increase the production and activation of MMP [Capobianco et al., 2012; Sugiura et al., 2012]. We have previously found increased MMP9 levels and activation in embryos from severe diabetic rats [Higa et al., 2011]. In this work, we also found increased MMP9 levels in the embryos from mild diabetic rats, which were reduced by idebenone. In pathological conditions, the excess of peroxynitrites can alter the mitochondrial chain function via the oxidative inactivation of complexes I, II and V and can also nitrate cytochrome c [Cassina and Radi, 1996; Cassina et al., 2000]. In pathological conditions, peroxynitrites can also increase the K_m for oxygen of COX [Cooper et al., 2003]. Maternal diabetes in rats has been shown to induce alterations in mitochondrial morphology in embryos from 9 to 11 days of gestation [Yang et al., 1995]. In the present study, by evaluating embryos on day 10.5 of gestation, we found decreased expression of COX in embryos from diabetic rats, and that

idebenone treatment prevented this alteration, suggesting that, despite their dependence on glycolytic pathways at the developmental stage evaluated here, embryos may benefit from idebenone treatment. An increase in COX expression mediated by idebenone has been previously seen in the hippocampus and cerebellum of a mouse model of Angelman syndrome with mitochondrial dysfunction [Llewellyn et al., 2015]. Interestingly, the mechanisms of COX upregulation seem to depend on upstream regulators, given that the mRNA levels of both *Nrf1* (which regulates COX IV transcription) and its coactivator, *Pgc-1α* [Puigserver and Spiegelman, 2003; Scarpulla, 2008], are upregulated with the maternal idebenone treatment in the embryos from diabetic rats. The relevance of maternal idebenone treatment as an upregulator of *Pgc-1α* expression in the embryos is supported by recent findings showing that maternal diabetes induces downregulation of *Pgc-1α* expression in the embryos, changes related to an increased rate of neural tube defects [Chen et al., 2017].

The nuclear receptor PPAR δ participates in cell proliferation and differentiation in adult tissue as well as in fetal tissues and has anti-inflammatory properties [Coll et al., 2010; Rotman et al., 2006]. We have previously found that aberrant PPAR δ signalling is involved in embryo dysmorphogenesis and that PPAR δ is reduced in embryos from severe diabetic rats [Higa et al., 2007, 2014]. In the present study, we also found reduced *Pparδ* mRNA levels in the embryos from mild diabetic rats and that idebenone was able to modulate *Pparδ* transcription, increasing the reduced mRNA levels of this nuclear receptor. Although the mechanism remains unclear, it has been recently suggested that PPARs are epigenetically regulated [Lendvai et al., 2016] and that reactive oxygen species participate in many important epigenetic processes [Afanas'ev,

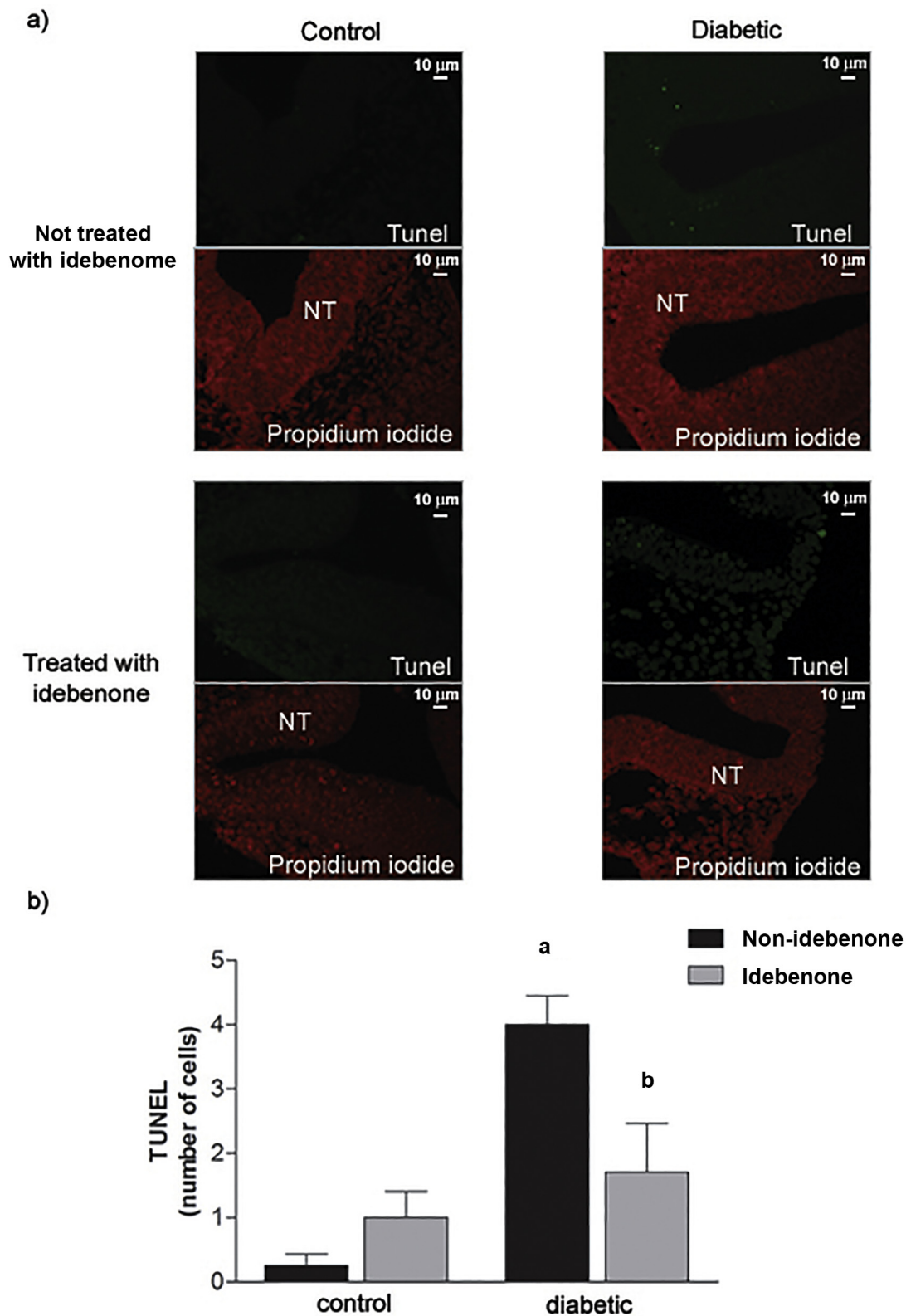


Figure 7 – Effect of maternal idebenone treatment on apoptosis in embryos from diabetic rats on day 10.5 of gestation. a) shows representative photomicrographs of a transverse section from the cephalic zone of the embryo (400X). Apoptotic cells were labelled in green and nuclei in red by propidium iodide; b) the number of TdT (terminal deoxynucleotidyl transferase)-mediated dUTP nick-end-labelling-positive cells is shown. Values are means \pm SEM ($n = 5$ rats). Two-way analysis of variance in conjunction with Bonferroni's test was performed. ^a $P < 0.001$ versus control rats not treated with idebenone; ^b $P < 0.05$ versus diabetic rats not treated with idebenone.

2015). The upregulation of *Pparδ* by idebenone observed in the embryos from diabetic rats may contribute to a decrease in the prooxidant/proinflammatory environment as PPARs regulate the expression of antioxidant enzymes and modulate the expression of the inducible isoform of nitric oxide synthase and MMPs [Barlaka et al., 2015; Chao et al., 2014]. PPARδ is also involved in inhibiting apoptosis in different tissues [Barlaka et al., 2015; Schuster et al., 2010].

An optimal balance between cell proliferation and apoptosis is required for proper development. An increased apoptotic rate related to a prooxidant–proinflammatory environment has been reported in murine embryos exposed to a diabetic environment [Gareskog et al., 2007; Wu et al., 2016; Yang et al., 2008]. In the present study, idebenone administration reduced the increased apoptotic rate in embryos from diabetic rats, probably by increasing PPARδ levels, reducing the prooxidant–proinflammatory processes, and preventing the alteration in mitochondrial function and biogenesis parameters. As a limitation, we were not able to measure idebenone levels in the embryo. Therefore, it is not clear whether the effects of maternal idebenone treatment on embryos is attributable to a direct effect on the embryos or to its impact on maternal tissues.

We conclude that oxidative stress is a main inductor of aberrant pathways in the embryo during early organogenesis and that the use of an antioxidant that targets mitochondria ameliorates several mitochondrial, nuclear, cytoplasmic and extracellular parameters, thus precluding embryonic changes associated with maternal diabetes. Idebenone is currently used in Friedreich's ataxia, Alzheimer's disease and Leber hereditary optic neuropathy disease [Gutzmann and Hadler, 1998; Meyerson et al., 2015; Parkinson et al., 2013]. Therefore, it would be interesting to study whether maternal idebenone treatment may improve diabetes-induced neural alterations in the offspring. As our data suggest a potential role of idebenone in preventing maternal diabetes-induced embryo damage, clinical studies are needed to address the potential use of this drug in pregestational diabetic patients.

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