

Electro-fluid dynamics of aqueous humor production: simulations and new directions

Aurelio Giancarlo Mauri¹, Lorenzo Sala², Paolo Airoldi¹, Giovanni Novielli³, Riccardo Sacco¹, Simone Cassani⁴, Giovanna Guidoboni^{4,2}, Brent Siesky⁵, Alon Harris⁵

¹Dipartimento di Matematica, Politecnico di Milano, Milano, Italy, ²Institut de Recherche Mathématique Avancée CNRS and Université de Strasbourg 7, Strasbourg Cedex, France, ³ST Microelectronics, Agrate Brianza (MB) Italy, ⁴Indiana University and Purdue University at Indianapolis, Department of Mathematical Sciences, IN, USA, ⁵Eugene and Marilyn Glick Eye Institute, Indiana University School of Medicine, IN, USA

Abstract

Purpose: to theoretically investigate the role of bicarbonate ion (HCO_3^-) on the nonpigmented transepithelial potential difference V_m , the sodium potassium pump (Na/K) and the active secretion of aqueous humor.

Methods: a three-dimensional mathematical model is proposed to isolate the roles of HCO_3^- and Na^+ , which are difficult to investigate experimentally. The model combines the velocity-extended Poisson-Nernst-Planck equations to describe ion electrodiffusion and the Stokes equations to describe aqueous humor flow into the basolateral space adjacent to the nonpigmented ephitelial cells.

Results: Computations show that V_m is close to baseline experimental measurements (on monkeys) in the range [-2.7, -2.3] mV only if HCO₃⁻ is included in the simulation. The model is also capable of reproducing the flow of Na⁺ exiting the cell and the flow of K⁺ entering the cell, in accordance with the physiology of the Na/K pump. The simulated Na/K ratio is 1.53, which is in very good agreement with the theoretical value of 1.5.

Conclusion: Model simulations suggest that HCO_3^- inhibition may prevent physiologically correct baseline values of the nonpigmented transepithelial potential difference and Na/K ATPase function. This may provide useful indication in the design of medications that decrease the active secretion of aqueous humor, and supports the

Correspondence: Aurelio Giancarlo Mauri, Dipartimento di Matematica, Politecnico di Milano, Piazza Leonardo da Vinci 32, 20133 Milano, Italy.

E-mail: aureliogiancarlo.mauri@polimi.it

advantage of using mathematical models as a noninvasive complement of animal models.

Keywords: aqueous humor production, electrochemical transport, mathematical modeling, numerical simulation, sodium-potassium pump

Abbreviations/Nomenclature: HCO_3^- : bicarbonate; Na^+ : sodium; K^+ : potassium; Cl^- : chloride; ATPase: adenosine triphosphatase; IOP: intraocular pressure; AH: aqueous humor; VE-PNP: velocity-extended Poisson-Nernst-Planck; NPE: nonpigmented epithelial cell; PDE: partial differential equation.

1. Introduction

Elevated intraocular pressure (IOP) is a recognized risk factor for glaucoma. Evidences show that the steady-state value of IOP is determined by the balance between production and drainage of aqueous humor (AH), and that sodium (Na⁺) and bicarbonate (HCO₃⁻) secretion into the basolateral space between nonpigmented epithelial cells (NPEs) contribute to AH production (see¹ and references cited therein). A possible strategy to reduce IOP via topical medications is to reduce AH production via beta-antagonists, alpha-agonists and carbonic anhydrase inhibitors^{2,3}. The connection between HCO₃⁻, Na⁺ and topical medications in the regulation of AH production is still controversial and difficult to study experimentally. Thus, the development and use of a mathematical model appears to be a promising approach to help unravel such a connection through simulation and comparison of different predicted scenarios.

In the present work, we propose a mathematical model capable of describing the relationship between nonpigmented epithelial transmembrane ion flow and AH production. The velocity-extended Poisson-Nernst-Planck (VE-PNP) equations are used to describe ion electrodiffusion through a moving electrolyte medium, whereas the Stokes equations are used to describe the motion of the incompressible electrolyte fluid. The illustration and discussion of the theoretical properties and algorithms to numerically solve the coupled VE-PNP/Stokes system can be found in^{4–9}. Here, the model is implemented within the three-dimensional object-oriented software platform MP-FEMOS (Multi-Physics Finite Element Modeling Oriented Simulator^{9–13}).

The proposed model is validated against baseline values of the nonpigmented transepithelial potential difference V_m . Then, the model is used to predict ion current flow under different working conditions characterized by the absence or presence of bicarbonate ions in the electrolyte fluid. Model simulations suggest that HCO_3^- inhibition may prevent to attain physiologically correct baseline values of V_m and Na/K ATPase function.

2. Methods

AH production within the ciliary body is driven by three main mechanisms: (a) transport of aqueous components by the ciliary circulation; (b) ultrafiltration and diffusion of clear fluid across the capillary-stroma barrier, and (c) osmotic efflux of clear fluid into the posterior chamber³. In this paper, we focus on the osmotic efflux, which is due to the active secretion of ions into the basolateral space between nonpigmented epithelial cells. Interestingly, ion accumulation in the basolateral space generates a transepithelial potential difference V_m measured experimentally in various animal species, as reported in Table 1.

Table 1. Reference data for the transepithelial potential difference V_m for several animal species.

$V_m [mV]$	animal species	reference
3.80 ± 0.26	ох	14
5.53 ± 0.41	ох	15
3.83 ± 0.16	rabbit	15
-3.7 ± 0.3	toad	16
-1.2 ± 0.1	rabbit	17
-1.35 ± 0.08	dog	18
-2.5 ± 0.2	monkey	19



Fig. 1. Left panel: electrical scheme of the ciliary body system. PE: pigmented epithelial cellular layer. The various compartments are connected through resistances. Black bullets identify circuit nodes whose associated electrical variable is an electric voltage. The yellow box identifies the nonpigmented epithelial membrane region treated by the model. *Right panel:* geometry of a transmembrane channel in the yellow region. Side A represents the intracellular NPE region, Side B represents the extracellular region in the basolateral space. The thickness of the channel is 5nm.

The resistive path between ciliary capillaries and posterior chamber is schematized in Fig. 1 (left panel). Based on the ionic concentrations reported in¹⁹ for the various segments in the path, it appears that the voltage drop between capillaries and NPE is 5 orders of magnitude smaller than the values in Tab. 1 so that, as a consequence, most of the transepithelial voltage drop occurs across the membrane of the NPE. Thus, in this work we focus on the NPE membrane, as indicated by the yellow square in Fig. 1 (left panel), and we consider a cylindrical ion channel whose inlet (SIDE A) and outlet (SIDE B) sections denote the intracellular NPE side and the basolateral space side, respectively, as depicted in Fig. 1 (right panel). We remark that the difference in potential between SIDE A and SIDE B corresponds to V_m . In particular, here we consider a particular type of ion channel, namely a sodium/potassium pump, since it has been proposed by many authors as important player in AH production^{1,3,20}.

As mentioned in Section 1, we describe the electro-fluid dynamics within an ion channel in the NPE membrane by combining two main mathematical blocks:

- Velocity-Extended Poisson-Nernst-Planck (VE-PNP) equations to treat ionic transport into the basolateral space;
- Stokes equations to treat water movement down the resultant osmotic pressure gradient into the posterior chamber.

The VE-PNP equations express the conservation of each ionic charge moving across the ion channel and of the associated electrical current⁴. The Stokes equations express the conservation of mass and momentum of the electrolyte charged fluid flowing across the channel and transporting the ions at low Reynolds number^{4,5,7,8}. The partial differential equations (PDEs) constituting the VE-PNP and the Stokes model blocks are illustrated in detail in Appendix A. The two blocks are mutually coupled through the volume force density due to the electric pressure exerted by the ions on each infinitesimal fluid element²¹. The solution of the VE-PNP/Stokes coupled system yields as output the spatio-temporal number density n_i of the *i*-th ion species, $i = 1, \ldots, M$, the electric potential φ , the electrolyte fluid velocity **u** and the fluid pressure *p*. Let *t* denote the time variable, $t_0 = 0$ the starting simulation time and *T* the total simulation time. The numerical approximation of the VE-PNP/Stokes coupled system is based on the following three steps:

- 1. We partition the time observational window $[t_0, t_0 + T]$ into $N_T \ge 1$ time intervals $\tau_k = [t_k, t_{k+1}]$, $k = 0, \ldots, N_T 1$, of uniform width $\Delta t = T/N_T$;
- 2. For each $k = 0, \ldots, N_T 1$, we advance in time using the Backward Euler time discretization method and we solve the PNP nonlinear block using a fixed point algorithm for a given velocity \mathbf{u}^k . This step returns the VE-PNP solution pair $(n_i^{k+1}, \varphi^{k+1}), i = 1, \ldots, M$, at the next time level;
- 3. We use n_i^{k+1} , $i = 1, \ldots, M$, and φ^{k+1} to compute the electric pressure exerted by the ions on the fluid and we solve the Stokes linear block. This step returns the Stokes solution pair $(\mathbf{u}^{k+1}, p^{k+1})$ at the next time level.

Numerical experiments for the validation of the model and the solution map schematically illustrated above are reported and discussed in⁹ and²² in the study of several nanoscale ionic channels of biophysical interest.

3. Results and discussion

In this section we use the model illustrated in Section 2 to investigate the role of the NPE transmembranal sodium-potassium pump on the osmotic efflux of AH into the basolateral space. Boundary conditions on the surface of the cylindrical domain of Fig. 1 (right panel) are enforced in such a way that: (i) the K⁺ ions enter the NPE at the intracellular side (A); (ii) the Na⁺ ions enter the basolateral space at the aqueous side (B), with the ratio Na^+/K^+ = 3:2 as in physiological Na/K ATPase; (iii) the electrostatic potential φ is grounded at side A; and *(iv)* the electric field flux is zero at side B. The values of all model parameters and of the initial and boundary conditions for the VE-PNP system are summarized in Table 2. The longitudinal dimension of the simulated ion channel is 5nm whereas the radius of the channel is 0.4nm. The computer simulations with the MP-FEMOS software platform run on a (rather coarse) grid made of 2000 tetrahedral elements and 600 vertices, with about 3000 degrees of freedom for the VE-PNP system and more than 10000 degrees of freedom for the Stokes system. Two distinct sets of ions are considered in the description of ion electrodiffusion, the first set including the species K^+ , Na^+ and Cl^- (M = 3), the second set including also the HCO_3^- ion (M = 4). The baseline ion concentrations are set according to the values reported in¹⁹.



Fig. 2. Left panel: spatial distribution of φ in the NPEC channel. Black curve: HCO₃⁻ is inhibited. Red curve: HCO₃⁻ is included. *Right panel:* spatial distributions of K⁺ (black) and Na⁺ (red) steady-state current densities.

Fig. 2 (left panel) illustrates the results of model validation against physiological reference data by plotting the steady-state spatial distribution of the electric potential (in mV). The computed nonpigmented transepithelial potential difference, $V_{m,c}$, is equal to the value of φ at side B. The negative value of $V_{m,c}$ indicates local net anion accumulation. We notice that $V_{m,c}$ is close to experimental baseline measurement (for monkeys) in the range [-2.7, 2.3]mV¹⁹ (cf. Table 1, last row) only if bicarbonate is included in the simulation (red curve). Fig. 2 (right panel) illustrates model predictions showing the spatial distributions of the Na⁺ and K⁺ steady-state current densities (in A cm⁻²). The positive value of the Na⁺ current indicates that Na⁺ ions exit the NPE whereas the negative value of the K⁺ current indicates that K⁺ ions enter the NPE, in accordance with the physiology of the Na/K pump. The ratio between the (module) of the two currents is 1.53 in very good agreement with the theoretical value of 1.5.

4. Conclusions and future perspectives

In the present article we have carried out the theoretical study of the osmotic efflux of aqueous humor into the posterior chamber of the eye due to the active secretion of ions into the basolateral space between nonpigmented epithelial cells. The research is motivated by the fact that it is rather difficult in experimental investigations to isolate the role of a single electrolyte in regulating aqueous humor production. To this purpose, we have devised a mathematical model localized at the nonpigmented transepithelial cellular scale with the aim of describing the ionic flux of sodium, potassium, chloride and bicarbonate across a single membrane ion channel to the basolateral space which is believed to lead ultimately to a net H₂O efflux into the posterior eye chamber. The model is constituted by a system of partial differential equations representing balance laws for ionic charge, fluid mass, ionic momentum density, fluid momentum, mobile charge and electric field at each spatial position in the channel and at each time level. Simulation results tested against experimental measurements in monkeys suggest that bicarbonate inhibition may prevent to attain physiologically correct baseline values of the transepithelial membrane potential. Simulations accounting the presence of bicarbonate also predict the possibility of reproducing a physiologically correct Na/K ATPase function. These results may provide useful indication in the design of IOP lowering medications to decrease aqueous humor production on an effective patient-specific basis, and support the advantage of using mathematical modelling as a noninvasive complement of the animal model. Future effort will be devoted to investigating the role of permanent surface charge and of protein conformation²³ on the modulation of the electrophysiological response of the transepithelial membrane channel.

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Parameter	value	units
z_{K^+}	+1	[-]
z_{Na^+}	+1	[-]
$z_{\rm Cl}$ –	-1	[-]
$z_{\rm HCO_3^-}$	-1	[-]
μ_{K^+}	$1.957 \cdot 10^{-5}$	$cm^{2}V^{-1}s^{-1}$
μ_{Na^+}	$1.334 \cdot 10^{-5}$	$cm^{2}V^{-1}s^{-1}$
$\mu_{\rm Cl}$ –	$2.033 \cdot 10^{-5}$	$cm^{2}V^{-1}s^{-1}$
$\mu_{\rm HCO_3^-}$	$1.185 \cdot 10^{-5}$	$cm^{2}V^{-1}s^{-1}$
$\overline{f}_{\mathrm{K}^+ _{\mathrm{SIDE A}}}$	$2 \cdot 10^{19}$	${\rm cm}^{-2}{\rm s}^{-1}$
$\overline{n}_{\rm Na^+} _{\rm SIDE A}$	$7.82 \cdot 10^{19}$	$\rm cm^{-3}$
$\overline{n}_{\rm Cl^-} _{\rm SIDE A}$	$6.44 \cdot 10^{19}$	$\rm cm^{-3}$
$\overline{n}_{\mathrm{HCO}_{2}^{-}} _{\mathrm{SIDE A}}$	$1.81 \cdot 10^{19}$	cm^{-3}
$\overline{n}_{\mathrm{K}^+ _{\mathrm{SIDE B}}}$	$2.41 \cdot 10^{18}$	$\rm cm^{-3}$
$\overline{f}_{\rm Na^+} _{\rm SIDE B}$	$3 \cdot 10^{19}$	$cm^{-2}s^{-1}$
$\overline{n}_{\rm Cl^-} _{\rm SIDE B}$	$7.89 \cdot 10^{19}$	$\rm cm^{-3}$
$\overline{n}_{\mathrm{HCO}_{2}^{-}} _{\mathrm{SIDE B}}$	$1.2 \cdot 10^{19}$	$\rm cm^{-3}$
$n_{\rm K^+}^0$	$2.41 \cdot 10^{18}$	$\rm cm^{-3}$
$n_{\mathrm{Na}^+}^0$	$8.19 \cdot 10^{19}$	$\rm cm^{-3}$
$n_{\rm Cl^-}^0$	$7.17 \cdot 10^{19}$	$\rm cm^{-3}$
$n_{\rm HCO_{-}}^{0}$	$1.51 \cdot 10^{19}$	$\rm cm^{-3}$
T	300	K
ϵ_f	$708.32 \cdot 10^{-10}$	$\rm Fcm^{-1}$
$\overline{\overline{\varphi}}$ SIDE A	0	V
$\sigma_{ m fixed}$	0	$\rm Ccm^{-2}$
μ_f	10^{-2}	$g cm^{-1} s^{-1}$
$ ho_f$	1	$ m gcm^{-3}$
g	0	$\mathrm{cm}\mathrm{s}^{-1}$
\mathbf{u}^0	0	$\mathrm{cm}\mathrm{s}^{-1}$
$\overline{p} _{\text{SIDE A}}$	0	$\mathrm{gcm^{-1}s^{-2}}$
$\overline{p} _{ ext{SIDE B}}$	0	$ m gcm^{-1}s^{-2}$

Table 2. Model parameters, boundary and initial conditions for the VE-PNP model. All values are taken as in^{22} .

A. Mathematical Model

In Sect. A.1 we introduce the mathematical description of the problem geometry. In Sect. A.2 we describe the PNP equations for ion flow and in Sect. A.3 we describe the Stokes equations for the intrachannel electrolyte fluid.

A.1 Geometry

Let \mathbf{x} and t denote the spatial and temporal coordinates, respectively. The simulation domain $\Omega \subset \mathbb{R}^3$ (the ion channel) is shown in Fig. 1 (right panel). The channel is filled with an incompressible, viscous and Newtonian fluid moving with velocity $\mathbf{u} = \mathbf{u}(\mathbf{x},t)$, in which $M \geq 1$ chemical species are dissolved. Each chemical has effective charge z_i , i = 1, ..., M, and concentration (number density) $n_i = n_i(\mathbf{x},t)$ [cm⁻³]. Cations have $z_i > 0$, anions have $z_i < 0$. We denote by $\partial\Omega$ the boundary of the domain Ω and by \mathbf{n} the unit outward normal vector on $\partial\Omega$. With each dependent variable U, we associate a partition of the domain boundary $\partial\Omega$ into the union of (generally different) subsets. We indicate by Γ_D^U the subset of $\partial\Omega$ where a Dirichlet condition and by Γ_N^U the subset where a Neumann condition is applied, in such a way that $\Gamma_D^U \cup \Gamma_N^U = \partial\Omega$ and $\Gamma_D^U \cap \Gamma_N^U = \emptyset$.

A.2 The Velocity-Extended Poisson-Nernst-Planck equation system

The Velocity-Extended Poisson-Nernst-Planck system to describe the electrodiffusion of M ion species n_i , i = 1, ..., M, throughout a fluid in motion with velocity \mathbf{u} consists of the following PDEs:

$$\frac{\partial n_i}{\partial t} + \operatorname{div} \mathbf{f}_i = 0, \tag{1a}$$

$$\mathbf{f_i} = \frac{z_i}{|z_i|} \mu_i n_i \mathbf{E} - D_i \nabla n_i + n_i \mathbf{u}, \tag{1b}$$

$$\operatorname{div} \mathbf{D} = q \sum_{i=1}^{M} z_i n_i, \tag{1c}$$

$$\mathbf{D} = \epsilon_f \mathbf{E} = -\epsilon_f \nabla \varphi. \tag{1d}$$

We refer to^{4,24–26} for a detailed derivation of (1), to^{5,7} for the study of existence of its solution and to^{11,27} for recent applications to ion channel simulations. In (1), the symbol f_i denotes the ion particle flux $[\text{cm}^{-2}\text{s}^{-1}]$, μ_i and D_i are the ion electrical mobility and diffusivity and T is system temperature. The third term in the right-hand side of the linear momentum balance equation (1b) expresses the coupling between the motion of the ion species and the moving electrolyte fluid. In the Poisson equation (1c), **D** is the electric displacement, **E** and φ are the electric field $[\text{Vcm}^{-1}]$ and electric potential [V], respectively, whereas $q = 1.602 \cdot 10^{-19} [\text{C}]$ and ϵ_f denote the electron charge and the electric premittivity, respectively. The diffusion coefficient

 D_i and the mobility μ_i are proportional through the Einstein relation

$$D_i = \mu_i \frac{k_B T}{q|z_i|},\tag{2}$$

where $k_B = 1.38 \cdot 10^{-16} [\text{cm}^2 \text{ g s}^{-2} \text{ K}^{-1}]$ is Boltzmann's constant and T is system temperature. Initial conditions for ion concentrations are $\forall i = 1, ..., M$:

$$n_i(\mathbf{x}, 0) = n_i^0(\mathbf{x}) \qquad \qquad \text{in } \Omega, \tag{3a}$$

where the functions n_i^0 are positive given data. The boundary conditions for the VE-PNP system are $\forall i = 1, ..., M$:

$$\varphi = \overline{\varphi}$$
 on $\Gamma_{\mathcal{P}}^{\varphi}$, (4a)

$$\mathbf{D} \cdot \mathbf{n} = \sigma_{\text{fixed}} \qquad \qquad \text{on } \Gamma_N^{\varphi}, \qquad \qquad (4b)$$

$$\mathbf{n} = f_i$$
 on $\Gamma_N^{n_i}$, (4d)

where $\overline{\varphi}$ is the electrostatic potential of the side Γ_D^{φ} , σ_{fixed} [C cm⁻²] is a fixed surface charge density on side Γ_N^{φ} due to the lipid membrane bilayer and \overline{n}_i is a given concentration of the chemical species i on side $\Gamma_D^{n_i}$. Conditions (4a) and (4c) enforce a given voltage and a given concentration on the associated side, respectively, whereas (4b) and (4d) enforce nonhomogeneous Neumann conditions on each associated side.

A.3 The Stokes equation system

The Stokes system describing the slow motion of an incompressible viscous fluid with a constant density $\rho_f \, [g \, cm^{-3}]$ consists of the following PDEs:

$$\operatorname{div} \mathbf{u} = 0, \tag{5a}$$

$$\rho_f \frac{\partial \mathbf{u}}{\partial t} = \operatorname{div} \underline{\underline{\sigma}}(\mathbf{u}, p) + q \sum_{i=1}^M z_i n_i \mathbf{E},$$
(5b)

$$\underline{\underline{\sigma}}(\mathbf{u},p) = 2\mu_f \underline{\underline{\epsilon}}(\mathbf{u}) - p\underline{\underline{\delta}},\tag{5c}$$

$$\underline{\underline{\epsilon}}(\mathbf{u}) = \underline{\underline{\nabla}}_{s} \mathbf{u} = \frac{1}{2} (\nabla \mathbf{u} + (\nabla \mathbf{u})^{T}).$$
(5d)

We refer to²⁸ for a complete mathematical and numerical treatment of the Stokes and Navier-Stokes equations for incompressible and compressible fluids. In (5), u is the fluid velocity $[cm s^{-1}]$, p the fluid pressure [Pa], μ_f the fluid shear viscosity $[g m^{-1} s^{-1}], \underline{\sigma}$ the stress tensor $[g cm^{-1} s^{-2}]$ and $\underline{\epsilon}$ the strain rate tensor $[s^{-1}]$. The symbol $\underline{\delta}$ is the second-order identity tensor of dimension 3 and the second-order tensor $\overline{\nabla}_{\mathbf{u}} \mathbf{u}$ is the symmetric part of the gradient of \mathbf{u} . Notice that in accordance

with the assumption of slow fluid motion, the quadratic convective term in the inertial forces has been neglected in the momentum balance equation (5b). The second term in the right-hand side of the momentum balance equation (5b) physically corresponds to the electric pressure exerted by the ionic charge on the electrolyte fluid, and mathematically represents the coupling between electrodiffusive ion transport and electrolyte fluid motion. The initial condition for electrolyte fluid velocity is

$$\mathbf{u}(\mathbf{x},0) = \mathbf{u}^0(\mathbf{x}) \qquad \qquad \text{in } \Omega, \tag{6a}$$

where the function \mathbf{u}^0 is a given datum, usually set equal to zero. The boundary conditions for the Stokes system are:

$$\mathbf{u} = \mathbf{g} \qquad \text{on } \Gamma_D^{\mathbf{u}}, \qquad (7a)$$
$$\underline{\sigma}(\mathbf{u}, p)\mathbf{n} = \mathbf{h} \qquad \text{on } \Gamma_N^{\mathbf{u}}, \qquad (7b)$$

$$\mathbf{p}(\mathbf{u},p)\mathbf{n} = \mathbf{h}$$
 on $\Gamma_N^{\mathbf{u}}$, (7b)

where g is the imposed velocity on the side $\Gamma_D^{\mathbf{u}}$ and $\mathbf{h} = -\overline{p}\mathbf{n}$ is the value of the traction on $\Gamma_N^{\mathbf{u}}$. Relation (7a) is the wall adherence condition (typically $\mathbf{g} = \mathbf{0}$) and (7b) is the application of the action-reaction principle at the boundary.

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