

1 **Immunohistochemical investigations on *Brucella ceti*-infected, neurobrucellosis-**  
2 **affected striped dolphins (*Stenella coeruleoalba*)**

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4 Gabriella Di Francesco, Antonio Petrini, Anna Rita D'Angelo, Ludovica Di Renzo, Mirella Luciani,  
5 Tiziana Di Febo, Enzo Ruggieri, Antonio Petrella, Carla Grattarola, Barbara Iulini, Osvaldo  
6 Matteucci, Giuseppe Lucifora, Eva Sierra, Antonio Fernández, Roberto Giacomini, Roberto Stuffer,  
7 Clotilde Angelucci, Marina Baffoni, Giovanni Di Guardo, Manuela Tittarelli

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10 **Keywords:** *Brucella ceti*, Immunohistochemistry, Monoclonal antibody, Neurobrucellosis, *Stenella*  
11 *coeruleoalba*.

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13 **Summary**

14 Bacteria of the genus *Brucella* cause brucellosis, an infectious disease common to humans as well as  
15 to terrestrial and aquatic mammals. Since 1994 several cases of *Brucella* spp. infection have been  
16 reported in marine mammals worldwide. Indeed, since human brucellosis ranks as one of the most  
17 common bacterial zoonotic infections on a global scale, it is necessary to increase our knowledge  
18 about it also in the marine environment. *Brucella ceti*, which is phenotypically similar to other smooth  
19 brucellae as *B. abortus* and *B. melitensis*, shares with the latter two the same surface antigens that are  
20 routinely used for the serological diagnosis of *Brucella* spp. infection. Marine mammal *Brucella* spp.  
21 infections are characterized by a pathogenicity similar to their terrestrial counterparts, with the  
22 occurrence of abortion, stillbirth and orchitis and an involvement of the host's central nervous system  
23 (CNS), similarly to what happens in mankind. While sero-epidemiological data suggest that *Brucella*  
24 spp. infection is widespread globally, detecting *Brucella* spp.-associated antigens by  
25 immunohistochemistry (IHC) in tissues from infected animals is often troublesome. The present study  
26 was aimed at investigating, by means of IHC based upon the utilization of an anti*Brucella* LPS  
27 monoclonal antibody (MAb), the CNS immunoreactivity (IR) shown by *B. ceti*-infected,  
28 neurobrucellosis-affected striped dolphins.

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33 **Riassunto**

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35 I batteri del genere *Brucella* causano la brucellosi, una malattia infettiva comune all'uomo e ai  
36 mammiferi terrestri e acquatici. Dal 1994 diversi casi d'infezione da *Brucella* sono stati segnalati nei  
37 mammiferi marini in tutto il mondo. Poiché la brucellosi umana è considerata una delle più comuni  
38 zoonosi su scala mondiale, è necessario migliorare le nostre conoscenze anche in ambiente marino.  
39 *Brucella ceti*, fenotipicamente simile ad altre brucelle lisce come *B. abortus* e *B. melitensis*, ha in  
40 comune con queste ultime i medesimi antigeni di superficie, che vengono peraltro utilizzati nella  
41 diagnosi sierologica dell'infezione. I mammiferi marini infetti ad opera di *Brucella* spp. mostrano  
42 reperti lesivi analoghi a quelli osservati nei mammiferi terrestri, uomo compreso, con presenza di  
43 aborti, natimortalità, orchite e neurobrucellosi. Se da un lato i dati siero-epidemiologici suggeriscono  
44 che l'infezione da *Brucella* spp. è praticamente cosmopolita, la rilevazione mediante tecniche  
45 d'indagine immunostochimica di antigeni brucellari nei tessuti di animali infetti è spesso  
46 problematica. Obiettivo del presente studio è stato quello di valutare, mediante l'impiego di un  
47 anticorpo monoclonale nei confronti dell'antigene LPS di *Brucella* spp, l'immunoreattività del  
48 sistema nervoso centrale in esemplari di stenella striata (*Stenella coeruleoalba*) *B. ceti*-infetti affetti  
49 da neurobrucellosi.

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52 **Parole chiave:** Anticorpo monoclonale *Brucella ceti*, Immunostochimica, Neurobrucellosi, *Stenella*  
53 *coeruleoalba*.

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55 *Brucella ceti* was first isolated from an aborted bottlenose dolphin (*Tursiops truncatus*) fetus in 1994  
56 (Ewalt *et al.*, 1994) and, since then, several cases of infection have been reported among free-ranging  
57 cetaceans worldwide (Guzman-Verri *et al.*, 2012). The first case of *B. ceti* infection in the  
58 Mediterranean was recorded only in 2009 (Isidoro-Ayza *et al.*, 2014), with the first case of *B. ceti*  
59 infection having been recorded along the Italian coastline in 2012 (Alba *et al.*, 2013). Besides the  
60 “classical” species, some recently discovered *Brucella* species have also demonstrated a zoonotic  
61 potential, as in the case of *B. ceti* (Whatmore *et al.*, 2008, De Massis *et al.*, 2019). *Brucella* spp.  
62 infection in marine mammals is characterized by a pathogenicity similar to that of natural infection  
63 models in terrestrial mammals. In addition, a documented involvement of the central nervous system  
64 (CNS) in the striped dolphin (*Stenella coeruleoalba*), similarly to what described in the human  
65 species, has been reported (Guzman-Verri *et al.*, 2012), with neurobrucellosis having not been  
66 recorded in bovine, caprine, ovine, swine, or canine hosts. Nevertheless, this syndrome is a relatively  
67 common feature in non-treated human brucellosis-affected patients (Obiako *et al.*, 2010) Therefore,  
68 cetacean neurobrucellosis may serve as an interesting comparative neuropathology and  
69 neuropathogenesis model to understand how the bacterium is capable to cross the blood-brain barrier,  
70 thereby giving rise to host’s CNS invasion. While sero-epidemiological data suggest that *Brucella*  
71 infection is widespread globally (Nymo *et al.*, 2011), detecting *Brucella* spp.-associated antigens by  
72 immunohistochemistry (IHC) in tissues from naturally or experimentally infected animals is often  
73 troublesome The present study was aimed at investigating, by means of an anti- *Brucella* LPS  
74 monoclonal antibody (MAb), the CNS immunoreactivity (IR) shown by *B. ceti*-infected,  
75 neurobrucellosis-affected striped dolphins, along with its comparative evaluation in a range of fetal  
76 tissues from *B. abortus*- and *B. melitensis*- infected ruminants.

77 A MAb raised against *Brucella* LPS was produced at Istituto Zooprofilattico Sperimentale  
78 dell’Abruzzo e Molise “G. Caporale” (IZSAM) and characterized by Western blotting (WB) and  
79 indirect ELISA according to Di Febo *et al* (2012) and Portanti *et al* (2006) , being subsequently

80 characterized against *B. abortus* RB51, *B. pinnipedialis* and *B. ceti*, which were not tested in the past  
81 experiments. Samples of lung, liver and placental tissues from 16 ovine fetuses originating from 15  
82 ewes experimentally infected with *B. melitensis* biotype 3, along with samples of lung, liver and  
83 placental tissues from 6 additional aborted fetuses carried by sheep belonging to *Brucella*-free flocks,  
84 were preliminarily investigated, 20 years ago, against *Brucella* spp. The study was subsequently  
85 enhanced, in recent years, through the inclusion of 9 cases of *B. ceti* infection in striped dolphins, 8  
86 of which were found stranded between 2012 and 2019 along the Italian coastline, while the remaining  
87 individual was found beached ashore in Canary Islands (Spain) in 2004.

88 The dolphin tissues were collected during *post mortem* examination, in tight agreement with the  
89 investigation protocols to be performed in the framework of the Italian National Stranding Network  
90 (INSN) for standard laboratory investigations on stranded cetacean specimens.

91 Positive and negative controls were included in each IHC run, with the positive ones being  
92 represented by the lung, liver and placental tissues from ovine and bovine fetuses either naturally or  
93 experimentally infected by *Brucella* spp. The brain from a *Dolphin Morbillivirus*-infected striped  
94 dolphin was additionally used as negative control tissue. Further negative controls were represented  
95 by tissue sections obtained from the 7 immunohistochemically positive, *B. ceti*-infected striped  
96 dolphins under study, from which the primary anti-*Brucella* Ab was omitted. More in detail, *Brucella*  
97 IHC was carried out using the MAbs 4B5A against LPS *Brucella* diluted 1:10 to 1:100. Tissue sections  
98 were previously heat treated for antigen retrieval (at 121°C for 8 minutes) in 0.01 M citrate buffer,  
99 pH 6.0. Immunoreactions were then visualized by means of a peroxidase technique (Envision Plus  
100 Kit, Dako at IZSAM and Vectastain *elite* ABC kit standard Vector at the Faculty of Veterinary  
101 Medicine, University of Teramo, Italy).

102 The *Brucella* spp. isolation and identification procedures were performed in accordance with the  
103 technique described in the OIE Manual of Diagnostic Tests and Vaccines (World Organisation for  
104 Animal Health, 2017). With the only exception of the two individuals in which *B. ceti* infection was

**Commentato [R1]:** Numbering is not indicated in the Figure files in my hand. No figure legends are present in the manuscript. Please provide this information. Moreover, you mention Fig. 2 before Fig. 1 in the text. Numbering of figures should be therefore revised.

105 diagnosed only by means of biomolecular and IHC techniques (ID 1,5 Table III). All the tissue  
106 samples were routinely processed for histopathology and *Brucella* immunohistochemistry (IHC),  
107 whose reliability and reproducibility were also evaluated by means of an “inter-laboratory  
108 comparison”, which involved two independent Pathologists (based at IZSAM and at Faculty of  
109 Veterinary Medicine, University of Teramo, Italy respectively) (Table III).

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111 The results of the characterization of 4B5A MAb are shown in Table I (i-ELISA) and Figure 1 (WB).  
112 Both i-ELISA and WB confirmed that the aforementioned MAb reacted with *Brucella* smooth  
113 strains, with the typical LPS-ladder pattern exhibited in WB analysis. Conversely, the same MAb did  
114 not react with rough *Brucella* strains (Table I). The results of IHC investigations are reported in  
115 Tables II and III. More in detail, *Brucella* spp.-associated antigens were detected in pulmonary  
116 necrotic cell debris as well as in the cytoplasm of both alveolar macrophages and neutrophils from  
117 the *B. abortus*-infected bovine fetuses (Fig. 2) as well as in liver cells from the *B. melitensis*-infected  
118 ovine fetuses under study (Fig. 3). Within the CNS from *B. ceti*-infected dolphins, macrophage-like  
119 cells were seen harbouring more or less consistent loads of microbial antigen (Figs 4 and 5). Neither  
120 background staining nor artifacts or positive IR were observed in negative control tissues.

121 The results obtained in the present study clearly showed a strong IR against *Brucella* LPS in tissues  
122 from all the *Brucella* spp.-infected, herein investigated bovine, ovine and striped dolphins (7 out 9  
123 individuals) (Fig. 3,4). In this respect, the negative IHC results observed in the CNS from 2 *B. ceti*-  
124 infected dolphins may be due either to the low sensitivity of *Brucella* spp. IHC when low bacterial  
125 concentrations are present in infected tissues, or to the different neuro-topographical concentrations  
126 of *B. ceti* organisms, not coincident with that of the microscopic field under study. An additional factor  
127 to be considered refers to the experimental conditions used in a portion of this work, that are  
128 counterparted by the “field conditions” under which *post mortem* examinations are routinely carried

**Commentato [R2]:** Table II: I would suggest to merge the columns “Tested” and “IHC POS” in order to present results as “16/26” for positives. These 26 ovine tissues were previously tested as *Brucella* positive (so how do you explain negative results for 10?) or they were tested for the first time in this study?

**Commentato [R3]:** Table III: I would suggest to make explicit in the table the country and year of beaching (now it can only be inferred by the ID).

129 out on stranded cetacean specimens, including also the adverse effects exerted by *post mortem*  
130 autolysis.

131 Based upon the herein presented results, *Brucella* spp. IHC should be regarded as a laboratory  
132 procedure which is useful not only when analyzing ovine and bovine infected tissues, but also in  
133 the case of *B. ceti*-infected, neurobrucellosis-affected striped dolphin CNS tissue specimens (in which  
134 macrophage-like cells were seen harbouring more or less consistent loads of bacterial antigen),  
135 providing a method capable of achieving a direct and reliable IHC diagnosis of *Brucella* infection.  
136 Furthermore, and not less important, the consistent background and the non-specific reactions  
137 observed when using an anti-*Brucella* -polyclonal Ab were not seen when MAb 4B5A was used.  
138 The additional knowledge provided by this study on the detection of *Brucella* infection in cetaceans  
139 may be helpful not only from a diagnostic standpoint but also for increasing our awareness on the  
140 (neuro)pathogenesis of *Brucella* infection in aquatic mammals and, not less important, also from a  
141 public health viewpoint, considering the documented zoonotic potential of *Brucella* microorganisms.  
142 Moreover, MAb 4B5A anti *Brucella* LPS could represent a diagnostic and research laboratory  
143 reagent, whose use may be highly recommended also for the IHC diagnosis and pathogenetic  
144 characterization of *B. ceti* and *B. pinnipedialis* infections in cetaceans and in pinnipeds, respectively.

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## Conflicts of Interest

151 The Authors declare that they have no competing interests in regard to this study.

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## Figures' Legends

200 Western blotting of Mab 4B5A vs *Brucella abortus* S99 (Lane 1), *Brucella melitensis* 16M (Lane  
201 2), *Brucella suis* biotipo 1 (Lane 3), *Brucella ceti* (Lane 4), *Brucella pinnipedialis* (Lane 5) (Fig.1)

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203 *Brucella* spp.-associated antigen positive immunohistochemical labeling in bovine fetal lung (Fig. 2),  
204 in ovine fetal liver (Fig. 3) as well as in CNS (cervical spinal cord) tissues (Figs 4 and 5) from  
205 neurobrucellosis-affected, *B. ceti*-infected striped dolphins. *Brucella* spp. IHC with MAb 4B5A,  
206 Mayer's hematoxylin counterstain, different magnifications.

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