## Aerial Monitoring of Rice Crop Variables using an UAV Robotic System

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- Keywords: UAV, Precision Agriculture, Image Processing, Vegetative Indices, Multispectral Imagery, Machine Learning.
- Abstract: This paper presents the integration of an UAV for the autonomous monitoring of rice crops. The system integrates image processing and machine learning algorithms to analyze multispectral aerial imagery. Our approach calculates 8 vegetation indices from the images at each stage of rice growth: vegetative, reproductive and ripening. Multivariable regressions and artificial neural networks have been implemented to model the relationship of these vegetation indices against two crop variables: biomass accumulation and leaf nitrogen concentration. Comprehensive experimental tests have been conducted to validate the setup. The results indicate that our system is capable of estimating biomass and nitrogen with an average correlation of 80% and 78% respectively.

### **1 INTRODUCTION**

The use of Unmanned Aerial Vehicles (UAV) in the solution of agriculture-related problems spans a wide range of operations such as crop variable measurement (Gevaert et al., 2015),(Gago et al., 2015) fruit detection (Carrijo et al., 2017), crop plot detection (Hongli et al., 2017), crop yield improvement (Arroyo et al., 2017) and crop mapping (Guo et al., 2012), (Khanna et al., 2015). In (Dongyan Zhang, Xingen Zhou, Jian Zhang, Linsheng Huang and Zhao, 2017) the sheath blight fungus disease of rice was detected using a Phantom 2 UAV coupled with a highresolution RGB multispectral camera (Micasense). Pix4D software was used to generate 2D and 3D georeferenced maps and to compute different vegetation indices. In (Yong et al., 2016), (Lu et al., 2015), UAVs were used for nitrogen estimation and chlorophyll quantification (Uto et al., 2013) using hyperspectral sensors.

Multispectral images have been used in plant analysis for some time now (Naito et al., 2017). Initially, few vegetation indices were commonly used, mainly the Normalized Difference Vegetation Index (NDVI) for estimating above ground biomass. However, due to the presence of external factors such as water, soil backgrounds and the difference of the crop at each stage of growth, different vegetative indices were introduced, such as the Soil-Adjusted Vegetation Index (SAVI), the Modified SAVI (MSAVI), the Modified Chlorophyll Absorption Ratio Index (MCARI) and the Modified Triangular Vegetation Index (MTVI), among others (Gnyp et al., 2014).

The development of an UAV-based crop monitoring system involves multiple challenges. In hardware terms the UAV requires sufficient autonomy to cover the crop area, also adequate on-board store capacity for high-resolution data, as well as low-weight sensors. The integration of the native UAV hardware with external sensors requires a significant interplay between hardware and software in order to guarantee data reliability, online processing and simple end-used experience. In previous work (Devia et al., 2019), we tackled these challenges by developing an autonomous UAV robotic system to monitor biomass dynamics based on NIR information. In this paper, we present a preliminary approach to also monitor leaf nitrogen concentration during the three main stages of plant growth: vegetative, reproductive and ripening. Our goal is to estimate biomass dynamics and nitrogen variations by calculating several vegetation indices from multispectral data. To this purpose, we present the robotic system architecture detailed in Figure 1. Two UAVs have been setup with both NIR and thermal cameras (upcoming work is oriented towards cooperative-UAV monitoring). For instance,

each robot captures a dataset of images independently and the processing is done in a PC base-station.

### 2 METHODS

### 2.1 UAV Robotic System

As mentioned, two UAVs were integrated: The AS-CTEC Pelican<sup>1</sup> and the DJI Phantom<sup>2</sup>. Also, the following equipment was used:

- ASCTEC Pelican:
  - ASCTEC autopilot.
  - Mastermind board for image processing and data acquisition.
  - Tetracam ADC-Lite multispectral camera: this camera captures visible light wavelengths longer than 520nm and near-infrared (NIR) wavelengths up to 920nm. The multispectral camera is located in the bottom of the drone and aligned with the center of mass.
- DJI Phantom:
  - DJI autopilot.
  - Parrot Sequoia multispectral camera<sup>3</sup>: his camera comes with 4 different sensor to capture image in different wavelengths: red, green, NIR and Red-Edge.
  - Flir duo R thermal camera<sup>4</sup> with  $640 \times 512$  in resolution with  $32^{\circ}C$  field of view. It incorporates a radiometric sensor operating at sampling frequency of 30Hz and capable of sensing the crop temperature between  $-20^{\circ}C$  and  $50^{\circ}C$  with a resolution of  $\pm 5^{\circ}C$ .

The standard GPS waypoint navigation for both drones comes with a graphical user interface -GUI that allows for autonomous take-off, waypoint trajectory following and landing. In this work, the flight planning was enhanced with an additional module created for the image capturing planning process. This new module creates a GPS grid with the positions were the images must be taken in order to ensure appropriate image resolution, crop plot coverage and sufficient overlapping of images for ensuring reliable data-mapping of the crop. In this sense. our photo planner algorithm uses geometric parameters of the

<sup>1</sup>http://www.asctec.de/en/uav-uas-drones-rpas-roav/ asctec-pelican/ NIR/thermal camera and the crop plot area to generate the cartesian points.

The required UAV altitude is also estimated in the photo planning algorithm. This is done by using the camera's field of view in each axis and the desired image resolution. Figure 2 presents experimental results regarding aerial crop coverage and geo-mapping of crop plots by applying classical image mosaicing techniques. The aforementioned methods were presented in previous works reported in (Rojas et al., 2017), (Jose et al., 2016), and (Rojas et al., 2018).

#### 2.2 Crop Variable Estimation

This section addresses the challenges associated with NIR and thermal image processing to properly compute Vegetative Indices (VI). It mainly consists in three different stages:

- 1. Image processing: By taking raw images from the NIR camera, the system determines the crop area (plots) that are suitable for VI analysis. In this process we apply perspective corrections to the images (using drone's IMU information) and filtering strategies to remove the background and noise. Finally, NIR image segmentation is applied to extract the crop plot (parcel) of interest, as detailed by Figure 3(a).
- 2. Vegetation index computation: VI are well-known formulas that use the reflectance of the plants in different wavelengths to provide information about the health state of the plant. Table 1 details the VI used in this work and their corresponding formulas.
- 3. Machine Learning: Once the vegetative indices are computed, we applied Multivariable Regression models (MR) to calculate the accumulated biomass. Only polynomial models were considered using the vegetative indices as the independent variables. Furthermore, we also used Artificial Neural Networks (ANN) to estimate leaf nitrogen concentration by training our system during the three main stage of rice growth: vegetative, reproductive and ripening. Both MR an ANN models required a ground-truth dataset for training. In this sense, biomass and nitrogen were directly measured from the rice crop by following the traditional sampling method. For biomass, 1 linear meter of plants were cut from the ground, as detailed by Figure 3(b). Plants were sampled and weighted, then put in the oven at 65 degrees Celsius for 4 days or until a constant weight was reached. For nitrogen, we used a SPAD sensor (Soil-Plant Analyses Development) to directly measured leaf nitrogen.

<sup>&</sup>lt;sup>2</sup>https://www.dji.com/phantom-4

<sup>&</sup>lt;sup>3</sup>https://www.parrot.com

<sup>&</sup>lt;sup>4</sup>https://www.flir.com/products/duo/

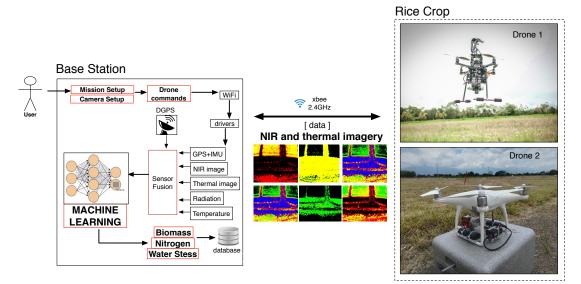


Figure 1: Multi-UAV robotic system architecture for crop monitoring.

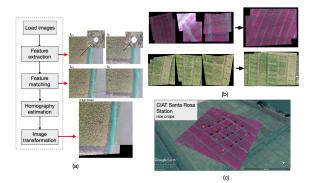


Figure 2: (a) image mosaicing flow diagram: SURF and ORB algorithms were implemented for feature extraction. The FLANN Fast (Library for Approximate Nearest Neighbors) algorithm was used for matching, whereas RANSAC (Random Sample Consensus) algorithm was used to create the homography for coordinate frame transformations. (b) Geo-referenced mosaicing results of applying the steps from (a) to both NIR and RGB images. (c) resulting NIR digital map of the crop and the trajectory followed by the drone.

### **3 EXPERIMENTAL RESULTS**

Experiments were carried out during 2017 and 2018 in the rice farms of the Center of International Agriculture -CIAT, located in the Department of Meta, Colombia: latitude  $4^{o}1'37.85''N$  with Longitude  $73^{o}28'28.65''W$ . Figure 4 shows experimental results for several weeks of in-field testing. Both MR and ANN methods have been used for the estimation of the crop variables based on NIR imagery. On average, the former, denoted as *Estimation* 1, Table 1: NIR Vegetation Indices used ( $\rho_f$  denotes the reflectance of the for the frequency f) (Gnyp et al., 2014).

Name	Equation
Normalized	$\frac{\rho_{780}-\rho_{670}}{\rho_{780}+\rho_{670}}$
Difference Veg-	1780 1070
etation Index	
-NDVI	
Green Normal-	$\frac{\rho_{780} - \rho_{500}}{\rho_{780} + \rho_{500}}$
ized Difference	. 700 . 500
Vegetation	
Index -GNDVI	
Soil-Adjusted	$(1+L)\left(\frac{\rho_{800}-\rho_{670}}{\rho_{800}+\rho_{670}+L}\right)$ with L = 0.5
Vegetation	
Index -SAVI	
Modified SAVI	$\frac{1}{2}\left(2\rho_{800}+1-\sqrt{(2\rho_{800}+1)^2-8(\rho_{800}-\rho_{670})}\right)$
-MSAVI	· · · · · · · · · · · · · · · · · · ·
Corrected	$\frac{\frac{\text{NDVI}+0.5}{\text{NDVI}+0.5}}{\sqrt{\text{NDVI}+0.5}}$
Transformed	
Vegetation	
Index - CTVI	
Difference Veg-	$ ho_{780} -  ho_{670}$
etation Index -	
DVI	

achieves more accurate estimation compared against the ground-truth value, denoted as *Measurement*. The results are consistent for the three stages evaluated: vegetative, reproductive and ripening. The left plots of Figure 4 show the biomass and nitrogen estimation values respectively. Abrupt variations of the signals (e.g. sampled image 225 in the horizontal axis) indicates a change of crop stage from vegetative to reproductive. In this sense, the time evolution window of both crop variables is about 3 months.

Using the VIs and the metadata of the corresponding images, the multilinear regression models

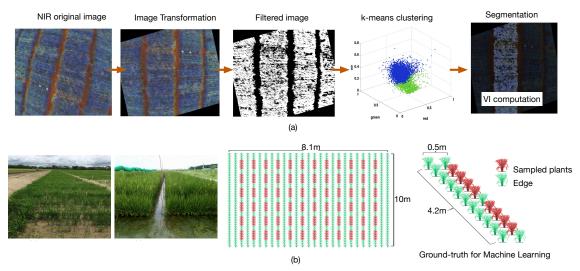


Figure 3: (a) Experimental results regarding NIR image pre-processing algorithms. Classical image clustering based on kmeans has enabled to separate NIR pixels into two cluster: plant and no-plant. This allows the image background subtraction to properly extract the parcel of interest. (b) Ground-truth dataset for machine learning training.

Plot/parcel	Repetition	Fresh weight [g]	Dry weight [g]	Water Content [g]	SPAD/nitrogen
13B1	1	656	118	82.01	47.70
38B4	1	394.00	76.00	80.71	45.30
63B1	1	450.00	88.00	80.44	34.27
88B4	1	814.00	100.00	87.71	41.23
13B5	2	792.00	136.00	82.83	44.37
38B8	2	420.00	76.00	81.90	46.53
63B5	2	646.00	120.00	81.42	43.07
88B8	2	550.00	104.00	81.09	40.30
13B9	3	626.00	126.00	79.87	36.63
38B12	3	656.00	110.00	83.23	44.10
63B9	3	428.00	76.00	82.24	38.13
88B12	3	540.00	108.00	80.00	43.63

Table 2: Numerical values for biomass and nitrogen/SPAD in vegetative stage directly measured from the crop (ground-truth values).

were performed for every possible combination of vegetative indices. Strong linear dependencies were found between the VIs from Table 1 and the dynamics of biomass and nitrogen. For instance, linear multivariable regressions of the form:  $\beta_c \alpha_c + \beta_{SR} SR \alpha_{SR} +$  $\beta_{\text{NDVI}}$ NDVI $\alpha_{NDVI}$  +  $\beta_{\text{GNDVI}}$ GNDVI $\alpha_{GNDVI}$  +  $\beta_{\text{CTVI}}$ CTVI $\alpha_{CTVI}$  $\beta_{SAVI}SAVI\alpha_{SAVI}$ ++ $\beta_{\text{DVI}} \text{DVI} \alpha_{DVI} + \beta_{\text{MSAVI}} \text{MSAVI} \alpha_{MSAVI}$  were used to estimate both crop variables, computing the 7 VIs with a constant coefficient, where  $\alpha_c$ ,  $\alpha_{SR}$ ,  $\alpha_{NDVI}$ ,  $\alpha_{GNDVI}$ ,  $\alpha_{CTVI}$ ,  $\alpha_{SAVI}$ ,  $\alpha_{DVI}$  and  $\alpha_{MSAVI}$  take the value of 0 or 1. For each coefficient combination (VIs and the constant term), two images were taken randomly from each plot. This was done since not all the plots had the same number of photos, so if all the photos were used, the resulting regression could have some bias.

For each stage of the crop (vegetative, reproduc-

tive and ripening), we performed 20 flights, capturing around 2,000 images per stage, yielding a dataset of 6,000 images per trial. Overall, around 18,000 images were processed for the estimation of biomass and nitrogen. Both MR and ANN methods used a training set of NIR images accounting for the 60% of the entire database, whereas the remaining 40% for testing and validation. On the other hand, the correlations between estimated data and the ground-truth measurements are shown in the histograms depicted in the right plots of Figure 4. On average, our system is capable of estimating biomass and nitrogen with a correlations above 80% were achieved during the first vegetative stage of the crop. During this stage, the

<sup>&</sup>lt;sup>5</sup>The following video illustrates the steps performed during the experiments: https://youtu.be/BTwD4GduXDo

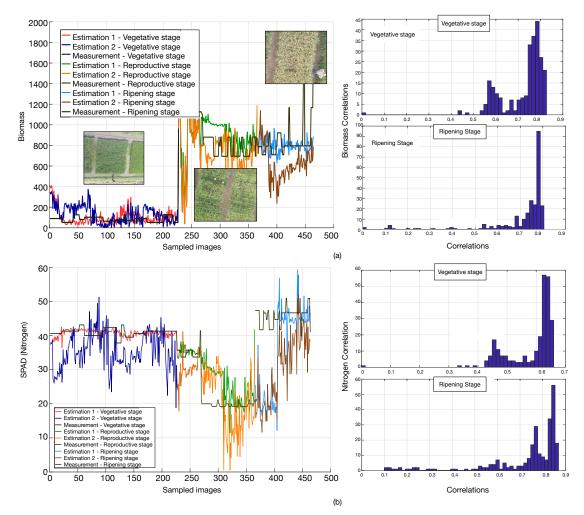


Figure 4: Experimental results (a) Biomass estimation (dry weight), (b) Nitrogen estimation. Left plots compare the estimated variables (labels Estimation 1 and 2) against the ground-truth labeled as Measurements. For nitrogen, this values is directly measured using a SPAD device. The time evolution window of both crop variables is about 3 months: vegetative, reproductive and ripening. Right plots contain the histogram information regarding the correlations achieved during the experiments.

green color of the plants is predominant, as observed by the inset in Figure 4(a). Therefore, our methods can be properly calibrated based on accurate VIs calculated from NIR image reflectances. Numerical values of the ground-truth measurements for the vegetative stage are consigned in Table 2. The reproductive stage is the more critical for our estimations algorithms since panicle formations yield yellow colors in the images. The merge of both green and yellow colors difficult the training, specially during the clustering phase where only two clusters are conformed: plants (green pixels) and no-plant (soil) cf. Figure 3(a). Larger fluctuations in biomass and nitrogen estimations occur in this crop stage, as observed in both Figure 4(a)(b) left plots. Lastly, in ripening stage, the yellow color becomes predominant, allowing a proper calibration and therefore accurate estimation of the crop variables.

### **4** CONCLUSIONS

This paper presented the integration and deployment of an UAV system for rice crop monitoring. By using multivariable regressions and neural networks, our system was able to estimation biomass and nitrogen dynamics during the time evolution of the crop, concretely for three stages: vegetative, reproductive and ripening. From the specialized literature, we identified the set of vegetation indices that were sensible to biomass and nitrogen variations according to NIR image reflectance properties. We combined those indices to achieve the estimations of both values. A comprehensive field testing of the proposed system enabled us to calculate correlations between the estimations and the in-field measurements of the crop variables. Since the vegetation indices tend to be evolve linearly during the crop growth, we achieved accurate correlations using multivariable regressions; on average, correlations of 80% for biomass and 78% for nitrogen were achieved. Upcoming work is oriented towards improving the correlations by including more sophisticated image classification and clustering algorithms to consider several feature spaces for the NIR pixels. By now, our system is not reliable during the reproductive stage of the crop due to the mixed plant color in between yellow and green. Also, different genotypes of rice varieties are planted in the same plot area. In this sense, we also expect to improve on the estimation, since the biomass and nitrogen readings are highly dependent of the plant variety.

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