



**UNIVERSITI PUTRA MALAYSIA**

***PATHOGENICITY OF STREPTOCOCCUS AGALACTIAE IN JUVENILE  
RED TILAPIA (OREOCHROMIS SP.) FROM A FISH FARM IN  
SELANGOR, MALAYSIA***

**ALI FARAG ABUSELIANA**

**FPV 2011 2**

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**MASTER OF VETERINARY SCIENCE  
UNIVERSITI PUTRA MALAYSIA**

**2011**

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RED TILAPIA (*OREOCHROMIS SP.*) FROM A FISH FARM IN  
SELANGOR, MALAYSIA**

**By**

**ALI FARAG ABUSELIANA**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfilment of the Requirements for the  
Degree of Master of Veterinary Science**

**March 2011**

## DEDICATION

*With appreciation and respect, this thesis is dedicated*

*To my parents and family,*

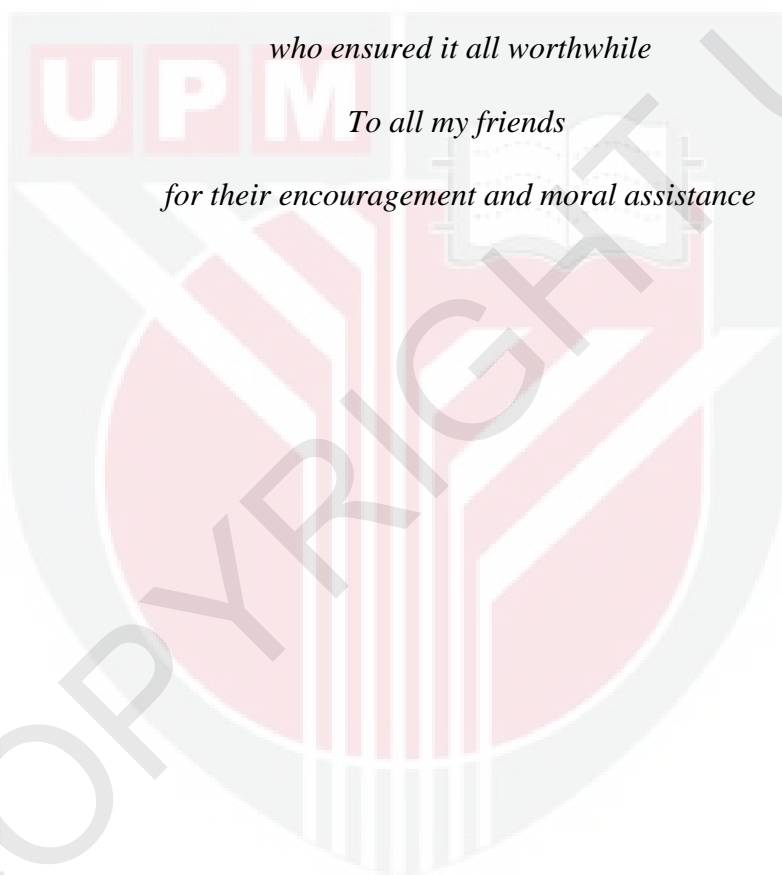
*for their understanding, encouragement and constant support*

*To all my supervisors,*

*who ensured it all worthwhile*

*To all my friends*

*for their encouragement and moral assistance*



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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Veterinary Science

**PATHOGENICITY OF *STREPTOCOCCUS AGALACTIAE* IN JUVENILE RED TILAPIA (*OREOCHROMIS SP.*) FROM A FISH FARM IN SELANGOR, MALAYSIA**

By

**ALI FARAG ABUSELIANA**

**March 2011**

**Chairman: Prof. Madya Hassan Hj Mohd Daud, PhD**

**Faculty : Veterinary Medicine**

Streptococcal infection is one of the emerging bacterial diseases that was reported to cause significant mortality and high economical loss in freshwater and saltwater fish species including *Tilapia sp.*, worldwide. Recently, a streptococcosis outbreak affecting Red tilapia (*Oreochromis sp.*) farm in Selangor was investigated. Affected fish showed loss of appetite, cornea opacity and serpentine swimming. Healthy and morbid fish were clinically examined. Samples from brain, liver, spleen and kidney were collected for causal agent isolation. Pure bacteria isolates were successfully isolated on trypticase soy agar (TSA) blood agar (BA) and brain heart infusion agar (BHIA). The colonies were of grayish white color, circular, convex, pin-head size and  $\beta$ -haemolytic. All isolates were gram-positive cocci, oxidase-negative and catalase-negative. They were identified as group B *Streptococcus agalactiae* (GBS) using a commercial identification kit (Streptococcal grouping kit, RapID™ STR System and BBL Crystal GP ID kit). Specific polymerase chain reaction (PCR) and 16S rRNA sequencing technique confirmed the isolates as GBS. The isolates were sensitive to amoxicillin, ampicillin, erythromycin, chloramphenicol, linomycin,

rifampicin, vancomycin, gentamicin, sulfamethoxazole + trimethoprim and tetracycline. On contrary, they were resistant to neomycin, amikacin, kanamycin and streptomycin. The 120 hours median lethal dose (LD<sub>50</sub>) value in juvenile tilapia injected intraperitoneally (IP) was  $1.5 \times 10^5$  cfu/mL. Experimental infections were carried out by bathing the fish for 30 minutes in water containing the bacteria and by intraperitoneal (IP) injection. It was observed that IP route was more potent to cause mortality to juvenile Red tilapia and produced clear clinical signs within five days. It was noted that the mortality started to reduce after five days and fish recovered after nine days post inoculation. In contrast, immersion route did not induce mortality, but produced moderate clinical signs such as lethargy and loss of appetite, and fish started to recover after six days. The findings of the current study indicated that *S. agalactiae* infection started to become an issue in tilapia farms and warrants focusing to formulate a suitable measure to prevent and control the disease before it becomes endemic in the future.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk jatah Master Sains Veterinar

**KEPATOGENAN *STREPTOCOCCUS AGALACTIAE* TERHADAP IKAN  
TILAPIA MERAH JUVENIL (*OREOCHROMIS SP.*) DARIPADA SEBUAH  
LADANG IKAN DI SELANGOR, MALAYSIA**

Oleh

**ALI FARAG ABUSELIANA**

**Mac 2011**

**Pengerusi: Prof. Madya Hassan Hj Mohd Daud, PhD**

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Jangkitan Streptococcal merupakan salah satu penyakit bakteria yang dilaporkan menyebabkan kematian dan kerugian ekonomi yang tinggi dalam spesies ikan air tawar dan masin termasuk spesies tilapia di seluruh dunia. Baru-baru ini, satu wabak penyakit streptokokosis melibatkan kultur ikan tilapia merah (*Oreochromis sp.*) di Selangor telah disiasat. Ikan yang dijangkiti menunjukkan tanda kehilangan selera makan, berenang bengkok-bengkok dan kekaburan kornea. Ikan yang sihat dan sakit disiasat secara klinikal dan sampel diambil dari otak, hati, limfa dan ginjal untuk pemencilan agen etiologiknya. Isolat kultur tulen berjaya dikultur di atas agar TSA dan darah. Koloni-koloni di atas agar TSA adalah berwarna putih kekelabuan, bulat, cembung, bersaiz pin dan  $\beta$ -hemolitik. Semua isolat adalah kokus, Gram positif, oksidase dan katalase negatif. Ia dikenalpasti sebagai kumpulan B *Streptococcus agalactiae* (GBS) menggunakan kit pengenalan komersil (Streptococcal grouping kit, RapID™ STR System dan BBL Crystal GP ID kit). Tindakbalas berantai polimerase (PCR) dan jujukan 16S rRNA mengesahkan isolat-

isolat sebagai GBS. Isolat-isolat adalah sensitif terhadap amoksisilin, ampicillin, eritromisin, kloramfenikol, linomisin, rifampisin, vancomisin, gentamisin, sulfamethoxazol + trimethoprim dan tetrasiklina. Sebaliknya, ia rentang terhadap neomisin, amikasin, kanamisin dan streptomisin. Nilai dos kematian median 120 jam ( $LD_{50}$ ) dalam anak ikan tilapia merah yang disuntik secara intraperitoneal (IP) adalah  $1.5 \times 10^5$  cfu/mL. Jangkitan eksperimen dicapai dengan merendam ikan selama 30 minit dalam air mengandungi bakteria dan secara suntikan intraperitoneal. Ia dapat dilihat bahawa jangkitan IP lebih berkuasa menyebabkan kematian terhadap anak ikan tilapia merah dan menunjukkan tanda-tanda klinikal yang jelas dalam masa lima hari. Ia dapat dilihat juga kematian mulai berkurangan selepas lima hari dan ikan pulih selepas sembilan hari pasca inokulasi. Sebaliknya, rendaman tidak mengaruhkan kematian, tetapi menghasilkan tanda-tanda klinikal sederhana seperti kelesuan dan kehilangan selera makan, dan ikan mulai pulih selepas enam hari. Hasil daripada kajian ini menunjukkan bahawa jangkitan *S. agalactiae* telah mula menjadi isu pada ternakan tilapia dan memerlukan kita mengambil langkah yang sesuai untuk mengelak dan mengawal jangkitan ini sebelum menjadi endemik pada masa hadapan.



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I certify that a Thesis Examination Committee has met on 24 March 2011 to conduct the final examination of Ali Farag Abuseliana on his thesis entitled "Pathogenicity of *Streptococcus agalactiae* in Juvenile Red Tilapia (*Oreochromis sp.*) from a Fish Farm in Selangor, Malaysia" in accordance with the Universities and University College Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Veterinary Science.

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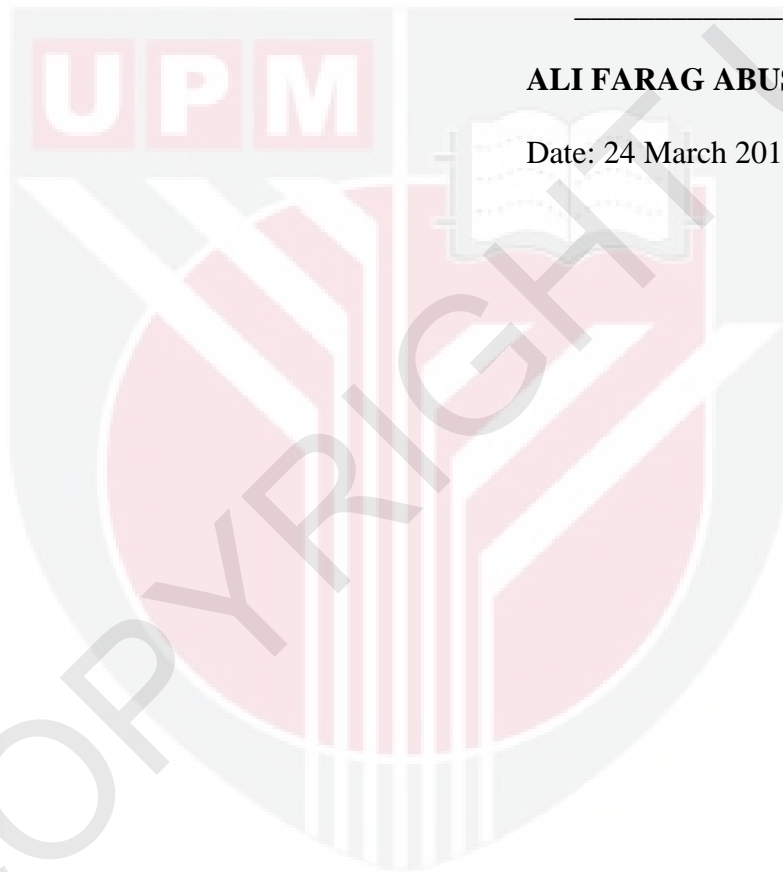
## DECLARATION

I declare that the thesis is my original work except for quotations and citation which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.

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**ALI FARAG ABUSELIANA**

Date: 24 March 2011



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## LIST OF ABBREVIATIONS

BA	Blood agar
B.C.	Before Christ
BHIA	Brain heart infusion agar
bp	Base pair
°C	Degree centigrade
CAMP	Christie, Atkins and Munch-Petersen
cfu	Colony-forming unit
cm	Centimeter
DDW	Double distilled water
DO	Dissolved oxygen
DNA	Deoxyribonucleic acid
dNTPs	Deoxynucleotide triphosphate
dpi	Days post inoculation
EB	Ethidium bromide
EDTA	Ethylenediaminetetraacetic acid
FAO	Food and Agriculture Organization of the United Nations
g	Gram
GBS	Group B <i>Streptococcus agalactiae</i>
H&E	Hematoxylin and eosin
hpi	Hours post inoculation
IP	Intraperitoneal
L	Liter
LD <sub>50</sub>	Median lethal dose

mg	Milligram
MIC	Minimum inhibitory concentration
PCR	Polymerase chain reaction
ppt	Parts per thousand
rRNA	Ribosomal ribonucleic acid
TBE	Tris-borate-EDTA
TSA	Trypticase soy agar
UV	Ultra violet



## CHAPTER 1

### INTRODUCTION

Fish are an excellent source of food for people and considered as a healthy alternative to other animal meats. Fish flesh consists mainly of high grade protein and low in fat, besides, containing minerals, vitamins and essential fatty acids. Fish protein makes up around 15% of total world animal protein supplies, providing more than 2.9 billion people with 20% or more of their average daily protein intake. Nowadays, fish account for 30% human protein supply in Asia, 20% in Africa and 15% in Latin America and Caribbean (Bone and Moore, 2008; FAO, 2008).

Aquaculture is probably one of the most important and fastest growing food production sectors in the world. It is increasing much more rapidly than animal husbandry and capture fisheries (Lucas and Southgate, 2003). World aquaculture has grown significantly in all its forms during the last fifty years and now constitutes about 50% of world food fish production (De Silva and Davy, 2010). From an annual production of less than one million tonne in the 50's, production has increased to about 52 million tonnes in 2006, with an annual growth rate of nearly 7% and a value of around US\$ 79 billion (FAO, 2008). Asia and Pacific regions are by far the world's leader in aquaculture production, producing more than 89% of the world's total aquaculture output in 2006. China is reported to be the world number one producer by producing 67% of the total quantity and 49% of the total value of world aquaculture production (FAO, 2008). With the increase in population expected over the next two decades and captured seafood production from fisheries

is at or near its peak, aquaculture is considered as the best solution to meet the growing demand for fish and other aquatic products. The ratio of per capita consumption and demand of aquatic products need to be maintained as global aquaculture production must increase by at least 40 million tonnes by 2030 (De Silva and Davy, 2010; FAO, 2008).

The aquaculture industry in Malaysia in 2007 contributed 268,500 tonnes, about 16% to national seafood supply with a value of about US\$ 0.37 billion (Wing-Keong, 2009). Freshwater aquaculture contributed 70,064 tonnes and marine and brackish water contributed 198,449 tonnes of total Malaysian aquaculture production in 2007. Tilapia, catfish, carp, blood cockle, shrimp and seaweeds were considered the most important aquatic species cultured in Malaysia (Annual Fisheries Statistics, 2007). The first introduction of Red tilapia (*Oreochromis niloticus*) hybrid to Malaysia was in middle of 1980s and from that time Red tilapia production has increased dramatically (Siti-Zahrah *et al.*, 2008). In 2007, tilapia production occupied 46% of Malaysian total freshwater aquaculture production (Wing-Keong, 2009).

The rapid increase in global aquaculture industry have exposed to many diseases that were not known in aquaculture fields. In theory, the relationship between the fish, the pathogen and the manifestation of the disease is largely due to the phenomenon of stresses which causes negative affects on fish immune system and makes fish more susceptible to diseases. Similar to livestock, fish farms are also exposed to different types of diseases which can cause huge economical damages to farming activity. The etiology of these diseases may be of viral, bacterial, fungal,

parasitic, environmental, nutritional or genetic origin. However, bacteria were often associated with multifactorial nature of fish diseases which were responsible for most of the diseases that cause mortality and losses in fish farms (Yin, 2004; Plumb, 1999; Inglis *et al.*, 1993).

Fish streptococcosis is one of bacterial diseases affecting both captive and wild fish in freshwater, estuarine and marine environments. Moreover, it is one of the fish diseases that is reported in intensive aquaculture systems causing high economic loss to fish farmers, and can cause more than 50% mortality within one week (Yanong and Francis-Floyd, 2006; Lio-Po and Lim, 2002; Inglis *et al.*, 1993). It was noted that the occurrence of streptococcosis outbreak increased when fish were stressed such as due to changes in optimal culture environment (Buller, 2004; Shoemaker *et al.*, 2000). Furthermore, fish streptococcosis is reported to be potential zoonotic capable in causing disease in humans (Toranzo *et al.*, 2005).

Several species of Streptococci have been reported to cause infections in wild and cultured fish species. *Streptococcus agalactiae*, one of Streptococci species, has a broad host range, infecting both terrestrial and aquatic animals. The organisms have been isolated from numerous fish species in natural disease outbreaks and showed to be pathogenic to several fish species (Musa *et al.*, 2009; Toranzo *et al.*, 2005; Evans *et al.*, 2002; Elliott *et al.*, 1990).

Inevitably, streptococcosis in tilapia will be a major setback in fish production program in Malaysia especially with rapid growth in tilapia industry and intensification of culture system. Thus, there is a need to focus on effort to obtain

comprehensive knowledge about this infectious disease, its etiological agent and the infectivity status, so as to find the best biosecurity measures to prevent and control the disease before it becomes a major serious threat to aquaculture industry in Malaysia.

The hypothesis of this study was *Streptococcus agalactiae* isolated from infected fish is pathogenic to cultured Red tilapia (*Oreochromis niloticus*).

The objectives of the study were:

1. To isolate and identify *S. agalactiae* from diseased farmed tilapia.
2. To determine the antibiotic susceptibility patterns of *S. agalactiae* isolates.
3. To determine the median lethal dose (LD<sub>50</sub>) of *S. agalactiae* to juvenile Red tilapia (*Oreochromis sp.*).
4. To determine the pathogenicity of *S. agalactiae* in juvenile Red tilapia (*Oreochromis sp.*).

## REFERENCES

- Al-Harbi, A. H., & Uddin, N. (2005). Bacterial diversity of tilapia (*Oreochromis niloticus*) cultured in brackish water in Saudi Arabia. *Aquaculture* , 250: 566-572.
- Annual Fisheries Statistics (2007). Retrieved 2009, from Department of Fisheries Malaysia web site: <http://www.dof.gov.my>
- Arora, D. R., & Arora, B. (2008). Textbook of Microbiology. New Delhi: CBS Publishers & Distributors.
- Austin, B., & Austin, D. A. (2007). Bacterial Fish Pathogens. Diseases of Farmed and Wild Fish. Chichester: Springer/Prazis Publishing.
- Baeck, G. W., Kim, J. H., Gomez, D. K., & Park, S. C. (2006). Isolation and characterization of *Streptococcus* sp. from diseased flounder (*Paralichthys olivaceus*) in Jeju Island. *Journal of Veterinary Science* , 7(1): 53-58.
- Berridge, B., Bercovier, H., & Frelief, P. (2001). *Streptococcus agalactiae* and *Streptococcus difficile* 16S-23S intergenic rDNA: genetic homogeneity and species-specific PCR. *Vet Microbiol* , 78:165-173.
- Betsy, T., & Keogh, J. (2005). Microbiology Demystified. New York: McGraw-Hill Companies.
- Bigarre, L., Cabon, J., Baud, M., Heimann, M., Body, A., Lieffrig, F. (2009). Outbreak of betanodavirus infection in tilapia, *Oreochromis niloticus* (L.), in fresh water. *Journal of Fish Diseases* , 32(8): 667-673.
- Black, J. G. (2008). Microbiology: Principles and Explorations. Jefferson City: John Wiley & Sons.
- Bone, Q. & Moore, R. H. (2008). Biology of Fishes. Abingdon: Taylor & Francis Group.
- Bou, G., Figueira, M., Canle, D., Cartelle, M., Eiros, J. M., & Villanueva, R. (2005). Evaluation of Group B *Streptococcus* Differential Agar for detection and isolation of *Streptococcus agalactiae*. *Clinical Microbiology and Infection* , 11(8): 676-678.
- Brimil, N., Barthell, E., Heindrichs, U., Kuhn, M., Luttkick, R., & Spellerberg, B. (2006). Epidemiology of *Streptococcus agalactiae* colonization in Germany. *International Journal of Medical Microbiology* , 296: 39-44.
- Brochet, M., Couve, E., Zouine, M., Vallaes, T., Rusniok, C., Lamy, M. (2006). Genomic diversity and evolution within the species *Streptococcus agalactiae*. *Microbes and Infection* , 8: 1227-1243.
- Buller, N. B. (2004). Bacteria From Fish and Other Aquatic Animals. Oxfordshire: CABI.



- Calderon, C. B., & Sabundayo, B. P. (2007). Antimicrobial Classifications: Drugs for Bugs. In R. Schwalbe, L. Steele-Moore, & A. C. Goodwin, Antimicrobial Susceptibility Testing Protocols (pp. 7-52). Boca Raton: Tylor& Francis Group.
- Clinical and Laboratory Standards Institute (2006). Performance standards for antimicrobial disk susceptibility tests. Approved standard, M2-A9, 9<sup>th</sup> ed., CLSI. Wayne, PA.
- Darwish, A., & Ismaiel, A. A. (2003). Laboratory efficacy of amoxicillin for the control of *Streptococcus iniae* infection in sunshine bass. *Journal of Aquatic Animal Health* , 15 (3):209-214.
- Darwish, A., & Hobbs, M. (2005). Laboratory efficacy of amoxicillin for the control of *Streptococcus iniae* infection in blue tilapia. *Journal of Aquatic Animal Health* , 17 (2): 197-202.
- De-Silva, S. S., & Davy, F. B. (2010). Aquaculture Successes in Asia: Contributing to Sustained Development and Poverty Alleviation. In S. S. De-Silva, & F. B. Davy, Success Stories in Asian Aquaculture (pp. 1-14). New York: Springer.
- Duremdez, R., Al-Marzouk, A., Qasem, J. A., Al-Harbi, A., & Gharabally, H. (2004). Isolation of *Streptococcus agalactiae* from cultured silver pomfret, *Pampus argenteus* (Euphrasen), in Kuwait. *Journal of Fish Diseases* , 27: 307-310.
- Edwards, P., Lin, C. K., & Yakupitiyage, A. (2000). Semi-intensive Pond Aquaculture. In M. C. Beveridge, & B. J. McAndrew, Tilapias: Biology and Exploitation (pp. 377-403). Dordrecht: Kluwer Academic Publishers.
- Eknath, A. E. (1995). The Nile Tilapia. In J. Thorpe, G. Gall, J. Lannan, & C. Nash, Conservation of Fish and Shellfish Resources: Managing Diversity (pp. 177-194). London: Academic Press.
- Eldar, A., Bejerano, Y., Livoff, A., Horovitz, A., & Bercovier, H. (1995). Experimental Streptococcal meningoencephalitis in cultured fish. *Veterinary Microbiology* , 43: 33-40.
- Elliott, J. A., Facklam, R. R., & Richter, C. B. (1990). Whole cell protein patterns of nonhemolytic group B, type Ib Streptococci isolated from human, mice, cattle, frog and fish. *Journal of Clinical Microbiology* , 28(3): 628-630.
- El-Sayed, A. M. (2006). Tilapia Culture. Wallingford: CABI Publishing.
- Evans, J. J., Bohnsack, J. F., Klesius, P. H., Whiting, A. A., Garcia, J. C., & Shoemaker, C. A. (2008). Phylogenetic relationships among *Streptococcus agalactiae* isolated from piscine, dolphin, bovine and human sources: a dolphin and piscine lineage associated with a fish epidemic in Kuwait is also associated with human neonatal infections in Japan. *Journal of Medical Microbiology* , 57: 1369–1376.

- Evans, J. J., Pasnik, D. J., Klesius, P. H., & Al-Ablani, S. (2006). First Report of *Streptococcus agalactiae* and *Lactococcus garvieae* from a wild bottlenose dolphin (*Tursiops truncatus*). *Journal of Wild Life Diseases* , 42(3): 561-569.
- Evans, J. J., Wiedenmayer, A. A., Klesius, P. H., & Shoemaker, C. A. (2004). Survival of *Streptococcus agalactiae* from frozen fish following natural and experimental infections. *Aquaculture* , 233: 15–21.
- Evans, J., Klesius, P., Gilbert, P., Shoemaker, C., Al-Sarawi, M., Landsberg, J. (2002). Characterization of B-haemolytic group B *Streptococcus agalactiae* in cultured seabream, *Sparus auratus* L., and wild mullet, *Liza klunzingeri* (Day), in Kuwait. *Journal of Fish Diseases* , 25: 505-513.
- FAO, (2008). The State of World Fisheries and Aquaculture. Retrieved 2009, from Food and Agriculture Organization of the United Nations Web site: <http://www.fao.org/docrep/011/i0250e/i0250e00.htm>
- Ferguson, H. W., Morales, J. A., & Ostland, V. E. (1994). Streptococcosis in aquarium fish. *Diseases of Aquatic Organisms*, 19: 1-6.
- Filho, C. I., Muller, E. E., Pretto-Giordano, L. G., & Bracarense, A. F. (2009). Histological findings of experimental *Streptococcus agalactiae* infection in Nile tilapia (*Oreochromis niloticus*). *Brazilian Journal of Veterinary Pathology* , 2(1): 12-15.
- Fitzsimmons, K. M. (2008). A good year for tilapia producers and consumers in 2007. *AQUA Culture AsiaPacific Magazine* , pp. 4(4): 24-25.
- Green, B. W., & Duke, C. B. (2006). Pond Production. In C. E. Lim, & C. D. Webster, *Tilapia: Biology, Culture and Nutrition* (pp. 253-288). New York: Food Products Press.
- Hasson, K. W., Wyld, E. M., Fan, Y., Lingsweiller, S. W., Weaver, S. J., Cheng, J. (2009). Streptococcosis in farmed *Litopenaeus vannamei*: a new emerging bacterial disease of penaeid shrimp. *Diseases of Aquatic Organisms* , 86: 93-106.
- Hernandez, E., Figueroa, J., & Iregui, C. (2009). Streptococcosis on a red tilapia, *Oreochromis sp.*, farm: a case study. *Journal of Fish Diseases* , 32: 247-252.
- Inglis, V., Roberts, R. J., & Bromage, N. R. (1993). *Bacterial Diseases of Fish*. Oxford: Blackwell.
- Johri, A. K., Paoletti, L. C., Glaser, P., Dua, M., Sharma, P. K., Grandi, G. (2006). Group B *Streptococcus*: global incidence and vaccine development. *Nature Reviews Microbiology* , 4(12): 932-942.
- Josupeit, H. (2009). Tilapia Market Report-May 2009. Retrieved January 2010, from FAO Globefish: <http://www.globefish.org>
- Kawamura, Y., Itoh, Y., Mishima, N., & Ohkusu, K. (2005). High genetic similarity of *Streptococcus agalactiae* and *Streptococcus difficilis*: *S. difficilis* Eldar et

- al.* 1995 is a later synonym of *S. agalactiae* Lehmann and Neumann 1896 (Approved Lists 1980). *Journal of Systematic and Evolutionary Microbiology* , 55: 961–965.
- Kilian, M. (2002). *Streptococcus* and *Enterococcus*. In D. Greenwood, R. C. Slack, & J. F. Peutherer, *Medical Microbiology* (pp. 174-188). London: Churchill Livingstone.
- Lancefield, R. C. (1933). A serological differentiation of human and other groups of hemolytic *Streptococci*. *Journal of Experimental Medicine* , 57(4): 571–595.
- Lehman, D. C., Mahon, C. R., & Suvarna, K. (2007). *Streptococcus, Enterococcus, and Other Catalase-Negative Gram-Positive Cocci*. In C. R. Mahon, D. C. Lehman, & G. Manuselis, *Textbook of Diagnostic Microbiology* (pp. 382-409). St. Louis: Saunders/Elsevier.
- Lim, C. E., & Webster, C. D. (2006). *Tilapia: Biology, Culture and Nutrition*. New York: Food Products Press.
- Lio-Po, G. D., & Lim, L. H. (2002). Infectious Diseases of Warmwater Fish in Fresh Water. In P. T. Woo, D. W. Bruno, & L. H. Lim, *Diseases and Disorders of Finfish in Cage Culture* (pp. 231-281). Wallingford: CABI Publishing.
- Lucas, J. S., & Southgate, P. C. (2003). *Aquaculture: Farming Aquatic Animals and Plants*. Oxford: Blackwell.
- Madigan, M. T., Martinko, J. M., Dunlap, P. V., & Clark, D. P. (2009). *Brock biology of microorganisms*. San Francisco: Pearson Benjamin Cummings.
- Martinez, G., Harel, J., & Gottschalk, M. (2001). Specific detection by PCR of *Streptococcus agalactiae* in milk. *The Canadian Journal of Veterinary Research* , 65:68-72.
- Mata, A. I., Gibello, A., Casamayor, A., Blanco, M. M., Dominguez, L., & Fernandez-Garayzabal, J. F. (2004). Multiplex PCR Assay for detection of bacterial pathogens associated with warm-water streptococcosis in fish. *Applied and Environmental Microbiology* , 70(5): 3183-3187.
- McAndrew, B. J. (2000). Evolution, phylogenetic relationships and biogeography. In M. C. Beveridge, & B. J. McAndrew, *Tilapias: Biology and Exploitation* (pp. 1-32). Dordrecht: Kluwer Academic Publishers.
- Meyer, F. P. (1991). Aquaculture disease and health management. *Journal of Animal Science* . , 69:4201-4208.
- Mian, G. F., Godoy, D. T., Leal, C. A., Yahara, T. Y., Costa, G. M., & Figueiredo, H. C. (2009). Aspects of the natural history and virulence of *S. agalactiae* infection in Nile tilapia. *Veterinary Microbiology* , 136: 180-183.
- Miller, R. A., Walker, R. D., Baya, A., Clemens, K., Coles, M., Hawke, J. P. (2003). Antimicrobial Susceptibility Testing of Aquatic Bacteria: Quality Control Disk Diffusion Ranges for *Escherichia coli* ATCC 25922 and *Aeromonas*

*salmonicida* subsp. *salmonicida* ATCC 33658 at 22 and 28°C. *Journal of Clinical Microbiology* , 41(9): 4318-4323.

- Murray, P. R., Rosenthal, K. S., & Pfaller, M. A. (2005). *Medical Microbiology*. Philadelphia: Elsevier/Mosby.
- Musa, N., Wei, L. S., Hamdan, R. H., Leong, L. K., Wee, W. (2009). Short communication: Streptococcosis in red tilapia (*Oreochromis niloticus*) commercial farm in Malaysia. *Aquaculture Research* , 40: 630-632.
- Pasnik, D. J., Evans, J. J., & Klesius, P. H. (2005). Duration of protective antibodies and correlation with survival in Nile tilapia (*Oreochromis niloticus*) following *Streptococcus agalactiae* vaccination. *Diseases of aquatic organisms* , 66: 129-134.
- Perera, R. P., Johnson, S. K., & Lewis, D. H. (1997). Epizootiological aspects of *Streptococcus iniae* affecting tilapia in Texas. *Aquaculture* , 152: 25-33.
- Pillay, T. V., & Kutty, M. N. (2005). *Aquaculture: Principles and Practices*. Oxford: Blackwell Publishing.
- Plumb, J. A. (1999). *Health Maintenance and Principal Microbial Diseases of Cultured Fishes*. Ames: Iowa State University Press.
- Popma, T., & Masser, M. (1999). *Tilapia: Life History and Biology*. Retrieved May 2009, from Southern Regional Aquaculture Center publications: <http://srac.tamu.edu/>
- Pretto-Giordano, L. G., Muller, E. E., de-Freitas, J. C., & da-Silva, V. G. (2010). Evaluation on the Pathogenesis of *Streptococcus agalactiae* in Nile Tilapia (*Oreochromis niloticus*). *Brazilian Archives of Biology and Technology* , 53(1): 87-92.
- Pretto-Giordano, L. G., Muller, E. E., Klesius, P., & da-Silva, V. G. (2009). Efficacy of an experimentally inactivated *Streptococcus agalactiae* vaccine in Nile tilapia (*Oreochromis niloticus*) reared in Brazil. *Aquaculture Research* , 1-6.
- Rasheed, V., & Plumb, J. (1984). Pathogenicity of a non-haemolytic group B *Streptococcus* sp. in gulf killifish (*Fundulus grandis*, Baird and Girard). *Aquaculture* , 37: 97-105.
- Rattanachaikunsopon, P., & Phumkhachorn, P. (2009). Prophylactic effect of *Andrographis paniculata* extracts against *Streptococcus agalactiae* infection in Nile tilapia (*Oreochromis niloticus*). *Journal of Bioscience and Bioengineering* , 107(5): 579-582.
- Robinson, J. A., & Meyer, F. P. (1966). Streptococcal fish pathogen. *Journal of Bacteriology* , 92(2): 512.
- Romalde, J. L., Ravelo, C., Valdes, I., Magarinos, B., Fuente, E., Martin, C. S. (2008). *Streptococcus phocae*, an emerging pathogen for salmonid culture. *Veterinary Microbiology* , 130: 198-207.

- Ross, L. J. (2000). Environmental Physiology and Energetics. In M. C. Beveridge, & B. J. McAndrew, *Tilapias: Biology and Exploitation* (pp. 89-128). Dordrecht: Kluwer Academic Publishers.
- Russo, R., Mitchell, H., & Yanong, R. P. (2006). Characterization of *Streptococcus iniae* Isolated from *Ornamental Cyprinid* Fishes and Development of Challenge Models. *Aquaculture* , 256: 105-110.
- Salvador, R., Muller, E. E., Freitas, J. C., Leonhardt, J. H., Pretto-Giordano, L. G., & Dias, J. A. (2005). Isolation and characterization of *Streptococcus* sp. group B in Nile tilapias (*Oreochromis niloticus*) reared in hapas nets and earth nurseries in the northern region of Parana State, Brazil. *Ciência Rural* , 35(6): 1374-1378.
- Sheehan, B. (2009a). Streptococcosis in tilapia: A more complex problem than expected? *symposium Managing Streptococcus in Warmwater* (pp. 9-14). Veracruz, Mexico: Intervet International B.V
- Sheehan, B. (2009b). AquaVac® Strep Sa: A novel vaccine for control of *Streptococcus agalactiae* Biotype 2 infections in farmed tilapia, *symposium Managing Streptococcus in Warmwater* (pp. 21-26). Veracruz, Mexico: Intervet International B.V
- Shelton, W. L., & Popma, T. J. (2006). Biology. In C. E. Lim, & C. D. Webster, *Tilapia: Biology, Culture and Nutrition* (pp. 1-49). New York: Food Products Press.
- Shewmaker, P. L., Camus, A. C., Bailiff, T., Steigerwalt, A. G., Morey, R. E., & Carvalho, M. S. (2007). *Streptococcus ictaluri* sp. nov., isolated from channel catfish *Ictalurus punctatus* broodstock. *International Journal of Systematic and Evolutionary Microbiology* , 57: 1603-1606.
- Shoemaker, C. A., Evans, J. J., & Klesius, P. H. (2000). Density and Dose Factors Affecting Mortality of *Streptococcus iniae* Infected Tilapia (*Oreochromis niloticus*). *Aquaculture* , 229-235.
- Shoemaker, C. A., Vandenberg, G. W., Désormeaux, A., Klesius, P. H., & Evans, J. J. (2006 a). Efficacy of a *Streptococcus iniae* modified bacterin delivered using Oralject™ technology in Nile tilapia (*Oreochromis niloticus*). *Aquaculture* , 255: 151-156.
- Shoemaker, C. A., Xu, D., Evans, J. J., & Klesius, P. H. (2006 b). Parasites and Diseases. In C. Lim, & C. D. Webster, *Tilapia: Biology, Culture and Nutrition* (pp. 561-582). New York: Food Products Press.
- Siti-Zahrah, A., Misri, S., Padilah, B., Zulkafli, R., Kua, B.C., Azila, A. and Rimatulhana, R. (2004). Pre- disposing factors associated with outbreak of Streptococcal infection in floating cage-cultured tilapia in reservoirs, p. 129. In 7th Asian Fisheries Forum 04 Abstracts. The Triennial Meeting of the Asian Fisheries Society, 30th Nov.-4th Dec. 2004 (pp. 420). Penang, Malaysia.

- Siti-Zahrah, A., Padilah, B., Azila, A., Rimatulhana, R., & Shahidan, H. (2008). Multiple streptococcal species infection in cage-cultured red tilapia but showing similar clinical signs. In M. G. Bondad-Reantaso, C. V. Mohan, M. Crumlish, & R. P. Subasinghe, *Diseases in Asian Aquaculture VI* (pp. 313-320). Manila: Asian Fisheries Society.
- Smith, P. R., Breton, A. L., Horsberg, T. E., & Corsin, F. (2008). Guideline for antimicrobial use in aquaculture. In L. Guardabassi, L. B. Jensen, & H. Kruse, *Guide to antimicrobial use in animals* (pp. 207-218). Oxford: Blackwell Publishing.
- Specter, S., Hodinka, R. L., & Young, S. A. (2000). *Clinical Virology Manual*. Washington: ASM Press.
- Stickney, R. R. (2000). *Encyclopedia of Aquaculture*. New York: John Wiley & Sons.
- Suanyuk, N., Kanghear, H., Khongpradit, R., & Supamattaya, K. (2005). *Streptococcus agalactiae* infection in tilapia (*Oreochromis niloticus*). *Songklanakarin Journal of Science and Technology* , 27: 307-319.
- Suanyuk, N., Kong, F., Ko, D., Gilbert, G. L., & Supamattaya, K. (2008). Occurrence of rare genotypes of *Streptococcus agalactiae* in cultured red tilapia *Oreochromis sp.* and Nile tilapia *O. niloticus* in Thailand-Relationship to human isolates? *Aquaculture* , 284:35-40.
- Sun, W., Dong, L., Kaneyama, K., Takegami, T., & Segami, N. (2008). Bacterial diversity in synovial fluids of patients with TMD determined by cloning and sequencing analysis of the 16S ribosomal RNA gene. *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology and Endodontology* , 105(5):566-571.
- Toranzo, A. E., Magarinos, B., & Romalde, J. L. (2005). A review of the main bacterial fish disease in mariculture system. *Aquaculture* , 246: 37-61.
- Wendover N. (2009). Managing tilapia health in complex commercial systems, *symposium Managing Streptococcus in Warmwater* (pp. 4-8). Veracruz, Mexico: Intervet International B.V.
- Wheat, P. F. (2001). History and Development of Antimicrobial Susceptibility Testing Methodology. Retrieved May 2009, from British Society for Antimicrobial Chemotherapy:  
[http://www.bsac.org.uk/db/documents/Chapter\\_1.pdf](http://www.bsac.org.uk/db/documents/Chapter_1.pdf)
- Wing-Keong, N. (2009). The current status and future prospects for the aquaculture industry in Malaysia. *World Aquaculture* , pp. 40(3): 26-30.
- Yang, W., & Li, A. (2009). Isolation and characterization of *Streptococcus dysgalactiae* from diseased *Acipenser schrenckii*. *Aquaculture* , 294: 14-17.
- Yanong, R. P., & Francis-Floyd, R. (2006). Streptococcal Infection of Fish. Retrieved May 2009, from Department of Fisheries and Aquatic Sciences,

Florida Cooperative Extension Service, Institute of Food and Agricultural Sciences, University of Florida: <http://edis.ifas.ufl.edu/fa057>

- Yildirim, A. O., Lammler, C., & Weib, R. (2002). Identification and characterization of *Streptococcus agalactiae* isolated from horses. *Veterinary Microbiology*, 85: 31-35.
- Yin, L. K. (2004). Current Trends in the Study of Bacterial and Viral Fish and Shrimp Diseases. Singapore: World Scientific Publishing Co.
- Yuasa, K., Kamaishi, T., Hatai, K., Bahnnan, M., & Borisutpeth, P. (2008). Two Cases of Streptococcal Infections of Cultured Tilapia in Asia. In M. Bondad-Reantaso, C. Mohan, M. Crumlish, & R. Subasinghe, Diseases in Asian Aquaculture VI (pp. 259-268). Manila: Fish Health Section, Asian Fisheries Society.

