

ISSN: 1472-8214 (Print) 1744-7623 (Online) Journal homepage: https://www.tandfonline.com/loi/iemd20

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To cite this article: Adamantia Liapikou, Catia Cilloniz, Andrea Palomepue & Toni Torres (2019): Emerging antibiotics for Community-Acquired Pneumonia, Expert Opinion on Emerging Drugs, DOI: 10.1080/14728214.2019.1685494

To link to this article: https://doi.org/10.1080/14728214.2019.1685494

Accepted author version posted online: 28 Oct 2019.



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Publisher: Taylor & Francis & Informa UK Limited, trading as Taylor & Francis GroupJournal: *Expert Opinion on Emerging Drugs*DOI: 10.1080/14728214.2019.1685494

Emerging antibiotics for Community-Acquired Pneumonia

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Abstract

Introduction: Community-acquired pneumonia is the most common infection leading to hospitalization and death in all age groups, especially in elderly populations. Increasing antibiotic resistance among the common bacterial pathogens associated with community-acquired pneumonia, especially Streptococcus pneumoniae and staphylococci, has made its empirical treatment increasingly problematic, highlighting the need for effective antibiotic therapy.

Areas covered: We searched PubMed and ClinicalTrials.gov for English-language reports of phase III clinical trials conducted between 2000 and 2019 concerning the antibiotic treatment of community-acquired pneumonia. We provide a summary of the latest approved drugs for this indication and highlight emerging drugs with a potential indication.

Expert opinion: Ceftaroline (a new cephalosporine) and omadacycline (a cycline alternative), either parenterally or orally, are the only two new antibiotics to have been approved by the FDA for the treatment of community-acquired pneumonia in the last five years. Among the antimicrobials in development, Lefamulin (the first pleuromutilin), is currently in phase III development. Among the known antibiotic classes, solithromycin (a macrolide), nemonoxacin (a quinolone), and delafloxacin and zabofloxacin (both fluoroquinolones), have been studied in phase II and III in clinical trials. The availability of these new antibiotics may offer opportunities to improve the empirical treatment for community-acquired pneumonia.

Keywords: Community acquired pneumonia, emerging, antibiotics, omadacycline, lefamulin, solythromycin

1. Background

Community-acquired pneumonia (CAP) remains an important worldwide health problem. It is responsible for hospitalization, re-admissions, pulmonary and extrapulmonary complications, and short- and long-term mortality [1, 2, 3]. The clinical presentation of CAP ranges from mild or localized to severe or systemic disease, which may be associated with complications such as sepsis, septic shock, and respiratory failure, even leading to multiple organ dysfunction and death. CAP is especially dangerous in elderly and immunocompromised patients, in whom the mortality rate can reach 20%–40%, which is significantly higher than the 6%–10% reported for the general population [4, 5].

Recently, investigations have focused on the increasing prevalence of antibioticresistant pathogens causing CAP, especially in severe cases [6, 7, 8], and on the relation between cell-mediated immunity and the inflammatory response to pneumonia [9, 10]. *Streptococcus pneumoniae* remains the main causative pathogen in CAP [11], despite studies indicating that respiratory viruses play an important role [12, 13]. However, a small proportion (6%) are caused by multidrug-resistant (MDR) pathogens, including methicillin-resistant *Staphylococcus aureus* (MRSA), *Pseudomonas aeruginosa, Klebsiella pneumoniae,* and *Acinetobacter baumannii.* The antibiotic resistance of these pathogens makes them a challenge for clinical management and a key global public health concern.

The development of molecular diagnostic techniques over the last three decades has led to the importance of respiratory viruses being emphasized as causes of CAP and its associated mortality worldwide. Indeed, approximately 20%–30% of patients hospitalized with viral CAP are estimated to need intensive care unit admission. Recently, our group [11] reported that viral sepsis was present in 19% of patients admitted to intensive care with CAP, with influenza viruses being most common, and males and older patients being at greatest risk. An interesting recent study by Zhou et al. [12] that investigated non-influenza viruses causing CAP showed that 63%, 26%, and 10% of cases were caused by influenza viruses, non-influenza viruses,

and mixed viral infections, respectively. The authors found that complications were common in patients with non-influenza CAP, but that outcomes were similar to those in patients with influenza. Together, these data support the importance of respiratory viruses as causes of CAP.

There is an urgent clinical need for effective antimicrobial therapy to treat the range of etiologies of CAP, especially in terms of improving the initial therapy given to patients with MDR pathogens. In this review, new antibiotics belonging to both old and new antibiotic classes will be analyzed and discussed with these considerations in mind.

2. Methods

The PubMed database was searched using the following search string: "Pneumonia" AND English AND ("Community-acquired" OR "Community acquired" OR "CAP" OR "Hospitalized" OR "Hospitalized") AND ("Antibiotics" OR "Antimicrobial" OR "Resistance" OR "Anti-Bacterial Agents" OR "Antiviral Agents"). Search results were restricted to clinical trials, meta-analyses, observational studies, and systematic reviews. Titles and abstracts were initially screened to identify relevant citations, and these were then reviewed in full by two authors. We included all publications presenting data on CAP, and reviewed the study setting, methodology, and characteristics of the study population. All authors confirmed that the inclusion of the identified publications was appropriate.

3. Existing Treatment Options and Needs

3.1 Antibiotics

Antibiotic resistance in *S. pneumoniae* (pneumococcus), the main pathogen responsible for CAP, is a major global concern. The emergence of MDR pneumococcus due to overuse of antibiotics is a global concern that affects the main anti-pneumococcus drugs, β -lactams, macrolides, and fluoroquinolones. Last year a study about the burden of pneumococcal CAP in adults across Europe [1] showed that the overall incidence was 68–7000 per 100,000 and the incidence in hospitalized CAP cases was 16–3581 per 100,000. The authors reported that the

incidence increased with age, and that the greatest impact was observed in patients with more comorbidities. Interestingly, a study published by our group. [14] about the 20-year trend in mortality due to pneumococcal CAP in 1120 hospitalized adults in Spain failed to show any decrease in mortality over time. The recent report by the SENTRY Antimicrobial Surveillance Program [15] evaluated antimicrobial susceptibility for *S. pneumoniae* isolates worldwide over a 20-year period (1997–2016) indicated that MDR and extensively drug resistant isolates varied by region. The Asia–Pacific region showed the highest rates (41% in 1997–1998; 53% in 2007–2008; 39% in 2015–2016), whereas Latin America presented the lowest rates (10% overall). In North America, the data showed an increase from 9% in 1997–1998 to 24% in 2009–2010, followed by a decrease to 17% in 2015–2016. An increase was also seen in Europe from 17% in 1997–1998 to 24% in 2007–2008, but this also decreased in the last period (19% in 2015–2016).

The PES pathogen group (i.e., *P. aeruginosa*, extended-spectrum β-lactamase Enterobacteriaceae, and methicillin-resistant S. aureus) cause approximately 6% of cases of CAP [16]. Among them, P. aeruginosa and MRSA are the most frequently reported, and they require different antimicrobial therapy compared with typical pathogens. A recent multinational point-prevalence study [17] presented data from 3193 patients with CAP in 222 hospitals from 54 countries and reported the burden and risk factors associated with *P. aeruginosa* infection. The study showed that there was a low prevalence of CAP caused by *P. aeruginosa* (4%), corresponding to only 11% of patients with culture-positive pneumonia. The prevalence rates of antibioticresistant and MDR P. aeruginosa were also low at 2% and 1% respectively. By continent, the prevalence of *P. aeruginosa* in CAP was as follows: 3.8% in Europe, 4.3% in North America, 5.2% in Asia, 4.9% in South America, 5.5% in Africa, and 3.1% in Oceania. The corresponding prevalence of antibiotic-resistant P. aeruginosa was 1.6% (MDR, 0.9%) in Europe, 2.5% (MDR, 1.2%) in North America, 2.2% (MDR, 0.5%) in Asia, 3.0% (MDR, 2%) in South America, and 3.9% (MDR, 2.3%) in Africa, with no reported cases of *P. aeruginosa* antibiotic resistance in Oceania. Similarly, a Spanish prospective cohort study [18] into the clinical outcomes and risk factors for MDR and non-MDR P. aeruginosa reported a prevalence of 1.1% for MDR P. aeruginosa among 2023 culture-positive patients. CAP due to P. aeruginosa was shown to be an individual risk factor for mortality in that study population.

In two independent European studies of patients hospitalized with CAP, it was reported that MDR pathogens were causative in 3.3% (a Spanish cohort) and 7.6% (an Italian cohort), with MRSA pathogens being most common [8, 19]. The Global initiative for MRSA pneumonia (GLIMP) analyzed data for 3191 patients receiving microbiological tests within 24 h of admission for CAP and reported a 3% global incidence for MRSA as a cause. Risk factors for MRSA in CAP were prior MRSA infection or colonization, recurrent skin infection, and severe pneumonia. Other studies in the USA have reported incidence rates of 0.7%–2.4% [20, 21].

A. baumannii has been considered a rare cause of CAP, but one that retains clinical relevance. The great majority of CAP cases caused by *A. baumannii* are reported in tropical or sub-tropical countries, principally in Asia-Pacific regions, with the highest prevalence in Hong Kong, Singapore, Taiwan, South Korea, and Australia [22, 23]. In Europe and North America, however, *A. baumannii* is exceedingly rare. A recent USA report found that only 19 cases of severe CAP were caused by this pathogen [24]. Interestingly, in a case-control study in the USA that aimed to characterize the epidemiology of MDR *A. baumannii* colonization in high-risk nursing home residents, 15% (n = 25) of residents were noted to be colonized, with all isolates being resistant to cephalosporins, monobactam (aztreonam), and quinolones. They reported the main risk factors for MDR *A. baumannii* as functional disability, *Proteus mirabilis* colonization, and diabetes mellitus [25].

CAP caused by *K. pneumoniae* is reported in approximately 1%–7% of cases [8], with MDR strains accounting for only 5%–36%. In Asian countries, such as Taiwan, Cambodia, and Shanghai, *K. pneumoniae* has been described as a frequent cause of bacteremia [26, 27]. By contrast, CAP caused by *K. pneumoniae* is generally rare in Europe and USA, despite there being an increasing incidence in these regions over recent years [28, 29].

Empirical antibiotic therapy for severe CAP remains based on international guidelines that recommend a macrolide or a respiratory fluoroquinolone in combination with a β -lactam, with coverage for PES pathogens only given when certain risk factors are present. The research and development of new antibiotics is scientifically and economically challenging, yet this must remain a key goal given current global needs and the spread of antimicrobial resistance. Given that antibiotic therapies for MRSA and *P. aeruginosa* differ from typical empirical therapy, we must

recognize the risk factors and local epidemiology relevant to these resistant microorganisms in clinical contexts. Early recognition and prompt initiation of broadspectrum empirical antibiotic therapy is mandatory to limit the negative effects of these pathogens, but this is limited by a decline in the production of new antibiotics and a failure to keep pace with epidemiological changes over recent decades. Arguably the most important consideration with these new antimicrobials is that they should be broad spectrum to allow clinicians to treat CAP in the era of increasing resistance.

3.2 Antivirals

Currently, neuraminidase inhibitors (e. g., oseltamivir, zanamivir, peramivir) are well established in clinical practice for use in patients with influenza infection, but no randomized-controlled trial (RCT) of has been completed to confirm the efficacy of these antivirals in hospitalized patients. The data of several prospective observational studies and meta-analysis indicate that neuraminidase inhibitor therapy at the time of hospitalization reduces both the length of hospitalization and the related mortality [30, 31]. The main problem with neuraminidase inhibitors like oseltamivir, however, is the need for early treatment. Other antivirals approved by the Food and Drug Administration (FDA) include adamantanes (e.g., amantadine and rimantadine) are not recommended because of widespread resistance. The polymerase inhibitors baloxavir marboxil and favipiravir have shown efficacy against influenza A and B, and against strains resistant to neuraminidase inhibitors, but is known about their resistance profiles.

During influenza seasons, respiratory syncytial virus (RSV) is a major cause of moderate-to-severe respiratory infection, especially in immunocompromised patients. Ribavirin is the only approved treatment option for RSV, but this antiviral suffers from its limited efficacy and propensity for genotoxicity.

4. Market review and current research goals

The increase in antimicrobial resistance has led to the development of new antibiotics in recent years, with may being approved for the treatment of pneumonia. Unfortunately, the development of new resistant bacterial strains has outstripped this

development because of high costs, long drug development times, and difficulties in designing and performing adequate trials. A current estimate is that fewer than 10 of the 50 leading pharmaceutical companies have active antimicrobial drug development programs [32].

Omadacycline, is a modernized tetracycline that was specifically designed to overcome tetracycline resistance. It received FDA approval in October 2018 for the treatment of bacterial CAP and acute bacterial skin and skin structure infections (ABSSSIs). Most agents detailed in this text have primarily been designed to treat infections caused by MRSA, but these have only proven secondarily to be active against MDR *S. Pneumoniae*. New lipoglycopeptides (e.g., dalbavancin, oritavancin, and telavancin) and oxazolidinones (e.g., tedizolid) are therefore attractive options for treating infections due to gram-positive MDR bacteria [33,34].

Unlike MRSA, research and development into novel drug candidates for gramnegative resistant bacteria, especially extended spectrum beta-lactamase inhibitors or carbapenem-resistant Enterobacteriaceae, will take longer and incur higher costs. Despite this clear need for novel antibiotics without the potential cross-resistance, no new drug classes are close to market entry. Two new cephalosporins with or without a β -lactamase inhibitor have been approved for such infection, just not for CAP: ceftolozane with tazobactam and ceftazidime with avibactam.

This situation is by no means new. Indeed, nearly all antibiotic classes in use today were discovered during the Golden Age of antibiotic innovation in the 1940s and 1960s [32]. Efforts to modify the chemical structures then focused on circumventing emerging class-specific target or drug-modifying resistance mechanisms, or to lower the affinity for efflux pumps, as well as on improving the pharmacokinetics and extending the spectrum of activity. The holy grail of antibiotic progress—a truly novel class of antibiotics—has not been achieved since 1987, and current economic incentives will likely continue this trend. A major hope lies with several biological agents, mainly monoclonal antibodies, which are in phase I or II development for use as adjunctive therapy in staphylococcal infections (e.g., 514G3, AR-301, DSTA-46375, suvratoxumab, SAL-200, ASN-100, and CF-301).

Numerous agencies and professional societies have tried to draw attention to the lack of new antibiotics, especially for the treatment of MDR gram-negative

pathogens. Since January 2012, when the Generating Antibiotic Incentives Now (GAIN) act came into effect in the USA, development of a new antibiotic agent that is active against one or several pathogens on a central list allows a Qualified Infectious Disease Product (QIDP) designation to be obtained [35]. Consequently, the overall USA budget to fight antimicrobial resistance almost doubled to about 1.2 billion dollars by 2016, with more antibacterial molecules clearly being approved in recent years.

5. Newly approved antibiotics and drugs in development

Newly approved and phase III drugs against gram-positive and gram-negative bacteria are described in Table 1. There are also a number of promising antibiotics in development that should be able to widen the therapeutic armament against difficultto-treat gram-positive infections. Solithromycin, delafloxacin, zabofloxacin, and nemonoxacin have each demonstrated broad-spectrum antibacterial activity, together with other beneficial features such as intracellular accumulation, antiinflammatory effects, and inhibition of biofilm production. A new drug, lefamulin, is a pleuromutilin antibiotic that is currently in phase III of development.

In this section we look at the main drugs showing potential in each of the main drug classes.

5.1. Cephalosporins: the potential of ceftaroline

The cephalosporins are known for their broad spectrum of activity, proven efficiency, and favorable safety profile, making them among the most commonly prescribed antimicrobials. Ceftobiprole and ceftaroline are particularly noted for their activity against MRSA and MDR S. *Pneumoniae*, but only ceftaroline is approved by the FDA in the treatment of CAP [36].

Ceftaroline (Teflaro® in the USA and Zinforo® in Europe) is a fifth-generation cephalosporin with an in vitro spectrum of activity that includes most of the common bacterial pathogens associated with CAP. It has bactericidal activity against MDR *S. pneumoniae,* non-extended-spectrum β -lactamase producing *K. pneumoniae,* and *Escherichia coli,* and its high affinity for PBP2a grants coverage against MRSA [37].

In vitro and in vivo animal models: The effect of ceftaroline in both combination therapy and monotherapy has been examined in multiple in vitro pharmacokinetic models. It has been used in combination with daptomycin, vancomycin, and rifampin. Ceftaroline and daptomycin both provide activity against MRSA, and ceftaroline increases the binding of daptomycin. B-lactams are used in combination with vancomycin due to the seesaw effect, an in vitro phenomenon where, as daptomycin and vancomycin minimum inhibitory concentations (MICs) increase in MRSA, the MICs of β -lactams, including ceftaroline, decrease [38].

However, both low-level and high-level resistance to ceftaroline have been observed in MRSA strains; particularly concerning are recent reports of resistance discovered in clinical MRSA isolates from patients in geographic regions never exposed to the drug [39]. Little is known of ceftaroline's ecological impact, though it is expected to be minor, as the drug is not excreted in the faeces.

The approval of ceftaroline for the treatment of CAP was based on two phase III multinational RCTs conducted in hospitalized patients: FOCUS 1 (NCT00621504) and FOCUS 2 (NCT00509106). In these, 600 mg ceftaroline fosamil was given every 12 h and demonstrated noninferiority to 1 g ceftriaxone every 24 h, but with differences in clinical cure rates at the test-of-cure visit favoring ceftaroline fosamil in each trial [40,41]. With ceftaroline, clinical cure was achieved in up to 80% of cases and the agent was well tolerated, with only mild adverse events (e.g., diarrhea, headache, and insomnia). However, neither study included patients admitted to intensive care or who had comorbidities, and there were few MDR S. *pneumoniae* (n \leq 10) and MRSA (n = 1) isolates. Analysis of data from the FOCUS trials showed that ceftaroline was still associated with a shorter time to clinical response than ceftriaxone [42]. Although there are no data from RCTs supporting the use of ceftaroline fosamil for MRSA pneumonia, some research supports its efficacy in cases of MRSA bacteremia, MRSA endocarditis, and CAP [43,44].

Ceftaroline appeared to be well tolerated in these trials, but several post-marketing reports of severe myelotoxicity associated with prolonged exposure (>7 days) to ceftaroline have emerged [45]. At present, it is approved by the FDA and the European Medicines Agency (EMA) for the treatment of both CAP and complicated skin and skin structure infections [46].

5.2. Tetracyclines: the potential of omadacycline

Tetracyclines are a group of broad-spectrum antibiotics whose general usefulness has reduced with the onset of bacterial resistance. Despite this, they remain the treatment of choice for CAP due to their effectiveness against atypical bacteria.

Omadacycline is the lead compound of the novel aminomethylcycline subclass of tetracyclines. Similar to tetracyclines, omadacycline acts by inhibiting protein synthesis, but binds to 70 S ribosomes with greater affinity than tetracycline. The modification in the C-9 position arose from the finding that tigecycline, a C-9 substituted semisynthetic derivative of minocycline, was not affected by the most common resistance mechanisms, namely efflux and ribosomal protection, despite retaining clinical activity [47]. It is a broad-spectrum antibiotic with improved activity against tetracycline-resistant pathogens, including methicillin-susceptible *S. aureus* and MRSA, as well as various *streptococci* and *enterococci*. It is also more active than doxycycline and minocycline against Enterobacteriaceae and *A. baumannii*, with MICs of <4 µg/mL for 90% of strains [48]. Omadacycline is also active against many clinically important *Enterobacteriaceae* and a wide range of anaerobes [49], including the atypical organisms *Legionella pneumophila*, *Mycoplasma pneumoniae*, and *Chlamydia pneumoniae*.

Of clinical importance, omadacycline has also shown promise against fast-growing mycobacteria in a comparison with tigecycline and doxycycline, where it was demonstrated that omadacycline and tigecycline had similar activities against *M. abscessus*, (MIC₅₀ of 1 µg/mL and MIC₉₀ of 2 µg/mL. Against *M. chelonae*, the MIC₅₀ values for omadacycline and tigecycline were 0.125 µg/mL and 0.06 µg/mL, respectively, indicating that the latter was superior; however, both drugs had the same MIC₉₀ of 0.25µg/mL [50]. This notwithstanding, omadacycline has pharmacokinetic advantages of higher and more sustained concentrations in the plasma and epithelial lining fluid (ELF)compared with tigecycline, suggesting that it may be a promising antibacterial agent for the treatment of CAP caused by susceptible pathogens [51].

In the US, omadacycline was approved in October 2018 for the treatment of bacterial CAP and ABSSSIs . It is taken once daily, which offers an advantage over

minocycline and doxycycline, and is available in both oral and intravenous forms as a tosylate salt. However, its oral bioavailability is only 35%, with food reducing the oral bioavailability. It should therefore be administered in the fasted state. It is 20% protein bound in the serum and has a biologic half-life of 17 h [52].

The use of omadacycline was based on the results of a 750-patient phase III RCT (OPTIC Trial) that compared oral or intravenous omadacycline to oral or intravenous moxifloxacin for the treatment of mild, moderate, and severe bacterial CAP from November 2015 [53]. Patients were given 100 mg intravenous omadacycline twice a day (two doses), followed by 100 mg daily, intravenously, or 400 mg intravenous moxifloxacin once a day for three days. In both cases, there was an option to switch to oral administration. Omadacycline was statistically non-inferior to moxifloxacin for early clinical response, with rates of 81.1% and 82.7%, respectively [54]. Even though omadacycline had slightly lower success rates with *S. aureus*, it was still considered non-inferior to moxifloxacin [55]. Unfortunately, patients with severe CAP (graded class V in the Fine score) or those who presented in septic shock were excluded. Gastrointestinal side effects were also greater with omadacycline [56].

5.3. The ketolides: solithromycin, a next-generation macrolide

Solithromycin is the first next-generation macrolide of the fluoroketolide class [57]. Drug-binding studies indicate that it interacts with the 50S ribosomal site that either coincides or overlaps with that of other macrolides and ketolide [58]. Solithromycin binds to three distinct sites on the ribosome, whereas telithromycin binds to two sites. In terms of resistance selection, solithromycin has demonstrated a low tendency to select for resistant mutants, with little or no yields being detected in single-step studies [59]. In addition, it has potent in vitro activity against the most common bacterial pathogens in CAP, including macrolide-, penicillin-, and fluoroquinolone- resistant *S. pneumoniae* isolates, as well as *Haemophilus influenzae* and atypical bacteria.

Pharmacodynamically, solithromycin is 16-times more potent than either azithromycin, clarithromycin, or telithromycin against gram-positive aerobes and both atypical and gram-negative CAP pathogens [58]. It has good oral bioavailability (67%) when given orally and is not influenced by concomitant food intake, it serum protein binding is 81%, and its biologic half-life of 8.5 h, allowing once daily dosing [60]. The major metabolic pathway appears to involve cytochrome P450 (CYP) 3A4, with most of the metabolite undergoing biliary excretion[61].

A phase II trial evaluated the safety and efficacy of solithromycin in the treatment of bacterial CAP (ClinicalTrials.gov registration no. NCT01168713) for the treatment of uncomplicated urogenital gonorrhea [62]. Recently, two phase III RCTs were completed for the treatment of mild-to-moderate bacterial CAP (NCT01968733 and NCT01756339). Solithromycin demonstrated noninferiority to moxifloxacin within 72 h, meeting the FDAs primary endpoint, but was inferior at 5–10 days, as required by the EMA. The efficacy and safety of oral solithromycin versus oral moxifloxacin in the treatment of bacterial CAP was also assessed in a global noninferiority RCT [63]. In the separate SOLITAIRE-IV phase III trial, the efficacy and safety of intravenous-tooral solithromycin was assessed against intravenous-to-oral moxifloxacin for the treatment of bacterial CAP [64]. All patients began treatment with 400 mg intravenous solithromycin or moxifloxacin before switching to oral dosing when clinically indicated for a total treatment duration of 7 days. Oral dosing was 800 mg daily then 400 mg daily for solithromycin, or 400 mg daily for moxifloxacin. In this, 79.3% of patients who received solithromycin showed early clinical response compared with 79.7% of patients who received moxifloxacin. Finally, solithromycin is being tested in a phase II/III pivotal trial for bacterial CAP in a pediatric cohort (age 2 months to 17 years) (NCT02605122).

It appears that solithromycin does cause more adverse events (34%; mainly at the infusion site) compared with moxifloxacin (13%), though it does not prolong the cardiac QT interval. Solithromycin is chemically and biologically differentiated from telithromycin by its side chain, which does not significantly block nicotinic acetylcholine receptors. This could potentially reduce blurry vision, exacerbations of myasthenia gravis, loss of consciousness, and idiosyncratic hepatic failure reported with ketolides [65]. Nevertheless, hepatic safety remains a concern, with 5%–10% of patients experiencing mild transaminase elevations. Given the relatively small sample sizes in which solithromycin was studied, the FDA believes this may underestimate this particular safety risk. For this reason, on December 29th, 2016, the FDA recommended further studies, increasing the number of exposed patients from 924 to approximately 12,000 in an effort to evaluate the risk of hepatic toxicity

before formally granting approval [66, 67].

5.4. Quinolones: nemonoxacin, zabofloxacin, and delafloxacin

The quinolones levofloxacin and moxifloxacin have both been applied in the treatment of CAP, with each having antibacterial efficacy against major gram-positive and gram-negative respiratory tract pathogens. Novel quinolones therefore hold promise for the treatment of CAP even though fluoroquinolone resistant gram-negative and gram-positive respiratory tract pathogens are detected worldwide [68].

5.4.1 Nemonoxacin

Nemonoxacin is a novel C-8-methoxy non-fluorinated quinolone that targets DNA gyrase and topoisomerase IV, having a broader profile of activity and reduced resistance profile compared with other fluoroquinolones. TaiGen in-licensed nemonoxacin from Procter & Gamble Healthcare in 2011, when it obtained worldwide rights [69]. The drug benefits from a broad spectrum of activity against gram-positive, gram-negative, and atypical pathogens, including activity against MRSA (MIC₉₀ = 1 μ g/mL) and vancomycin-resistant pathogens, with similar activity to levofloxacin and moxifloxacin against most gram-negative bacteria [70]. It has also been shown to exhibit the best in vitro activity against *Nocardia spp*. among all tested antibiotics, including fluoroquinolones, carbapenems, tigecycline, and linezolid [71]. However, it exhibits poor activity against both MDR and non-MDR strains of *Mycobacterium tuberculosis*.

According to pharmacokinetic studies, the oral absorption of 500 mg nemonoxacin is rapid [70]. It has a mean time to maximum plasma concentration of 1–2 h, a half-life of 12.83–18.56 h, and a near 100% bioavailability. Moreover, it has low serum protein binding (16%) and no known interaction with cytochrome P450 isoenzymes, giving it minimal potential drug-drug interactions. Food significantly decreases both the maximum serum concentration (by 34%–46%) and the area under the curve (by 18%–27%) but increases the time to maximum serum concentration (by two- to three-fold). It is available in both oral and intravenous forms.

To date, phase II and III trials have compared oral formulations of nemonoxacin with

levofloxacin, but only a phase II trial has compared intravenous formulations of nemonoxacin with moxifloxacin. Despite these being complete, only one phase II trial of oral nemonoxacin has been published [72,73]. In a recent phase III noninferiority RCT in Taiwan, the efficacy and safety of nemonoxacin was compared with levofloxacin in 532 adults (≥ 70 years old) with mildly to moderately severe CAP (PORT scores II-IV) [74]. This showed nemonoxacin to be non-inferior to levofloxacin, with clinical cure rates of 94% in both groups and a favorable safety profile for nemonoxacin (adverse events of the gastrointestinal and nervous systems were most common). A second phase III multicenter, noninferiority RCT has completed enrollment and publication is anticipated (NCT02205112) [75].

In Taiwan, an oral formulation of nemonoxacin was approved in March 2014 for the treatment of bacterial CAP, and it had reached market in Taiwan, Russia, the Commonwealth of Independent States, Turkey, mainland China, and Latin America (brand name Taigexyn) by the end of 2016 [76]. It has already received priority review status by the FDA as a QIDP once further phase III studies are available documenting its safety and efficacy.

5.4.2 Zabofloxacin

Zabofloxacin is a broad-spectrum fluoroquinolone manufactured by Dong Wha Pharm. Co., Ltd. (Seoul, Republic of Korea). It has proven bactericidal efficacy both in vitro and in vivo against major gram-positive and gram-negative communityacquired pathogens of the respiratory tract, including *S. pneumoniae*, *S. aureus*, *H. influenzae*, *and M. catarrhalis*. By contrast, zabofloxacin has no activity against major pathogens associated with nosocomial pneumonia, such as *P. aeruginosa* and *A. baumannii* [77, 78]. The support for zabofloxacin's antibacterial efficacy and information about its pharmacokinetic profile have been gathered from three clinical trials. In a phase III, noninferiority RCT efficacy of 367 mg oral zabofloxacin once daily for 5 days was comparable to 400 mg moxifloxacin once daily for 7 days [79]. In a phase II RCT of the safety and efficacy of 400 mg oral zabofloxacin for 5 days compared to 500 mg oral levofloxacin for 7 days in CAP was terminated in 2012.

5.4.3 Delafloxacin

Delafloxacin, currently marketed as Baxdela in the US, is a fluoroquinolone with activity against a variety of gram-positive bacteria, including methicillin- and quinolone-resistant *S. aureus*, as well as quinolone-resistant strains of *P. aeruginosa* or *K. pneumonia* 80,81].

Its unique chemical structure facilitates improved cellular transmembrane penetration and potency in acidic environments common to most infectious sites. It has a novel mechanism of action that inhibits bacterial DNA replication by binding concurrently to both topoisomerase IV and DNA gyrase, in contrast to older fluoroquinolones that only inhibit either enzyme. The dual targeting of gyrase and topoisomerase IV decreases likelihood of resistance, which requires the accumulation of multiple mutations affecting both enzymes. This feature may contribute to the activity of delafloxacin against MRSA isolates, including those harboring mutations in the quinolone resistance determining region (QRDR) and to the low levels of resistance to delafloxacin among these MRSA isolates [82].

It also has greater affinity for DNA gyrase that older fluoroquinolones [83], which contributes to it having MICs that are consistently three- to five-fold lower. These features mean that delafloxacin not only exhibits a broader spectrum of activity but also confers less resistance compared to other fluoroquinolones.

Indeed, a 2014 surveillance study revealed that all European and 98% of USA isolates of *S. pneumoniae* were inhibited at <0.03 mg/L [84], with susceptibility rates among USA *E. coli and K. pneumoniae* isolates of 65% and 78%, respectively. Delafloxacin is also active against *P. aeruginosa* (MIC_{50/90} 0.25/4 mg/l; 65% susceptible; comparable to other anti-pseudomonal fluoroquinolones) [85] as well as anaerobes, atypical respiratory tract pathogens (e.g., *Legionella, Chlamydia*, and *Mycoplasma*), and even against *M. tuberculosis*. Moreover, the biofilm penetration capacity of has been reported to vary between 0.6% and 52%, with an acidic environment favoring antibiotic entry [86].

The bioavailability of delafloxacin is 58% when given orally and it can be dosed without regard to food. It is only 16% protein bound and has a half-life of 3.7 h following a single 300 mg intravenous dose, which ranged from 4.2 to 8.5 h following multiple oral doses per day. Delafloxacin shares the class characteristic of high pulmonary distribution with a 13:1 mean penetration ratio into the pulmonary ELF compared with the free plasma concentration [87].

The role of delafloxacin for the treatment of CAP has shown promise in two phase II studies. In a phase II RCT, 309 outpatients affected by CAP were treated with once daily oral administration of delafloxacin at different dosages (100 mg, 200 mg, or 400 mg) for 7 days, with overall clinical and bacteriologic cure rates achieved in up to 87% of patients. A phase III RCT (DEFINE-BCAP) has also compared delafloxacin with either moxifloxacin or linezolid for the treatment of adults with bacterial CAP, including MRSA [88]. To date, the incidence of adverse effects appears to be dose dependent, with diarrhea and nausea being most common; others include mild central nervous system effects, endocrine abnormalities, and increased serum liver function tests. However, no cases of clinically relevant prolongations of the QT/QTc interval have been reported in healthy volunteers [78]. It is also worth noting that delafloxacin is currently in phase III clinical development for the treatment of uncomplicated gonorrhea (as a single 900 mg oral dose) [89], and that phase III trials have determined that it is non-inferior to vancomycin or aztreonam for the treatment of ABSSSI [90,91].

In 2017, delafloxacin received FDA approval for the treatment of ABSSSIs caused by gram-positive and gram-negative bacteria, including MRSA, and is available in intravenous and oral formulations. The recommended dosage is 300 mg administered intravenously over 60 minutes or 450 mg administered orally every 12 h for a maximum of 14 days [92]. Delafloxacin has been designated a QIDP by the FDA and has been granted fast track status for CAP once phase III trials are complete.

5.5. The pleuromutilins: Lefamulin

Pleuromutilin antibiotics are semisynthetic derivatives of pleuromutilin, a natural product of the fungi *Pleurotus mutilus* (now called *Clitopilus scyphoides*). These function by inhibiting bacterial protein synthesis through binding to the peptidyl transferase site on 23S RNA of the 50S ribosome. Retapamulin was the first agent to be approved by the FDA in 2006, but this was only for topical use in impetigo. Another pleuromutilin, tiamulin, has been used with success as a veterinary drug in Europe and Canada [33]. However, lefamulin is the first pleuromutilin to be developed for oral or intravenous use in humans.

Lefamulin works by binding to the 50S subunit of the bacterial ribosome at four distinctive binding sites in a highly conserved core of the ribosomal peptidyl transferase center. This unique mechanism of action also results in a lack of cross-resistance to most currently marketed antibiotics and has inspired the development of analogs in an attempt to find clinically desirable derivatives [93]. Indeed, the drug has potent in vitro activity against organisms resistant to β -lactam antibiotics, fluoroquinolones, macrolides, tetracyclines, and vancomycin.

Lefamulin achieves extensive penetration and accumulation in pulmonary epithelial lining fluid, with epithelial lining fluid exposure to lefamulin being 5.7-fold higher than the unbound fraction in plasma [94]. It is 80% protein bound in the serum, has a biologic half-life of 12 hours, and is largely excreted unchanged via the gastrointestinal tract (86%), with the remainder eliminated via the kidneys (14%) [95]. Intravenous dosing is only required once per day, but the oral route requires dosing twice per day. Pharmacodynamically, it is highly potent with an MIC₉₀ that is four-times lower than either oxacillin, vancomycin, linezolid, or ceftaroline for *S. aureus* species (including MRSA) and many other common CAP-associated pathogens [96,97]. In addition, it has displayed potency against Neisseria spp., including MDR and extensively drug resistant N. gonorrhea isolates, supporting its potential use against gonorrhea [98].

Two phase III, noninferiority RCTs have compared the efficacy and safety of lefamulin with moxifloxacin for adult patients with mildly to moderately severe CAP, associated with PORT scores of II–IV (ClinicalTrials.gov identifier: NCT02559310 & NCT02813694) [99,100]. In LEAP 2, lefamulin met the primary endpoint for noninferiority (within 10%) set by the EMA based on investigator assessment of clinical response rates at 5–10 days following therapy completion in modified intent-to-treat and clinically evaluable at test-of-cure populations [101]. The reported side effects mainly consist of headaches (7%), nausea (7%), and diarrhea/vomiting (4%) [102]. Intravenous and oral formulations of lefamulin are currently undergoing FDA review for use in the treatment of CAP.

6. Antiviral options

6.1. Favipiravir

Favipiravir is a selective and potent inhibitor of influenza viral RNA polymerase [103] with efficacy against all subtypes and strains of influenza (A, B and C), including those that are sensitive or resistant to neuraminidase and M2 inhibitors. Interestingly, favipiravir has antiviral activity against other RNA viruses, including Bunyaviridae, Arenaviridae, Filoviridae, Flaviviridae, and Picornaviridae [104-108]. In 2010, Sleeman et al. evaluated the in vitro ability of favipiravir to block the proliferation of representative influenza viruses: 2009 A(H1N1) strains, A(H1N1) and A(H1N2) viruses of swine origin which were isolated from humans during 2007 and 2008, A(H2N2), A(H4N2), and A(H7N2), A(H5N1). This included strains that were resistant to either oseltamivir, zanamivir, or both, yet favipiravir retained antiviral activity against a wide range of influenza viruses, including the resistant ones. Currently, favipiravir is approved in Japan, where its use is restricted to patients infected with influenza virus resistant to neuraminidase inhibitors or in the event of a pandemic. Unfortunately, favipiravir is associated with a risk for teratogenicity and embryotoxicity.

6.2 Pimodivir

Pimodivir inhibits the protein basic 2 (PB2) subunit of the influenza A virus polymerase complex, thereby inhibiting viral replication[109]. It has in vitro activity against influenza A virus, including influenza pandemic 2009 H1N1, H7N9, H5N1, and strains resistant to neuraminidase and amantadine. Although it lacks activity against influenza virus B, in vitro studies have demonstrated synergy between pimodivir and oseltamivir.

In a phase IIa study [110] of 104 healthy volunteers infected with an influenza virus A (H3N2), 72 received pimodivir and 32 received placebo. The pimodivir was given once daily for 5 days from 24 h after viral inoculation at doses of 100 mg, 400 mg, 900/600 mg (as a loading dose), or 1,200/600 mg (as a loading dose). The authors significantly reduced viral shedding, influenza-like symptoms, and clinical symptoms in all dose groups compared with placebo, and that these doses were generally safe and well tolerated. In the subsequent phase IIb study [111], adults with acute uncomplicated influenza A infection were given one of the following treatments twice daily for 5 days: placebo, pimodivir 300 mg, pimodivir 600 mg, or pimodivir 600 mg

plus oseltamivir 75 mg. Pimodivir produced significant virologic improvements over placebo, with or without oseltamivir, showing a trend in clinical improvement and not serious adverse events. Currently, a phase III test among patients hospitalized with influenza A virus infection is ongoing.

6.3. Presatovir

Presatovir is an antiviral that inhibits the fusion of RSV with host cell membranes. Its efficacy has been demonstrated in preclinical and clinical studies [112–114], with in vitro activity against both major RSV strains (A and B). Studies have specifically reported lower viral loads after healthy volunteers infected with RSV received presatovir [113, 115]. Moreover, these studies have reported a favorable safety profile.

6.4. Other agents

Ziresovir is a fusion protein inhibitor with purported activity against RSV and is currently undergoing phase II clinical trials. Lumicitabine, a nucleoside analog prodrug, is also in clinical development. However, there is limited data on either of these new drugs.

7. Conclusion

CAP management requires prompt and adequate antibiotic therapy. Newly approved and investigational agents for the treatment of CAP hold promise that we can enhance our antibiotic armamentarium. However, we must first ensure that these new antibiotics are not only effective and well tolerated in patients but also that their use is kept appropriate so that we can avoid the emergence of resistance.

8. Expert opinion

Although the specific of antimicrobial therapy for CAP vary between countries, β lactams, fluoroquinolones, and macrolides are among the most commonly recommended agents. Ceftaroline, a new cephalosporine, may be considered a reasonable future option for empirical therapy in areas with a high prevalence of MRSA or drug-resistant pathogens, rather than being kept as a reserved drug. Hope exists with the development of novel agents from known antibiotic classes, such as solithromycin (a ketolide) and zabofloxacin and delafloxacin (fluoroquinolones).

Solithromycin benefits from having multiple binding sites in the 50S ribosome, making it more potent than other macrolides. Its oral formulation appears to be clinically effective, well tolerated, and to be suitable for once daily dosing over 5 days. We anticipate that its development will be pursued to treat infections in children and pregnancy given its desirable antibacterial activities and safety profile in preclinical studies. Omadacycline (a newly approved cycline) offers a single-agent parenteral or oral alternative to traditional empirical therapy in bacterial CAP. Additionally, it is significantly more active than doxycycline or minocycline against Enterobacteriaceae and *A. baumannii*, and meets the FDA criterion for susceptibility of *K. pneumoniae*. Unfortunately, patients with severe CAP (Fine class V) or those who presented in septic shock were excluded from the RCTs of solithromycin and omadacycline, so we lack clinical data for their use in those patients.

Importantly, delafloxacin could represent a highly promising option for the treatment of CAP based on broad-spectrum activity (including MRSA), oral formulation, diminished risk of resistance selection, and a favorable tolerability profile. The chemical structure of delafloxacin (lack of basic group in position C7) also makes it a non-zwitterion agent that facilitates it's in vivo activity in acidic mediums. Together with its in vitro sensitivity against anaerobes, lipid solubility, and pulmonary diffusion, this could make it a viable empirical treatment in cases of aspiration pneumonia or bacterial pulmonary abscesses. However, fluoroquinolones have increasingly fallen from favor because of rare adverse events, including tendinopathies, neuropathies, and aortic dissection. On this basis, omadacycline probably represents the most appropriate first-choice empirical treatment for CAP.

A major limitation of the antimicrobials described in this review is a lack of potency against pathogens responsible for nosocomial pneumonia. Infections caused by MDR *P. aeruginosa, A. baumannii,* and *Stenotrophomonas maltophilia* will therefore continue to pose clinical challenges because of the limited availability of potent antimicrobials against these gram-negative pathogens. Although there are several promising antibiotics in development, regulatory approval over the next five years will be key to any return to the availability of a more comprehensive antimicrobial

armamentarium that can cope with the broad spectrum of bacterial disease presenting in CAP.

Funding

This paper was not funded.

Declaration of interest

The authors have no relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript. This includes employment, consultancies, honoraria, stock ownership or options, expert testimony, grants or patents received or pending, or royalties.

Reviewer disclosures

Peer reviewers on this manuscript have no relevant financial or other relationships to disclose.

References

Papers of special note have been highlighted as:

* of interest

** of considerable interest

1. Torres A, Cillóniz C, Blasi F, Chalmers JD, Gaillat J, Dartois N, Schmitt H-J, Welte T. Burden of pneumococcal community-acquired pneumonia in adults across Europe: A literature review. *Respir Med* 2018; 137: 6–13.

2. Feldman C, Anderson R. Pneumonia as a systemic illness. *Curr Opin Pulm Med* 2018; 24: 237–243.

3. Eurich DT, Marrie TJ, Minhas-Sandhu JK, Majumdar SR. Ten-year mortality after community-acquired pneumonia. A prospective cohort. Am J Respir Crit Care Med 2015; 192:597–604

4. Hayes BH, Haberling DL, Kennedy JL, et al. Burden of pneumonia-associated hospitalizations: United States. Chest 2018; 153:427–437.

*Provides trends in hospitalizations related to pneumonia and their changing patterns.

 Cillóniz C, Rodríguez-Hurtado D, Torres A. Characteristics and Management of Community-Acquired Pneumonia in the Era of Global Aging. *Med Sci (Basel)* 2018;
6.

6. Cillóniz C, Dominedò C, Torres A. Multidrug Resistant Gram-Negative Bacteria in Community-Acquired Pneumonia. *Crit Care* 2019; 23: 79.

7. Cillóniz C, Dominedò C, Nicolini A, Torres A. PES Pathogens in Severe Community-Acquired Pneumonia. *Microorganisms* 2019; 7.

8. Aliberti S, Cilloniz C, Chalmers JD, Zanaboni AM, Cosentini R, Tarsia P, et al. Multidrug-resistant pathogens in hospitalised patients coming from the community with pneumonia: a European perspective. Thorax. 2013 Nov;68(11):997-9.

9. Bermejo-Martin JF, Cilloniz C, Mendez R, Almansa R, Gabarrus A, Ceccato A, Torres A, Menendez R, NEUMONAC group. Lymphopenic Community Acquired Pneumonia (L-CAP), an Immunological Phenotype Associated with Higher Risk of Mortality. *EBioMedicine* 2017; 24: 231–236. 10. Méndez R, Menéndez R, Amara-Elori I, Feced L, Piró A, Ramírez P, et al. Lymphopenic community-acquired pneumonia is associated with a dysregulated immune response and increased severity and mortality. *J. Infect.* 2019;.

11. Cillóniz C, Ewig S, Polverino E, Marcos MA, Esquinas C, Gabarrús A, et al. Microbial aetiology of community-acquired pneumonia and its relation to severity. *Thorax* 2011; 66: 340–346.

12 Jain S, Self WH, Wunderink RG, Fakhran S, Balk R, Bramley AM, Reed C, Grijalva CG, et al, et al. Community-Acquired Pneumonia Requiring Hospitalization among U.S. Adults. *N. Engl. J. Med.* 2015; 373: 415–427.

13. Radovanovic D, Sotgiu G, Jankovic M, Mahesh PA, Marcos PJ, Abdalla MI, et al; GLIMP Study Group. An international perspective on hospitalized patients with viral community-acquired pneumonia. *Eur. J. Intern. Med.* 2019; 60: 54–70.

14. Cillóniz C, Liapikou A, Martin-Loeches I, García-Vidal C, Gabarrús A, Ceccato A, Magdaleno D, et al. Twenty-year trend in mortality among hospitalized patients with pneumococcal community-acquired pneumonia. *PLoS ONE* 2018; 13: e0200504.

15. Sader HS, Mendes RE, Le J, Denys G, Flamm RK, Jones RN. Antimicrobial Susceptibility of Streptococcus pneumoniae from North America, Europe, Latin America, and the Asia-Pacific Region: Results From 20 Years of the SENTRY Antimicrobial Surveillance Program (1997-2016). *Open Forum Infect Dis* 2019; 6: S14–S23.

16. Prina E, Ranzani OT, Polverino E, Cillóniz C, Ferrer M, Fernandez L, Puig de la Bellacasa J, Menéndez R, Mensa J, Torres A. Risk factors associated with potentially antibiotic-resistant pathogens in community-acquired pneumonia. *Ann Am Thorac Soc* 2015; 12: 153–160.

17. Restrepo MI, Babu BL, Reyes LF, Chalmers JD, Soni NJ, Sibila O, et al;, GLIMP. Burden and risk factors for *Pseudomonas aeruginosa* community-acquired pneumonia: a multinational point prevalence study of hospitalised patients. *Eur. Respir. J.* 2018; 52.

18. Cillóniz C, Gabarrús A, Ferrer M, Puig de la Bellacasa J, Rinaudo M, Mensa J, et al. Community-Acquired Pneumonia Due to Multidrug- and Non-Multidrug-Resistant Pseudomonas aeruginosa. *Chest* 2016; 150: 415–425.

19. Obed M, García-Vidal C, Pessacq P, Mykietiuk A, Viasus D, Cazzola L, et al. [Clinical features and outcome of community-acquired methicillin-resistant *Staphylococcus aureus* pneumonia]. *Enferm. Infecc. Microbiol. Clin.* 2014; 32: 23– 27.

20. Self WH, Wunderink RG, Williams DJ, Zhu Y, Anderson EJ, Balk RA, et al. *Staphylococcus aureus* Community-acquired Pneumonia: Prevalence, Clinical Characteristics, and Outcomes. *Clin. Infect. Dis.* 2016; 63: 300–309.

21. Moran GJ, Krishnadasan A, Gorwitz RJ, Fosheim GE, Albrecht V, Limbago B, Talan DA, EMERGEncy ID NET Study Group. Prevalence of methicillin-resistant staphylococcus aureus as an etiology of community-acquired pneumonia. *Clin. Infect. Dis.* 2012; 54: 1126–1133.

22. Kim YA, Kim JJ, Won DJ, Lee K. Seasonal and Temperature-Associated Increase in Community-Onset Acinetobacter baumannii Complex Colonization or Infection. *Ann Lab Med* 2018; 38: 266–270.

23. Ong CWM, Lye DCB, Khoo KL, Chua GSW, Yeoh SF, Leo YS, Tambyah PA, Chua AC. Severe community-acquired *Acinetobacter baumannii* pneumonia: an emerging highly lethal infectious disease in the Asia-Pacific. *Respirology* 2009; 14: 1200–1205.

24. Serota DP, Sexton ME, Kraft CS, Palacio F. Severe Community-Acquired Pneumonia due to *Acinetobacter baumannii* in North America: Case Report and Review of the Literature. *Open Forum Infect Dis* 2018; 5: ofy044.

25. Mody L, Gibson KE, Horcher A, Prenovost K, McNamara SE, Foxman B, Kaye KS, Bradley S. Prevalence of and risk factors for multidrug-resistant *Acinetobacter baumannii* colonization among high-risk nursing home residents. *Infect Control Hosp Epidemiol* 2015; 36: 1155–1162.

26. Lin Y-T, Jeng Y-Y, Chen T-L, Fung C-P. Bacteremic community-acquired pneumonia due to *Klebsiella pneumoniae*: clinical and microbiological characteristics in Taiwan, 2001-2008. *BMC Infect. Dis.* 2010; 10: 307.

27. Tseng C-P, Wu H-S, Wu T-H, Lin Y-T, Fung C-P. Clinical characteristics and outcome of patients with community-onset *Klebsiella pneumoniae* bacteremia requiring intensive care. *J Microbiol Immunol Infect* 2013; 46: 217–223.

28. Rafat C, Messika J, Barnaud G, Dufour N, Magdoud F, Billard-Pomarès T, et al. Hypervirulent *Klebsiella pneumoniae*, a 5-year study in a French ICU. *J. Med. Microbiol.* 2018; 67: 1083–1089.

29. Decré D, Verdet C, Emirian A, Le Gourrierec T, Petit J-C, Offenstadt G, et al. Emerging severe and fatal infections due to *Klebsiella pneumoniae* in two university hospitals in France. *J. Clin. Microbiol.* 2011; 49: 3012–3014.

30. Venkatesan S, Myles PR, Bolton KJ, Muthuri SG, Al Khuwaitir T, Anovadiya AP, et al. Neuraminidase Inhibitors and Hospital Length of Stay: A Meta-analysis of Individual Participant Data to Determine Treatment Effectiveness Among Patients Hospitalized With Nonfatal 2009 Pandemic Influenza A(H1N1) Virus Infection. *J. Infect. Dis.* 2019;.

31. Muthuri SG, Venkatesan S, Myles PR, Leonardi-Bee J, Lim WS, Al Mamun A, et al. Impact of neuraminidase inhibitors on influenza A(H1N1) pdm09-related pneumonia: an individual participant data meta-analysis. *Influenza Other Respir Viruses* 2016; 10: 192–204.

32. U. Theuretzbacher. Antibiotic innovation for future public health needs. Review. Clinical Microbiology and Infection 23 (2017) 713e717.

33. Prabhavathi Fernandes, Evan Martens. Antibiotics in late clinical development. Biochemical Pharmacology 133 (2017) 152–163.

34. Matteo Bassetti, Maria Merelli, Chiara Temperoni and Augusta Astilean. New antibiotics for bad bugs: where are we?Annals of Clinical Microbiology and Antimicrobials 2013, 12:22.

35.Mark A.T. Blaskovich, Mark S. Butler and Matthew A. Cooper. Polishing the tarnished silver bullet: the quest for new antibiotics. Review Article. Essays in Biochemistry (2017) 61 103–114.

36. Adamantia Liapikou, Catia Cillóniz, and Antonio Torres. Ceftobiprole for the treatment of pneumonia: a European perspective. Drug Des Devel Ther. 2015; 9: 4565–4572.

37. Carreno JJ, Lodise TP. Ceftaroline Fosamil for the treatment of community acquired pneumonia: from FOCUS to CAPTURE. Infect Dis Ther 2014; 3:123–132.

38. Barber KE, Ireland CE, Bukavyn N. Observation of "seesaw effect" with vancomycin, teicoplanin, daptomycin and ceftaroline in 150 unique MRSA strains. *Infect Dis Ther* 2014; 3:35-43.

39. Mendes RE, Tsakris A, Sader HS, Jones RN, Biek D, McGhee P, et al. Characterization of methicillin-resistant *Staphylococcus aureus* displaying increased MICs of ceftaroline. J Antimicrob Chemother 2012;67:1321e4.

40. File TM, Low DE, Eckburg PB, et al. FOCUS 1: a randomized, double blinded, multicentre, Phase III trial of the efficacy and safety of ceftaroline fosamil versus ceftriaxone in community-acquired pneumonia. J Antimicrob Chemother 2011; 66(Suppl 3):iii19–32.

41. Low DE, File TM, Eckburg PB, et al. FOCUS 2: a randomized, double blinded, multicentre, Phase III trial of the efficacy and safety of ceftaroline fosamil versus ceftriaxone in community-acquired pneumonia. J Antimicrob Chemother 2011;66(Suppl 3): iii33–44.

42odise TP, Anzueto AR, Weber DJ, et al. Assessment of time to clinical response, a proxy for discharge readiness, among hospitalized patients with community-acquired pneumonia who received either ceftaroline fosamil or ceftriaxone in two phase III FOCUS trials. Antimicrob Agents Chemother 2015; 59:1119–1126.

43. Johnson LB, Ramani A, Guervil DJ.Use of Ceftaroline Fosamil in Osteomyelitis: CAPTURE Study Experience.BMC Infect Dis. 2019 Feb 21;19(1):183.

44. Destache CJ, Guervil DJ, Kaye KS. Ceftaroline fosamil for the treatment of Gram-positive endocarditis: CAPTURE study experience. Int J Antimicrob Agents. 2019 May;53(5):644-649.

45. Furtek KJ, Kubiak DW, Barra M, Varughese CA, Ashbaugh CD, Koo S. High incidence of neutropenia in patients with prolonged ceftaroline exposure. J Antimicrob Chemother 2016; 71:2010e23.

46. Shirley DA, Heil EL, Johnson JK. Ceftaroline fosamil: a brief clinical review. Infect Dis Ther. 2013; 2:95–110.

47. Durães F, Sousa E. Omadacycline: A Newly Approved Antibacterial from the Class of Tetracyclines. Pharmaceuticals (Basel). 2019 Apr 21;12(2).

48. Pfaller MA, Rhomberg PR, Huband MD, Flamm RK. Activity of omadacycline tested against *Streptococcus pneumoniae* from a global surveillance program (2014). Diagn Microbiol Infect Dis 2018; 90:143–147.

*Activity of omadacycline for the key community-acquired pneumonia pathogens is presented.

49. Noel, G.J.; Draper, M.P.; Hait, H.; Tanaka, S.K.; Arbeit, R.D. A randomized, evaluator-blind, phase 2 study comparing the safety and efficacy of omadacycline to those of linezolid for treatment of complicated skin and skin structure infections. *Antimicrob. Agents Chemother.* 2012, *56*, 5650–5654.

50. Shoen, C.; Benaroch, D.; Sklaney, M.; Cynamon, M. In vitro activities of omadacycline against rapidly growing mycobacteria. *Antimicrob. Agents Chemother.* 2019, AAC.02522-02518.

51. Gotfried MH, Horn K, Garrity-Ryan L, et al. Comparison of omadacycline and tigecycline pharmacokinetics in the plasma, epithelial lining fluid, and alveolar cells of healthy adult subjects. Antimicrob Agents Chemother 2017; 61:e01135–e1217.

52 Villano S, Steenbergen J, Loh E. Omadacycline: development of a novel aminomethylcycline antibiotic for treating drug-resistant bacterial infections. Future microbiology 2016; 11: 1421-1434.

53.

https://www.clinicaltrials.gov/ct2/show/results/NCT02531438?term=omadacycline&ra nk=4

54. Paratek Announces Positive Phase 3 Study of Omadacycline in Community-Acquired Bacterial Pneumonia". *www.globenewswire.com*. April 3, 2017. Retrieved 16 May 2017.

55. Stets R, Popescu M, Gonong JR, Mitha I, Nseir W, Madej A, et al. Omadacycline for

Community-Acquired Bacterial Pneumonia. N Engl J Med. 2019 Feb 7;380(6):517-527. 51. 56.https://www.drugs.com/history/nuzyra.html

57. Llano-Sotelo B, Dunkle J, Klepacki D, et al. Binding and action of CEM-101, a new

fluoroketolide antibiotic that inhibits protein synthesis. Antimicrob Agents Chemother 2010; 54(12): 4961-70.

58. Zhanel GG, Hartel E, Adam H, et al. Solithromycin: A Novel Fluoroketolide for the

Treatment of Community-Acquired Bacterial Pneumonia. Drugs 2016; 76:1737-1757.

* This paper thoroughly describes solithromycin and all clinical trials that have been performed by solithromycin in community- acquired bacterial pneumonia

59. McGhee P, Clark C, Kosowska-Shick KM, Nagai K, Dewasse B, Beachel L, et al. In vitro activity of CEM-101 *against Streptococcus pneumoniae and Streptococcus pyogenes* with defined macrolide resistance mechanisms. Antimicrob Agents Chemother. 2010; 54(1):230–8.

60. Jamieson BD, Ciric S, Fernandes P. Safety and pharmacokinetics of solithromycin in subjects with hepatic impairment. Antimicrob Agents Chemother. 2015; 59:4379–4386.

61. Rodvold KA, Gotfried MH, Still JG, et al. Comparison of plasma, epithelial lining fluid, and alveolar macrophage concentrations of solithromycin (CEM-101) in healthy adult subjects. Antimicrob Agents Chemother 2012; 56(10): 5076-81.

* Excellent study outlining the concentrations of solithromycin in epithelial lining fluid and alveolar macrophages.

62. Oldach D, Clark K, Schranz J, et al. Randomized, double-blind, multicenter phase 2

study comparing the efficacy and safety of oral solithromycin (CEM-101) to those of oral levofloxacin in the treatment of patients with community-acquired bacterial pneumonia. Antimicrob Agents Chemother2013;57(6):2526-34.

63. Barrera CM, Mykietiuk A, Metev H, et al; SOLITAIRE-ORAL Pneumonia Team. Efficacy and safety of oral solithromycin versus oral moxifloxacin for treatment of community-acquired bacterial pneumonia: a global, double-blind, multicentre, randomized, active-controlled, non-inferiority trial (SOLITAIRE-ORAL). Lancet Infect Dis 2016; 16(4): 421-30. * Large randomized control trial demonstrating the clinical efficacy of solithromycin in the management of patients with BCAP.

64. File Jr. TM, Rewerska B, Tanaseanu CM, et al. SOLITAIRE-IV: A Randomized, Double-Blind, Multicenter Study Comparing the Efficacy and Safety of Intravenousto-Oral Solithromycin to Intravenous-to-Oral Moxifloxacin for Treatment of Community-Acquired Bacterial Pneumonia. Clin Infect Dis. 2016 Oct 15;63(8):1007-1016.

65. Fernandes P, Martens E, Bertrand D, Pereira D. The solithromycin journey-it is all in the chemistry. Bioorg Med Chem. 2016. Dec 15; 24(24):6420-6428.

66. FDA Briefing document Solithromycin Oral Capsule and Injection Meeting of the Antimicrobial Drugs Advisory Committee (AMDAC). https://www.fda.gov November, 4 2016.

67. https://www.drugs.com/history/solithera.html

68. Rodríguez-Martínez JM, Machuca J, Cano ME, et al. quinolone resistance: two decades on. Drug Resist Updat. 2016;29:13–29.

69. Qin X, Huang H. Review of nemonoxacin with special focus on clinical development. Drug Des Devel Ther. 2014;8:765-74.

70. Guo B, Wu X, Zhang Y, et al. Safety and clinical pharmacokinetics of nemonoxacin, a

novel non-fluorinated quinolone, in healthy Chinese volunteers following single and multiple oral doses. Clinical drug investigation 2012; 32:475-486.

71. Lai CC, Tan CK, Lin SH, et al. Comparative in vitro activities of nemonoxacin, doripenem, tigecycline and 16 other antimicrobials against *Nocardia brasiliensis, Nocardia asteroides* and unusual *Nocardia species*. J Antimicrob Chemother 2009; 64: 73-8.

72. Liu Y, Zhang Y, Wu J, Zhu D, Sun S, Zhao L, Wang X, Liu H, Ren Z, Wang C, et al. A randomized, double-blind, multicenter Phase II study comparing the efficacy and safety of oral nemonoxacin with oral levofloxacin in the treatment of community-acquired pneumonia. J Microbiol Immunol Infect. 2017 Dec;50(6):811-820.

73. van Rensburg DJ, Perng RP, Mitha IH, et al. Efficacy and safety of nemonoxacin

versus levofloxacin for community acquired pneumonia. Antimicrob Agents Chemother 2010; 54: 4098-106.

74. Cheng S, Wu R, Hsu Z, et al. Efficacy and Safety of Oral Nemonoxacin in Treatment of Community-Acquired Pneumonia: Subgroup Analysis Results in Taiwanese Patients in a

Randomized, Double-Blind, Multi-Center, Phase III Comparative Study with Levofloxacin.

2015. Available at: www.atsjournals.org/doi/abs/10.1164/ajrccmconference

2015.191.1_MeetingAbstracts.A4063.

75.TaiGen Biotechnology. A Phase III Study to Evaluate the Efficacy and Safety of Intravenous Infusion of Nemonoxacin in Treating CAP. 2016. Available at: https://clinicaltrials.gov/ct2/show/NCT02205112

76. https://adisinsight.springer.com/drugs/800022726

77. Kwon AR, Min YH, Ryu JM, et al. In vitro and in vivo activities of DW-224 a, a novel fluoroquinolone antibiotic agent. J Antimicrob Chemother. 2006;58:684–688.

78. Kocsis B, Domokos J, Szabo D. Chemical structure and pharmacokinetics of novel

quinolone agents represented by avarofloxacin, delafloxacin, finafloxacin, zabofloxacin

and nemonoxacin. Annals of clinical microbiology and antimicrobials 2016; 15:34.

79.Rhee CK, Chang JH, Choi EG, et al. Zabofloxacin versus moxifloxacin in patients with COPD exacerbation: a multicenter, double-blind, double-dummy, randomized, controlled, phase III, non-inferiority trial. Int J Chron Obstruct Pulmon Dis. 2015;10:2265–2275.

• A Phase III clinical trial investigated zabofloxacin versus moxifloxacin in patients with COPD exacerbation

80. Marra A, Bortolon E, Molstad D, et al: Evaluation of Delafloxacin in Rat Granuloma Pouch Infections Caused by Gram- Negative Pathogens. In Program and Abstract of 50th Interscience Conference on Antimicrobial Agents and Chemotherapy. Chicago, USA: Abstract; 2011. A1-680. 81. Remy JM, Tow-Keogh CA, McConnell TS, Dalton JM, Devito JA: Activity of

delafloxacin against methicillin-resistant Staphylococcus aureus: resistance selection and characterization. J Antimicrob Chemother 2012, 67(12):2814–2820.

82. Remy JM, Tow-Keogh CA, McConnell TS, Dalton JM, Devito JA. Activity of delafloxacin

against methicillin-resistant *Staphylococcus aureus*: resistance selection and characterization. J Antimicrob Chemother 2012; 67:2814–20,

83. Jorgensen SCJ, Mercuro NJ, Davis SL, Rybak MJ. Delafloxacin: place in therapy and review of microbiologic, clinical and pharmacologic properties. *Infect Dis Ther.* 2018;7(2):197-217.

84. Pfaller MA, Sader HS, Rhomberg PR, Flamm RK. *In Vitro* Activity of Delafloxacin against Contemporary Bacterial Pathogens from the United States and Europe, 2014. Antimicrob Agents Chemother. 2017 Mar 24; 61(4).

85. Sheikh J. Clinical microbiology review. Delafloxacin. NDA#208610, 208611. Melinta Therapeutics, Inc. Division of Anti-Infective Products. Center for Drug Evaluation and Research. US Food and Drug Administration; 2017.

86. Bauer J, Siala W, Tulkens PM, et al. A combined pharmacodynamics quantitative and qualitative model reveals the potent activity of daptomycin and delafloxacin against *Staphylococcus aureus* biofilms. Antimicrob Agents Chemother. 2013; 57:2726–2737.

** This study investigated delafloxacin against Staphylococcus aureus biofilms

87. Thabit AK, Crandon JL, Nicolau DP. Pharmacodynamic and pharmacokinetic profiling of delafloxacin in a murine lung model against community-acquired respiratory tract pathogens. Int J Antimicrob Agents. 2016;48(5):535–41.

88.https://www.clinicaltrials.gov/ct2/results?cond=&term=NCT02679573&cntry=&stat e=&city=&dist=

89. A comparative evaluation of the single dose efficacy of oral delafloxacin versus the single-dose efficacy of an intramuscular injection of ceftriaxone in subjects with uncomplicated urogenital gonorrhea. Identifier

NCT02015637https://www.clinicaltrials.gov/ct2/show/

NCT02015637? term=NCT02015637&rank=1 [Accessed 12 November 2014]

90.Pullman, J., Gardovskis J, Farley B, Sun E, Quintas M, Lawrence L, et al; PROCEED Study Group. et al., Efficacy and safety of delafloxacin compared with vancomycin plus aztreonam for acute bacterial skin and skin structure infections: a Phase 3, double blind, randomized study. J Antimicrob Chemother, 2017. 72(12): p. 3471-3480.

91. O'Riordan, W., McManus A, Teras J, Poromanski I, Cruz-Saldariagga M, Quintas M, et al., A Comparison of the Efficacy and Safety of Intravenous Followed by Oral Delafloxacin With Vancomycin Plus Aztreonam for the Treatment of Acute Bacterial Skin and Skin Structure Infections: A Phase 3, Multinational, Double- Blind, Randomized Study. Clin Infect Dis, 2018. 67(5): p. 657-666.

92. Tulkens PM, Van Bambeke F, Zinner SH.Profile of a Novel Anionic Fluoroquinolone-Delafloxacin. Clin Infect Dis. 2019 Apr 8;68(Supplement_3):S213-S222.

93. Novak R. 2011. Are pleuromutilin antibiotics finally fit for human use? Ann. N. Y. Acad. Sci. 1241:71–81.

94. Zeitlinger M, Schwameis R, Burian A, et al. Simultaneous assessment of the pharmacokinetics of a pleuromutilin, lefamulin, in plasma, soft tissues and pulmonary epithelial lining fluid. J Antimicrob Chemother 2016; 71:1022–1026.

95. Sader HS, Paukner S, Ivezic-Schoenfeld Z, et al. Antimicrobial activity of the novel pleuromutilin antibiotic BC-3781 against organisms responsible for community-acquired

respiratory tract infections (CARTIs). The Journal of antimicrobial chemotherapy 2012;

67:1170-1175.

96. Paukner S, Gelone SP, Arends SJR, Flamm RK, Sader HS Antibacterial Activity of Lefamulin against Pathogens Most Commonly Causing Community-Acquired Bacterial Pneumonia: SENTRY Antimicrobial Surveillance Program (2015-2016).Antimicrob Agents Chemother. 2019 Mar 27;63(4).

97. Prince WT, Ivezic-Schoenfeld Z, Lell C, et al. Phase II clinical study of BC-3781,

pleuromutilin antibiotic, in treatment of patients with acute bacterial skin and skin structure

infections. Antimicrobial agents and chemotherapy 2013; 57:2087-2094.

98. Jacobsson, S., et al., In Vitro Activity of the Novel Pleuromutilin Lefamulin (BC-3781)

and Effect of Efflux Pump Inactivation on Multidrug-Resistant and Extensively Drug-

Resistant Neisseria gonorrhoeae. Antimicrob Agents Chemother, 2017, 61(11).

99. File TM Jr, Goldberg L, Das A, Sweeney C, Saviski J, Gelone SP, et al. Efficacy and Safety of IV-to-Oral Lefamulin, a Pleuromutilin Antibiotic, for Treatment of Community-Acquired Bacterial Pneumonia: The Phase 3 LEAP 1 Trial. Clin Infect Dis. 2019 Feb 4.

100. Seltzer E. Study to Compare Lefamulin to Moxifloxacin (With or Without Linezolid) for

the Treatment of Adults With Pneumonia (LEAP). 2016. Available at:

https://clinicaltrials.gov/ct2/show/NCT02559310.

101. Gasink L. Study to Compare Lefamulin to Moxifloxacin for the Treatment of Adults With

Pneumonia (LEAP2). 2016. Available at: https://clinicaltrials.gov/ct2/show/NCT02813694.

102. Eyal Z, Matzov D, Krupkin M, et al. A novel pleuromutilin antibacterial compound, its

binding mode and selectivity mechanism. Scientific reports 2016;6:39004.

103. Furuta Y, Takahashi K, Kuno-Maekawa M, Sangawa H, Uehara S, Kozaki K, Nomura N, Egawa H, Shiraki K. Mechanism of action of T-705 against influenza virus. *Antimicrob. Agents Chemother.* 2005; 49: 981–986.

104. Smither SJ, Eastaugh LS, Steward JA, Nelson M, Lenk RP, Lever MS. Postexposure efficacy of oral T-705 (Favipiravir) against inhalational Ebola virus infection in a mouse model. *Antiviral Res.* 2014; 104: 153–155.

105. Gowen BB, Wong M-H, Jung K-H, Sanders AB, Mendenhall M, Bailey KW,

Furuta Y, Sidwell RW. In vitro and in vivo activities of T-705 against arenavirus and bunyavirus infections. *Antimicrob. Agents Chemother.* 2007; 51: 3168–3176.

106.Gowen BB, Smee DF, Wong M-H, Hall JO, Jung K-H, Bailey KW, Stevens JR, Furuta Y, Morrey JD. Treatment of late stage disease in a model of arenaviral hemorrhagic fever: T-705 efficacy and reduced toxicity suggests an alternative to ribavirin. *PLoS ONE* 2008; 3: e3725.

107.Mendenhall M, Russell A, Smee DF, Hall JO, Skirpstunas R, Furuta Y, Gowen BB. Effective oral favipiravir (T-705) therapy initiated after the onset of clinical disease in a model of arenavirus hemorrhagic fever. *PLoS Negl Trop Dis* 2011; 5: e1342.

108.Safronetz D, Rosenke K, Westover JB, Martellaro C, Okumura A, Furuta Y, et al. The broad-spectrum antiviral favipiravir protects guinea pigs from lethal Lassa virus infection post-disease onset. *Sci Rep* 2015; 5: 14775.

109. Clark MP, Ledeboer MW, Davies I, Byrn RA, Jones SM, Perola E, et al. Discovery of a novel, first-in-class, orally bioavailable azaindole inhibitor (VX-787) of influenza PB2. *J. Med. Chem.* 2014; 57: 6668–6678.

110. Trevejo JM, Asmal M, Vingerhoets J, Polo R, Robertson S, Jiang Y, et al. Pimodivir treatment in adult volunteers experimentally inoculated with live influenza virus: a Phase IIa, randomized, double-blind, placebo-controlled study. *Antivir. Ther. (Lond.)* 2018; 23: 335–344.

111.Finberg RW, Lanno R, Anderson D, Fleischhackl R, van Duijnhoven W, Kauffman RS, et al. Phase 2b Study of Pimodivir (JNJ-63623872) as Monotherapy or in Combination With Oseltamivir for Treatment of Acute Uncomplicated Seasonal Influenza A: TOPAZ Trial. *J. Infect. Dis.* 2019; 219: 1026–1034.

109.Perron M, Stray K, Kinkade A, Theodore D, Lee G, Eisenberg E, et al. GS-5806 Inhibits a Broad Range of Respiratory Syncytial Virus Clinical Isolates by Blocking the Virus-Cell Fusion Process. *Antimicrob. Agents Chemother.* 2015; 60: 1264– 1273.

110. DeVincenzo JP, Whitley RJ, Mackman RL, Scaglioni-Weinlich C, Harrison L, Farrell E, et al. Oral GS-5806 activity in a respiratory syncytial virus challenge study. *N. Engl. J. Med.* 2014; 371: 711–722.

111. Jordan R, Shao M, Mackman RL, Perron M, Cihlar T, Lewis SA, et al. Antiviral Efficacy of a Respiratory Syncytial Virus (RSV) Fusion Inhibitor in a Bovine Model of RSV Infection. *Antimicrob. Agents Chemother.* 2015; 59: 4889–4900.

112. Mackman RL, Sangi M, Sperandio D, Parrish JP, Eisenberg E, Perron M, et al. Discovery of an oral respiratory syncytial virus (RSV) fusion inhibitor (GS-5806) and clinical proof of concept in a human RSV challenge study. *J. Med. Chem.* 2015; 58: 1630–1643.

Accepted Manuts

TABLES 1: Antibacterial agents in the pipeline for CAP

Drug Class	Drug Name	Spectrum	Dose	Development	Indications
				phase for CAP	
			7 00 (7 4)		
Fluoroquinolone	Nemonoxacın	MRSA, VRE,	500mg /24h	Phase III	ABSSI, BCAD diabati
		A. Duumunnii		i v	c foot ulcor
				formulation.	infections
				under US IND:	
				phase III oral	
				formulation	
				complete	
			-	complete	
	Delafloxacin	Gram+ and –,	Dosage for IV	Phase III for	Approved in
	Baxdela	including	use is 300	VAP	2017 IOI Acute
		MRSA	and 450 mg		bacterial skin
		1111071	/12h p.o.		and skin
					structure
					infection
	Zabofloxacin	Gram+ and –,	367mg/24h	Phase II for	AECOPD
			p.o. for 5 days	САР	BCAP
Macrolide	Solithromycin	Gram(+),	800mg	Phase III for	ABSSSI;
		including	loading dose	CABP and	Prophylaxis
		macrolide	and 400mg	uncomplicated	for N.
		resistant	once a day for	gonorrnoea, in	meningitidis;
		strains	live days	phase if for	urethritis
				paediatric use	and
					other
					urogenital
C					infections
Tetracycline	Omadacycline	Broad	100mg/24h	Phase II	FDA
		spectrum	(i.v.)	complete;	ACCEPTED
		incl MDR	()	phase III	FOR: ABSSSI
			150mg/24n	planned	AND BCAP
			p.o.		
Pleuromutilin	Lefamulin	Broad	600mg/12h	Phase III	BCAP
		spectrum	p.o.		
Cephalosporine	Ceftaroline	Gram (+),	600mg/12h		FDA
	Teflaro	gram (-), and	for 10 days		ACCEPTED

	MRSA			FOR :BCAP
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ABSSI: Acute bacterial skin and skin structure infection, BCAP: Bacterial community-acquired pneumonia