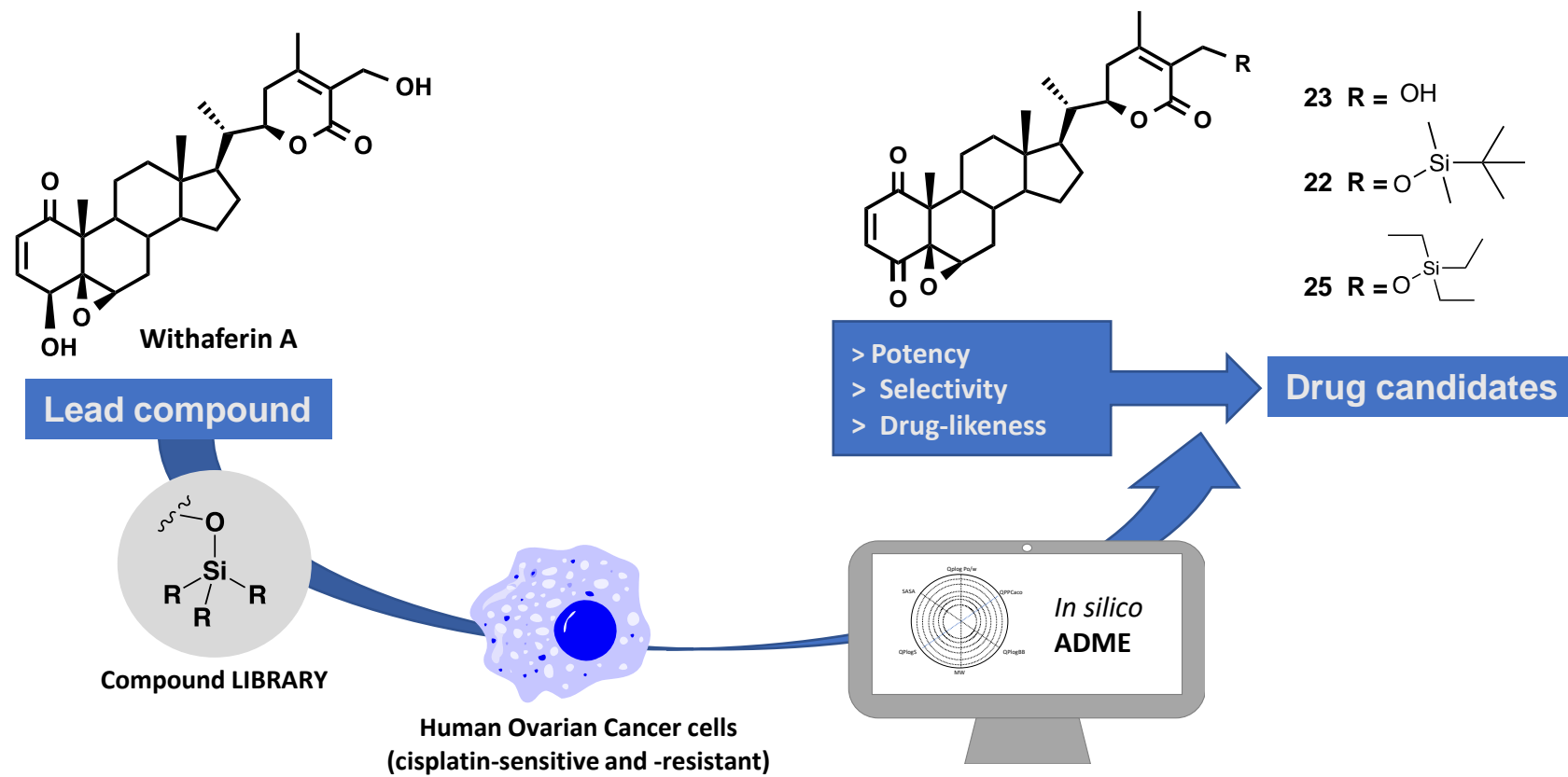


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**Expanding the Chemical Space of Withaferin A by
Incorporating Silicon to Improve its Clinical Potential on
Human Ovarian Carcinoma Cells**

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4 Expanding the Chemical Space of Withaferin A by
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8 Incorporating Silicon to Improve its Clinical
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12 Potential on Human Ovarian Carcinoma Cells
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3 ABSTRACT: Ovarian cancer represents the seventh most commonly diagnosed cancer
4 worldwide. Currently, platinum-based chemotherapy resistance and multiple severe side
5 effects represent a major hurdle for ovarian cancer patients who relapse. Therefore, there is a
6 need to enlarge the chemical space of medicinal chemistry, and in this regard, the
7 incorporation of silicon in a drug scaffold offers great potential. Herein, we report on the
8 development of a withaferin A-silyl ether library, 30 analogues are reported for the first time.
9 Cytotoxicity assays on human epithelial ovarian carcinoma cisplatin-sensitive and -resistant
10 cell lines identified several analogues displaying nanomolar-potency, higher than the lead
11 compound and reference drug. This cytotoxic potency is also coupled with a good selectivity
12 index on a non-tumoral cell line. Cell cycle analysis of two potent analogues revealed cell
13 death by apoptosis without indication of a cell cycle arrest in G0/G1 phase. The structure-
14 activity relationship (SAR) and *in silico* ADME studies demonstrated that the incorporation
15 of silicon and a carbonyl group at C-4 in the WA-framework enhances potency, selectivity
16 and drug-likeness. These findings reveal analogues **22**, **23** and **25** as potential candidates for
17 clinical translation in patients with relapsed ovarian cancer.
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INTRODUCTION

Ovarian cancer (OC) represents the seventh most commonly diagnosed cancer worldwide.¹ Current treatment entails cytoreductive surgery followed by platinum- or taxane-based chemotherapy.² Initially, OC responds positively in 70 to 80% of the cases. However, nearly 70% of patients suffer a relapse within 6 months of the last chemotherapeutic cycle, which is attributed to patients eventually developing resistance to carboplatin and paclitaxel adjuvant chemotherapy.³ Therefore, chemotherapy resistance, whether primary (i.e., intrinsic) or secondary (i.e., acquired), represents a major hurdle in OC treatment.⁴ Additionally, platinum-based chemotherapy is associated with multiple severe side effects (e.g. nausea, myelosuppression, neurotoxicity, nephrotoxicity, hepatotoxicity and ototoxicity).⁵ Thus, there is an urgent need for new second-line therapies to improve the prognosis of patients with relapsed OC. As an alternative treatment strategy to reduce the side effects and resistance caused by cis-platinum-based chemotherapy a number of combinations with other compounds have been explored. In this sense, Natural Products (NPs) are ideal candidates for OC chemoprevention or adjuvants of conventional chemotherapy. Recently, Pistollato and co-workers⁶ have reviewed NPs targeting OC, describing the molecular mechanisms underlying their effects. These NPs, which include curcumin, epigallocatechin 3-gallate, resveratrol, sulforaphane and Withaferin A, are characterized by long-term safety and negligible and/or inexistent side effects, and have been proposed as possible adjuvants to traditional chemotherapy.

Withaferin A (WA), a natural steroidal lactone, is a promising drug candidate multi-targeting various cancer hallmarks.⁷ WA down-regulates the Notch, Akt and bcl-2 pathways and causes growth inhibition and apoptosis induction in ovarian carcinoma cell lines, CaOV3 and SKOV3.⁸ Studies conducted on various epithelial cancer cell lines (cisplatin-sensitive A2780, cisplatin-resistant variant A2780/CP70, and p53 mutant CaOV3) revealed a

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3 synergetic effect of WA in combination with doxorubicin⁹ and cisplatin¹⁰ on cell death
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5 through the generation of ROS-mediated autophagy, leading to DNA damage and induction
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7 of apoptosis. The authors suggested this synergetic therapy could minimize/eliminate the side
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9 effects and induction of drug resistance associated with high drug doses. In addition, WA in
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11 combination with liposomal preparation of doxorubicin targets aldehyde dehydrogenase I
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13 positive cancer stem cells in OC.¹¹ Moreover, WA alone and in combination with cisplatin
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15 targets putative cancer stem cells,^{12, 13} suggesting that this may present more efficacious
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17 therapy for OC. Despite the recognized potential of WA as an antitumor agent, its application
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19 in clinical oncology is non-existent.
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24 Furthermore, the medicinal applications of organosilicon molecules are particularly
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26 interesting because differences in their chemical properties can contribute to enhancing
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28 potency and improving pharmacological attributes.¹⁴ These differences offer the potential for
29
30 unique and/or specific interactions between an organosilicon molecule and a biological
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32 macromolecule. In fact, there are many examples demonstrating that the incorporation of silyl
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34 groups provides a general strategy to increase size and lipophilicity for drug design. Although
35
36 there are no marketed drugs containing silicon, some silicon compounds have entered human
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38 clinical trials.¹⁵ Though still a growing area, the incorporation of silicon in drug scaffolds
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40 may well offer great potential for enlarging the chemical space of medicinal chemistry.
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42 Moreover, computational methods have become a promising tool for identifying active lead
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44 compounds and are being used with the pipeline of drug discovery in most pharmaceutical
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46 companies.¹⁶ Thus, theoretical prediction of pharmacokinetic properties i.e. ADMET
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48 (absorption, distribution, metabolism, excretion, and toxicity) play a key role in drug
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50 discovery, since an unfavorable ADMET has been identified as the major cause of failure of
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52 candidate molecules in drug development.¹⁷
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3 Previously, we have reported that incorporation of silyl ether substituents in the WA-
4 framework enhance its cytotoxic effect on HeLa (carcinoma of the cervix), A-549 (lung
5 carcinoma), and MCF-7 (breast adenocarcinoma) human cancer cell lines, whereas a ketone
6 group at C-4 increases selectivity.^{18,19} Moreover, the induction of apoptosis by 27-*O*-(*tert*-
7 butyldimethylsilyl)-4-dehydroxy-4-oxo-withaferin A without necrosis under extreme
8 experimental conditions has drawn our attention to these organosilicon analogues.¹⁸
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12 Therefore, encouraged by the previous work on WA targets ovarian cancer cells, and the
13 expectation of bio-organosilicon in drug design, efforts to enlarge the chemical space of WA
14 to improve its clinical potential as anticancer agent are continuing. The current study reports
15 the design, synthesis and evaluation of a WA-silyl ether library with enhanced ovarian cancer
16 cell cytotoxicity compared to the lead compound and reference drug. Furthermore, two
17 analogues were investigated for their ability to induce apoptosis, confirming previous studies
18 on this type of scaffold.¹⁹ In addition, extensive structure–activity relationship (SAR) and *in*
19 *silico* ADME studies were employed to understand the pharmacokinetic properties of this
20 series of WA-analogues.
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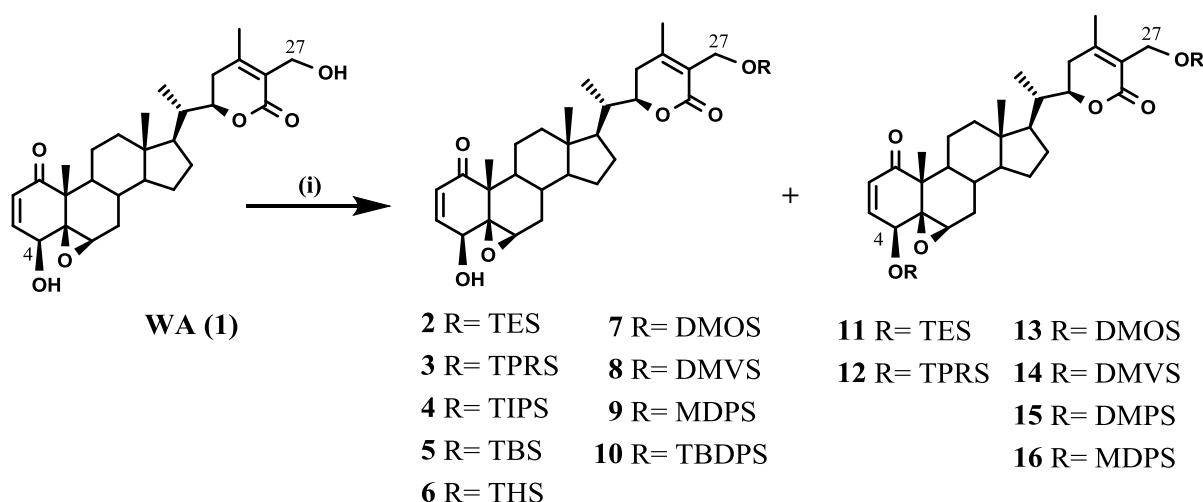
40 RESULTS AND DISCUSSION

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42 **1. Chemistry.** When silicon is incorporated into an organic compound, the chemical and
43 physical differences contributed by the silyl group can provide compounds with unique
44 properties that are relevant for medicinal chemistry.¹⁴
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49 Withaferin A is a C28 ergosterane-type steroid with a δ -lactone ring between C-22 and C-
50 26 in the side chain. To refine structural features and enhance the anticancer profile of WA, a
51 suitable starting material from *Withania aristata*,¹⁸ a library of WA-silyl ether analogues (**2**-
52 **22** and **24-34**) were designed and synthesized (Schemes 1-4 and detailed in the Supporting
53 Information S32-S46).
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The synthesis of this WA-library was carried out using silyl chloride with different electronic and steric properties, such as hydrophobicity, size and aromaticity. The first step in this task was to investigate the modification of the hydroxyl groups at C-4 and C-27 by converting them into silyl ethers. Thus, 27-silyl ether (**2-10**) and 4,27-disilyl ether (**11-16**) analogues were synthesized following the strategy outlined in Scheme 1.

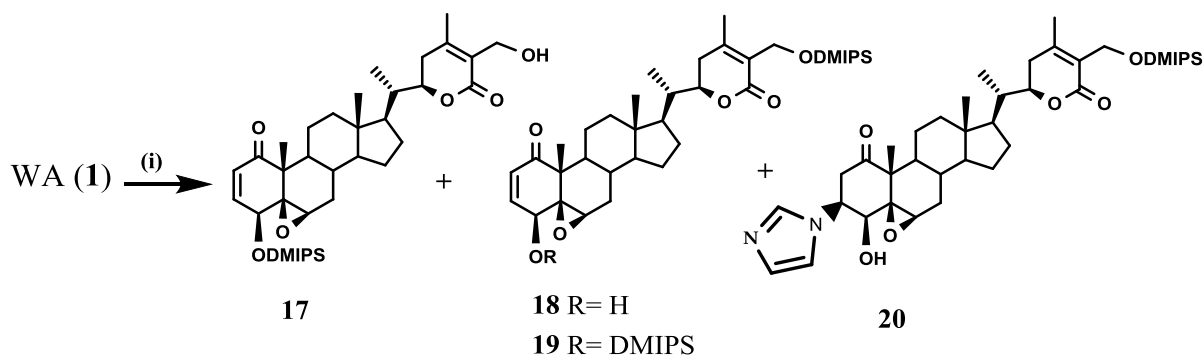
Scheme 1. Synthesis of Withaferin A-silyl Ether Analogues 2-16^a



M, methyl; E, ethyl; IP, *iso*-propyl; PR, propyl; B, butyl; TB, *tert*-butyl; H, hexyl; O, octyl; P, phenyl; V, vinyl
^aReagents and conditions: (i) R₁R₂R₃SiCl, DMAP, CH₂Cl₂, rt.

It is worth noting that treatment of WA (**1**) with DMIPSiCl afforded, in addition to the expected silyl ethers **18** and **19**, analogues **17** and **20**, which were formed by selective silylation of the secondary alcohol at C-4 and Michael addition of imidazole to the enone system in the WA-framework, respectively (Scheme 2).

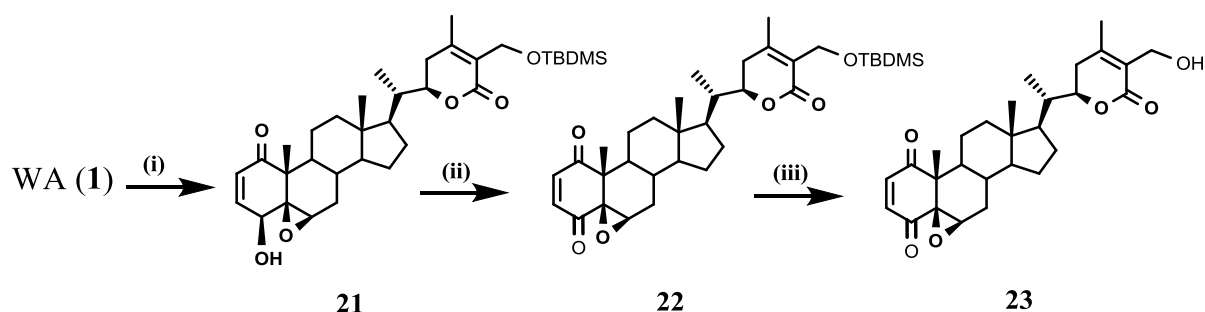
Scheme 2. Synthesis of Withaferin A-silyl Ether Analogues 17-20^a



^aReagents and conditions: DMIPSiCl, imidazole, DMAP, CH₂Cl₂, rt.

Previously reported structure-activity relationship (SAR) studies on withanolides,¹⁶ indicated that compounds bearing a ketone at C-4 have a selective pharmacological profile. Encouraged by these results, the synthesis of analogue **23** was carried out following the strategy outlined in Scheme 3. Firstly, selective protection of the primary alcohol in WA (**1**) with TBDMSiCl yielded the corresponding silyl ether derivative **21**, whose oxidation by treatment with Collins reagent afforded the ketone analogue **22**. Further cleavage of the protecting group in **22** with carboxylic acid resin led to the 4-dehydro-WA analogue **23**.

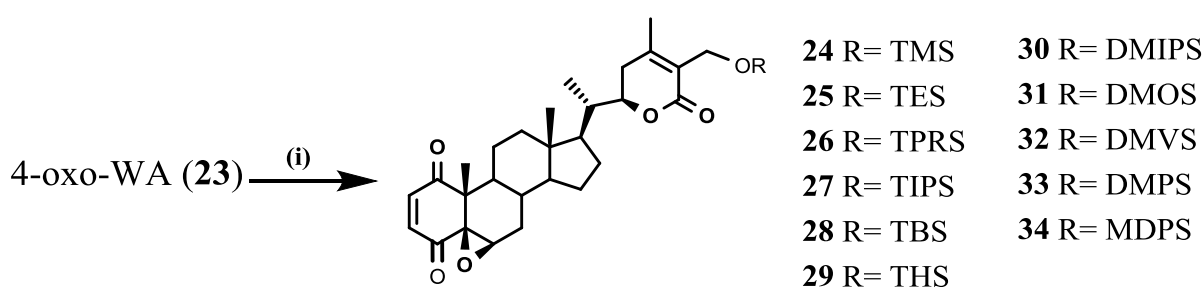
Scheme 3. Synthesis of Withaferin A-analogues 21-23^a



^aReagents and conditions: (i) TBDMSiCl, imidazol, DMAP, CH₂Cl₂, r.t.; (ii) CrO₃, py, CH₂Cl₂, rt, 15 min; (iii) Dowex (50WX8-200), acetone, rt, 24 h.

Derivatives **24-34** bearing both a 4-ketone and a 27-silyl ether group were prepared from compound **23** by silylation at C-27 with different silyl chloride reagents as shown in Scheme 4 (Supporting Information, S41-S46).

Scheme 4. Synthesis of 4-Oxo-withaferin A-silyl Ether Analogues 24-34^a



^aReagents and conditions: (i) R₁R₂R₃SiCl, imidazol, DMAP, CH₂Cl₂, rt.

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3 Among the former synthetic analogues, 30 out of 33 are reported for the first time. The
4 structures of the new compounds were elucidated by HRMS and NMR analysis (Supporting
5 information, S2-S31), whereas those of the previous reported analogues, compounds **21-23**
6 were elucidated by comparison of their spectral data with those reported in the literature.¹⁸
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15 **2. Biological Evaluation.**

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17 **Antiproliferative activity.** The *in vitro* antiproliferative activity of lead compound **1** and
18 derivative **23** (4-oxo-WA), and their silyl-analogues **2-22** and **24-34**, respectively, were
19 evaluated on two human epithelial ovarian tumor cell lines, cisplatin-sensitive (A2780) and
20 cisplatin-resistant (A2780/CP70) cells, and on the non-carcinoma cell line ARPE-19 (human
21 retinal pigment epithelial), the latter used to test for selectivity.
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28 Cytotoxic evaluation (Table 1, and Supporting Information, S47) against the cisplatin-
29 sensitive cell line revealed that the cytotoxicity of 11 analogues (compounds **3-5**, **7-9**, **13**, **15**,
30 **20**, **22**, and **23**) was higher than the widely known anticancer WA (**1**, IC₅₀ 32.7 nM) and the
31 reference drug (carboplatin, IC₅₀ 2.6 μM), exhibiting IC₅₀ values ranging from 7.3 to 32 nM.
32 Moreover, it is noteworthy that silyl ether analogues bearing a dimethyloctyl (**7** and **13**, IC₅₀
33 10 and 1.5 nM, respectively) or a dimethylphenyl (**15**, IC₅₀ 2.9 nM) moiety as well as
34 oxidation at C-4 (**23**, IC₅₀ 7.3 nM) are favorable trends for optimal cytotoxicity against
35 A2780 cells, improving activity by 3.2- to 21.8-fold compared to lead compound **1**.
36 Curiously, potency of the imidazole derivative **20** (IC₅₀ 20 nM) was slightly higher than the
37 lead compound **1**, which is opposite to previously reported SAR of withanolides in which the
38 enone system is essential for the anticancer activity.²⁰ On the other hand, functional group
39 interconversion of alcohols in WA by a trihexylsilyl (analogues **6** and **29**) or a tripropylsilyl
40 (derivative **12**) moiety was particularly detrimental furnishing completely inactive analogues.
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3 Drug resistance is a major obstacle for first line chemotherapy in ovarian cancer treatment.⁴
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5 Taking into consideration the promising results obtained for WA-silyl-analogues assayed on
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7 A2780 cisplatin-sensitive cell line, this series of compounds was tested for efficacy on a
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9 human ovarian carcinoma cisplatin-resistant (A2780/CP70) cell line. The results (Table 1)
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11 indicated that 15 analogues showed from similar (**7-10**, **14**, **15**, **18**, **27** and **30**) to slightly
12
13 improved (**5**, **13**, **17**, **20**, **25** and **26**) profile than the lead compound, WA (IC₅₀ 32 nM).
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15 Moreover, potencies of target compounds **2**, **3** and **23** on A2780/CP70 cells, exhibiting IC₅₀
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17 values ranging from 1 to 12.8 nM, were significantly improved from 2- to 30-fold compared
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19 to compound **1**. Regarding analogues with a drastic loss of activity, again analogues **6** and **29**
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21 and the disilyl ether derivatives **12** and **16** were from 794- to 364-fold less active than WA.
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26 After confirming that some of the newly-synthesized analogues showed potent activity
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28 against both human epithelial ovarian tumor cell lines, cytotoxicity against a non-carcinoma
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30 cell line (ARPE19, human retinal pigment epithelial cells) was evaluated for all derivatives to
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32 test for selectivity (Table 1). We assume that a selectivity index (SI) value higher than two
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34 indicates a good selectivity for inducing cytotoxicity in tumor cell lines as compared to those
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36 in non-cancerous cells, according to Suffness.²¹ Among the evaluated compounds, 25 of them
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38 showed selectivity to a some extent (SI > 2) in the non-carcinoma (ARPE19) cell line with
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40 respect to the A2780 cell line. Selectivity was observed for the most potent analogues, **7**, **8**,
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42 **13**, **15**, **20**, **22** and **23** (SI ranging from 2.6 to 212.0), but also for those compounds equipotent
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44 to WA, analogues **18**, **25**, and **34** (SI 191.2, 53.5 and 58.6, respectively). Similarly, SI was
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46 higher than two for 21 analogues regarding the A2780/CP70 cell line. The most active
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48 analogues on this cell line, compounds **2**, **3**, **17**, **20**, **23**, **25** and **26**, showed SI values from 2.3
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50 to 93.5. In addition, analogues **10**, **18** and **30** with a similar profile to WA were not cytotoxic
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52 on the non-carcinoma cell line (SI 79.1, 185.7 and 62.9, respectively).
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Table 1. Cytotoxic Activity (IC₅₀, nM)^a of WA-analogues^b on Human Ovarian Carcinoma Cell Lines,^c and on a non-Carcinoma Cancer Cells (ARPE19).

compd	A2780	A2780/CP70	ARPE19	SI ^d A2780	SI ^d A2780/CP70
1	32.7 ± 0.2	32.0 ± 2.0	37.0 ± 14.0	1.1	1.2
2	> 100	12.8 ± 2.0	30.0 ± 10.0	-	2.3
3	32.0 ± 2.0	3.6 ± 1.4	30.0 ± 0.6	0.9	8.3
4	30.0 ± 2.0	62.0 ± 20.0	62.0 ± 30.0	2.1	1.0
5	27.0 ± 4.0	22.0 ± 0.3	260.0 ± 210.0	9.6	11.8
7	10.0 ± 0.7	33.0 ± 10.0	32.0 ± 5.0	3.2	1.0
8	27.0 ± 10.0	31.0 ± 2.0	86.0 ± 16.0	3.2	2.8
9	22.0 ± 5.0	28.3 ± 0.3	70.0 ± 10.0	3.2	2.5
10	46.0 ± 9	29.0 ± 6.0	2295.0 ± 25.0	49.9	79.1
13	1.5 ± 0.5	24.9 ± 10.0	318.0 ± 61.0	212.0	12.8
14	33.0 ± 0.5	27.5 ± 5.0	92.0 ± 30.0	2.8	3.4
15	2.9 ± 1.0	29 ± 0.005	309.0 ± 40.0	106.6	10.7
17	34.0 ± 1.0	23.0 ± 9.0	310.0 ± 200.0	9.1	13.5
18	34.0 ± 0.6	35.0 ± 0.4	6500.0 ± 150.0	191.2	185.7
20	20.0 ± 7.0	19.0 ± 10.0	52.0 ± 10.0	2.6	2.7
21	47.5 ± 19.0	> 100	12.4 ± 1.0	0.3	0.1
22	17.0 ± 10.0	> 100	1660.0 ± 90.0	97.7	11.3
23	7.3 ± 6.0	< 1	32.0 ± 2.0	4.3	>32.0
24	41.0 ± 5.0	68.8 ± 8.0	820.0 ± 300.0	20.0	11.9
25	35.0 ± 10.0	20.0 ± 0.2	1870.0 ± 70.0	53.5	93.5
26	69.0 ± 20.0	21.0 ± 6.0	1740.0 ± 340.0	25.2	82.9
27	35.0 ± 6.0	35.0 ± 1.0	37.5 ± 4.0	1.1	1.1
28	53.0 ± 40.0	> 100	617.0 ± 500.0	11.6	-
30	43.0 ± 0.2	34.0 ± 0.2	2140.0 ± 100.0	49.8	62.9
33	59.0 ± 10.0	> 100	2305.0 ± 270.0	39.1	-
34	35.0 ± 10.0	> 100	2050.0 ± 350.0	58.6	-

^a IC₅₀ values (nM) of WA-silyl analogues were determined as described in the Biological Studies section. Carboplatin was used as a reference drug (IC₅₀ 2.6, 44.9 and 4.6 μM on A2780, A2780/CP70 and ARPE19 cell lines, respectively). Results are expressed as the mean ± standard deviation of three independent experiments performed in duplicate. ^b WA-analogues exhibiting IC₅₀ values ≤ 100 nM. ^c Human ovarian carcinoma cisplatin-sensible (A2780) and cisplatin-resistant (A2780/CP70) cell lines ^dSI, Selectivity Index.

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3 The overall results of the biological assays identified analogues **13**, **15**, **22** and **23** on the
4 cisplatin-sensitive cell line, and even more noteworthy, analogues **3**, **17**, **23**, **25** and **26** on the
5 cisplatin-resistant cell line as having significantly improved activity profiles compared to lead
6 compound **1**. These profiles were coupled with remarkable selectivity on the non-tumoral cell
7 line and, therefore, are suitable for further studies.
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17 **Cell Cycle Assay.** Our previous studies have indicated an apoptotic effect associated with
18 withanolide-type steroids, and reported the first examples of WA-silyl ether analogues, **21**
19 and **22**¹⁸ in HeLa cells. Moreover, WA has been reported to trigger the apoptotic cascade by
20 extrinsic or intrinsic pathways, e.g., in promyelocytic leukemia HL-6022 and U937 cells,²³
21 prostate cancer cells,²⁴ head and neck squamous carcinoma cells (HNSCC)²⁵ and Caki cells.²⁶
22 In the present study, the cell cycle event mediated by two potent analogues, **21** and **22**, was
23 investigated on human ovarian cancer cell line A2780. It should be noted that **22** showed a
24 very good selectivity on non-cancerous cells (SI 97.68) (Table 1, and Supporting
25 Information, S47). Cell cycle analysis was carried out using NucleoCounter® NC-3000™
26 system by rapid quantification of DNA content which was measured using fluorescent 4',6-
27 diamidino-2-phenylindole dihydrochloride (DAPI) stained cells. This assay will determine
28 cell sorting at different phases of cell cycle. Results showed that both analogues, **21** and **22**,
29 induce a dose-dependent increase in DNA fragmentation as evidenced by the increase in the
30 number of cells with low intensity DAPI signal in sub-G0/G1 as compared to control cells
31 (Figure 1, and Supporting Information, S49). This is suggestive of cell death by apoptosis
32 without indication of a cell cycle arrest in G0/G1 over 48 h contact time with the compounds.
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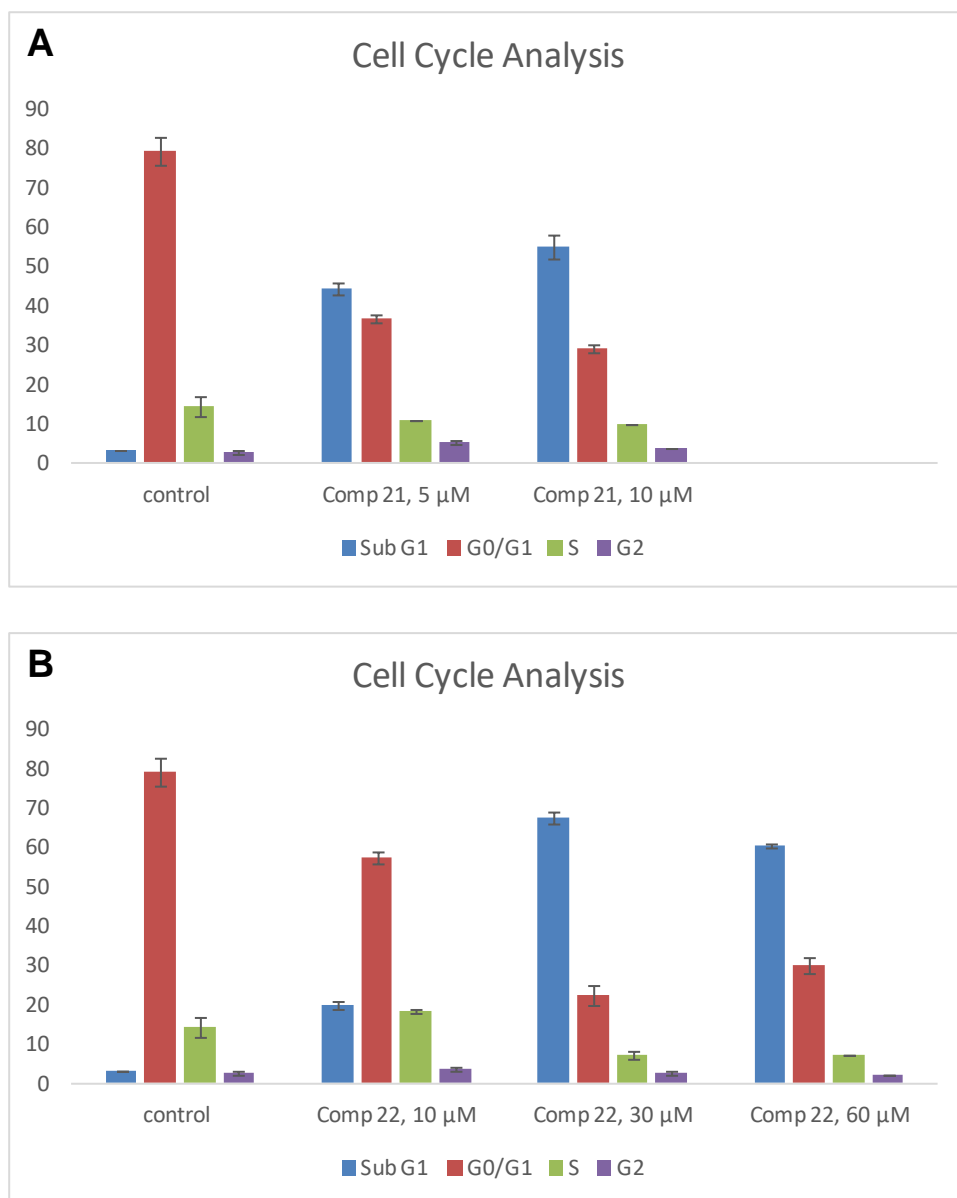


Figure 1. Percentage of cells in sub-G1, G0/G1, S and G2 phases after performing two-step cell cycle assay on human ovarian cancer cells (A2780 cell line) with compounds **21** (A) and **22** (B) at different concentrations. Data are based on 48 h exposure to compounds or vehicle control. Each column represents mean \pm s. e. mean of $n = 2$.

3. Structure-Activity Relationship Analysis. The previous reported SAR studies agree that an α,β -unsaturated ketone on ring A, a $5\beta,6\beta$ -epoxide in ring B and an α,β -unsaturated δ -lactone on side-chain in the WA-framework²⁰ are antitumor structural feature requirements, and more recently, acylation is reported to enhance cytotoxicity.¹⁹

In this work, chemical modulation by incorporation of silicon on the WA-framework was investigated. Thus, taking into consideration the IC_{50} values against both cancer cell lines

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3 assayed, the effect of silyl ether substituents was analyzed for each synthesized analogue (**2-**
4 **34**), according to the nature of the group attachment to silicon. The trends of the SAR study
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6 from this series of WA-silyl analogues on the A2789 cell line were as follows. Compounds
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8 carrying heterogeneous alkyl substituents on the silyl ether (**7-10**) displayed higher
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10 cytotoxicity than those with a homogeneous silyl ether (**2-6**). Indeed, replacement of
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12 dimethyloctyl or dimethylphenyl groups in the potent analogues **13** and **15** by a trihexyl,
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14 triisopropyl, or methyldiphenyl moiety led to the loss of activity (**6**, **12** and **29**). Previous
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16 work revealed that oxidation of the secondary alcohol at C-4 of the WA framework plays an
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18 important role in cytotoxicity.¹⁸ Surprisingly, the 4-oxo-WA derivative **23** (7.3 nM) showed a
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20 4.4-fold increase in activity as compared to **1** (32.7 nM). Furthermore, to explore the effect of
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22 replacement the primary hydroxyl group at C-27 by a silyl ether, derivatives **24-34** were
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24 prepared from **23** (Scheme 4). These analogues showed a broad profile of inhibitory activities
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26 with IC₅₀ values ranging from 35 nM to 25.17 μM, although all of them were significantly
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28 less potent than the congener **23** (IC₅₀ 7.3 nM) and lead compound **1** (IC₅₀ 32.7 nM). In
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30 general, silyl-analogues with a hydroxyl group at C-4 (**3-9** and **18**) are more potent than those
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32 with a ketone group (**26-29**, **31** and **32**) at this position.

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35 Regarding the cisplatin-resistant cell line (A2780/CP70), SAR studies of this series of
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37 analogues revealed that compounds carrying a heterogeneous alkyl substituent on the silyl
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39 ether (**7-10** and **13-15**) displayed similar profiles (IC₅₀ 24.9-33.0 nM) to **1**, indicating that
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41 activity was not greatly influenced by their corresponding silyl ether moiety on the
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43 withanolide skeleton. Analogues with a homogeneous alkyl substituent on the silyl ether (**2-6**,
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45 **11** and **12**) showed a great range of cytotoxic activity. In fact, replacement of the triethyl or
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47 tripropyl substituent in potent compounds **2** and **3** by a trihexyl, di-tripropyl, or di-
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49 methyldiphenyl moiety (compounds **6**, **12**, and **16**) led to significant loss of activity.
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51 Moreover, analogue **23** was 30-fold more potent than the parent against the cisplatin-resistant
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3 cell line, whereas silyl-analogues **24-34** (Scheme 4) were significantly less potent than their
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5 congener **23**, as occurred on the A2780 cell line. Therefore, a heterogeneous alkyl substituent
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7 on the silyl ether is favorable *versus* a homogenous one for the A2789 cell line, with the
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9 DMOS and DMPS the best functional groups. On the other hand, the homogenous alkyl
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11 substituents, TES and TPRS are the best functional groups on the A2789/CP70 cell line.
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13 Compounds with a hydroxyl group at C-4 are favorites *versus* those oxidized at C-4 on both
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15 cell lines, except for compound **23**.
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19 These results reveal that silylation of the WA-framework leads to a significant range of
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21 cytotoxic activity. Moreover, minor modifications on the silyl ether substituent, affecting
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23 spatial occupation at this site, had noteworthy repercussions on compound activity.
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25 Furthermore, silylation of the WA-framework leads to selectivity on both tumor cell lines.
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27 Moreover, taking into consideration that compounds with a silyl ether group increases
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29 hydrophobicity,²⁷ WA-silyl analogues could present advantages as drug candidates.
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36 **4. *In silico* ADME Predictions.** Understanding pharmacokinetic properties (ADMET,
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38 absorption, distribution, metabolism, and excretion molecular properties) is an important step
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40 in drug discovery to select new lead/drug candidates, since potent *in vitro* activity along with
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42 enhanced ADMET profiles increase the probability of clinical success.²⁸ Moreover, despite a
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44 great deal of research conducted on the potential anticancer properties of WA-related
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46 withanolides, there are only two reports of ADMET studies.^{29,30}
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50 The QikProp module of Schrödinger software³¹ was used for analyzing physicochemical
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52 and pharmacokinetic descriptors (ADME properties) of selected compounds (IC₅₀ values ≤ 1
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54 μM on A2780 and A2780/CP70 cell lines) with the aim of increasing the success rate of
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56 compounds reaching further stages of development. A detailed account of these parameters is
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58 given in Table S50 of the Supporting Information. These parameters provide insights into key
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3 aspects such as drug likeness, solubility, permeability, bioavailability predictions, oral
4 absorption, metabolism, etc. One of the primary descriptors that were taken into account was
5 #stars. The #stars descriptor informs about the number of properties of each compound that
6 fail to remain within the recommended ranges, therefore, a lower number of #stars denotes a
7 better drug-like molecule.³¹
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12 Thus, taking into consideration the IC_{50} values of the assayed series of analogues with
13 higher cytotoxic effect than WA ($IC_{50} < 34.7$ nM on A2780 cell line), and #stars values (0 or
14 1), compounds **2**, **8**, **17**, **20**, **22**, **23** and **25** were selected to analyze their predicted
15 pharmacokinetic properties (Table 2). Analogue **23** and the silyl ether analogue **25** as well as
16 WA lie within the recommended range of known drugs for all the analyzed parameters (#
17 stars = 0), whereas silyl ether analogue **2**, **8**, **17**, **20** and **22** fail in the QPlogS displaying low
18 aqueous solubility, although values for these analogues were near the upper limit of the
19 recommended range. Nevertheless, these silyl analogues showed predicted intestinal
20 absorption rate (QPPCaco) and apparent cell permeability (QPPMDK) greater than the lead
21 compound WA, thus predicting good oral bioavailability. The lipophilicity is also an
22 important physicochemical property requirement for a potential drug. It is expressed as
23 QPlogPo/w and play a crucial role in absorption, bioavailability, hydrophobic drug-receptor
24 interactions, metabolism, and toxicity.³² All the selected compounds log Po/w values lay
25 within the permissible range. The predicted values for properties such as octanol/water
26 partition (QPlogPo/w), gut-blood barrier permeability (QPPCaco, QPlogBB and
27 QPPMDCK), human serum albumin binding (QPlogKhsa) and percent of human oral
28 absorption (> 83%) were within ideal ranges. Therefore, compounds under study were
29 predicted to have good drug-likeness, since they have mostly favorable pharmacokinetic
30 properties, especially regarding membrane permeability and oral absorptivity.
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Table 2. *In silico* ADME profile predictions of selected WA-analogues^a and their range/recommended values.^b

Property/ Descriptor	1	2	8	17	20	22	23	25	Range/ recommended values
#stars	0	1	1	1	1	1	0	0	0-5
QPlogBB	-1.311	-1.072	-1.113	-1.253	-1.258	-087	-1.267	-1.156	-3.0 to 1.2
QPPCaco	258.201	1060.23	759.53	550.181	600.427	996.046	243.685	711.663	<25 poor, >500 great
QPPMDCK	114.484	526.993	367.48	259.339	285.031	492.595	107.544	342.512	<25 poor, >500 great
QPlogKhsa	0.329	1.189	1.017	1.136	1.1	0.892	-0.13	0.827	-1.5 to 1.5
QPlogPo/w	3.046	5.959	5.291	5.752	5.573	5.464	2.519	5.391	-2.0 to 6.5
QPlogKp	-3.842	-2.271	-2.635	-3.075	-2.816	-2.615	-3.977	-2.74	-8.0 to -1.0
QPlogS	-4.999	-7.586	-7.336	-7.379	-7.844	-6.713	-3.974	-6.489	-6.5 to 0.5
#metab	4	4	4	4	5	3	3	3	1 to 8
%HOA	87.948	90.068	83.565	83.76	83.39	86.682	84.413	83.643	>80% high <25% poor
PSA	96.36	85.36	85.36	85.36	103.18	82.2	93.2	82.2	7.0 to 200.0
SASA	718.234	906.765	874.31	88.418	949.884	877.558	711.717	882.911	300.0 to 1,000.0
Mol MW	470.605	584.867	554.8	570.84	638.918	582.851	468.589	582.851	130.0 to 725.0
#rotor	5	9	7	7	7	6	4	8	0 to 15
donorHB	1	1	1	0	1	0	0	0	0.0 to 6.0
acceptHB	9.4	9.55	9.55	8.55	11.55	9.85	9.7	9.85	2.0 to 20.0
volume	1396.689	1803.47	1709.1	1757.66	1903.72	1778.773	1383.64	1785.65	500.0 to 2,000.0

^a WA-analogues exhibiting IC₅₀ values lower than those for WA on A2780 and/or A2780 cell lines (IC₅₀ < 32.7 nM), and #stars values between 0-1. ^b #star (number of property values that fall outside the 95% range of similar values for known drugs), QPlogBB (predicted brain/blood partition coefficient), QPPCaco2 (predicted human epithelial colorectal adenocarcinoma cell line permeability in nm/s), QPPMDCK (predicted Madin-Darby canine kidney permeability in nm/s), QPlogKhsa (predicted binding to human serum albumin), QPlogPo/w (predicted octanol/water partition coefficient), QPlogKp (skin permeability), QPlogS (predicted aqueous solubility), #metab (number of likely metabolic reactions), % HOA (predicted human oral absorption on 0 to 100%), PSA (Van der Waals surface area of polar nitrogen and oxygen atoms and carbonyl atoms), SASA (total solvent accessible surface area), MW (molecular weight), #rotor (number of non-trivial, non-hindered rotatable bonds).

CONCLUSIONS

The current study reports on our efforts to find new drug candidates for OC resistant to current treatments. Therefore, the synthesis and evaluation of a WA-silyl ether library, together with structure-activity relationship and *in silico* ADME studies, were employed to find drug candidates for the treatment of ovarian cancer. We have successfully identified a new generation of potent and selective WA-analogues with a significantly improved cytotoxic profile. In fact, ten WA-analogues exhibited higher potency than the lead compound and reference drug on the cisplatin-sensitive cell line, and more notable, fifteen analogues enhanced cytotoxic profile on the cisplatin-resistant cell line. Cell cycle analysis of two potent analogues revealed cell death by apoptosis without cell cycle arrest in G0/G1. Furthermore, the predicted pharmacokinetic properties highlight three analogues with great potential to become drug candidates: the 4-oxo-WA (**23**) exhibiting single-digit nanomolar potency on both cancer cell lines, and two potent silyl-ether analogues, **22** and **25**, on A2780 and A2780/CP70 cells, respectively. This potency is accompanied by an excellent selectivity index and favorable drug-likeness. Thus, analogues reported herein are promising candidates in anticancer drug development for OC that deserve further investigation.

In summary, the current study provides an insight into the anticancer potential of WA-analogues on OC, whether alone or in combination with clinical drugs, and support the increasingly important role that silicon will play in drug design.

EXPERIMENTAL SECTION

General Methods for Chemistry. Optical rotations were measured on a Perkin Elmer 241 automatic polarimeter, in CHCl₃ at 25 °C, the $[\alpha]_D$ values are given in units of 10⁻¹ deg cm² g⁻¹. ¹H (500 MHz) and ¹³C (125 MHz) NMR spectra were recorded on a Bruker Avance 400 spectrometer; chemical shifts are given in ppm and coupling constants in Hz. Solutions were

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3 typically prepared in CDCl_3 with chemical shifts referenced to deuterated solvent as an
4 internal standard. EIMS and HREIMS were measured on a Micromass Autospec
5 spectrometer, and ESIMS and HRESIMS (positive mode) were measured on a LCT Premier
6 XE Micromass Autospec spectrometer. Silica gel 60 for column chromatography (particle
7 size 15-40 and 63-200 μm), Polygram Sil G/UV₂₅₄ used for analytical and preparative TLC,
8 and HPTLC-Platten Nano-Sil 20 UV₂₅₄ were purchased from Macherey-Nagel. Reactions
9 were monitored by TLC, the spots were visualized by UV light and heating silica gel plates
10 sprayed with $\text{H}_2\text{O}-\text{H}_2\text{SO}_4-\text{AcH}$ (1:4:20). Varian high-performance liquid chromatography
11 (HPLC) equipment consisted of a ProStar 210 solvent delivery module, ProStar 335
12 photodiode array detector, using an analytical Pursuit C18 column (2.0 x 100 mm, 3 μm)
13 with a flow rate of 0.3 mL/min, and mixtures of acetonitrile- H_2O as eluent. The degree of
14 purity of the compounds was over 95% as indicated by the appearance of a single peak using
15 HPLC. Unless otherwise noted, solvents and reagents were obtained from commercial
16 suppliers and used without further purification. Anhydrous THF and Cl_2CH_2 were distilled
17 from sodium/ benzophenone and calcium hydride ketyl under nitrogen, respectively. All
18 solvents used were analytical grade from Panreac, and the reagents were purchased from
19 Sigma Aldrich. Withaferin A (WA, **1**), used as starting material, was isolated from the leaves
20 of *W. aristata* as previously described.¹⁶
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47 **Materials for Biological Studies.** MTT and media for growing cell lines and all
48 supplements were purchased from Sigma Aldrich, UK, and cell lines were purchased from
49 ATCC. Non-cancerous cells were kindly donated by Prof Roger Phillips and Dr Simon
50 Allison at the University of Huddersfield. Phosphate buffered saline (PBS), 50 $\mu\text{g}/\text{mL}$
51 Annexin V-CF488A conjugate, Annexin V binding buffer (10 x concentrate), Solution 15
52 (500 $\mu\text{g}/\text{mL}$ Hoechst 33342), Solution 16 (500 $\mu\text{g}/\text{mL}$ Propidium Iodide), Solution 10 (Lysis
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3 buffer), Solution 11 (stabilization buffer), Solution 12 (500 $\mu\text{g}/\text{mL}$ DAPI), NC-Slide A8™,
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5 NC-Slide A2™ glass slides and via-1 cassettes were bought from ChemoMetec, Denmark.
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7 NC-3000™ image cytometer was used to perform the assays.
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12 **Cell Culture and Viability Assay.** Cells were grown and maintained in DMEM or RPMI
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14 1640 medium supplemented with 10% fetal bovine serum, 5% penicillin streptomycin at 37
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16 °C, 5% CO₂/95% air as instructed by the suppliers. The cells were plated in 96-well culture
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18 plates at a density of 1×10^4 cells mL⁻¹ and allowed to adhere at 37 °C for 24 h. The
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20 following day, different doses of S186 or vehicle were added to the cells in varying
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22 concentrations of compounds and were further incubated for 96 hours. Following the
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24 aforementioned incubation time the supernatant was removed and MTT (3-(4,5-
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26 dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) was added for 4 h. The ability of
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28 cells to form formazan crystals by active mitochondrial respiration was determined using a
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30 Microplate reader after dissolving the crystals in DMSO. Cytotoxicity was expressed as a
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32 relative percentage of the absorbance measured at 540 nm in the control and extract-treated
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34 cells. Data were presented as the mean \pm s.e. mean.
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43 **Two-step Cell Cycle Assay.** Cells were seeded into T25 flasks containing 5 mL of
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45 complete media and were incubated for 24 h at 37 °C. After 24 h elapsed cells were treated
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47 with vehicle control or compounds at different concentrations and left in the incubator for a
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49 further 48 h. Cells were then subjected to two-step cell cycle assay according to the
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51 manufacture's instructions. Briefly, 1 mL of cells (1×10^6 cells/mL) was transferred to
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53 eppendorf tubes. In a separate eppendorf a mixture of 1960 μL of Lysis buffer (Solution 10)
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55 plus 40 μL of 500 $\mu\text{g}/\text{mL}$ DAPI (Solution 12) was prepared. Eppendorfs containing cells
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57 were centrifuged at 400 g for 5 minutes, supernatant was removed and cells were re-
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3 suspended in 250 μL of the above mixture, mixed well and incubated at 37 $^{\circ}\text{C}$ for 5 minutes.
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5 250 μL of stabilization buffer (Solution 11) was then added to the cells and mixed well. 10
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7 μL of each sample was then loaded on A8 slide and subjected to the two-step cell cycle assay
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9 using NC-3000TM.
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14 **Statistical Analysis.** All results were expressed as means \pm s.e. mean. Significant
15 differences between groups were determined using unpaired Student's *t*-test. Significance
16 was set at $p < 0.05$.
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23 **ADME Properties Predictions of WA-analogues.** Prediction of descriptors related to
24 adsorption, distribution, metabolism and excretion (ADME) properties of the compounds
25 were predicted using the QikProp program (QikProp, version 5.3)³¹ in Fast mode and based
26 on the method of Jorgensen^{33,34}. Preparation of compounds and the 2D-to-3D conversion was
27 performed using LigPrep tool, a module of the Small-Molecule Drug Discovery Suite in
28 Schrödinger software package, followed by a MacroModel 12.01 Monte Carlo
29 conformational search to locate the lowest energy conformation of each ligand. The program
30 computes pharmacokinetic relevant properties such as octanol/water partitioning coefficient,
31 aqueous solubility, brain/blood partition coefficient, Caco-2 cell permeability, serum protein
32 binding, number of likely metabolic reactions, and others. Drug likeness (#stars), number of
33 property descriptors from the full list of descriptors computed by the QikProp that fall outside
34 the range of values determined for 95% of known drugs, was used as additional compound
35 selection filter.
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3 ASSOCIATED CONTENT
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5 **Supporting Information.** NMR spectra, experimental data and procedures for preparation of
6 analogues **2-34**, cytotoxicity data, and cell cycle scatter plots are available free of charge via
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8 Internet at <http://pubs.acs.org>.
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15 AUTHOR INFORMATION

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22 **Notes**

23
24 The authors declare no competing financial interest.
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47 ABBREVIATIONS USED

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49 SAR, structure-activity relationship; NPs, natural products; WA, withaferin A; Et₃N,
50 triethylamina; DMAP, 4-(*N,N*-dimethylamino)pyridine; rt, room temperature; DMIPSiCl,
51 dimethylisopropylsilyl chloride; py, pyridine; TBDMSCl, *tert*-butyldimethylsilyl chloride;
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54 MTT, 3-(4,5-dimethyl thiazol-2-yl)-2,5-diphenyltetrazolium bromide; DMSO, dimethyl
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56 sulfoxide; PBS, phosphate buffered saline.
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