



Induction of autophagy and activation of SIRT-1 deacetylation mechanisms mediate neuroprotection by the pomegranate metabolite urolithin A in BV2 microglia and differentiated 3D human neural progenitor cells

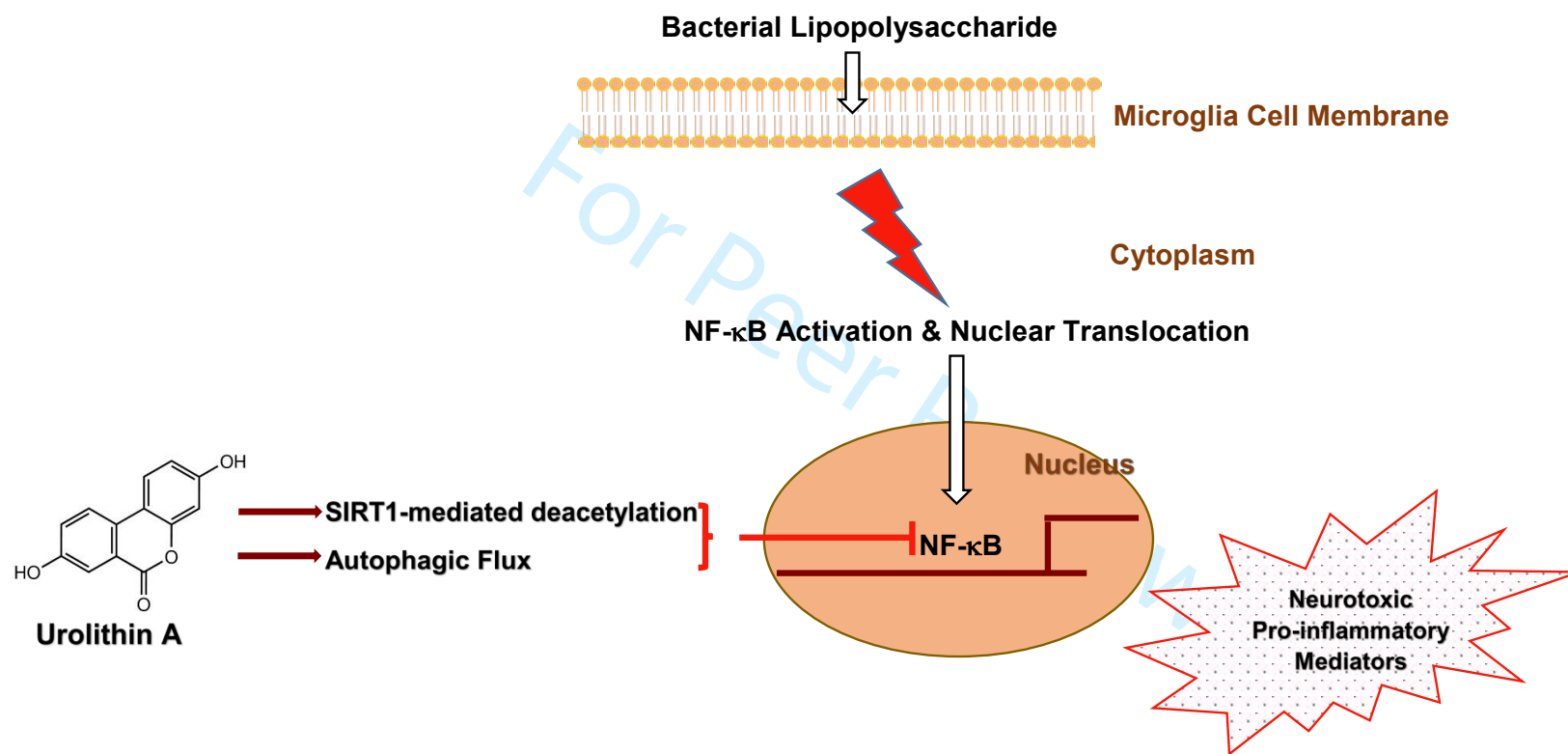
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Graphical Abstract

Urolithin A is a gut-derived metabolite of fruits such as pomegranate. In this study we investigated molecular mechanisms involved in the neuroprotective actions of this compound. The outcome of the experiments showed that urolithin A produced inhibition of neuroinflammation and amyloid beta neurotoxicity by activating DNA deacetylation mechanisms and induction of autophagy in BV2 microglia and human neural progenitor cells.

For Peer Review



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3 **Induction of autophagy and activation of SIRT-1 deacetylation mechanisms**
4 **mediate neuroprotection by the pomegranate metabolite urolithin A in BV2**
5 **microglia and differentiated 3D human neural progenitor cells**
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Abstract

1. Scope

Urolithin A is an anti-inflammatory and neuroprotective gut-derived metabolite from ellagitannins and ellagic acid in pomegranate, berries and nuts. We investigated the roles of SIRT-1 and autophagy in the neuroprotective activity of urolithin A.

2. Methods and results

Analyses of culture supernatants from lipopolysaccharide-stimulated BV2 microglia showed that urolithin A (2.5-10 μ M) produced significant reduction in the production of nitrite, tumour necrosis factor (TNF) α and interleukin (IL)-6. Anti-inflammatory effect of the compound was reversed in the presence of sirtuin (SIRT)-1 and the autophagy inhibitors EX527 and chloroquine, respectively. Protein analyses revealed reduction in p65 and acetyl-p65 protein. Treatment of BV2 microglia with urolithin A resulted in increased SIRT-1 activity and nuclear protein, while induction of autophagy by the compound was demonstrated using autophagy fluorescent and autophagy LC3 HiBiT reporter assays. Viability assays revealed that urolithin A produced neuroprotective effect in APPSwe-transfected ReNcell VM human neural cells, which was reversed in the presence of EX527 and chloroquine. Increase in both SIRT-1 and autophagic activities were also detected in these cells following treatment with urolithin A.

3. Conclusions

We propose that SIRT-1 activation and induction of autophagy are involved in the neuroprotective activity of urolithin A in brain cells.

1. Introduction

Neuroinflammation is progressively becoming a critical process in many neurodegenerative diseases including Alzheimer's (AD) and Parkinson's diseases (PD). Neuroinflammation is activated by mechanisms such as oxidative stress and activation of brain resident macrophages known as microglia^[1]. Furthermore, mediators, such as pro-inflammatory cytokines, released during neuroinflammation are critical in neurodegeneration^[1].

Sirtuin-1 (SIRT-1), an NAD⁺-dependent class III histone deacetylase, is known to play a significant role in suppressing the release of pro-inflammatory cytokines^[2,3]. In microglia, over-expression of SIRT-1 has been demonstrated to inhibit microglia activation^[4]. Furthermore, the SIRT-1 activator resveratrol has been reported to inhibit neuroinflammation in LPS-activated rat primary microglia^[5] and in cultured astrocytes^[6]. These observations were further confirmed by recent findings which have shown that SIRT-1 activation is neuroprotective against neuronal apoptosis, neuroinflammation and oxidative stress^[7]. Autophagy is a catabolic process that is critical in the degradation of cytoplasmic proteins in multicellular organisms. Several lines of evidence have linked autophagy to neuroinflammation^[8] as well as to neurodegeneration^[9]. Consequently, pharmacological modulation of brain SIRT-1 and autophagy with nutritional factors is a promising strategy in neurodegenerative diseases.

Urolithins are bioavailable metabolites that are produced by the human gut microbiota from ellagitannins and ellagic acid found in food products such as pomegranate, berries, and nuts. Several biological activities have been reported for the urolithins; they were reported to inhibit inflammation *in vivo* in rat model of inflammatory bowel disease^[10], as well as *in vitro* in human colonic fibroblasts^[11], RAW 264.7 murine macrophages^[12, 13] and BV2 microglia^[14]. Studies by DaSilva et al. have also shown that urolithins inhibited neuroinflammation in LPS-stimulated BV2 microglia and protected SH-SY5Y cells from oxidative stress-induced toxicity^[15]. Furthermore, urolithin A was shown to extend the lifespan of *C. elegans* and improves muscle function in mice through upregulation of mitophagy^[16]. In this study, we aimed to establish the roles of SIRT-1-mediated deacetylation and autophagic flux in the neuroprotective activity of urolithin A, the most active urolithin.

2. Materials and methods

2.1 Cell culture and treatments

The BV2 mouse microglia cell line (ICLC ATL03001; Interlab Cell Line Collection, Banca Biologica e Cell Factory, Italy) was cultured in RPMI1640 medium with 10% fetal bovine serum, 2 mM L-glutamine, 1 mM sodium pyruvate, 100 U/ml penicillin and 100 mg/ml streptomycin in a 5% CO₂ incubator.

HEK293 (ECACC 85120602) cells were obtained from the European Collection of Cell Cultures (ECCAC) (Salisbury, UK) and were cultured in MEM-eagle's medium (Thermo Scientific). MEM medium was supplemented with 10% FBS, 2 mM L-glutamine, 1 mM sodium pyruvate, 2 mM L-glutamine, 100 U/ml penicillin, and 100 mg/ml streptomycin in a 5% CO₂ incubator.

Immortalised neural progenitor cells (ReNcell VM cells) were acquired commercially from Millipore (Hertfordshire, UK) and maintained as earlier described^[17]. Cells were differentiated by replacing the culture medium with ReNcell NSC maintenance medium without the growth factors EGF and FGF-2 for 14 days. Differentiation into neurons was confirmed by immunocytochemical staining for β III-tubulin and MAP2^[17] (Supplementary Figure S1-S3).

Urolithin A (Santa Cruz), EX527 (Sigma) and chloroquine (Arcos Organics) were prepared in dimethylsulfoxide (DMSO) for pharmacological experiments. The percentage of DMSO in cell culture was 0.2%. For all experiments, 5 x 10⁴ cells/ml were seeded out in culture plates.

2.2 Dosage information

Treatment of these cells with urolithin A (2.5-10 μ M) with LPS stimulation for 24 h did not affect cell viability (Supplementary Figure S4).

2.3 Determination of pro-inflammatory mediator release

BV2 microglia were pre-treated with urolithin A (2.5-10 μ M) or vehicle (0.2% DMSO). After 30 min, cells were stimulated with LPS (100 ng/ml) for 24 h. Levels of nitrite in culture medium were determined using the Griess reagent (Promega), according to the manufacturer's instructions. Production of TNF α and IL-6 was determined using mouse ELISA kits (Biolegend), according to the manufacturer's instructions.

Detection limits were 1.5 pg/ml and 2 pg/ml for TNF α and IL-6, respectively. These

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3 experiments were also carried out in cells treated with SIRT-1 inhibitor
4 (EX527; 1 μ M) or autophagy inhibitor (chloroquine; 40 μ M) 30 min before addition of
5 urolithin A (10 μ M) and stimulation with LPS (100 ng/ml).
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8 9 **2.4 Transfection of ReNcell VM human neural stem cells with APPSwe** 10 **plasmids**

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12 Differentiated ReNcell VM cells were transfected with APPSwe plasmids as
13 described and reported earlier^[17]. Transfected cells were treated with urolithin A
14 (2.5-10 μ M) for 48 h. Cell viability was evaluated using MTT and LDH viability
15 assays. Levels of A β ₄₂ in culture medium of cells treated with urolithin A for 14 days
16 were determined by using a human A β ₄₂ ELISA kit (Life Technologies). Cell viability
17 experiments were also carried out in cells treated with SIRT-1 inhibitor
18 (EX527; 1 μ M) or autophagy inhibitor (chloroquine; 40 μ M) 30 min prior to treatment
19 with urolithin A (10 μ M).
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27 **2.5 Immunofluorescence microscopy**

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29 Immunofluorescence detection for NF- κ Bp65, acetyl-p65, SIRT-1, nestin, β III-tubulin,
30 and MAP2 proteins was carried with Alexa Fluor 488-conjugated donkey anti rabbit
31 IgG secondary antibody (Life Technologies; 1:500) and images were obtained using
32 EVOS® FLoid® cell imaging station as described elsewhere^[15, 16].
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37 **2.6 In-cell western for quantitative immunocytochemistry measurement of** 38 **protein levels**

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40 This is a rapid method for quantification of proteins in cells. The method is similar to
41 immunoblotting and immunocytochemistry and combines the specificity achieved
42 with immunoblotting with the quantification capability of ELISA. BV2 microglia were
43 seeded into a 96-well plate at a density of 5 x 10⁴ cells/ml. Thereafter, cells were
44 treated with urolithin A (2.5-10 μ M), and then stimulated with LPS (100 ng/ml) for
45 60 min. Cell fixation was achieved with 8% paraformaldehyde solution, followed by
46 overnight incubation with primary antibodies. The following antibodies were used:
47 phospho-p65 NF- κ B (1:200; Cell Signalling), acetyl-p65 NF- κ B (1:200; Cell
48 Signalling). This was followed by incubation with anti-rabbit HRP secondary antibody
49 for 2 h at room temperature. Then, 100 μ L HRP substrate was added to the plate
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3 and signal measured at 450nm with a microplate reader. Readings were normalised
4 with Janus Green stain (Abcam).
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6 7 **2.7 SIRT-1 ELISA and activity assay** 8

9 Cultured BV2 microglia or differentiated ReNcell VM human neural cells were treated
10 with urolithin A (2.5-10 μ M) for 24 h. Thereafter, nuclear extracts were collected for
11 analysis of SIRT-1 using either mouse or human SIRT-1 ELISA kit (Abcam),
12 according to the manufacturer's instructions. SIRT-1 activity in cultured cells was
13 also evaluated using the luminescent SIRT-Glo™ Assay (Promega), according to the
14 manufacturer's instructions and described earlier^[18, 19].
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20 21 **2.8 Autophagy fluorescent assay** 22

23 This experiment was carried out using an autophagy assay kit (Sigma), which
24 provides a simple and direct approach for detecting autophagy in cells by employing
25 a fluorescent autophagosome marker. BV2 or differentiated ReNcell VM cells were
26 treated with urolithin A (2.5-10 μ M) for 24 h. Thereafter, medium was removed and
27 the cells were incubated with autophagosome detection reagent at 37°C with 5%
28 CO₂ for 1 h. Fluorescence intensity ($\lambda_{\text{ex}} = 360/\lambda_{\text{em}} = 520$ nm) was measured using a
29 Polarstar Optima Plate microplate reader.
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35 36 **2.9 Autophagy LC3 HiBiT reporter assay** 37

38 The autophagy LC3 HiBiT reporter vector (Promega) has a HiBiT tag and contains a
39 sequence encoding the MAP1LC3B gene. On induction of autophagy, LC3 reporter
40 proteins are trapped in autophagosomes and degraded. HEK293, BV2 microglia, or
41 differentiated ReNcell VM cells were transfected with the autophagy LC3 HiBiT
42 reporter vector using Fugene 6 transfection reagent (Promega) and incubated for
43 16 h at 37°C in 5% CO₂. Thereafter, transfected cells were plated in white 96-well
44 plates and treated with urolithin A (2.5-10 μ M) for 6 h. Then, 100 μ l of Nano-Glo®
45 HiBiT lytic reagent (Promega) was added to each well and the samples were mixed
46 using an orbital shaker for 2 min. Luminescence was measured on a Polarstar
47 Optima Plate microplate reader.
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2.10 Statistical analysis

Values of all experiments were represented as a mean \pm SEM of at least 3 experiments. Values were compared using one-way ANOVA followed by a post hoc Tukey test.

3. Results and discussion

3.1 Urolithin A inhibits neuroinflammation and acetylation of NF- κ B in LPS-activated BV2 microglia

Firstly, we were interested in establishing an anti-inflammatory activity for urolithin A in LPS-activated BV2 microglia. Our results showed that treatment with LPS (100 ng/ml) resulted in a significant ($p < 0.001$) elevation in the levels of TNF α , IL-6, and nitrite in cell culture supernatants (Figure 3A). However, pre-treatment with urolithin A (2.5-10 μ M) resulted in a significant ($p < 0.01$) and concentration-dependent reduction in the release of TNF α and nitrite. Levels of IL-6 was also decreased significantly and but not in a concentration-dependent manner. Inhibition of neuroinflammation by the compound was further confirmed using quantitative in cell ELISA and immunofluorescence experiments showing inhibition of LPS-induced phosphorylation (Figure 2B) nuclear accumulation (Supplementary Figure S5A) of NF- κ B in BV2 cells. Similar anti-inflammatory effects have been reported for urolithin A and other urolithins in BV2 microglia^[14, 15].

It has been suggested that phase II glucuronide and sulfate conjugate metabolites of urolithins may be more physiologically relevant than the urolithins in order to produce pharmacological actions *in vivo* ^[20]. However, in addition to demonstrating *in vivo* neuroprotective activity of urolithins, Yuan et al used *in silico* computational studies to predict blood-brain barrier permeability of the compounds ^[21]. The relative *in vivo* neuroprotective activities of urolithins and their phase II metabolites therefore warrant further investigation.

In neuroinflammation, activation of NF- κ B and subsequent transcriptional regulation of pro-inflammatory genes involves post-translational modifications such as reversible acetylation of the NF- κ B-p65 subunit^[22]. Our investigations demonstrated that treatment with urolithin A reduced protein levels of acetyl-p65 in BV2 microglia stimulated with LPS (Figure 2C and Supplementary Figure S5B). We report here for

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3 the first time that inhibition of LPS-induced acetylation of p65 by urolithin A is a
4 critical mechanism in the anti-inflammatory activity of the compound, thus suggesting
5 a role for nuclear histone deacetylation mechanisms.
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8 9 **3.2 The role of SIRT-1 activation in the inhibition of neuroinflammation by** 10 **urolithin A in BV2 microglia**

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12 Sirtuins (SIRT) are class III histone deacetylases (HDACs), and have been strongly
13 linked to regulation of neuroinflammation and neurodegeneration^[23, 24]. Specifically,
14 SIRT-1 activation prevents NF- κ B activation through mechanisms involving
15 deacetylation of the p65 subunit^[25]. Furthermore, inhibition of SIRT-1 in
16 LPS-activated microglia has been observed to correlate with increased production of
17 inflammatory cytokines^[26, 27].
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20 Encouraged by our results showing that urolithin A reduced protein levels of
21 acetyl-p65 in LPS-stimulated microglia, we were interested in evaluating whether this
22 compound produces a direct activation of SIRT-1 in BV2 microglia. ELISA
23 experiments showed that at 2.5 μ M of urolithin A, the increase in nuclear SIRT-1
24 protein was not significant when compared with untreated cells (Figure 3A).
25 However, nuclear SIRT-1 protein levels in cells treated with 5 and 10 μ M of the
26 compound were significantly ($p < 0.05$) higher than the levels in untreated cells.
27 Similarly, significant ($p < 0.01$) SIRT-1 activity was observed in BV2 cells treated with
28 5 and 10 μ M of urolithin A (Figure 3B). These results were further confirmed by
29 immunofluorescence microscopy for the detection of SIRT-1 in BV2 cells
30 (Supplementary Figure S6).
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33 Experiments were thereafter carried out to confirm a direct involvement of SIRT-1 in
34 the anti-inflammatory activity of urolithin A in LPS-activated microglia. In this case,
35 BV2 cells were treated with EX527 (1 μ M) prior to urolithin A (10 μ M) followed by
36 LPS stimulation for 24 h. Analyses of cell culture supernatants revealed that urolithin
37 A treatment significantly ($p < 0.05$) diminished LPS-induced release of TNF α , IL-6,
38 and nitrite (Figure 4). However, in the presence of EX527, the anti-inflammatory
39 activity observed by urolithin A treatment was reversed (Figure 4). A similar outcome
40 is depicted in Supplementary Figure S7, showing a partial reversal in the inhibition of
41 LPS-induced nuclear accumulation of NF- κ B p65 sub-unit in BV2 microglia in the
42 presence of EX527.
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3 Taken together, these results show for the first time that urolithin A produces a direct
4 activation of SIRT-1 in BV2 microglia. The outcome of these experiments also
5 suggests that SIRT-1 is required for the anti-inflammatory activity of urolithin A in
6 BV2 microglia. These are interesting observations as neuroprotective dietary
7 compounds in fruits and vegetables such as kaempferol, quercetin, acacetin,
8 apigenin, rutin and luteolin have been previously shown to produce NF- κ B-mediated
9 inhibition of neuroinflammation and to provide health benefits through the activation
10 of SIRT-1^[28]. Recently, we showed that compounds such as thymoquinone and
11 agathisflavone produced inhibition of neuroinflammation in LPS-activated BV2
12 microglia through mechanisms involving SIRT-1^[18, 19]. There have also been reports
13 linking SIRT-1 activation to anti-inflammatory and neuroprotective actions of
14 resveratrol^[29] further confirms our results. SIRT-1-mediated inhibition of
15 neuroinflammation by dietary phytochemicals and their metabolites is a remarkable
16 development with significant implications in nutritional strategies for
17 neuroinflammation-mediated neurodegeneration in the ageing brain.

3.3 Induction of autophagic flux contributes to inhibition of neuroinflammation by urolithin A

34 Recent reports have suggested that urolithin A increases autophagy^[13, 16]. We were
35 therefore interested in evaluating whether this compound induces autophagy in BV2
36 microglia, and whether increasing autophagic flux contributes to its anti-inflammatory
37 activity in LPS-activated BV2 microglia. To answer the first question, we investigated
38 the effect of urolithin A treatment in an autophagy fluorescent assay, which is assay
39 is based on the fluorescent activity of an autophagosome marker. Results in
40 Figure 5A show that treatment with urolithin A (5 and 10 μ M) resulted in a significant
41 ($p < 0.05$) increase in autophagic activity, when compared to untreated cells.

42 Next, we transfected both HEK293 and BV2 cells with the autophagy LC3 HiBiT
43 reporter vector followed by urolithin A treatment. This approach was used due to low
44 transfection efficiency normally observed in BV2 microglia. Results obtained from
45 easy-to-transfect cells such as HEK293 cells are valuable in confirming the outcome
46 of experiments with BV2 cells. As expected, results in figures 5B and 5C show that
47 luminescent signals from HEK293 cells were stronger than those in BV2 cells, as a
48 result of a more efficient transfection of HEK293 cells. The differences in
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luminescence in HEK293 and BV2 microglia notwithstanding, we observed significant ($p < 0.05$) and concentration-dependent decrease in luminescent signals in both HEK293 and BV2 cells treated with urolithin A in comparison to untreated control cells, thus demonstrating lower levels of the LC3 reporter as a result of treatment with the compound.

This is the first report showing that urolithin A increases autophagy in microglia and it seems to reflect earlier observations in peripheral macrophages^[13]. These results as well as reports proposing a role for autophagy in inflammation in macrophages and microglia^[30, 31] urged us to study whether increasing autophagic flux in BV2 microglia by treatment with urolithin A might contribute to its inhibitory effects on neuroinflammation. Incubation of BV2 microglia with chloroquine (40 μM) followed by urolithin A (10 μM) prior to stimulation with LPS, resulted in a significant diminishment of the inhibition of LPS-induced release of pro-inflammatory mediators (TNF α , IL-6, and nitrite) by urolithin A (Figure 6). This observation led us to conclude that the ability of this compound to induce autophagy is a contributory factor in its ability to produce anti-inflammatory activity in BV2 microglia and in macrophages^[13].

Studies have linked autophagy to neuroinflammatory events in microglia. Autophagic processes have been shown to mediate the release of pro-inflammatory cytokines such as IL-1 β in the microglia^[32]. In addition, a *Ginkgo biloba* extract was shown to activate autophagy and inhibit microglia-mediated neuroinflammation in AD mouse brain model and primary microglia^[33, 34]. This suggests that induction of autophagy by nutritional phytochemicals may be a critical strategy in preventing neuroinflammation in brain ageing and neurodegenerative diseases.

3.4 Urolithin A prevented neurotoxicity and A β production in APPSwe-transfected ReNcell VM human neural cells

The 3D human neural cell culture system is a valuable model for AD^[35, 36]. We have therefore investigated the neuroprotective effects of urolithin A in these cells following transfection with the human APPSwe plasmid to produce the characteristic amyloid beta-induced neurotoxicity. Figures in S1-S3 (supplementary data) show differentiation of the ReNcell VM human neural cells into neurons as shown by immunostaining for β III-tubulin and MAP2. Figures 7A and 7B show that transfection of differentiated ReNcell VM human neural cells with the APPSwe plasmid for 48 h

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3 resulted in significant ($p < 0.001$) reduction in cell viability with a corresponding
4 significant ($p < 0.01$) increase in LDH released by the cells. However, in the presence
5 of urolithin A, a significant ($p < 0.05$) and concentration-dependent neuroprotective
6 activity was observed. In addition, treatment with the compound resulted in a
7 significant ($p < 0.05$) reduction in the production of the neurotoxic amyloid beta by the
8 cells (Figure 7C). These results are consistent with an earlier report which showed
9 that urolithin A produced neuroprotection in SH-SY5Y neuronal cells^[15]. Similar
10 results have also been reported in *C. elegans* following $A\beta_{(1-42)}$ -induced neurotoxicity
11 and paralysis^[21].
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19 **3.5 Neuroprotection by urolithin A was reversed by inhibition of SIRT-1 and** 20 **autophagy in APPSwe-transfected ReNcell VM human neural cells** 21

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23 Further experiments demonstrated that urolithin A increases the activity and protein
24 levels of SIRT-1 in differentiated ReNcell VM human neural cells (Figures 8A-C), and
25 that its ability to protect neural cells from amyloid-induced toxicity is dependent on
26 SIRT-1 (Figures 9A and 9B). This is demonstrated by a loss of neuroprotective
27 activity in the presence of EX527. SIRT-1-mediated neuroprotection by urolithin A
28 reflects earlier reports showing that SIRT-1 is neuroprotective against $A\beta$ plaque
29 formation in the APPswe/PS1dE9 mouse model^[37]. The observed SIRT-1-mediated
30 neuroprotective effect of urolithin A has significant implications for its potential
31 benefits in AD, as it has been reported that SIRT-1 reduced $A\beta$ levels, oxidative
32 stress, and consequently neuronal loss^[38]. The effects of urolithin A on SIRT-1 in
33 both the microglia and neural cells clearly suggest that the compound might prevent
34 neurodegeneration through the inhibition of neuroinflammation and $A\beta$ -mediated
35 neurotoxicity
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46 Studies have suggested that induction of autophagy and neuroprotection involve
47 mechanisms related to activation of SIRT-1^[39]. Following our observation that
48 urolithin A activates neuroprotective mechanisms such as activation of SIRT-1 in
49 differentiated ReNcell VM human neural cells, we were interested in evaluating
50 whether the compound also induces autophagy in these cells. Treating the cells with
51 urolithin A resulted in the induction of autophagy as shown in Figures 10A and 10B.
52 We further demonstrate that in the presence of chloroquine (an autophagy inhibitor),
53 the neuroprotective effect of urolithin A against amyloid-induced neurotoxicity was
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3 significantly reversed (Figures 11A and 11B). These results suggest a possible link
4 between the activation of SIRT-1 and the induction of autophagy by urolithin A and
5 its neuroprotective effects in neural cells.
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8 9 **4. Concluding remarks**

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11 This study to our knowledge shows for the first time that SIRT-1 activation and
12 induction of autophagy are two potentially linked mechanisms involved in the anti-
13 inflammatory activity of urolithin A in LPS-activated BV2 microglia. These
14 mechanisms also determine the neuroprotective activity of the compound in a 3D
15 model of AD. It is not currently clear if autophagic induction and SIRT-1 activation by
16 this compound are coupled or if they contribute independently to the neuroprotective
17 effects of urolithin A in microglia and neurons.
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Figure Legends

Figure 1

Structure of urolithin A

Figure 2

Effect of urolithin A (2.5-10 μM) on NF- κB -mediated pro-inflammatory mediator release in LPS-stimulated BV2 microglia. Treatment resulted in a reduction in the production of TNF α , IL-6 and nitrite in culture supernatants following incubation with LPS (100 ng/ml) for 24 h (A). In cell western ELISA in LPS-activated BV2 microglia for phospho-p65 NF- κB (B) and acetyl-p65 NF- κB (C). Values are mean \pm SEM for at least 3 independent experiments (** $p < 0.01$; *** $p < 0.001$, compared with LPS only; one-way ANOVA with post-hoc Tukey test).

Figure 3

Urolithin A directly activates SIRT-1 in BV2 microglia. BV2 cells were treated with urolithin A (2.5-10 μM) for 24 h. This was followed by ELISA to determine levels of SIRT-1 protein in nuclear extracts (A), luminescent SIRT-Glo assay for SIRT-1 activity in live cells (B). Values are mean \pm SEM for at least 3 independent experiments (ns: not significant; * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$, compared with untreated control; one-way ANOVA with post-hoc Tukey test).

Figure 4

Inhibition of neuroinflammation by urolithin A is dependent on SIRT-1. Levels of nitrite, TNF α and IL-6 in culture supernatants in BV2 microglia stimulated with LPS (100 ng/ml) were reduced in the presence of urolithin A (10 μM). However, in the presence of EX527 (1 μM), anti-inflammatory activity of urolithin A was diminished. Values are mean \pm SEM for at least 3 independent experiments (** $p < 0.01$; *** $p < 0.001$ compared with untreated control. ⁰⁰ $p < 0.01$; ⁰⁰⁰ $p < 0.001$, compared with LPS control. ^{&&} $p < 0.01$; ^{&&&} $p < 0.001$ compared with urolithin A +LPS treatment only; one-way ANOVA with post-hoc Tukey test).

Figure 5

Autophagic induction by Urolithin. Treatment of BV2 microglia resulted in induction of autophagy measured with a colorimetric autophagy assay kit (A). Transfection of HEK293 cells (B) and BV2 microglia (C) with the LC3 HiBiT reporter followed by treatment with urolithin A (2.5-10 μM) resulted in an increase in reporter activity as measured by luminescence. Values are mean \pm SEM for at

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3 least 3 independent experiments (ns: not significant; * $p < 0.05$; *** $p < 0.001$,
4 compared with untreated control; one-way ANOVA with post-hoc Tukey test).

6 **Figure 6**

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8 Autophagy induction plays a role in the anti-inflammatory activity of urolithin A in
9 LPS-activated BV2 microglia. Levels of TNF α , IL-6 and nitrite in culture
10 supernatants of BV2 microglia stimulated with LPS (100 ng/ml) were reduced in
11 the presence of urolithin A (10 μ M). However, in the presence of chloroquine
12 (40 μ M), anti-inflammatory activity of urolithin A was diminished. Values are
13 mean \pm SEM for at least 3 independent experiments (*** $p < 0.001$ compared with
14 untreated control. ⁰⁰⁰ $p < 0.001$ compared with LPS only. &&& $p < 0.001$ compared
15 with urolithin A +LPS treatment; one-way ANOVA with post-hoc Tukey test).

22 **Figure 7**

23
24 Urolithin A produces neuroprotective effect in differentiated ReNcell VM human
25 neural stem cells. Treatment of APPSwe-transfected ReNcell VM human neural
26 stem cells with urolithin A (2.5-10 μ M) prevented neuronal death measured with
27 MTT (A) and LDH (B) assays. This was accompanied by a reduction in the
28 production of amyloid beta (C). Values are mean \pm SEM for at least 3
29 independent experiments (* $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$, compared with
30 APPSwe-transfected cells; one-way ANOVA with post-hoc Tukey test).

36 **Figure 8**

37
38 Treatment of differentiated ReNcell VM human neural stem cells with urolithin A
39 increased nuclear concentration of SIRT-1 protein (A) and increased SIRT-1
40 activity in live cells (B). Immunofluorescent microscopy also shows an induction
41 in nuclear accumulation of SIRT-1 in the cells (C). Values are mean \pm SEM for at
42 least 3 independent experiments (** $p < 0.01$; *** $p < 0.001$, compared with untreated
43 control; one-way ANOVA with post-hoc Tukey test).

48 **Figure 9**

49
50 Loss of neuroprotection by urolithin A (10 μ M) in the presence of EX527 (1 μ M) in
51 APPSwe-transfected ReNcell VM human neural stem cells. Neural viability was
52 measured using the MTT (A) and LDH (B) assays. Values are mean \pm SEM for at
53 least 3 independent experiments (** $p < 0.01$; *** $P < 0.001$, compared with untreated
54 cells. ⁰ $p < 0.05$; ⁰⁰⁰ $p < 0.001$, compared with APPSwe control. && $p < 0.01$;
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3 $p < 0.001$, compared with urolithin A + APPSwe control; one-way ANOVA with
4 post-hoc Tukey test).

6 **Figure 10**

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8 Treatment of differentiated ReNcell VM human neural stem cells resulted in
9 induction of autophagy measured with a colorimetric autophagy assay kit (A).
10 Transfection of differentiated ReNcell VM human neural stem cells with the LC3
11 HiBiT reporter followed by treatment with urolithin A (2.5-10 μM) resulted in an
12 increase in reporter activity as measured by luminescence (B). Values are mean
13 \pm SEM for at least 3 independent experiments (ns: not significant; * $p < 0.05$;
14 ** $p < 0.01$; *** $p < 0.001$, compared with untreated control; one-way ANOVA with
15 post-hoc Tukey test).

22 **Figure 11**

23
24 Reversal of neuroprotection by urolithin A (10 μM) in APPSwe-transfected
25 ReNcell VM human neural stem cells in the presence of chloroquine (40 μM) was
26 measured with MTT (A) and LDH (B) assays. Values are mean \pm SEM for at least
27 3 independent experiments (*** $p < 0.001$; compared with untreated cells.

28
29 $p < 0.001$, compared with APPSwe control. $p < 0.01$; $p < 0.001$, compared
30 with urolithin A + APPSwe control; one-way ANOVA with post-hoc Tukey test).

34 **Supplementary Figure 1 (S1)**

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36 Immunofluorescence detection of nestin in undifferentiated ReNcell VM human
37 neural stem cells.

39 **Supplementary Figure 2 (S2)**

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41 Immunofluorescence detection of β III tubulin in differentiated ReNcell VM human
42 neural stem cells.

45 **Supplementary Figure 3 (S3)**

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47 Immunofluorescence detection of MAP2 in differentiated ReNcell VM human
48 neural stem cells.

50 **Supplementary Figure 4 (S4)**

51
52 Treatment of BV2 microglia with urolithin A (2.5-10 μM), followed by stimulation
53 with LPS (100 ng/ml) for 24 h did not affect cell viability as determined by MTT
54 viability assay.

57 **Supplementary Figure 5 (S5)**

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3 Immunofluorescent analyses showing that pre-treatment with urolithin A (2.5-
4 10 μ M) resulted in inhibition of nuclear accumulation of NF- κ B p65 sub-unit (A)
5 and acetylation (B) in BV2 microglia stimulated with LPS (100 ng/ml) for 60 min.
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8 **Supplementary Figure 6 (S6)**
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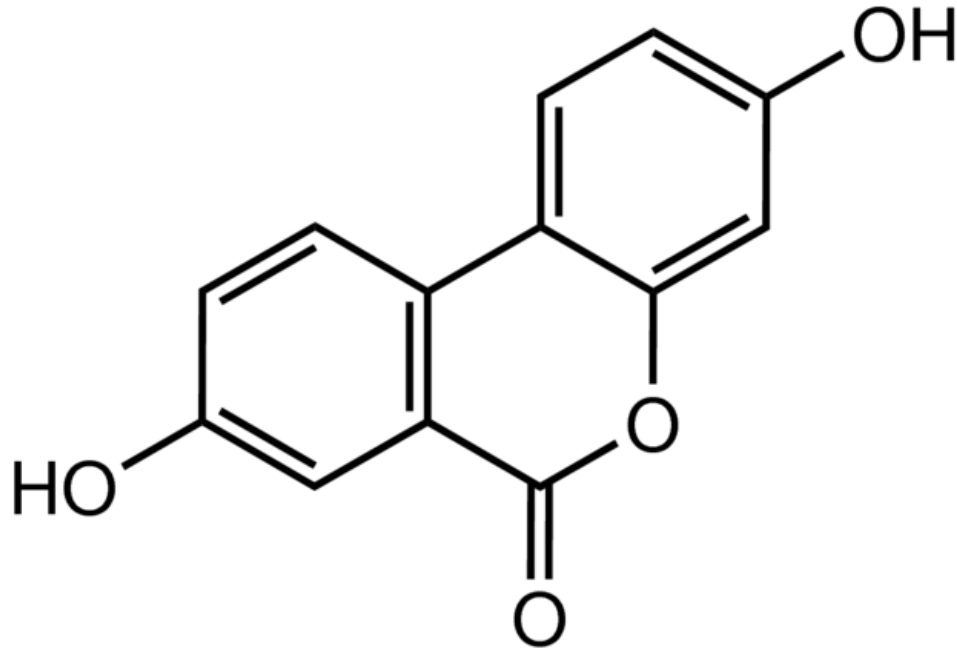
10 Immunofluorescent detection of SIRT-1 protein following treatment of BV2
11 microglia with urolithin A (2.5-10 μ M).
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13 **Supplementary Figure 7 (S7)**
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15 Immunofluorescence showing SIRT-1-dependent inhibition of nuclear localisation
16 of NF- κ B in LPS-stimulated BV2 microglia by urolithin A.
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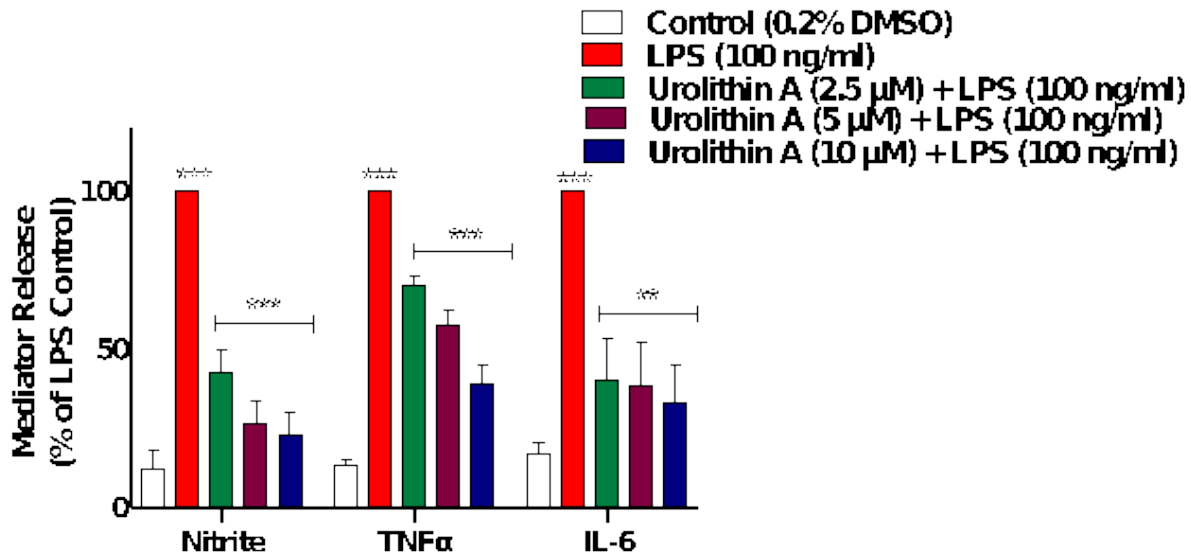
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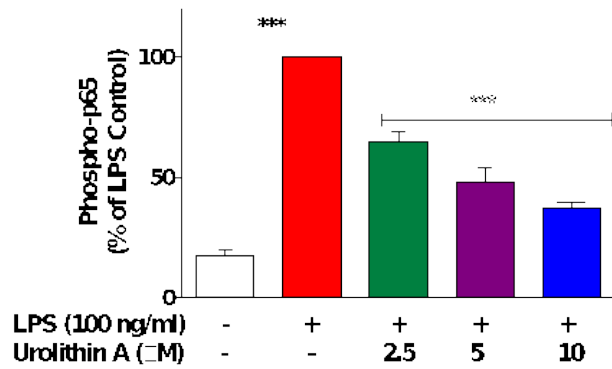
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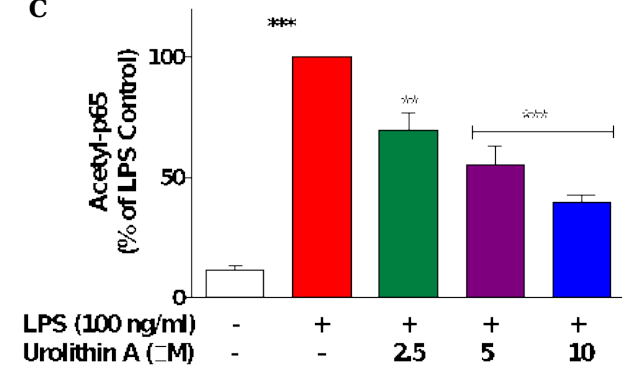
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Figure 2

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Figure 3

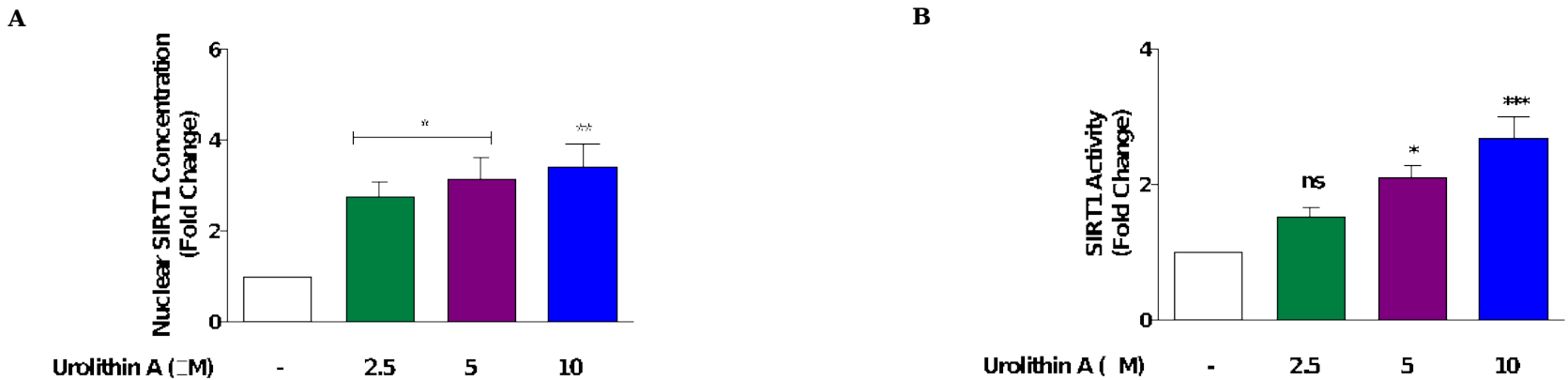
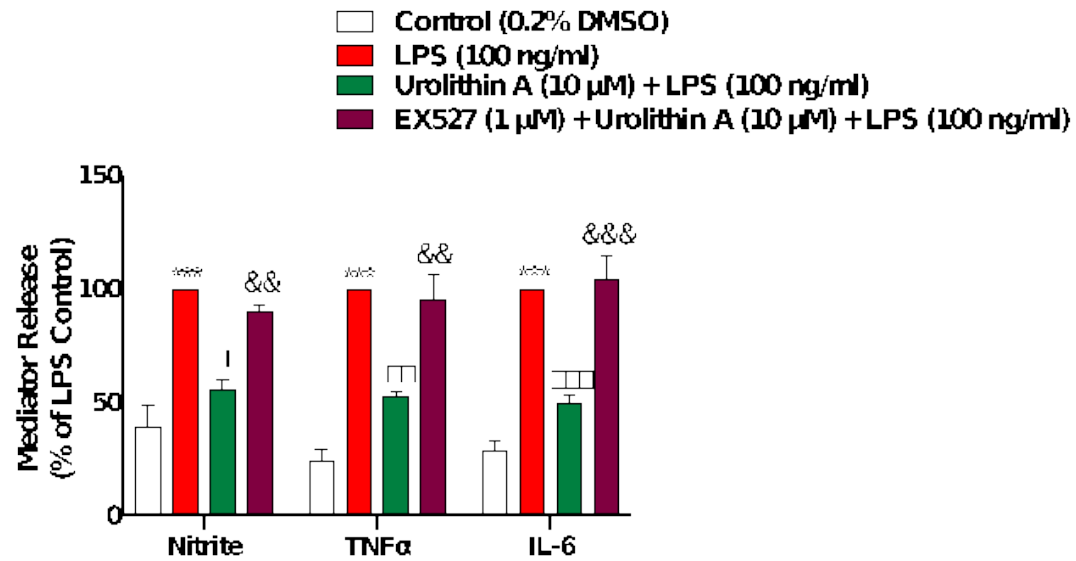


Figure 4



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Figure 5

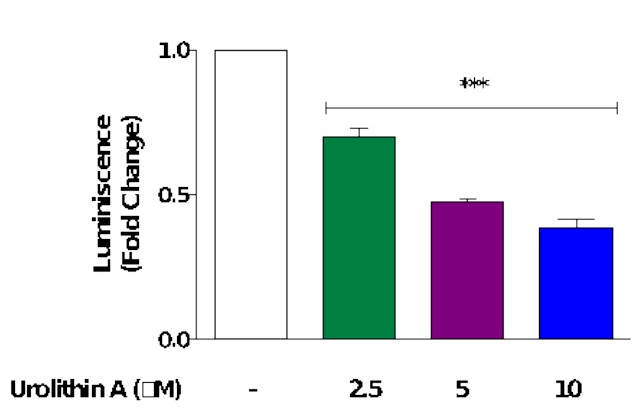
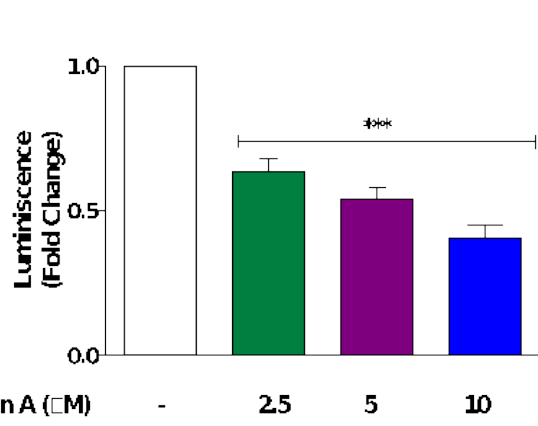
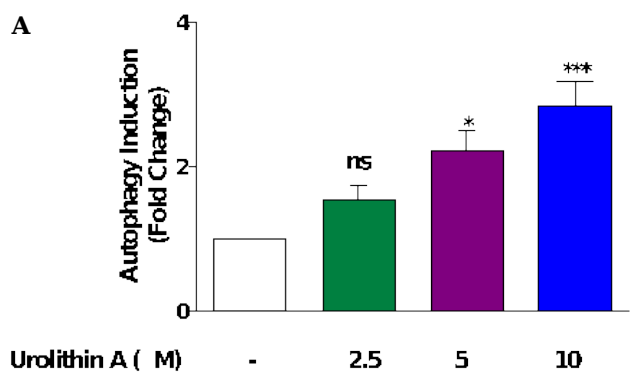
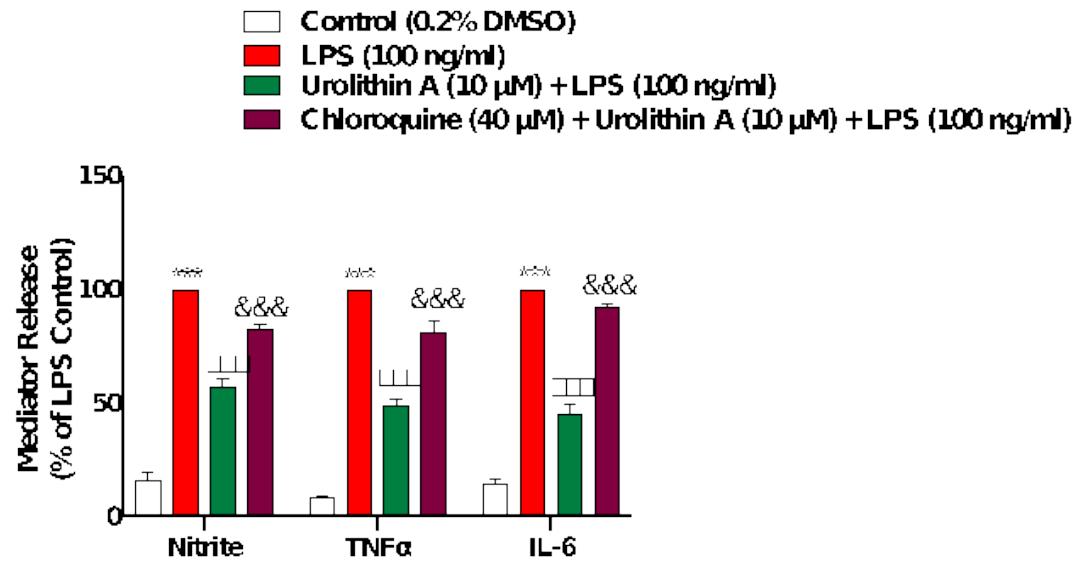
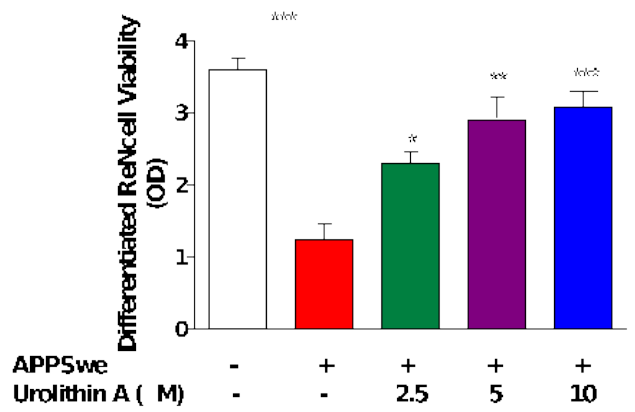


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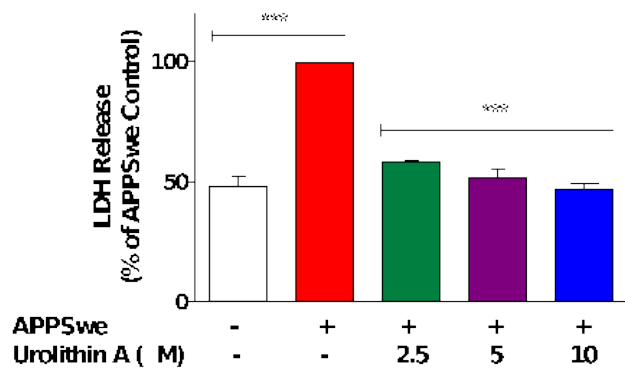


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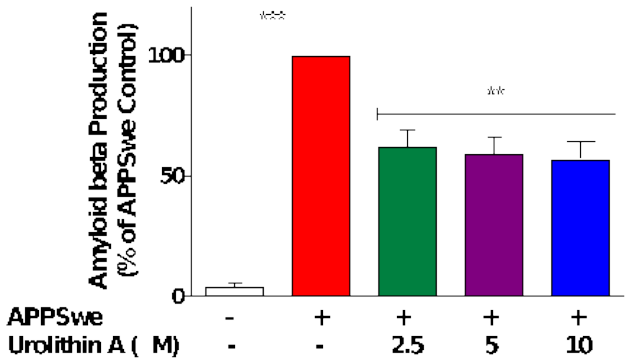
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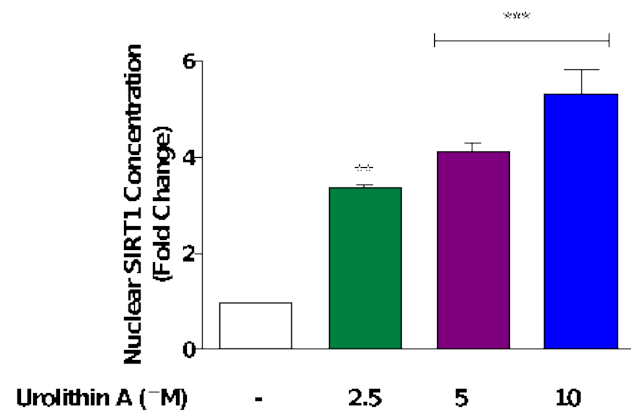
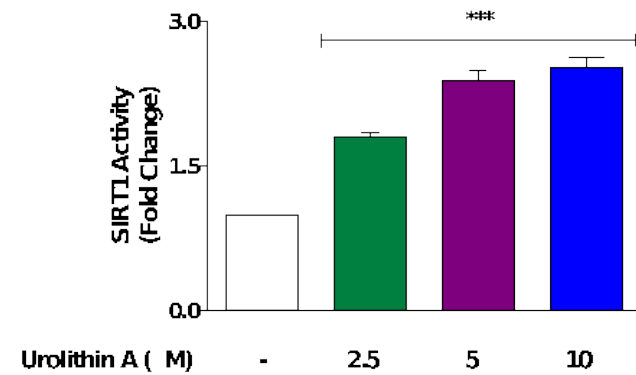


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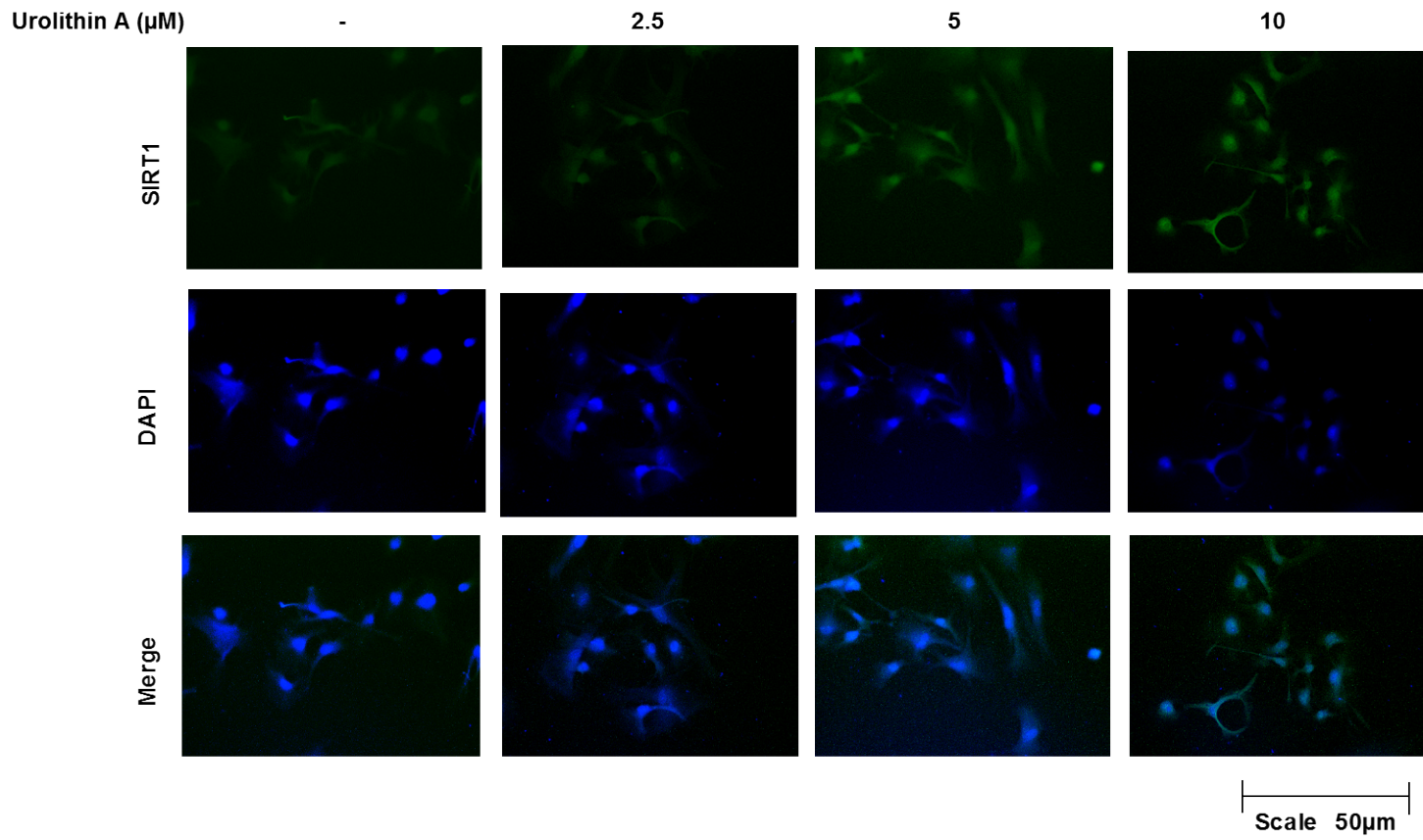


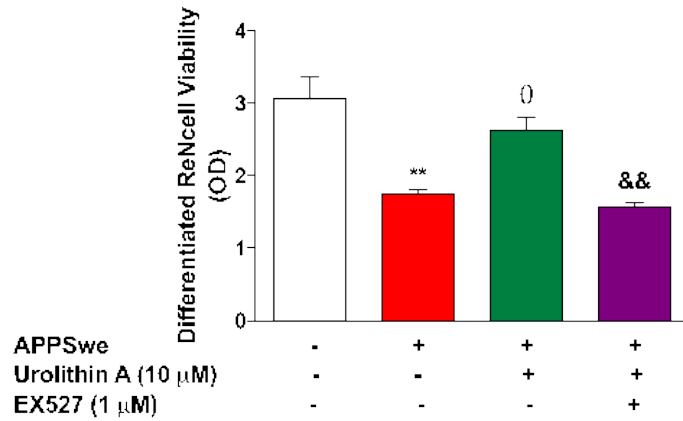
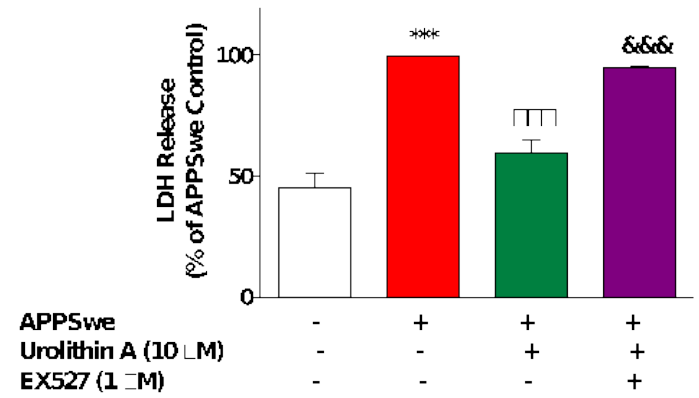
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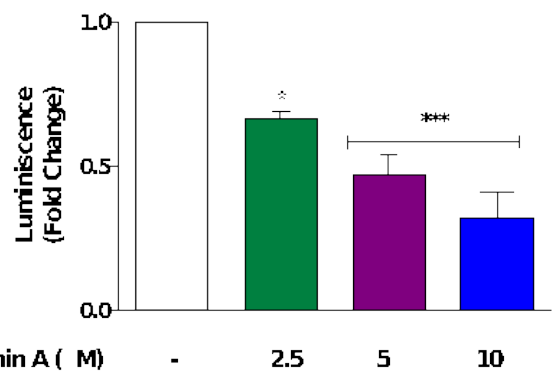


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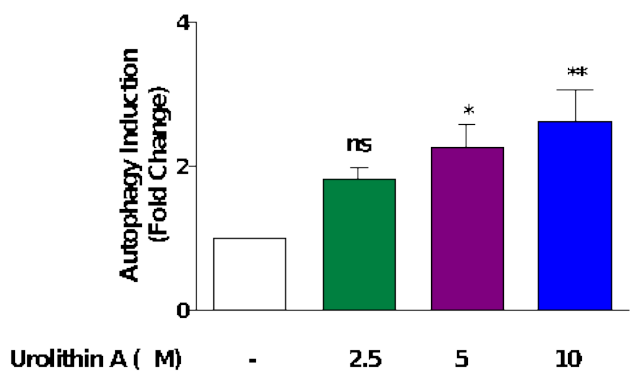
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Figure 10

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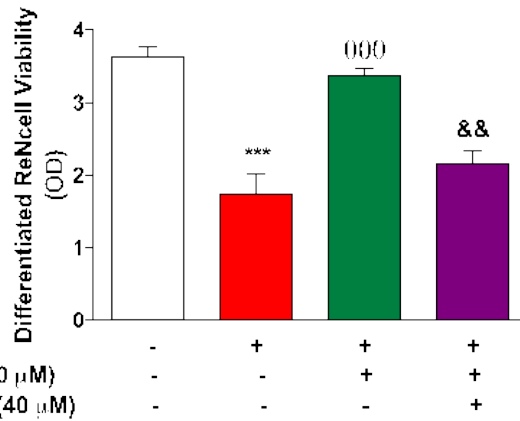
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