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Half-sandwich d<sup>6</sup> metal complexes comprising of 2-substituted-1,8-napthyridine ligands with unexpected bonding modes: Synthesis, structural and anti-cancer studies

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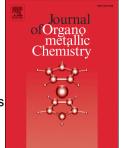
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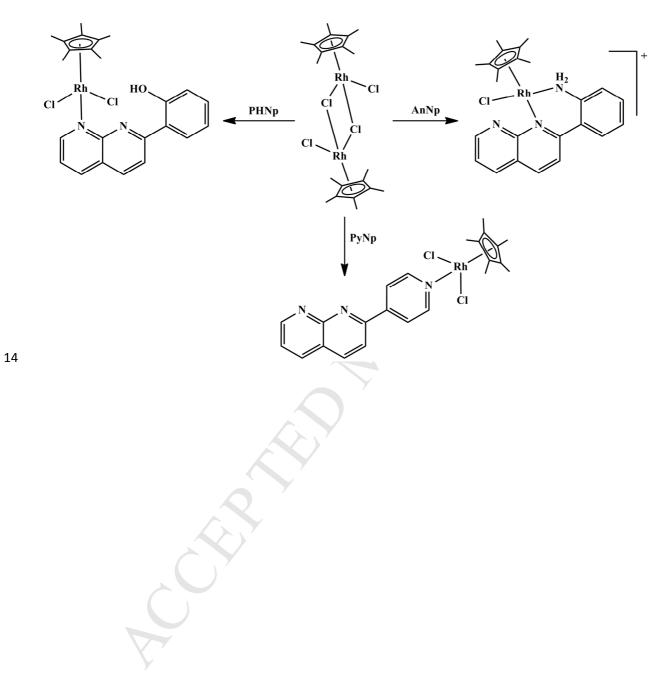
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## 12 Graphical abstract



## 15 Abstract

Friedländer condensation reaction between 2-aminonicotinealdehyde and 16 The acetophenone derivatives in presence of potassium hydroxide yielded 2-substituted-1,8-17 napthyridine derivatives viz. 2-(1,8-napthyridin-2-yl)phenol (PHNp), 2-(1,8-napthyridin-2-18 yl)aniline (AnNp) and 2-(pyridine-4-yl)-1,8-napthyridine (PyNp). Treatment of the chloro-19 bridged dimers  $[(arene)MCl_2]_2$  [arene = p-cymene, Cp\*; M = Ru, Rh and Ir] with two 20 21 equivalents of napthyridine ligands (PHNp, AnNp and PyNp) allowed the formation of mononuclear napthyridine complexes having formula [(arene)M(PHNp)Cl<sub>2</sub>] 22 (1-3). [(arene)M(AnNp)Cl]PF<sub>6</sub> (4-6) and [(arene)M(PyNp)Cl<sub>2</sub>] (7-9). These napthyridine compounds 23 were isolated as neutral and cationic complexes which were further characterized by analytical 24 and spectroscopic techniques. The molecular structures of some of the respective napthyridine 25 26 complexes were established by carrying out the single crystal X-ray analysis. Single crystal Xray studies revealed the coordination of the napthyridine ligands to the metal center wherein 27 AnNp ligand coordinated metal in a bidentate chelating NN' manner and PHNp and PyNp ligand 28 coordinated metal in a monodentate fashion. In case of PHNp complexes the coordination occurs 29 through napthyridine nitrogen N(1) whereas in case of PyNp complexes the coordination takes 30 place through pyridine nitrogen N(1). These napthyridine complexes possessed cytotoxicity 31 32 against HCT-116 (human colorectal cancer) and MIA-PaCa-2 (pancreatic carcinoma) cancer cell lines as compared to non-cancer cell line ARPE-19. 33

34 Keywords: Ruthenium, rhodium, iridium, napthyridine's, cytotoxicity

#### 35 Introduction

Arene ruthenium complexes are a well-known class of organometallic compounds 36 generally referred to as half-sandwich complexes having potential applications in many arenas 37 38 [1]. These organometallic compounds have been found to possess clinical as well as industrial applications [2]. Ruthenium complexes have also displayed remarkable activity in medicinal 39 chemistry and these complexes have the potential to act as metal based anti-cancer drugs [3, 4]. 40 Two such complexes in particular namely  $[Ru(\eta^6-arene)Cl(en)]^+$  (en = ethylenediamine) and 41 [Ru(cymene)Cl<sub>2</sub>(PTA)], known as RAPTA-C (PTA = 1,3,5-triaza-7-phosphaadamantane), have 42 been found to possess excellent cytotoxic and anti-cancer activity both in vitro and in vivo 43 including activity against cisplatin resistant cancer cells [5, 6]. Analogous to arene ruthenium 44 complexes, Cp\* rhodium and Cp\* iridium complexes are also being studied as an alternative to 45 platinum based drugs because of the inert facial co-ligand Cp\* which is expected to offer several 46 advantages such as water solubility and lability [7]. These complexes also serve as catalysts for 47 various organic transformations such as hydrogenation and C-H activation [8, 9]. 48

1,8-napthyridines represents an important class of ligands containing two fused pyridine 49 rings and whose structure is closely related to bipyridines and phenanthrolines. These ligands 50 possess several donor sites thus allowing them to act as monodentate, bidentate chelating and 51 bridging coordinating ligand [10]. Substitution of an appropriate donor groups such as pyridyl, 52 thiazolyl, furyl and pyrrole at the 2-position of 1,8-napthyridine provides a ligand, which can 53 coordinate metal with the substituted ring in addition to the napthyridine ring [11]. The 54 55 napthyridine ligands acts as a bridge to communicate between the two metal centers to come 56 close to each other and which is expected to alter the bonding magnetic or energy transfer 57 interactions. In cases where the donor groups attached at the 2-position of the napthyridine ring were substituted phenyl (F and OMe) and N-methyl pyrrole groups, interesting coordinating 58

chemistry has been observed for diruthenium complexes [12]. Despite having a rich diversified 59 chemistry of napthyridine metal complexes, it is noteworthy that only a few half-sandwich 60 platinum group metal napthyridine complexes have been reported to date [10, 13]. Based on the 61 reactivity and different bonding modes of napthyridine ligands we anticipated that what would 62 be the outcome if we substitute donor group such as phenol, aniline and pyridine at the 2-63 position of the napthyridine ring. We assumed that the formed ligand would display distinctly 64 unusual bonding modes and we have therefore explored this possibility in our present work. 65 Ligands used in the present study are 2-(1,8-napthyridin-2-yl)phenol (PHNp), 2-(1,8-66 napthyridin-2-yl)aniline (AnNp) and 2-(pyridine-4-yl)-1,8-napthyridine (PyNp). To the best of 67 68 our knowledge the coordination chemistry of these ligands has not been explored previously.

Herein, we report the synthesis and anti-cancer studies of ruthenium, rhodium and
iridium half-sandwich complexes bearing phenol, aniline and 4-pyridyl groups substituted at 2position of 1,8-napthyridine moiety.

## 72 Experimental

#### 73 *Physical methods and materials*

The reagents used were of commercial quality and used without further purification. 74 Metal chloride's RuCl<sub>3</sub>.*n*H<sub>2</sub>O, RhCl<sub>3</sub>.*n*H<sub>2</sub>O and IrCl<sub>3</sub>.*n*H<sub>2</sub>O were purchased from Arora Matthey 75 Limited.  $\alpha$ -phellandrene, pentamethylcyclopentadiene and 2'-hydroxyacetophenone were 76 purchased from Sigma-Aldrich. 2-aminonicotinaldehyde and 4-acetyl pyridine were obtained 77 from Alfa Aesar and 2'-aminoacetophenone was obtained from Spectrochem. The solvents were 78 dried and distilled prior to use according to standard procedures [14]. Precursor metal complexes 79  $[(p-cymene)RuCl_2]_2$  and  $[Cp*MCl_2]_2$  (M = Rh/Ir) were prepared according to the published 80 procedures [15, 16]. <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on a Bruker Advance II 400 MHz 81

spectrometer using CDCl<sub>3</sub> and DMSO-d<sub>6</sub> as solvents; chemical shifts were referenced to TMS.
Infrared spectra (KBr pellets; 400-4000 cm<sup>-1</sup>) were recorded on a Perkin-Elmer 983
spectrophotometer. Mass spectra were obtained from Waters ZQ 4000 mass spectrometer by ESI
method using acetonitrile as solvent. Elemental analyses of the complexes were carried out on a
Perkin-Elmer 2400 CHN/S analyzer.

#### 87 Structure determination by X-ray crystallography

Solvent diffusion method was used for growing single crystals of compounds by layering 88 solutions of the compounds in dichloromethane or chloroform with a fourfold excess of hexane 89 and allowing them to stand undisturbed for one week. Suitable single crystal of compounds were 90 91 chosen and glued onto the tip of glass fiber which was centered in the X-ray beam. The data for the complexes was collected on an Oxford Diffraction Xcalibur Eos Gemini diffractometer using 92 graphite monochromated Mo-K $\alpha$  radiation ( $\lambda = 0.71073$  Å). The strategy for the data collection 93 94 was evaluated using the CrysAlisPro CCD software. Crystal data were collected by standard "phi-omega scan" techniques and were scaled and reduced using CrysAlisPro RED software. 95 The structures were solved by direct methods using SHELXS-97 and refined by full-matrix least 96 squares with SHELXL-97 refining on  $F^2$  [17, 18]. The positions of all the atoms were obtained 97 by direct methods. Metal atoms in the complex were located from the E-maps and all non-98 hydrogen atoms were refined anisotropically by full-matrix least-squares. Hydrogen atoms were 99 100 placed in geometrically idealized positions and constrained to ride on their parent atoms with C---H distances in the range 0.95-1.00 Angstrom. Isotropic thermal parameters U<sub>eq</sub> were fixed such 101 that they were 1.2U<sub>eq</sub> of their parent atom Ueq for CH's and 1.5U<sub>eq</sub> of their parent atom U<sub>eq</sub> in 102 case of methyl groups. Crystallographic and structure refinement parameters for the complexes 103

are summarized in Table 1 and selected bond lengths and bond angles are presented in Table 2.Figures 1-5 were drawn with ORTEP3 program [19].

106 The crystal structure of complex (4) contains disordered DCM molecule. Crystal
107 structure of complexes (8 and 9) contains CHCl<sub>3</sub> molecule in their solved structure.

#### 108 Cell line testing

The biological importance of the naphthyridines prompted us to study the cytotoxic 109 activity of the synthesized napthyridine ligands and complexes. The cytotoxicity was tested 110 against the HCT-116 colorectal carcinoma and MIA-PaCa-2 pancreatic carcinoma cell lines with 111 the  $IC_{50}$  result shown in Table 3. Both cell lines were originally purchased from the American 112 Type Culture Collection (ATCC) and all other reagents were purchased from Sigma Aldrich Co. 113 Ltd (Dorset, UK) unless otherwise stated. The ARPE-19 cell line which is a non-cancer, human 114 epithelial cell line derived from the retina and this was also obtained from ATCC. Cytotoxicity 115 116 of ligands and compounds were evaluated using the standard MTT 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide cellular viability assay as follows. Cells were inoculated into 117 96 well plates at 1.5 x  $10^3$  cells per well and incubated at 37 °C in an atmosphere of 5% CO<sub>2</sub> 118 prior to drug exposure. The napthyridine ligands (Scheme 1) and complexes (1-9) (Scheme 2) 119 were all dissolved in DMSO at a concentration of 25 mM and diluted further with medium to 120 obtain drug solutions ranging from 0.5 to 150 µM. Cisplatin was dissolved in phosphate buffered 121 saline at a stock concentration of 25 mM. The final DMSO concentration was 0.2% (v/v), which 122 is nontoxic to cells. Cells were exposed to drug for 96 hours and cell survival was determined 123 using the MTT assay [20, 21]. Following drug exposure, 20 µL of MTT (0.5 mg/ml) in 124 phosphate buffered saline was added to each well and it was further incubated at 37 °C for 4 125 hours in an atmosphere of 5% CO<sub>2</sub>. The solution was then removed and the formed formazan 126

127 crystals was dissolved in 150  $\mu$ M DMSO and the absorbance of the solution was recorded at 550 128 nm using an ELISA spectrophotometer. Percentage cell survival was calculated by dividing the 129 true absorbance of treated cell by the true absorbance for controls (exposed to 0.2% DMSO). The 130 IC<sub>50</sub> values were determined from plots of % survival against drug concentration. Each 131 experiment was repeated three times and a mean value obtained and stated as IC<sub>50</sub> ( $\mu$ M) ± SD.

### 132 Synthesis of 2-substituted-1,8-napthyridine ligands

The ligands 2-(1,8-napthyridin-2-yl)phenol (PHNp), 2-(1,8-napthyridin-2-yl)aniline 133 (AnNp) and 2-(pyridine-4-yl)-1,8-napthyridine (PyNp) were synthesized according to 134 Friedländer condensation (Scheme 1) [22]. Herein a detailed procedure is provided only for 2-135 (1,8-napthyridin-2-yl)phenol (L1). A stirred solution of 2'-hydroxy acetophenone (408 mg, 3 136 mmol), 2-aminonicotinealdehyde (366 mg, 3 mmol) and KOH (336 mg, 6 mmol) in 50% 137 aqueous methanol (10 mL) was heated to 60 °C overnight. The reaction mixture was quenched 138 by the addition of water (20 mL) and the resulting precipitate was isolated by filtration. The 139 crude product was washed with water, dried in vacuum and collected. 140

- 141 2-(1,8-napthyridin-2-yl)phenol (PHNp)
- 142 Color: Orange crystals; Yield: 82 %; IR (KBr, cm<sup>-1</sup>): 3521(b), 3224(m), 2961(m), 2870(w),
- 143 1644(s), 1469(m), 1358(m); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 14.7 (s, 1H, OH), 8.26 (d, 1H, J =
- 144 12 Hz), 8.15 (d, 1H, J = 8 Hz), 8.08 (d, 1H, J = 8 Hz), 7.90 (d, 1H, J = 8 Hz), 7.43-7.46 (m, 1H),
- 145 7.31-7.34 (m, 1H), 7.24 (d, 1H, *J* = 8 Hz), 7.06 (d, 1H, *J* = 8 Hz), 6.88-6.92 (m, 1H); Anal. Calc
- 146 for C<sub>14</sub>H<sub>10</sub>N<sub>2</sub>O (222.24); C, 75.66; H, 4.54; N, 12.60. Found: C, 75.76; H, 4.67; N, 12.73 %.
- 147 2-(1,8-napthyridin-2-yl)aniline (AnNp)
- 148 Color: Yellow crystals; Yield: 86 %; IR (KBr, cm<sup>-1</sup>): 3402(s), 3293(m), 3058(w), 2922(w),
- 149 1611(s), 1566(m), 1541(m), 1499(m), 1257(m); <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>):  $\delta = 8.84$  (dd,

- 150 1H, J = 4 and 4 Hz), 8.28 (d, 1H, J = 8 Hz), 8.25 (dd, 1H, J = 4 and 4 Hz), 8.00 (d, 1H, J = 8
- 151 Hz), 7.70 (d, 1H, *J* = 8Hz), 7.38-7.41 (m, 1H), 7.34 (s, 2H, NH<sub>2</sub>), 6.98 (t, 1H, *J* = 8 Hz), 6.65 (d,
- 152 1H, J = 8 Hz), 6.46 (t, 1H, J = 8 Hz); Anal. Calc for C<sub>14</sub>H<sub>11</sub>N<sub>3</sub> (221.25); C, 76.00; H, 5.01; N,
- 153 18.99. Found: C, 76.09; H, 5.12; N, 19.08 %.
- 154 2-(pyridine-4-yl)-1,8-napthyridine (PyNp)
- 155 Color: White powder; Yield: 65 %; IR (KBr, cm<sup>-1</sup>): 3021(m), 2927(m), 2811(m), 1634(m),
- 156 1587(m), 1494(m), 1340(s); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 9.21$  (dd, 1H, J = 4 and 4 Hz),
- 157 8.82 (d, 2H, J = 8 Hz), 8.38 (d, 1H, J = 8 Hz), 8.28 (dd, 1H, J = 8 and 8 Hz), 8.20 (d, 2H, J = 8
- 158 Hz), 8.07 (d, 1H, J = 8 Hz), 7.54-7.57 (m, 1H); Anal. Calc for C<sub>13</sub>H<sub>19</sub>N<sub>3</sub> (207.23); C, 75.35; H,
- 159 4.38; N, 20.28. Found: C, 75.47; H, 4.46; N, 20.41 %.

### 160 General procedure for preparation of neutral napthyridine complexes (1-3)

- A mixture of metal precursor  $[(arene)MCl_2]_2$  (arene = *p*-cymene, Cp\*; M = Ru, Rh and Ir) (0.1 mmol) and ligand 2-(1,8-napthyridin-2-yl)phenol (PHNp) (0.2 mmol) were dissolved in dichloromethane and the reaction mixture was stirred overnight at room temperature. This solution was then filtered over celite and the solvent was evaporated under reduced pressure to afford yellow solid which was washed with diethyl ether (2 x 10 mL) and air dried (Scheme 2).
- 166 [(*p-cymene*)*Ru*(*PHNp*)*Cl*<sub>2</sub>] (1)
- 167 Yield: 69 mg (75%); IR (KBr, cm<sup>-1</sup>): 3411(b), 3056(m), 1611(m), 1585(m), 1548(m), 1508(m),
- 168 1473(m), 1249(m), 1155(s); <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>):  $\delta$  = 14.92 (s, 1H, OH), 8.89 (d, 1H,
- 169 J = 4 Hz), 8.48 (d, 1H, J = 8 Hz), 8.31 (t, 2H, J = 8 Hz), 8.07 (d, 1H, J = 8 Hz), 7.47-7.50 (m,
- 170 1H), 7.22 (t, 1H, J = 8 Hz), 6.77-6.82 (m, 2H), 5.61 (d, 2H, J = 4 Hz,  $CH_{(p-cym)}$ ), 5.57 (d, 2H, J = 4 Hz, J = 4 Hz,  $CH_{(p-cym)}$ ), 5.57 (d, 2H, J = 4 Hz,  $CH_{(p-cym)}$ ), 5.57
- 171 8 Hz,  $CH_{(p-cym)}$ ), 2.61 (sept, 1H,  $CH_{(p-cym)}$ ), 1.86 (s, 3H,  $CH_{(p-cym)}$ ), 0.98 (d, 6H, J = 8 Hz,  $CH_{(p-cym)}$ )
- 172 <sub>cym</sub>); <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>):  $\delta$  (ppm) = 160.60, 160.29, 155.19, 149.35, 138.65,

- 173 135.10, 133.83, 128.16, 121.69, 121.11, 119.92, 118.16, (C-PHNp), 105.51, 103.91, 87.71,
- 174 84.21, 30.31, 23.81, 17.41 (C-*p*-cym); ESI-MS (m/z) [Found (Calcd)]: [456.96 (457.08)] [M-Cl-
- 175  $HCl]^+$ ; Anal. Calc for C<sub>24</sub>H<sub>24</sub>Cl<sub>2</sub>N<sub>2</sub>ORu (528.43); C, 54.55; H, 4.58; N, 5.30. Found: C, 54.64; H,
- 176 4.67; N, 5.39 %.

#### 177 [*Cp*\**Rh*(*PHNp*)*Cl*<sub>2</sub>] (2)

Yield: 82 mg (77%); IR (KBr, cm<sup>-1</sup>): 3434(b), 2137(m), 1608(m), 1503(m), 1475(m), 1384(m), 178 1137(m); <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>):  $\delta = 15.12$  (s, 1H, OH), 9.10 (d, 1H, J = 4 Hz), 8.68 179 (d, 1H, J = 8 Hz), 8.51 (d, 2H, J = 8 Hz), 8.26 (d, 2H, J = 4 Hz), 7.67-7.70 (m, 1H), 7.42 (t, 1H, J 180 = 8 Hz), 6.97-7.02 (m, 1H), 1.60 (s, 15H,  $CH_{(Cp^*)}$ ); <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>):  $\delta$  = 160.61, 181 182 160.29, 154.49, 152.35, 139.65, 137.30, 132.83, 128.16, 122.69, 121.11, 118.92, 118.83, 118.18 (C-PHNp), 98.68 (Cp\*<sub>ipso</sub>), 8.51 (Cp\*<sub>Me</sub>); ESI-MS (m/z) [Found (Calcd)]: [459.02 (459.09)] [M-183 Cl-HCl]<sup>+</sup>; Anal. Calc for C<sub>24</sub>H<sub>25</sub>Cl<sub>2</sub>N<sub>2</sub>ORh (531.27); C, 54.26; H, 4.74; N, 5.27. Found: C, 54.37; 184 185 H, 4.87; N, 5.41 %.

#### 186 [*Cp*\**Ir*(*PHNp*)*Cl*<sub>2</sub>] (3)

Yield: 98 mg (78%); IR (KBr, cm<sup>-1</sup>): 3401(b), 2918(w), 2150(m), 1632(m), 1608(m), 1503(m), 187 1475(m), 1137(m); <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>):  $\delta = 15.12$  (s, 1H, OH), 9.10 (dd, 1H, J = 4188 and 4 Hz), 8.68 (d, 1H, J = 8 Hz), 8.51 (t, 2H, J = 8 Hz), 8.27 (d, 1H, J = 8 Hz), 7.67-7.70 (m, 189 1H), 7.42 (t, 1H, J = 8 Hz), 6.98-7.02 (m, 2H), 1.61 (s, 15H, CH<sub>(Cp\*)</sub>); <sup>13</sup>C NMR (100 MHz, 190 DMSO-d<sub>6</sub>):  $\delta = 160.62, 160.30, 154.49, 152.36, 139.65, 137.30, 132.83, 128.17, 122.69, 121.11, 122.69, 121.11, 122.69, 121.11, 122.69, 121.11, 122.69, 121.11, 122.69, 121.11, 122.69, 121.11, 122.69, 121.11, 122.69, 121.11, 122.69, 121.11, 122.69, 121.11, 122.69, 121.11, 122.69, 121.11, 122.69, 121.11, 122.69, 121.11, 122.69, 121.11, 122.69, 122.69, 121.11, 122.69, 122.$ 191 118.92, 118.26, 118.18 (C-PHNp), 92.01 (Cp\*<sub>ipso</sub>), 8.17 (Cp\*<sub>Me</sub>); ESI-MS (m/z) [Found 192 (Calcd)]: [549.10 (549.15)] [M-Cl-HCl]<sup>+</sup>; Anal. Calc for C<sub>24</sub>H<sub>25</sub>Cl<sub>2</sub>N<sub>2</sub>OIr (620.59); C, 46.45; H, 193 4.06; N, 4.51. Found: C, 46.58; H, 4.14; N, 4.62 %. 194

195

#### 196 General procedure for preparation of cationic napthyridine metal complexes (4-6)

A mixture of metal precursor  $[(arene)MCl_2]_2$  (arene = *p*-cymene, Cp\*; M = Ru, Rh and Ir) (0.1 mmol) and 2-(1,8-napthyridin-2-yl)aniline (AnNp) were dissolved in dry methanol (5 mL) and stirred at room temperature for 1 hour. Then 4 equivalents of NH<sub>4</sub>PF<sub>6</sub> dissolved in dry methanol (2 mL) was added dropwise to the reaction mixture and stirring continued for further 5-6 hours whereupon a yellow solid precipitated out from the reaction mixture. The precipitate was filtered, washed with cold methanol (2 x 5 ml) and diethyl ether (2 x 10 ml) and air dried (Scheme 2).

#### 204 $[(p-cymene)Ru(AnNp)Cl]PF_6(4)$

Yield: 101 mg (79%); IR (KBr, cm<sup>-1</sup>): 3314(m), 3125(m), 1604(m), 1494(w), 1462(w), 1111(m), 205 842(s); <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>):  $\delta = 9.86$  (d, 1H, J = 12 Hz), 8.97 (d, 1H, J = 4 Hz), 8.70 206 (d, 1H, J = 8 Hz), 8.27 (d, 1H, J = 8 Hz), 8.00 (d, 1H, J = 4 Hz), 7.86 (t, 1H, J = 8 Hz), 7.51-7.57 207 (m, 2H), 7.31 (t, 2H, J = 8 Hz), 5.99 (dd, 2H, J = 4 and 4 Hz, CH<sub>(p-cvm)</sub>), 5.59 (d, 1H, J = 8.0 Hz, 208  $CH_{(p-cym)}$ ), 5.54 (d, 1H, J = 8 Hz,  $CH_{(p-cym)}$ ), 2.19 (sept, 1H,  $CH_{(p-cym)}$ ), 1.98 (s, 3H,  $CH_{(p-cym)}$ ), 209 0.98 (d, 3H, J = 8 Hz, CH<sub>(p-cym)</sub>), 0.89 (d, 3H, J = 4 Hz, CH<sub>(p-cym)</sub>); <sup>13</sup>C NMR (100 MHz, DMSO-210  $d_6$ ):  $\delta = 161.78, 158.61, 155.19, 148.13, 138.12, 137.32, 134.18, 129.14, 123.41, 120.05, 119.18, 129.14, 123.41, 120.05, 119.18, 129.14, 123.41, 120.05, 119.18, 129.14, 123.41, 120.05, 119.18, 129.14,$ 211 117.16, (C-AnNp), 106.14, 103.41, 89.51, 87.11, 85.22, 30.71, 23.81, 20.21, 16.41 (C-*p*-cym); 212 ESI-MS (m/z) [Found (Calcd)]: [492.06 (492.07)] [M-PF<sub>6</sub>]<sup>+</sup>, ESI-MS (m/z) [Found (Calcd)]: 213 [456.0 (456.10)] [M-PF<sub>6</sub>-HCl]<sup>+</sup>; Anal. Calc for C<sub>24</sub>H<sub>25</sub>ClN<sub>3</sub>F<sub>6</sub>PRu (636.96); C, 45.25; H, 3.96; N, 214 6.60. Found: C, 45.38; H, 4.07; N, 6.71 %. 215

#### 216 $[Cp*Rh(AnNp)Cl]PF_6(5)$

217 Yield: 93 mg (72%); IR (KBr, cm<sup>-1</sup>): 3269(w), 2923(w), 1606(m), 1491(w), 1462(w), 843(s); <sup>1</sup>H

218 NMR (400 MHz, DMSO-d<sub>6</sub>):  $\delta$  = 9.23 (d, 1H, *J* = 4 Hz), 8.81 (d, 1H, *J* = 8 Hz), 8.63 (d, 1H, *J* =

8 Hz), 8.25 (d, 1H, J = 8 Hz), 7.93 (d, 1H, J = 8 Hz), 7.77-7.80 (m, 1H), 7.65 (t, 1H, J = 4 Hz), 7.47-7.54 (m, 2H), 7.40 (t, 1H, J = 8 Hz), 6.71 (d, 1H, J = 4 Hz), 1.34 (s, 15H, CH<sub>(Cp\*)</sub>); <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>):  $\delta = 161.34$ , 157.13, 153.61, 142.01, 140.24, 139.21, 135.47, 133.45, 126.14, 124.16, 123.12, 120.33, 118.23 (C-AnNp), 89.38 (Cp\*<sub>ipso</sub>), 8.13 (Cp\*<sub>Me</sub>); ESI-MS (m/z) [Found (Calcd)]: [494.05 (494.08)] [M-PF<sub>6</sub>]<sup>+</sup>, ESI-MS (m/z) [Found (Calcd)]: [458.06 (458.11)] [M-PF<sub>6</sub>-HCl]<sup>+</sup>; Anal. Calc for C<sub>24</sub>H<sub>26</sub>ClN<sub>3</sub>F<sub>6</sub>PRh (639.80); C, 45.05; H, 4.10; N, 6.57. Found: C, 45.15; H, 4.21; N, 6.63 %.

### 226 $[Cp*Ir(AnNp)Cl]PF_6(6)$

Yield: 110 mg (75%); IR (KBr, cm<sup>-1</sup>): 3316(w), 3254(w), 3065(w), 2923(w), 1606(m), 1462(w), 227 1154(w), 842(s); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 9.34$  (d, 1H, J = 12 Hz), 8.85 (d, 1H, J = 8228 Hz), 8.49 (d, 1H, J = 8 Hz), 8.18 (d, 1H, J = 4 Hz), 8.10 (d, 1H, J = 8 Hz), 8.01 (d, 1H, J = 8 Hz), 229 7.80 (d, 1H, J = 8 Hz), 7.57 (t, 1H, J = 4 Hz), 7.34 (t, 1H, J = 8 Hz), 7.23 (d, 1H, J = 4 Hz), 6.69 230 (d, 1H, J = 12 Hz), 1.39 (s, 15H, CH<sub>(Cp\*)</sub>); <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>):  $\delta = 160.04, 153.63,$ 231 152.89, 142.01, 141.33, 137.57, 131.89, 131.45, 125.84, 123.78, 123.62, 123.36, 119.22 (C-232 AnNp), 86.38 (Cp\*<sub>ipso</sub>), 8.07 (Cp\*<sub>Me</sub>); ESI-MS (m/z) [Found (Calcd)]: [584.07 (584.14)] [M-233 PF<sub>6</sub>]<sup>+</sup>, ESI-MS (m/z) [Found (Calcd)]: [548.15 (548.16)] [M-PF<sub>6</sub>-HCl]<sup>+</sup>; Anal. Calc for 234 C<sub>24</sub>H<sub>26</sub>ClN<sub>3</sub>F<sub>6</sub>PIr (729.11); C, 39.54; H, 3.59; N, 5.76. Found: C, 39.66; H, 3.68; N, 5.82 %. 235 General procedure for preparation of neutral napthyridine metal complexes (7-9) 236

A mixture of metal precursor  $[(arene)MCl_2]_2$  (arene = *p*-cymene, Cp\*; M = Ru, Rh and Ir) (0.1 mmol) and ligand 2-(pyridine-4-yl)-1,8-napthyridine (PyNp) (0.2 mmol) were dissolved in dry methanol (5 mL) and stirred at room temperature for 6 hours. A yellow compound precipitated out from the reaction mixture. The precipitate was collected washed with cold methanol (2 x 5 mL) and diethyl ether (3 x 10 mL) and air dried (Scheme 2).

#### 242 $[(p-cymene)Ru(PyNp)Cl_2](7)$

- 243 Yield: 72 mg (70%); IR (KBr, cm<sup>-1</sup>): 2965(m), 2870(m), 1637(m), 1615(m), 1384(w), 1124(m);
- <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 9.14 (s, 3H), 8.35 (d, 1H, *J* = 8 Hz), 8.24 (d, 1H, *J* = 8 Hz),
- 245 8.08 (d, 2H, J = 8 Hz), 7.95 (d, 1H, J = 8 Hz), 7.53-7.57 (m, 1H), 5.45 (d, 2H, J = 4 Hz, CH<sub>(p-</sub>
- cym)), 5.22 (d, 2H, J = 4 Hz CH<sub>(p-cym)</sub>), 2.94 (sept, 1H, CH<sub>(p-cym)</sub>), 2.06 (s, 3H, CH<sub>(p-cym)</sub>), 1.28 (d,
  6H, J = 8 Hz, CH<sub>(p-cym)</sub>); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ = 155.78, 154.16, 153.19, 148.23,
  139.12, 136.21, 135.11, 130.14, 128.06, 123.05, 119.18, 118.16, (C-PyNp), 106.18, 103.41,
- 249 90.13, 88.41, 30.20, 21.84, 17.80 (C-*p*-cym); ESI-MS (m/z) [Found (Calcd)]: [478.21 (478.06)]
- 250  $[M-Cl]^+$ , ESI-MS (m/z) [Found (Calcd)]: [442.15 (443.09)]  $[M-2Cl]^+$ ; Anal. Calc for
- 251 C<sub>23</sub>H<sub>23</sub>Cl<sub>2</sub>N<sub>3</sub>Ru (513.42); C, 53.80; H, 4.52; N, 8.18. Found: C, 53.93; H, 4.63; N, 8.29 %.

## 252 $[Cp*Rh(PyNp)Cl_2](8)$

Yield: 92 mg (89%); IR (KBr, cm<sup>-1</sup>): 2985(m), 2860(m), 1635(m), 1609(m), 1378(w), 1127(m); 253 <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 9.19$  (s, 1H), 9.15 (d, 2H, J = 4 Hz), 8.45 (d, 1H, J = 8 Hz), 254 8.31 (dd, 1H, J = 4 and 4 Hz), 8.21 (d, 1H, J = 8 Hz), 8.04 (d, 2H, J = 8 Hz), 7.56-7.59 (m, 1H), 255 1.63 (s, 15H,  $CH_{(Cp^*)}$ ); <sup>13</sup>C NMR (100 MHz,  $CDCl_3$ ):  $\delta = 156.31$ , 155.42, 154.34, 153.18, 256 148.33, 139.24, 137.14, 123.43, 123.35, 123.07, 118.14 (C-PyNp), 87.12 (Cp\*<sub>ipso</sub>), 8.63 (Cp\*<sub>Me</sub>); 257 ESI-MS (m/z) [Found (Calcd)]: [480.07 (480.15)] [M-C1]<sup>+</sup>, ESI-MS (m/z) [Found (Calcd)]: 258 [444.16 (445.10)] [M-2C1]<sup>+</sup>; Anal. Calc for C<sub>23</sub>H<sub>24</sub>Cl<sub>2</sub>N<sub>3</sub>Rh (516.26); C, 53.51; H, 4.69; N, 8.14. 259 Found: C, 53.62; H, 4.76; N, 8.23 %. 260

- 261 [*Cp*\**Ir*(*PyNp*)*Cl*<sub>2</sub>] (9)
- 262 Yield: 86 mg (71%); IR (KBr, cm<sup>-1</sup>): 2991(m), 2875(m), 1631(m), 1611(m), 1374(w), 1125(m);
- 263 <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 9.20 (s, 1H), 9.13 (d, 2H, *J* = 8 Hz), 8.45 (d, 1H, *J* = 8 Hz),
- 264 8.31 (dd, 1H, *J* = 4 and 4 Hz), 8.19 (d, 2H, *J* = 4 Hz), 8.06 (d, 1H, *J* = 8 Hz), 7.57-7.60 (m, 1H),

265 1.58 (s, 15H,  $CH_{(Cp^*)}$ ); <sup>13</sup>C NMR (100 MHz,  $CDCl_3$ ):  $\delta = 155.70$ , 155.59, 154.74, 153.88, 266 147.30, 139.32, 137.14, 123.56, 123.10, 123.02, 119.83 (C-PyNp), 86.0 ( $Cp^*_{ipso}$ ), 8.58 ( $Cp^*_{Me}$ ); 267 ESI-MS (m/z) [Found (Calcd)]: [570.21 (570.12)] [M-Cl]<sup>+</sup>, ESI-MS (m/z) [Found (Calcd)]: 268 [534.23 (535.15)] [M-2Cl]<sup>+</sup>; Anal. Calc for  $C_{23}H_{24}Cl_2N_3Ir$  (605.07); C, 45.62; H, 3.99; N, 6.94. 269 Found: C, 45.79; H, 4.08; N, 7.05 %.

- 270 Results and discussion
- 271 Synthesis of napthyridine ligands and metal complexes

The napthyridine ligands were synthesized according to Friedländer condensation 272 reaction and the napthyridine metal complexes (1-9) were synthesized by the reaction of metal 273 274 precursors with respective napthyridine ligands. At first the reaction of PHNp with metal precursor was carried out using triethylamine as base with an aim of deprotonating phenolic 275 hydrogen which would result in formation of NO chelated complex but no deprotonation 276 occurred as evidenced from the <sup>1</sup>H NMR spectra of the complexes which showed the presence of 277 phenolic proton. So the reaction was carried out without using triethylamine and which resulted 278 in formation of the same compounds. X-ray studies revealed that the PHNp ligand coordinated 279 metal in a monodentate manner through napthyridine nitrogen. Reaction of AnNp with precursor 280 afforded mononuclear bidentate chelated complexes. Surprisingly substituting 4-pyridyl group at 281 the 2-position of 1,8-napthyridine in PyNp yielded complexes where PyNp is coordinated to 282 metal through pyridine nitrogen rather than napthyridine nitrogen as in case of PHNp complexes. 283 The three-napthyridine ligands used in this work exhibited interesting coordination modes. 284 Complexes of PHNp and PyNp ligands were isolated as neutral complexes whereas with AnNp 285 ligand, complexes were isolated as ionic salts with PF<sub>6</sub> counter ion. The napthyridine ligands and 286 metal complexes were obtained in good yields. These complexes are soluble in organic solvents 287

such as chloroform, dichloromethane, acetonitrile and DMSO but insoluble in diethyl ether and hexane. The napthyridine ligands and the napthyridine metal complexes were characterized spectroscopically and the molecular structures of some of the complexes were established by single crystal X-ray analysis.

## 292 Spectroscopic characterization of napthyridine ligands

The infrared spectra of PHNp and AnNp ligands exhibited stretching frequencies for OH 293 at 3521 cm<sup>-1</sup> and for the NH<sub>2</sub> group at 3402 and 3293 cm<sup>-1</sup>. The C=N and C=C stretching 294 frequencies of the napthyridine ligands were observed in the range of 1450-1650 cm<sup>-1</sup>. The 295 proton NMR spectra of the napthyridine ligands displayed signals in the range of 6.46-9.21 ppm 296 for the protons of the napthyridine moiety and substituted rings. In addition to these resonances 297 the PHNp ligand displayed a singlet at 14.7 ppm for the phenolic proton and the AnNp ligand 298 exhibited a broad singlet for the NH<sub>2</sub> proton at 7.34 ppm respectively. The respective <sup>1</sup>H NMR 299 300 spectra of the napthyridine ligands are presented in the SI.

## 301 Spectroscopic characterization of napthyridine metal complexes

## 302 IR studies of metal complexes

The IR spectra of the metal complexes suggest the coordination of the napthyridine 303 ligands to the metal atom. The presence of a broad band for complexes (1-3) in the lower 304 frequency region around 3401-3434 cm<sup>-1</sup> for the phenolic OH group suggests that the phenolic 305 group is not involved in coordination to the metal center. This was further supported by NMR 306 and single crystal analysis. Complexes (4-6) displayed characteristic stretching frequencies for 307 NH<sub>2</sub> group at lower frequency region around 3125-3326 cm<sup>-1</sup> as compared to the free ligand at 308 3402 and 3293 cm<sup>-1</sup>. This shift to lower frequency suggests the coordination of the NH<sub>2</sub> group. 309 Further the C=N stretching frequencies of the complexes decreases slightly and was observed in 310

the range of 1603-1615 cm<sup>-1</sup> as compared to the free ligand which suggest the coordination of the napthyridine and pyridine nitrogen's. In addition to these bands a sharp band for the cationic complexes (**4**-**6**) was observed around 842-843 cm<sup>-1</sup> attributed to the P-F stretching frequency of the counter ion [23].

315

## <sup>1</sup>H NMR studies of metal complexes

The <sup>1</sup>H NMR spectra of the metal complexes further support the formation of the 316 complexes. The resulting spectra are depicted in the supplementary information and ensuing data 317 318 are summarized in the experimental section. The metal complexes exhibited signals associated with the ligand protons and signals due to *p*-cymene and Cp\* ring protons. The signals 319 associated with the napthyridine and substituted ring protons were observed in the downfield 320 region around 6.65-9.86 ppm. This shifting of the ligand resonances indicates the coordination of 321 the napthyridine ligands to the metal atom. Also the appearance of the phenolic proton signals in 322 complexes (1-3) around 14.9-15.1 ppm confirms that the phenolic OH group is not involved in 323 coordination to the metal atom. Further confirmation of the binding of napthyridine ligands was 324 confirmed by the appearance of the *p*-cymene and Cp\* ring proton signals. The aromatic protons 325 of the *p*-cymene ligand consisted of doublets around 5.22-5.99 ppm. The isopropyl group of the 326 *p*-cymene ligand displayed doublet for complexes (1 and 7) whereas for complex (4) it showed 327 two doublets around 0.89-1.28 ppm. The methyl and methine protons of the *p*-cymene ligand 328 were observed as a singlet around 1.98-2.07 ppm and septet around 2.19-2.94 ppm. The methyl 329 protons of the Cp\* moiety was observed as a singlet around 1.34-1.61 ppm. 330

331

## <sup>13</sup>C NMR studies of metal complexes

The <sup>13</sup>C NMR spectra further justify the formation of the complexes. The <sup>13</sup>C NMR spectra of the complexes displayed signals associated with the napthyridine carbons and the

appearance of the *p*-cymene moiety carbons, methyl carbon of Cp\* and ring carbon of Cp\* 334 confirms the formation of these complexes. In the <sup>13</sup>C NMR spectra of the complexes the carbon 335 resonances associated with the napthyridine ligand were observed in the region around 118-161 336 ppm. The aromatic carbon resonances of the *p*-cymene ligand were observed in the region 337 around 84-106 ppm. The methine, isopropyl and methyl carbon resonances of the *p*-cymene 338 moiety were observed in the region around 16-31 ppm. In addition to these carbon resonances 339 the ring carbons of the Cp\* ligand displayed signal around 86.0-98.6 ppm and the signal for the 340 methyl protons of the Cp\* moiety was observed around 8.07-8.63 ppm. Overall results from 341 NMR spectral studies strongly support the formation of the metal complexes. 342

### 343 Mass spectral studies of metal complexes

The composition and formation of napthyridine metal complexes (1-9) have further been 344 justified by ESI-mass spectral studies. The mass spectra of the complexes are presented in the 345 346 supplementary information and the values are listed in the experimental section (2.5). The mass spectra of the complexes (1-3) displayed their molecular ion peaks at m/z: 456.96, m/z: 459.02 347 and m/z: 549.10 corresponding to [M-Cl-HCl]<sup>+</sup> ion. Complexes (4-6) displayed their molecular 348 ion peaks at m/z: 492.06, m/z: 494.05 and m/z: 584.07 which corresponds to  $[M-PF_6]^+$  ion peak. 349 Also peaks were observed at m/z: 456.0, m/z: 458.06 and m/z: 548.15 which is due to [M-PF<sub>6</sub>-350 HCl]<sup>+</sup> ion. Similarly complexes (7-9) displayed peaks at m/z: 478.21, m/z: 480.15 and m/z: 351 570.21 which is due to the loss of one chloride  $[M-C1]^+$  ion and also it exhibited peaks at m/z: 352 442.15, m/z: 444.16 and m/z: 534.23 which is due to the loss of both the chloride  $[M-2C1]^+$  ion. 353

## 354 Description of the crystal structures of complexes

355 Apart from spectroscopic analysis, the molecular structures of some of the respective 356 complexes were established by single crystal X-ray analysis. Our attempt to isolate the single

crystal for all the complexes was unsuccessful; however we obtained single crystals for the 357 PHNp ligand and complexes 3, 4, 5, 8 and 9 respectively. By carrying out the single crystal 358 analyses we were able to confirm the various coordination modes of the napthyridine ligand and 359 the geometry of the complexes. The data and molecular structure of complexes 3 and 9 presented 360 here is to only confirm the structure and composition of the molecule. The ORTEP plot of PHNp 361 and the complexes along with atom numbering scheme are shown in (Figures 1-5) respectively. 362 363 The summary of the crystal data including data collection and structure refinement parameters are summarized in Table 1 and geometrical parameters including bond lengths, bond angles and 364 metal atom involving ring centroid values are listed in Table 2. The molecular structure of PHNp 365 366 ligand is shown in (Figure 1). Complexes (1-3) and (7-9) have the neutral species with formula  $[(arene)M(PHNp)Cl_2]$  and  $[(arene)M(PyNp)Cl_2]$  [(arene) = p-cymene, M = Ru (1), (7); Cp\*, M = 367 Rh (2), (8) and Ir (3), (9)]. Complexes (4-6) have the cationic species with general formula 368 [(arene)M(AnNp)Cl] [(arene) = p-cymene, M = Ru (4); Cp\*, M = Rh (5) and Ir (6)] and counter 369 anion PF<sub>6</sub>. These complexes adopts a familiar three legged piano-stool geometry around the 370 metal center with coordination sites occupied by arene/Cp\* ring (arene = p-cymene and Cp\*) in 371 a  $\eta^6/\eta^5$  manner, nitrogen donor atoms from napthyridine ligand and terminal chloride. The 372 geometry at the metal atom is pseudo-octahedral wherein the polycyclic arene ligand acts as seat 373 and napthyridine ligand and terminal chloride form the legs. In complex (3) iridium metal is 374 coordinated through Cp\* ring, PHNp ligand in a monodentate fashion coordinating iridium 375 through napthyridine nitrogen (N1) and two chlorides thus possessing a three-legged piano-stool 376 structure (Figure 2). In complexes 4 and 5 the metal is coordinated through *p*-cymene and Cp\* 377 ring, AnNp ligand in a bidentate manner and one chloride. The AnNp ligand acts as a chelating 378 ligand coordinating metal through aniline nitrogen N(1) and napthyridine nitrogen (N2) forming 379

380 a six membered metallacycle (Figure 3). The napthyridine nitrogen N(3) is not involved in coordination. Similarly in complex 8 the metal is coordinated through two chloride's, Cp\* 381 moiety and PyNp ligand wherein it acted as a neutral monodentate ligand coordinating rhodium 382 through pyridine nitrogen (N1) (Figure 4 and 5). The distance between the metal to centroid of 383 the arene/Cp\* ring are {1.775 (3), 1.799 (4), 1.803 (5) and 1.773 (8) and 1.778 (9) Å}. The Metal 384 to nitrogen (M-N) bond distances in these complexes is found to be in the range of 2.11-2.17 Å 385 which are in close agreement for reported complexes with napthyridine ligands (Table 2) [10, 386 13]. It is to be noted that in complexes (4 and 5) the M-N bond distance {2.143(3) and 2.138(2), 387 Å} from aniline N(1) is comparatively shorter than the napthyridine N(2) nitrogen-metal 388 distances {2.164(3) and 2.179(3) Å}. The M-Cl bond lengths in these complexes shows no 389 significant differences and was found to be in the range of 2.405-2.447 Å which are comparable 390 with earlier reported complexes [24]. The bite angle values in in these napthyridine complexes 391 were observed in the range of 80.0-91.3° which are consistent with the piano stool arrangement 392 of various groups around the metal center (Table 2) [25, 26]. 393

Further from the crystal packing diagram it was observed that complex (4) formed a 394 dimeric unit via intermolecular van der Waal N-H·····Cl (2.647 Å) interaction between the 395 terminal chloride and amino hydrogen (Figure 6). Also in complex (5) two different types of 396 weak intermolecular van der Waal N-H·····Cl (2.548 Å) between chloride and amino group of 397 AnNp and C-H·····Cl (2.815 Å) interactions between chloride and napthyridine hydrogen was 398 observed (Figure 7). Interestingly the crystal packing in complex (8) formed a dimeric unit via 399 intermolecular van der Waal C-H·····Cl (2.723 and 2.809 Å) interaction between the chloride ion 400 and H-atom from napthyridine and pyridine of PyNp (Figure 8). These weak interactions play a 401 significant role in the formation of supramolecular motifs with interesting features. 402

#### 403 *Chemosensitivity studies*

The response of HCT-116 and MIA-PaCa-2 cells to the napthyridine ligands and the 404 respective metal complexes (1-9) are presented in Table 3. PHNp ligand was found to be 405 moderately active against both HCT-116 and MIA-PaCa-2 cell lines with IC<sub>50</sub> values of 406 39.90±1.60 and 62.48±6.28 µM respectively. In contrast, both AnNp and PyNp were found to be 407 inactive with IC<sub>50</sub> value > 100  $\mu$ M against both the cell lines. Upon complexation of napthyridine 408 ligands complexes (1-3) of PHNp ligand displayed less activity whereas PyNp complexes did not 409 410 display any cytotoxicity and iridium complex (6) with AnNp ligand possessed the highest activity against HCT-116 cells. Complexes (1-3) were found to be active against HCT-116 cell 411 line with IC<sub>50</sub> value in the range of  $37.79\pm4.90$  to  $82.09\pm3.55$  µM as compared to PHNp ligand 412 whose IC<sub>50</sub> value is  $39.90\pm1.60 \mu$ M. Against the Mia-PaCa-2 cell line, complex (2) was the most 413 active with an IC<sub>50</sub> of 25.13 $\pm$ 6.07  $\mu$ M. Complexes (3) and (6) were moderately active against the 414 MIA-PaCa-2 cell line with IC<sub>50</sub> values of 52.13±12.28 and 82.09±2.55 µM. Differences between 415 the response of HCT116 and Mia-PaCa-2 cells exist suggesting that these complexes are 416 targeting inherent biochemical differences between these cells as opposed to having general 417 cytotoxic properties. However, all the compounds were found to be less cytotoxic as compared to 418 cisplatin whose  $IC_{50}$  value is 2.78  $\mu$ M. In contrast to cisplatin however, none of the novel 419 compounds tested exhibited a cytotoxic effect on the non-cancer ARPE-19 cell line (IC<sub>50</sub> values 420 were > 100  $\mu$ M). The selectivity ratio for cisplatin (defined as the IC<sub>50</sub> of ARPE-19 divided by 421 the IC<sub>50</sub> of cancer cell lines) was 1.23 for HCT-116 cells whereas for complex (3), the selectivity 422 423 index was >2.64. Similar values for the selectivity index were obtained for other complexes tested suggesting that whilst potency is reduced compared to cisplatin, selectivity for cancer cells 424 in vitro under identical experimental conditions is enhanced. 425

#### 426 Conclusion

In this work, we report the coordination chemistry of 2-substituted-1,8-naphthyridine 427 ligands towards ruthenium, rhodium and iridium half-sandwich complexes. Complexes of PHNp 428 and PyNp were isolated as neutral complexes whereas AnNp complexes were isolated as ionic 429 salts with PF<sub>6</sub> counter anion. The molecular structures of neutral complexes revealed that PHNp 430 and PyNp ligands acted as monodentate ligand coordinating metal through napthyridine nitrogen 431 432 (N1) and pyridine nitrogen N(1). Whereas the X-ray structures of cationic complexes with AnNp ligand showed that it behaved as neutral chelating ligand coordinating metal atom in a bidentate 433 NN' fashion through aniline nitrogen N(1) and napthyridine nitrogen (N2) forming a six-434 435 membered chelated ring. Chemosensitivity activity of the complexes carried out against HCT-116 and MIA-PaCa-2cancer cell lines showed that PHNp and its complexes are cytotoxic; 436 however iridium-based AnNp complex possessed the highest activity among all other 437 438 napthyridine complexes. The work presented here exhibits interesting coordination modes of the 2-substituted-1,8-napthyridine ligands that have cytotoxic activity against cancer cells in vitro. 439 Whilst the potency of these compounds is less than cisplatin, there is evidence to suggest that 440 these complexes have greater selectivity for cancer as opposed to non-cancer cells in vitro than 441 cisplatin and therefore representing promising compounds for further development and 442 evaluation. 443

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448

#### 449 Supplementary material

450	CCDC 1568178 (4), 1568179 (5), 1568180 (8), 1568181 (PHNp), contains the
451	supplementary crystallographic data for this paper. These data can be obtained free of charge via
452	www.ccdc.cam.ac.uk/data_request/cif, by e-mailing data_request@ccdc.cam.ac.uk, or by
453	contacting The Cambridge Crystallographic Data Centre, 12, Union Road, Cambridge CB2 1EZ,
454	UK; Fax: +44 1223 336033.

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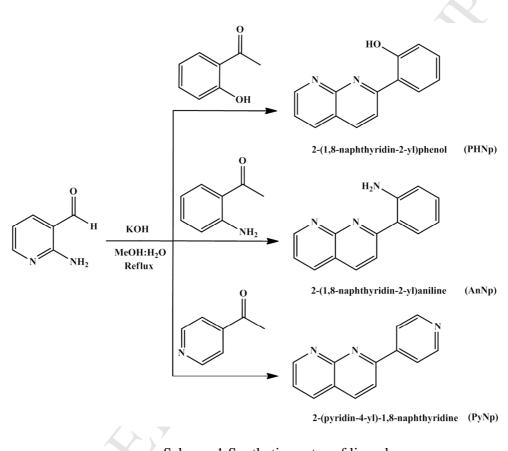
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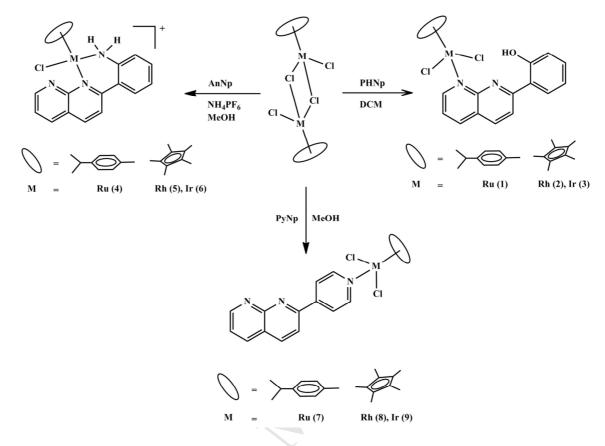
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Scheme 1 Synthetic routes of ligands



502 Scheme 2 Synthesis of neutral and cationic napthyridine metal complexes (1-9)

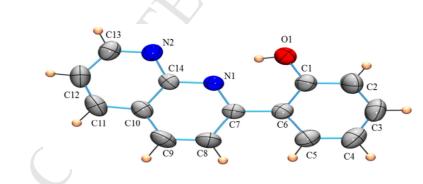
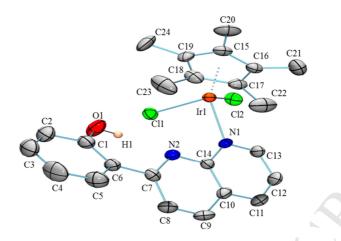


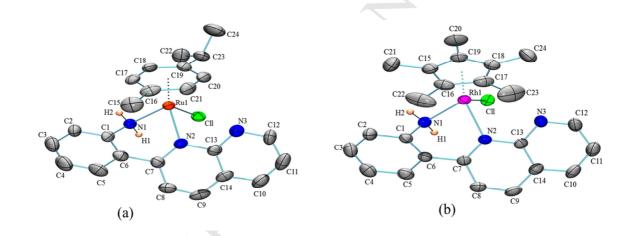
Figure 1 ORTEP plot of PHNp with 50% probability thermal ellipsoids.



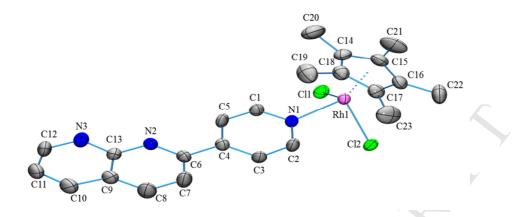
507 Figure 2 ORTEP plot of complex (3) with 50% probability thermal ellipsoids. Hydrogen atoms

508 (except on O1) are omitted for clarity.

509

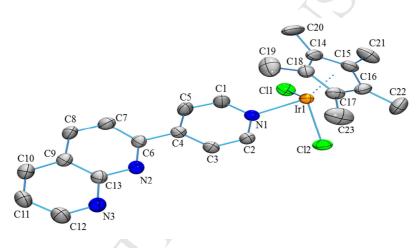


511 Figure 3 (a) ORTEP plot of complex (4) and (b) ORTEP plot of complex (5) with 50% 512 probability thermal ellipsoids. Counter anions, hydrogen atoms (except on N1) are omitted for 513 clarity.

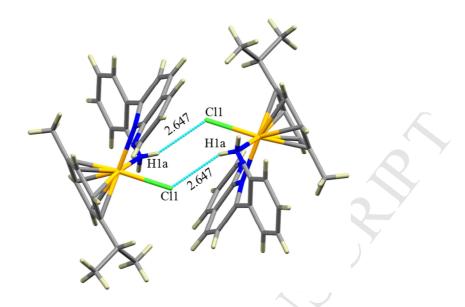


515 Figure 4 ORTEP plot of complex (8) with 50% probability thermal ellipsoids. Solvent molecules

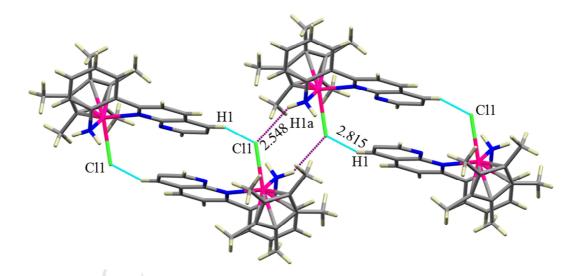
and hydrogen atoms are omitted for clarity.



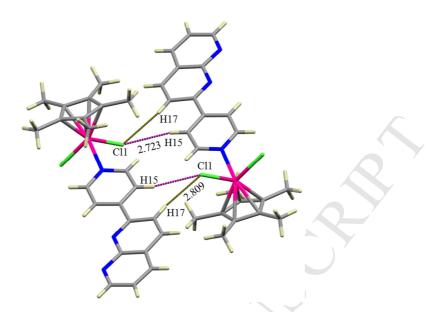
- 518 Figure 5 ORTEP plot of complex (9) with 50% probability thermal ellipsoids. Solvent molecule
- 519 and hydrogen atoms are omitted for clarity.



- 521 Figure 6 Packing diagram of complex (4) showing a dimeric unit formed via intermolecular van
- 522 der Waal N-H·····Cl (2.647 Å) interaction between the terminal chloride and amino hydrogen.



- 524 Figure 7 Packing diagram of complex (5) showing two different types of weak intermolecular
- 525 van der Waal N-H·····Cl and C-H·····Cl interactions.



527 Figure 8 Crystal packing diagram of complex (8) showing a dimeric unit formed via

528 intermolecular van der Waal C-H·····Cl interactions.

Complexes	PHNp	[3]	$[4] PF_6. CH_2Cl_2$	[ <b>5</b> ] PF <sub>6</sub>	[8] CHCl <sub>3</sub>	[ <b>9</b> ] CHCl <sub>3</sub>
Empirical formula	$C_{14}H_{10}N_2O$	$C_{24}H_{25}Cl_2N_2OIr$	$C_{25}H_{27}Cl_3F_6N_5PRu$	C24H25ClF6N3PRh	$C_{24}H_{25}Cl_5N_3Rh$	$C_{24}H_{25}Cl_5N_3Ir$
Formula weight	222.24	620.56	721.89	638.80	635.63	724.94
Temperature (K)	295(5)	293.6(4)	294(2)	293.6(2)	293.46(16)	295.8(5)
Wavelength (Å)	0.71073	0.71073	0.71073	0.71073	0.71073	0.71073
Crystal system	monoclinic	monoclinic	triclinic	triclinic	monoclinic	orthorhombic
Space group	$P2_1/n$	$P2_{l}/c$	PT	PT	$P2_{l}/c$	Pbca
a (Å)/α (°)	6.7873(9)/90	13.4948(7)/90	10.7040(6)/89.251	10.4457(6)/103.015(5)	14.6829(6)/90	14.5012(7)/90
b (Å)/β (°)	7.4846(8)/91.156(11)	7.9005(4)/100.679(6)	12.2053(7)/72.359(5)	11.3360(6)/109.254(5)	13.1266(5)/104.747	14.1704(9)/90
c (Å)/γ (°)	21.206(3)/90	21.1365(15)/90	13.0455(8)/64.432(5)	11.8077(6)/94.772(4)	14.4475(6)/90	26.0020(10)/90
Volume (Å <sup>3</sup> )	1077.1(2)	2214.5(2)	1451.22(15)	1267.08(13)	2692.84(19)	5343.1(5)
Z	4	4	2	2	4	11
Density (calc) (Mg/m <sup>-3</sup> )	1.371	1.861	1.652	1.674	1.568	1.802
Absorption coefficient $(\mu)$	0.089	0.832	0.931	0.906	1.148	5.516
(mm <sup>-1)</sup>				)		
F(000)	464	1208	724	642	1280	2816
Crystal size (mm <sup>3</sup> )	0.25 x 0.23 x 0.21	0.21 x 0.12 x 0.09	0.27 x 0.21 x 0.15	0.25 x 0.15 x 0.07	0.25 x 0.23 x 0.21	0.35 x 0.25 x 0.
Theta range for data collection	3.844 to 28.941°	3.240 to 29.075°	3.19 to 29.02°	3.332 to 29.115°	3.262 to 28.993°	3.5140 to 28.72
Index ranges	-9<=h<=5, -5<=k<=10, -	-18<=h<=15, -5<=k<=10,	-14<=h<=13, -	-14<=h<=14, -15<=k<=15,	-20<=h<=18, -	-19<=h<=17, -
	26<=l<=25	-28<=l<=27	16<=k<=16, -17<=l<=16	-40<=l<16	11<=k<=17, -19<=l<12	12<=k<=16, -
						32<=l<18
Reflections collected	3683	6905	10127	8777	11091	14307
Independent reflections	2370 [R(int) = 0.0209]	4465 [R(int) = 0.0265]	6534 [R(int) = 0.0445]	5707 [R(int) = 0.0267]	6178 [R(int) = 0.0309]	5588 [R(int) =
•						0.0317]
Completeness to theta = $25.00^{\circ}$	96.2 %	89.5 %	99.5 %	99.5 %	99.3 %	78.7 %
Absorption correction	Semi-empirical from	Semi-empirical from	Semi-empirical from	Semi-empirical from	Semi-empirical from	Semi-empirical
	equivalents	equivalents	equivalents	equivalents	equivalents	from equivalen
Max and min transmission	1.0000 and 0.67974	1.0000 and 0.36096	0.8730 and 0.7871	1.0000 and 0.69116	26.37 and 3.26	1.00000 and
					<b>D U U U</b>	0.31067
Refinement method	Full-matrix least-squares	Full-matrix least-squares	Full-matrix least-squares	Full-matrix least-squares	Full-matrix least-	Full-matrix leas
	on F <sup>2</sup>	on $F^2$	on F <sup>2</sup>	on F <sup>2</sup>	squares on F <sup>2</sup>	squares on F <sup>2</sup>
Data/restraints/parameters	2370/0/154	4465/0/270	6534/5/377	5707/0/325	6178/0/298	6178/0/298
Goodness-of-fit on F <sup>2</sup>	1.088	1.013	1.038	1.051	1.061	1.077
Final R indices [I>2sigma(I)]	R1 = 0.0566, wR2 =	R1 = 0.0346, wR2 =	R1 = 0.0552, $wR2 =$	R1 = 0.0361, wR2 =	R1 = 0.0505, wR2 =	R1 = 0.0402, w
	0.1361	0.0699	0.1360	0.0895	0.1047	= 0.0781
R indices (all data)	R1 = 0.0871, wR2 =	R1 = 0.524, wR2 =	R1 = 0.0691, wR2 =	R1 = 0.0423, wR2 =	R1 = 0.0664, wR2 =	R1 = 0.0588, w
	0.1615	0.0346	0.1469	0.0935	0.1127	= 0.0858
Largest diff. peak and hole	0.193 and -0.148	1.319 and -0.929	0.879 and -0.698	0.650 and -0.539	0.843 and -0.861	1.518 and -1.00
(e.Å <sup>-3</sup> )						
CCDC No.	1568181		1568178	1568179	1568180	

## 530 Table1 Crystal data and structure refinement parameters of complexes

Complex	3	4	5	8	9
M(1)-CNT	1.775	1.799	1.803	1.773	1.778
M(1)-N(1)	2.154(4)	2.143(3)	2.138(2)	2.128(3)	2.117(5)
M(1)-N(2)		2.164(3)	2.179(3)	2.427(1)	
M(1)-Cl(1)	2.405(1)	2.4165(11)	2.4478(7)	2.405(1)	2.409(2)
M(1)-Cl(2)	2.426(2)				2.404(1)
N(1)-M(1)-N(2)		82.29(14)	80.12(8)		
N(1)-M(1)-Cl(1)	87.7(1)	82.67(10)	82.90(6)	87.27(8)	87.6(1)
N(2)-M(1)-Cl(1)		83.55(10)	91.00(6)		
N(1)-M(1)-Cl(2)	89.0(1)			89.76(8)	85.9(1)
Cl(1)-M(1)-Cl(2)	85.13(5)			91.25(4)	89.46(5)

532 Table 2 Selected bond lengths (Å) and bond angles (°) of complexes

533 CNT represents the centroid of the arene/Cp\* ring and M = Ru, Rh and Ir.

Table 3 IC<sub>50</sub> values of napthyridine ligands and complexes (1-9) along with cisplatin against

535 HCT-116 and MIA-PaCa-2 cancer cell line and non-cancer ARPE-19 cell line. Each value

represents the mean  $\pm$  standard deviation from three independent experiments.

Compounds	IC <sub>50</sub> (µM)				
	HCT-116	MIA-PaCa-2	ARPE-19		
PHNp	39.90±1.64	62.48±6.28	>100		
AnNp	>100	>100	>100		
PyNp	>100	>100	>100		
Complex 1	82.09±3.54	>100	>100		
Complex 2	61.28±13.8	25.14±6.07	>100		
Complex <b>3</b>	37.79±4.89	52.13±12.28	>100		
Complex 4	>100	>100	>100		
Complex 5	>100	>100	>100		
Complex 6	$31.33 \pm 12.92$	82.09±2.55	>100		
Complex 7	>100	>100	>100		
Complex 8	>100	>100	>100		
Complex 9	>100	>100	>100		
Cisplatin	$2.78 \pm 1.40$	$3.15\pm0.10$	3.43±0.48		

537  $IC_{50}$  = concentration of the drug required to inhibit the growth of 50% of the cancer cells ( $\mu$ M).

## Highlights

- 2-substituted-napthyridine derivatives are synthesized by Friedländer condensation.
- ✤ Interesting bonding modes of napthyridine ligands were observed.
- ✤ Complexes were cytotoxic against cancer cell lines.