

Supporting Information

Directing single-molecule emission with DNA origami-assembled optical antennas

Kristina Hübner¹, Mauricio Pilo-Pais^{2,5}, Florian Selbach¹, Tim Liedl², Philip Tinnefeld^{1},
Fernando D. Stefani^{3,4*} and Guillermo P. Acuna^{5*}*

¹Department of Chemistry and Center for NanoScience, Ludwig-Maximilians-Universität München, Butenandtstr. 5-13 Haus E, 81377 München, Germany

²Faculty of Physics and Center for NanoScience, Ludwig-Maximilians-Universität München, Geschwister-Scholl-Platz 1, 80539 München, Germany

³Centro de Investigaciones en Bionanociencias (CIBION), Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), Godoy Cruz 2390, C1425FQD Ciudad Autónoma de Buenos Aires, Argentina

⁴Departamento de Física, Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires, Güiraldes 2620, C1428EHA Ciudad Autónoma de Buenos Aires, Argentina

⁵Department of Physics, University of Fribourg, Chemin du Musée 3, Fribourg CH-1700, Switzerland

1. Gold Nanoparticle Fabrication

Synthesis of gold nanoparticles (Au NPs): 60 nm spherical gold NPs were synthesized by a four-step seed growth and etching method^{1,2}. Small gold clusters were prepared by reduction of Au³⁺ ions with sodium borohydride in cetyltrimethylammonium bromide (CTAB) solution. These clusters were grown to small seed nanoparticles by further addition of Au³⁺ ions and ascorbic acid in CTAB solution. Further larger polyhedron nanoparticles were formed by anisotropic growth of seed particles in cetyltrimethylammonium chloride (CTAC) solution with Au³⁺ ions and ascorbic acid. Spherical NPs were obtained by etching the surface of nanopolyhedrons by addition of Au³⁺ ions in CTAB solution. The experimental details of the individual steps are specified below.

Clusters: HAuCl₄ solution (10 mM, 250 μL) was mixed with a CTAB solution (100 mM, 9.75 mL), followed by rapid injection of a freshly prepared ice-cold NaBH₄ solution (10 mM, 600 μL). The mixture was left undisturbed for 3 h at 30 °C.

Seeds: The prepared cluster solution (300 μL) was injected into a growth solution of CTAB (100 mM, 2.44 mL), deionized water (D.I. water; 47.5 mL), HAuCl₄ solution (10 mM, 1 mL), and ascorbic acid solution (100 mM, 3.75 mL). The reaction mixture was gently shaken and then left undisturbed for 3 h at 30 °C. The colloid was washed by centrifugation (11500 × g, 40 min) and redispersed in D.I. water (12.5 mL).

Nanopolyhedrons: The seed solution (3.6 mL) was added into a CTAC solution (25 mM, 300 mL). After the sequential addition of ascorbic acid solution (100 mM, 7.5 mL) and HAuCl₄ solution (10 mM, 15 mL), the mixture solution was kept undisturbed for 3 h at 30 °C and then washed by centrifugation (1750 × g, 30 min) and redispersion in CTAB solution (20 mM, 5 mL).

Au NPs: The nanopolyhedron solution (5 mL) was diluted by the addition of CTAB solution (20 mM, 250 mL), followed by the addition of HAuCl₄ solution (10 mM, 1.9 mL). After stirring (550 rpm, 2 h, 40 °C), the mixture was washed twice with D.I. water (centrifugation at 1750 × g for 30 min) and redispersed finally in D.I. water (10 mL).

2. Sample Preparation

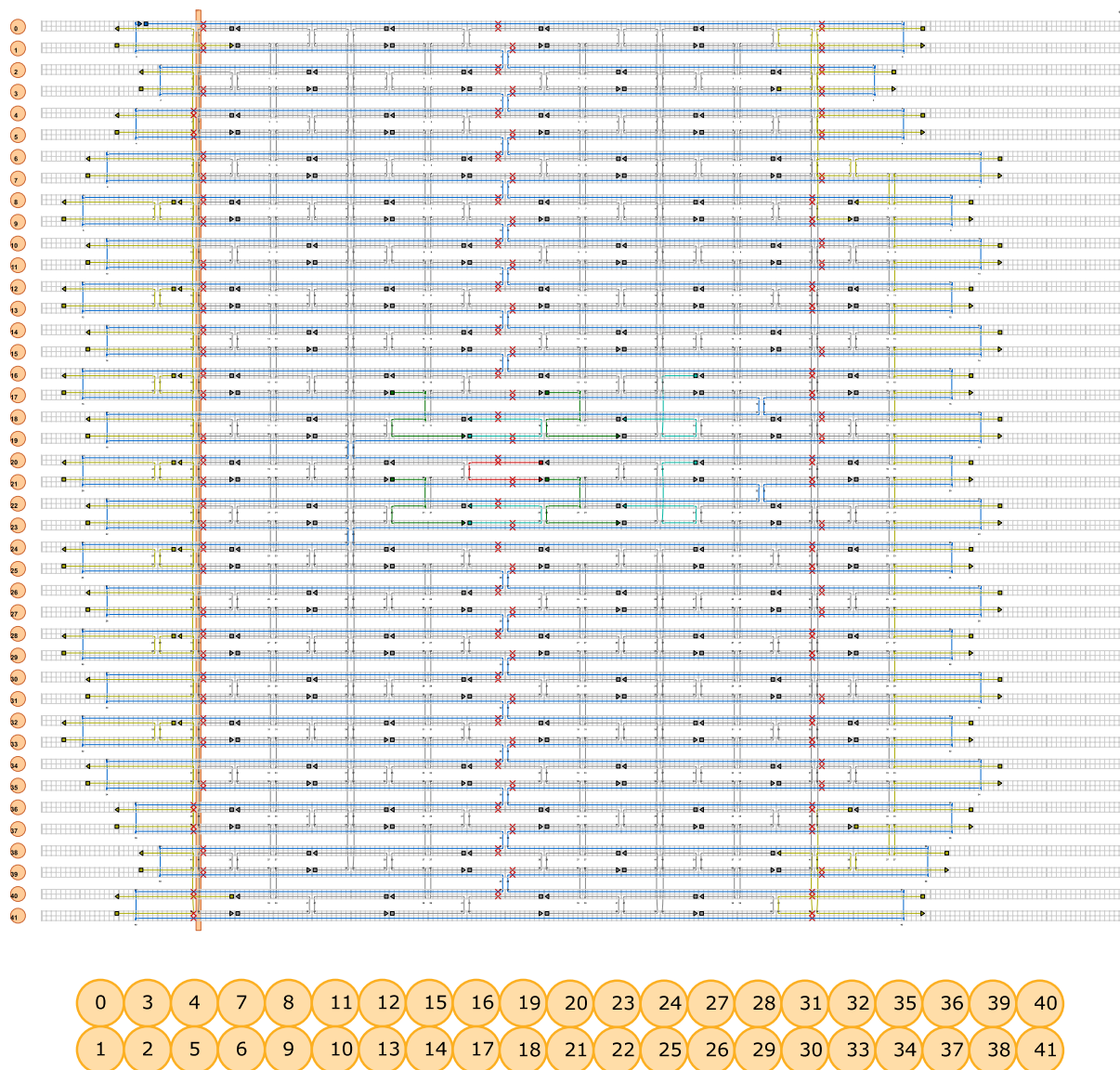


Figure S 1: 2LS DNA origami design. Scaffold (blue), staples (gray), Cy5 modified staples (red), handles (green and turquoise), and endcaps (yellow).

The DNA origami structure was designed using CaDNAo³ and visualized for twist correction using CanDo⁴. It consists of a square lattice design that folds onto a 2-layers sheet (2LS) of size ~60 nm×50 nm×5 nm. An attachment binding site with four 15-nuclotides long handles (15xA) is located at the center of each of the two layers of the 2LS origami. Each binding site is aimed to attach a single 60 nm Au NP. The CaDNAo design is shown in Figure S1, with the scaffold portrayed in blue, the core staples in gray, the handles on each layer in green and turquoise, the staple with Cy5 in red, and the endcaps in yellow. The endcaps contain CCCC extensions to prevent base-stacking interactions between origamis.

Handles	5' staple AAAAAAAAAAAAAAAAA 3'
Cy5	5' staple TTT Cy5 3'
Endcaps	5' CCCC staple CCCC 3' or 5' CCCC staple 3' or 5' staple CCCC 3'
Thiol-DNA	5' Thiol C6 TTTTTTTTTTTTTTTTTTTT 3'

Table S 1: *Modified DNA strands. Handles contain a 15xA extension, the Cy5 staple has a 3-T spacer, and the endcaps 4xC extension(s). Thiolated DNA has a 15xT which is complementary to the handles sequence.*

The 2LS DNA origami structure was folded using a p7249 scaffold (single-stranded M13mp18 bacteriophage genome, in-house produced), staple strands (Eurofins Genomics, Germany), and modified strands (Biomers.net GmbH, Germany and Eurofins Genomics, Germany) in a 1xTAE buffer (40 mM Tris, 40 mM acetic acid, 1 mM EDTA, pH 8) and 12 mM MgCl₂ using a 1:10 scaffold:staples ratio (10 nM scaffold final concentration). The solution was heated to 75 °C, held for 5 minutes, brought to 65 °C, and ramped down to 25 °C at a rate of 1 °C/ 20 mins. The folded DNA origami structure was purified from excess staple strands using gel electrophoresis. All gels were ran using a 1% agarose gel, 1xTAE buffer with 11 mM MgCl₂ for 2.5 hours at 4 V/ cm. The appropriate 2LS origami band was cut out and squeezed from the gel using cover slips wrapped in

parafilm. Only side lanes of the gel were post-stained and used as markers to cut the appropriate band to avoid staining the origami used on this study. The concentration was determined via UV-Vis absorption spectroscopy (Nanodrop).

Thiolated DNA (Thiol-C6-15xT, Biomers.net GmbH, Germany) was mixed with ultrasmooth Au NPs (see section 1), adjusted to 0.03% SDS and frozen for 30 mins⁵. Excess DNA was removed using gel electrophoresis. This step also ensures the removal of any self-aggregated dimer formed during the NP functionalization. The concentration was determined via UV-Vis absorption spectroscopy (Nanodrop).

The purified 2LS origami was mixed with the purified Au NPs using an excess of five Au NPs per binding site and adding NaCl to a final concentration of 500 mM. After overnight incubation, the excess of NPs was removed by gel electrophoresis and the band containing correctly formed dimers was extracted as described before.

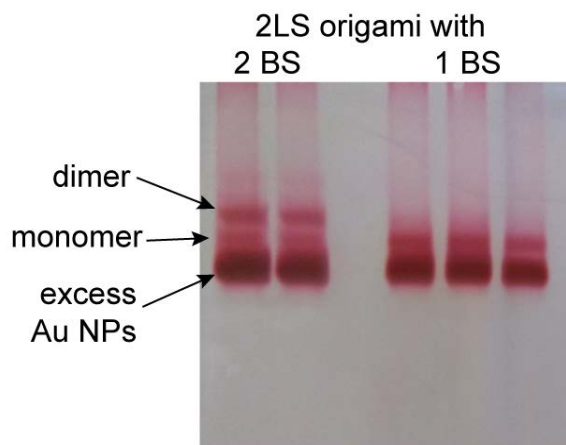


Figure S 2: Gel image after gel electrophoresis of the designed 2LS origami with two binding sites (2 BS) and one binding side (1 BS).

3. Surface Preparation and Sample Immobilization

For immobilization of the OA structures, the glass cover slips were first rinsed with water and then cleaned in a UV cleaning system (PSD Pro System, Novascan Technologies, USA). Afterwards, the surfaces were passivated with a BSA-biotin in PBS solution (1 mg/mL) for 1 h, neutrAvidin in PBS solution (0.5 mg/mL) for 20 min and single stranded DNA (7T for the reference and 7A for the monomer and dimer samples) with a biotin attached to the 5'-end (10 nM) for 20 min. In between all the steps, the surfaces were washed with 0.5xTE buffer containing 10 mM MgCl₂. The samples were incubated on the surface for 5 min and afterwards washed with buffer. Confocal and wide-field measurements were performed under oxygen removal and ROXS using trolox/troloxquinone in 1xTAE with 12 mM MgCl₂.^{6,7}

4. Imaging Systems

4.1 Wide-Field Setup

Wide-field measurements were performed on a custom-built wide-field setup based on an inverted Olympus IX71 microscope. As excitation source a red diode laser with a wavelength of 644 nm is used (ibeam smart, Toptica Photonics, Germany). The laser gets spectrally cleaned by a cleanup filter (Brightline HC 650/13, Semrock, USA) before passing the lambda quarter (AQWP05M 400-800 nm, Thorlabs, Germany)/ lambda half waveplate (AHWP05M 400-800 nm, Thorlabs, Germany) and afterwards being focused in the back focal plane of the objective (UApo N 60x, NA = 1.49, WD = 0.1 mm, Olympus, Germany). For stabilization of the sample an actively stabilized optical table (TS-300, JRS Scientific Instruments, Switzerland) and a nosepiece stage (IX2-NPS, Olympus, Germany) were implemented to the microscope. Then the light is directed

through the objective and the sample is illuminated. Fluorescence will get collected by the same objective and separated from the excitation light by a dichroic beamsplitter (Dual Line zt532/640 rpc, AHF Analysentechnik, Germany). Afterwards the fluorescence light is filtered with an emission filter (ET 700/75, Chroma, USA) and focused by a lens on an EMCCD camera (iXon X3 DU-897, Andor, North Ireland). The videos are recorded by the open source microscopy ImageJ software Micro-Manager⁸.

4.2 Confocal Setup

Confocal measurements were performed on a custom-build confocal setup based on an inverted Olympus IX81 microscope. The excitation source is a 78 MHz-pulsed laser (SuperK Extreme, NKT Photonics, Denmark), that can be tuned in a region from 400 to 2400 nm. In these experiments working at 639 nm to excite the Cy5 dye. For wavelength selection an AODS (20160 8R, Crystal Technology, Inc., USA) and an AOTF (AA.AOTF.ns:TN, AA-Opto-Electronic, France) are implemented that also clean up the spectrum of the laser. After wavelength selection a neutral density filter (ndF, OD 0-2, Thorlabs, Germany) is passed and the laser beam is coupled into a polarization maintaining single mode fiber (PM-Fiber, P1-488PM-FC-2, Thorlabs, Germany). The polarization of the laser can be controlled by the combination of a linear polarizer (LPVISE100-A, Thorlabs, Germany), an electro optical modulator (EOM, electro-optical modulator, L 0202, Qioptiq, Germany) and a lambda quarter waveplate (AQWP05M-600, Thorlabs, Germany). With these optics a circular polarization can be achieved by driving the EOM with a high frequency. Afterwards, the laser beam is coupled into the microscope body and focused on the sample placed on top of an immersion oil objective (UPlanSApo 100x, NA = 1.4, WD = 0.12 mm, Olympus, Germany). The fluorescence of the sample is collected by the same objective and separated from the excitation light by a dichroic mirror (DS, zt532/640rpc, Chroma,

USA). To filter out scattered laser light, the fluorescence is focused on a 50 μm pinhole (Linios, Germany) and afterwards directed through an emission filter (RazorEdge® 647, Semrock, USA) and again focused by a lens on the the APD (Avalanche Photo Diode, SPCM, AQR 14, Perkin Elmer, USA). Time Correlated Single Photon counting is achieved by an TCSPC system (Hydra Harp 400, PicoQuant, Germany). Scanning of the samples in x and y is realized by a piezo stage (P-517.3CL, E-501.00, Physik Instrumente GmbH&Co. KG, Germany). For data processing, a custom written LabVIEW software (National Instruments, USA) is used. To extract the fluorescence lifetime from the decays, a mono-exponential fit and a deconvolution from the instrumental response function using the program FluoFit (PicoQuant, Germany) were used.

5. Imaging

5.1 Wide-Field Imaging Series

To analyze the emission dipole orientation, the orientation of maximum absorption and the bleaching behavior, a set of different measurements were performed. In a first measurement, the laser power was set to a power density of 0.22 kW cm^{-2} and the objective was shifted towards the sample by approximately $1 \mu\text{m}$ to take defocused images of the single molecules. This shift in the objective position leads to a deliberate defocused signal on the camera chip that allows us to study the angular emission pattern. Circularly polarized light was used in order to attain uniform excitation of all OA-Cy5 photon emitter structures that are randomly oriented on the glass. Second, after completion of the defocused imaging, the sample is placed back to focus for single molecule studies. These measurements are performed while stepwise rotating the linearly polarized excitation light by spinning the lambda half waveplate for ten degrees nine times every five

seconds. Third, the incident polarization is switched back to circular and the laser power density is increased to 3.04 kW cm^{-2} in order to bleach in a reduced amount of time all the fluorophores present in the imaged field of view. This final step allows us to filter out the signals arising from aggregates of NPs or DNA origami structures and to only consider structures with a single bleaching step, a signature of the presence of a single molecule.

5.2 Confocal Measurements

In order to study the yield of dimer structures, we performed confocal measurements. We used the fluorescence lifetime of the different samples to distinguish dimer from monomer and reference structures. Samples were first scanned with a circularly polarized laser beam and afterwards spots were picked and measured until a single bleaching step occurred.

6. Simulations

Numerical simulations were performed using a Finite Difference Frequency Domain (FDFD) commercial software (CST STUDIO SUITE, Microwave module). The Cy5 fluorophores were modelled by a small (0.1 nm) current source oscillating at 447 THz, corresponding to the wavelength of maximum fluorescence emission of Cy5. The emission pattern generated by the dipolar current source was calculated for the experimental geometry: a glass ($n = 1.515$) – water ($n = 1.33$) interface, the current source (fluorophore) was placed on the water side at 40 nm from the glass-water interface, oriented parallel or perpendicular to the interface. The simulated images were calculated by plotting the electric field generated on each position of the selected image plane $\mathbf{r} = (\rho \cos \varphi; \rho \sin \varphi; z)$, integrating the fraction of the emission pattern collected by the objective lens:

$$E_j(\mathbf{r}) = C \int_0^{2\pi} \int_{\theta_{min}}^{\theta_{max}} \sqrt{\cos \theta} E_{0j}(\theta, \psi) e^{ik(\rho \sin \theta (\cos(\psi-\varphi)) + z \cos \theta)} \sin \theta d\theta d\psi$$

where j stands for x , y , or z (Cartesian components of the field). The solid angle is integrated in spherical coordinates. The azimuth angle is integrated all around the z -axis, and the focusing angle range (θ_{min} and θ_{max}) depends on the NA of the objective. $k = 2\pi/\lambda$. $E_{0j}(\theta, \psi)$ is the j component of the emission pattern field parametrized in the azimuthal and polar angles (θ, ψ).

For the quantum yield simulations we followed the procedure included in ⁹.

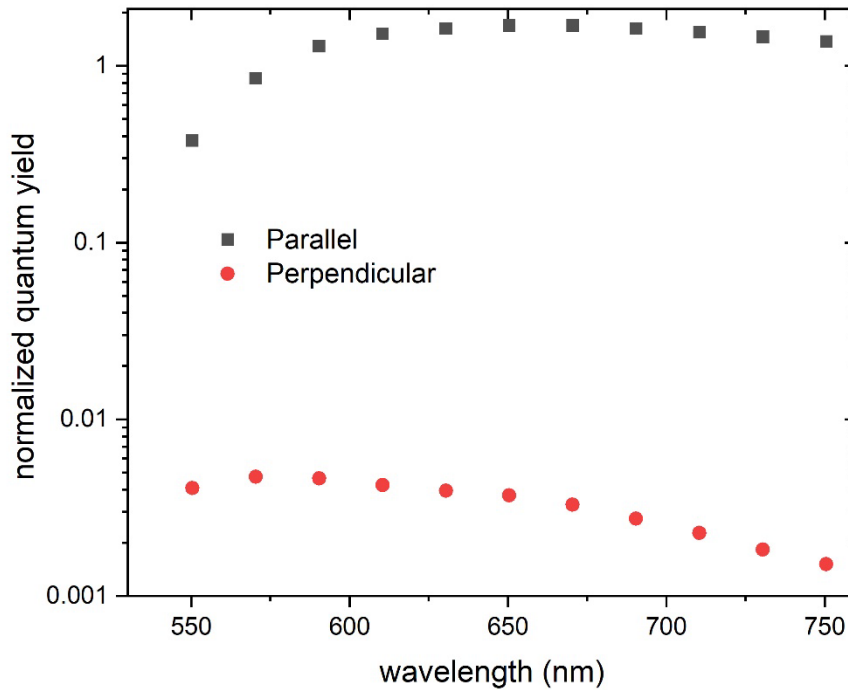


Figure S 3: Simulated quantum yield for the Cy5 fluorophore located at the hotspot of the OA normalized to the quantum yield without the OA (0.27) for two orientations, along the dimer main axis (parallel) and perpendicular.

7. Data Analysis

For extraction of the fluorescence enhancement, modulation and angle of maximum excitation the modulating time traces of single molecules are processed and fitted (Figure S 4). Therefore, a mean

fluorescence enhancement value for each excitation polarization is extracted. The extracted values are then fitted with a cosine square function.

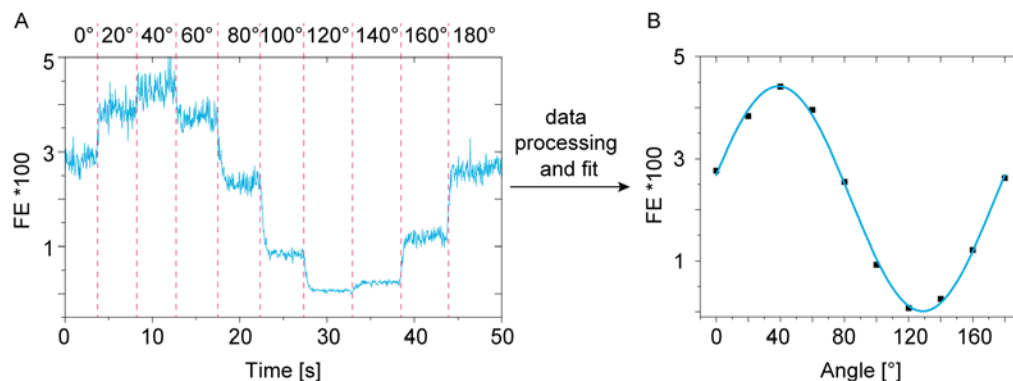


Figure S 4: Trace from the polarization resolved excitation measurements of a single Cy5 molecule (A). (B) Extracted mean intensity for all polarizations fitted by a cosine square function.

By this fitting, the maximum fluorescence enhancement that corresponds to the angle of maximum excitation can be extracted.

References

- (1) Darvishzadeh-Varcheie, M.; Guclu, C.; Ragan, R.; Boyraz, O.; Capolino, F. Electric Field Enhancement with Plasmonic Colloidal Nanoantennas Excited by a Silicon Nitride Waveguide. *Opt. Express* **2016**.
- (2) Yoon, J. H.; Selbach, F.; Langolf, L.; Schlücker, S. Ideal Dimers of Gold Nanospheres for Precision Plasmonics: Synthesis and Characterization at the Single-Particle Level for Identification of Higher Order Modes. *Small* **2018**, *14* (4), 1702754.
- (3) Douglas, S. M.; Marblestone, A. H.; Teerapittayanon, S.; Vazquez, A.; Church, G. M.; Shih, W. M. Rapid Prototyping of 3D DNA-Origami Shapes with CaDNAno. *Nucleic Acids Res.* **2009**.
- (4) Kim, D.-N.; Kilchherr, F.; Dietz, H.; Bathe, M. Quantitative Prediction of 3D Solution Shape and Flexibility of Nucleic Acid Nanostructures. *Nucleic Acids Res.* **2012**.
- (5) Liu, B.; Liu, J. Freezing Directed Construction of Bio/Nano Interfaces: Reagentless Conjugation, Denser Spherical Nucleic Acids, and Better Nanoflakes. *J. Am. Chem. Soc.* **2017**, *139* (28), 9471–9474.
- (6) Vogelsang, J.; Kasper, R.; Steinhauer, C.; Person, B.; Heilemann, M.; Sauer, M.; Tinnefeld, P. A Reducing and Oxidizing System Minimizes Photobleaching and Blinking of Fluorescent Dyes. *Angew. Chemie - Int. Ed.* **2008**.
- (7) Cordes, T.; Vogelsang, J.; Tinnefeld, P. On the Mechanism of Trolox as Antiblinking and Antibleaching Reagent. *J. Am. Chem. Soc.* **2009**.
- (8) Edelstein, A.; Amodaj, N.; Hoover, K.; Vale, R.; Stuurman, N. Computer Control of

- Microscopes Using Manager. *Current Protocols in Molecular Biology*. 2010.
- (9) Acuna, G. P.; Möller, F. M.; Holzmeister, P.; Beater, S.; Lalkens, B.; Tinnefeld, P. Fluorescence Enhancement at Docking Sites of DNA-Directed Self-Assembled Nanoantennas. *Science* (80-.). **2012**, 338 (6106), 506–510.