

# **UNIVERSITI PUTRA MALAYSIA**

UNIVERSITI PUTRA MALAYSIA<br>
MOLECULAR INSIGHTS ON THE EFFECTS OF ANIONS TOWARDS<br>
HYDROLASES IN [BMIM]-BASED IONIC LIQUIDS<br>
MUHAMMAD ALIF BIN MOHAMMAD LATIF<br>
FS 2014 68 *MOLECULAR INSIGHTS ON THE EFFECTS OF ANIONS TOWARDS HYDROLASES IN [BMIM]-BASED IONIC LIQUIDS* 

**MUHAMMAD ALIF BIN MOHAMMAD LATIF**

**FS 2014 68**



## **MOLECULAR INSIGHTS ON THE EFFECTS OF ANIONS TOWARDS HYDROLASES IN [BMIM]-BASED IONIC LIQUIDS**



By

# **MUHAMMAD ALIF BIN MOHAMMAD LATIF**

**Thesis Submitted to School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

**August 2014**

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of any material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

## **MOLECULAR INSIGHTS ON THE EFFECTS OF ANIONS TOWARDS HYDROLASES IN [BMIM]-BASED IONIC LIQUIDS**

By

## **MUHAMMAD ALIF BIN MOHAMMAD LATIF**

#### **August 2014**

#### **Chair: Professor Mohd Basyaruddin Abdul Rahman, PhD**

**Faculty: Science**

ITYDROLLASES IN [BMIM]-BASED IONIC LIQUIDS<br>
By<br>
MUHAMMAD ALIF BIN MOHAMMAD LATIF<br>
August 2014<br>
August 2014<br>
Chair: Professor Mohd Basyaruddin Abdul Rahman, PhD<br>
Faculty: Seicne<br>
Faculty: Seicne<br>
Chair: Professor Mohd Basy The use of non-aqueous solvents in biocatalysis has shown improvements in enzyme performance. A new type of non-aqueous solvent has attracted a lot of interests in biocatalysis, called the Room Temperature Ionic Liquid (RTIL). A lot of biocatalysis experiments have showed that RTILs can further increase the reaction rates and yields when used instead of conventional organic solvents. However, since there are many RTIL combinations available, selecting a specific RTIL for use in biocatalysis have proven to be quite challenging. A detailed understanding on the effects that different RTIL combination imposed on enzymes is therefore important. Here, the behavior of enzymes in RTILs was characterized by their effects when different RTIL anions were used. A series of molecular-level investigations were conducted using molecular dynamics (MD) and stochastic dynamics (SD) simulations in order to gain more information on the structural and dynamics properties of enzymes in RTILs. Four hydrolases, consisted of α-Chymotrypsin, thermolysin, *Candida Antarctica* Lipase B (CALB) and *Candida rugosa* Lipase (CRL) were studied. These hydrolases were solvated in aqueous and five, 1-butyl-3 methylimidazolium ([BMIM])-based RTILs with different anions such as hexafluorophosphate  $(\text{IPF}_6)$ . tetrafluoroborate  $(IBF<sub>4</sub>)$ <sup>-</sup>). chloride  $(ICII^{\dagger})$ . trifluoromethanesulfonate ([TfO] and bis-trifluoromethylsulfonylimide ( $[Tf_2N]$ ). The effects of water molecules in the systems were studied at 5%, 10%, 15%, 20% and 50% of water, based on the weight/weight percentages of the protein mass (w/w protein). All RTIL solvent models produced a liquid ordering at room temperature and an average density that was close to experimental data with a percentage error of below than 5%.

ditions at 125% modelle was lender of  $\approx 1$  in Fig. 1 and the measurement of the measur The structural stability of all hydrolases studied showed a dependency towards the water content, in which the minimum atomic displacements were observed around 10 to 20% of water. Around this water percentage region, [TfO]- anion rendered the most stable conformation for  $\alpha$ -Chymotrypsin, CALB and CRL. The smallest [Cl] $\bar{C}$ anion was found to produce the least stable conformations compared to other RTILs studied. In the case of thermolysin, the order of structural stability between the RTIL anions at 15% of water was  $[PF_6] > [TfQ] \sim [Tf_2N] > [Cl] \sim [BF_4]$  which was different from other hydrolases studied. Further investigations revealed that in  $[BMIM][PF_6]$ , thermolysin showed better structural stability than in aqueous, even when simulated at 90 °C. The effect of changing the RTIL anions towards the enzyme flexibility was only clearly visible at higher water content (20% and 50%  $w/w$  protein), especially for [PF<sub>6</sub>] and [Tf<sub>2</sub>N] anions. The analysis on local flexibility showed that only the surface of the protein was affected. For the lipases, the local flexibility was found significantly reduced in certain regions which were highly flexible in aqueous solution, particularly for the lid of the CRL. MD simulations revealed a structured ordering of RTIL anions around the enzymes while the water molecules were found localized at certain region of the protein surface. Hydrophobic anions such as  $[PF_6]$  covered more areas and were more organized at low water content while [Cl]- anion behave otherwise. Meanwhile, a number of water molecules were stripped off from the surface of  $\alpha$ -Chymotrypsin, CALB and CRL. RTILs with  $[PF_6]$  and  $[TfO]$  anions retained more water on the surface as compared to  $[BF_4]$ and [Cl]<sup>-</sup> anions, consistently for the three hydrolases. [Tf<sub>2</sub>N]<sup>-</sup> anion was found stripping the most number of water for the case ofα-Chymotrypsin and CALB while the least was found for CRL.

The solvation thermodynamics of amino acid side chain analogues in water and five [BMIM]-based RTILs was investigated using SD simulations. The solvation free energy was calculated using Bennett's Acceptance Ratio method. Results from the simulations in water were in agreement with published experimental and simulation data. RTILs showed better solvation capabilities when compared with water. Nonpolar analogues produced lower solvation free energy in hydrophobic anions such as  $[PF_6]$  and  $[Tf_2N]$  while the polar ones showed better solvation in hydrophilic anions such as [BF<sub>4</sub>], [Cl] and [TfO]. The solvation properties in [BMIM][Cl] also explained why the enzymes experienced more conformational distortions in this RTIL at low water content. Overall, computer simulations were able to explain several effects of RTIL anions on the structure and dynamics of enzymes at molecular level. The structural stability and flexibility of the enzymes were found affected by the water content, more than the types of the RTIL anions studied. MD simulation results were correlated with experimental reports. It was found that the behavior of anions and water at the protein surface played a major role towards the properties of enzymes in RTILs. The results also suggested that the surface properties of the biocatalyst and the physicochemical properties of the substrate should be taken into consideration when choosing a particular RTIL as the solvent system.

Abstrak tesis yang dikemukakan kepada Senat of Universiti Putra Malaysia Sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

## **PENDEKATAN MOLEKULAR BERKENAAN KESAN ANION TERHADAP HIDROLASE DI DALAM CECAIR IONIK BERASASKAN [BMIM]**

Oleh

## **MUHAMMAD ALIF BIN MOHAMMAD LATIF**

#### **Ogos 2014**

#### **Pengerusi: Profesor Mohd Basyaruddin Abdul Rahman, PhD**

**Faculti: Sains** 

ITIDROLLASE DI DALAM CECATIK IONIK BERASASKAN [BVIIST]<br>
(C)ch<br>
MUHAMMAD ALIF BIN MOHAMMAD LATIF<br>
(Ogss 2014)<br>
(C)ch<br>
MUHAMMAD ALIF BIN MOHAMMAD LATIF<br>
(Ogss 2014)<br>
Pengeunsia Francisco Trofteor Mohd Basyarruddin Abdul Rahm Penggunaan larutan bukan akues di dalam biokatalisis telah menunjukkan peningkatan prestasi enzim. Sejenis larutan bukan akues telah berjaya menarik perhatian di dalam bidang biokatalisis, bernama Cecair Ionik Suhu Bilik (RTIL). Terdapat banyak laporan eksperimen yang menunjukkan bahawa RTIL mampu menambah lagi kadar reaksi dan hasil apabila digunakan sebagai pelarut bukan akues menggantikan organik konvensional. Walaubagaimanapun, terdapat banyak kombinasi RTIL yang boleh digunakan, maka untuk memilih RTIL yang spesifik untuk biokatalisis adalah agak sukar. Pemahaman yang terperinci tentang kesan yang berbeza apabila RTIL yang berbeza digunakan ke atas enzim adalh amat penting. Di sini, perilaku enzim di dalam RTIL dicirikan oleh kesan mereka apabila menggunakan RTIL anion yang berbeza. Satu siri siasatan di tahap molekul telah dijalankan menggunakan simulasi dinamik molekul (MD) dan dinamik stokastik (SD) untuk mendapatkan maklumat lanjut mengenai sifat-sifat dinamik dan struktur enzim di dalam RTIL. Empat hidrolase, terdiri daripada α-Chymotrypsin, thermolysin, *Candida antartica* Lipase B (CALB) dan *Candida rugosa* Lipase (CRL) telah dikaji. Hidrolase-hidrolase ini telah dilarutkan di dalam akues dan di dalam lima RTILs berasaskan kation 1-butil-3-methylimidazolium ([BMIM]) dengan anion yang berbeza seperti hexafluorofosfat ([PF<sub>6</sub>] ), tetrafluoroborat ([BF<sub>4</sub>] ), klorida ([Cl] ), trifluorometanaesulfonat ([TfO] dan bis-trifluorometilsulfonilimida ([Tf2N]). Kesan molekul air di dalam setiap sistem dikaji pada 5%, 10%, 15%, 20% dan 50% air, berdasarkan peratusan berat/berat jisim protein (w/w protein). Kesemua model larutan RTIL menghasilkan aturan cecair pada suhu bilik dan kepadatan purata yang dekat dengan data eksperimen dengan ralat peratusan kurang daripada 5%. Kestabilan struktur semua hidrolase yang dikaji menunjukkan pergantungan kearah kandungan air, dan anjakan atom minimum diperhatikan pada kira-kira 10 hingga 20% air.

Di rantau peratusan air ini, anion [TfO]-memberikan bentuk yang paling stabil untuk α-Chymotrypsin, CALB dan CRL. Anion yang paling kecil [Cl]- didapati menghasilkan bentuk yang paling kurang stabil berbanding dengan RTIL lain yang dikaji. Di dalam kes thermolysin, susunan kestabilan struktur antara anion RTIL pada 15% air adalah  $[PF_6]$  >  $[TfO]$  ~  $[Tf_2N]$  >  $[C1]$  ~  $[BF_4]$  yang mana, berbeza dari hidrolases lain yang dikaji. Siasatan lanjut mendedahkan bahawa dalam [BMIM][PF6], thermolysin menunjukkan kestabilan struktur yang lebih baik daripada di dalam akueus, walaupun simulasi dijalankan pada suhu 90 °C. Kesan daripada mengubah anion RTIL terhadap fleksibiliti enzim hanya jelas kelihatan pada kandungan air yang tinggi (20% dan 50% w/w protein), terutamanya untuk anion [PF6] - dan [Tf2N]- . Analisa ke atas fleksibiliti setempat menunjukkan hanya permukaan protein yang terjejas. Bagi lipase, fleksibiliti didapati berkurangan di kawasan-kawasan tertentu yang amat fleksibel dalam larutan akues, terutamanya bagi bahagian penutup struktur CRL. Simulasi MD turut mendedahkan yang anion RTIL mempunyai aturan berstruktur di sekitar enzim manakala molekul air ditemui secara setempat di kawasan tertentu pada permukaan protein. Anion hidrofobik seperti [PF<sub>6</sub>] melindungi lebih banyak kawasan dan lebih teratur pada kandungan air yang rendah manakala anion [Cl]- berkelakuan sebaliknya. Sementara itu, beberapa molekul air telah dilucutkan dari permukaan α-Chymotrypsin, CALB dan CRL.

(1960)<br>Holly iPsi, metrimotyain metriminiskan keitolisia steuktur yang lebih nati<br>Jamin suku metrimotyain metrimotyain metrimotyai keitolisia steukturan metrimotyain metrimotyain metrimotyain metrimotyain metrimotyain metr RTIL dengan anion [PF<sub>6</sub>] dan [TfO] mengekalkan lebih banyak air di permukaan berbanding dengan anion [BF<sub>4</sub>] dan [Cl], secara konsisten untuk tiga hidrolase itu. Anion [Tf2N]- pula didapati melucutkan paling banyak air untuk kes α-Chymotrypsin dan CALB manakala sebaliknya berlaku untuk CRL. Termodinamik pensolvatan bagi analog-analog rantaian sisi asid amino di dalam air dan lima RTIL berasaskan [BMIM] telah dikaji dengan menggunakan simulasi SD. Tenaga bebas pensolvatan telah dikira menggunakan kaedah Penerimaan Nisbah Bennett. Keputusan dari simulasi di dalam air didapati bersetuju dengan data eksperimen dan simulasi yang telah diterbitkan. RTIL menunjukkan keupayaan pensolvatan yang lebih baik berbanding dengan air. Analog tak berkutub menghasilkan tenaga bebas pensolvatan yang lebih rendah di dalam anion hidrofobik seperti [PF6] dan [Tf2N] manakala analog berkutub menunjukkan pensolvatan yang lebih baik di dalam anion hidrofilik seperti [BF<sub>4</sub>], [Cl] dan [TfO]. Sifat-sifat pensolvatan di dalam [BMIM][Cl] juga menjelaskan mengapa enzim mengalami gangguan struktur yang lebih didalam RTIL ini pada kandungan air yang rendah. Secara keseluruhan, simulasi komputer dapat menjelaskan berbagai kesan anion RTIL kepada struktur dan dinamik enzim di peringkat molekul. Kestabilan struktur dan fleksibiliti enzim didapati dipengaruhi oleh kandungan air, lebih daripada jenis anion RTIL yang dikaji. Keputusan simulasi MD didapati berkait rapat dengan laporan eksperimen. Juga, didapati bahawa kelakuan anion dan air di permukaan protein memainkan peranan utama kearah sifatsifat enzim dalam RTIL. Hasil kajian juga menunjukkan bahawa sifat-sifat permukaan biomangkin dan sifat-sifat fizikokimia substrat perlu diambil kira apabila memilih RTIL tertentu sebagai sistem pelarut.

## **ACKNOWLEDGEMENT**

Individual thanks to Allah (SWT), the Creator and Nabi Muhammad (PBUH), thus I can complete my thesis dissertation as fulfillment of requirements for the degree of Doctor of Philosophy at Universiti Putra Malaysia.

Firstly, I would like to take this opportunity to express my huge appreciation towards<br>my man supervisor, Prof. Dr. Molod Bassyarddinn Abdul Rahman, for giving me<br>countless opportunities and for his enciles aspect througho Firstly, I would like to take this opportunity to express my huge appreciation towards my main supervisor, Prof. Dr. Mohd Basyaruddin Abdul Rahman, for giving me countless opportunities and for his endless support throughout my candidature. I am deeply grateful to have such great person to be my guide. My acknowledgement also goes to my co-supervisors with a special tribute to Dr. Nuno Miguel Da Silva Micaêlo for a wonderful and enlightening experience in Braga, Portugal. My sincere gratitude also goes towards my colleagues, students at Macromolecular Simulation Laboratory and at the faculty along with all members of Enzyme and Microbial Technology Research Centre, UPM. I also acknowledge the National Science Fellowship MOSTI for the financial support.

Last but not least, I would like to thank my beloved family and friends for their help and support throughout my candidature. Thank you for always being there for me.

#### **APPROVAL**

[BMM] Hoasted lone Lequels" in accordance with the Universities and Chromete Colleges Act 1971 and the Constitution of the Universiti Plant Malaysia [PU(A)<br>106] IS Match 1998. The Conmittee reconnends that the student be a I certify that a Thesis Examination Committee has met on 19<sup>th</sup> August 2014 to conduct the final examination of Muhammad Alif bin Mohammad Latif on his thesis entitled "Molecular Insights on the Effects of Anions towards Hydrolases In [BMIM]-based Ionic Liquids" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Examination Committee were as follows:

#### **Mahiran Basri, PhD**

Professor Faculty of Science Universiti Putra Malaysia (Chairman)

#### **Raja Noor Zaliha Raja Abdul Rahman, PhD**

Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Internal Examiner)

#### **Hishamuddin Zainuddin, PhD**

Associate Professor Faculty of Science Universiti Putra Malaysia (Internal Examiner)

#### **Romas J. Kazlauskas, PhD**

Professor College of Biological Sciences University of Minnesota United States of America (External Examiner)

the company of the contract of

**NORITAH OMAR, PhD**  Associate Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 19 September 2014

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

#### **Mohd Basyaruddin Abdul Rahman, PhD**

Professor Faculty of Science Universiti Putra Malaysia (Chairman)

#### **Roghayeh Abedi Karjiban, PhD**

Lecturer Faculty of Science Universiti Putra Malaysia (Member)

## **Bimo Ario Tejo, PhD**

Senior Lecturer Faculty of Science Universiti Putra Malaysia (Member)

# **Nuno Miguel da Silva Micaelo, PhD**

Franchistor<br>
Franchistor Internal Malaysia<br>
(Chairman)<br>
(Chairman)<br>
Chairman<br>
Chairman Malaysia<br>
Charman Philip Correct Internal Malaysia<br>
(Mernher)<br>
Engar Ario Tejor, Philip Chairman<br>
Similar Lecturer<br>
Franchistor Science Lecturer Department of Chemistry, School of Science Minho University, Campus de Gualtar Braga, Portugal (Member)

## **BUJANG BIN KIM HUAT, PhD**

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

 $\mathcal{L}_\text{max}$  and  $\mathcal{L}_\text{max}$  and  $\mathcal{L}_\text{max}$  and  $\mathcal{L}_\text{max}$  and  $\mathcal{L}_\text{max}$  and  $\mathcal{L}_\text{max}$ 

## **DECLARATION**

## **Declaration by graduate student**

I hereby confirm that:

- $\bullet$  this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- The thosis has no original work.<br>
The this this state is more of any order of the state is state to substitute provide<br>
the state state with the state state state of the state is a functions:<br>
The degree of any order metho • written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
	- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.



Name and Matric No.: Muhammad Alif Mohammad Latif, GS 27830

## **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.



# **TABLE OF CONTENTS**







# **5 SUMMARY, GENERAL CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH 114** 5.1 Summary 114 5.2 Effects of RTIL Anions towards Hydrolases in General 117 5.3 General Conclusion 118<br>5.4 Recommendations for Future Research 119 Recommendations for Future Research



# **LIST OF TABLES**



Asp102. Data averaged over the last 2 ns of MD simulations.



# **LIST OF FIGURES**





- 4.4 Calculated densities for each RTIL studied during 5 ns of MD simulation. Values were averaged from three MD simulations for each RTIL. 63
- 4.05 The average densities of RTILs from MD simulations plotted against experimental measurements (Brennecke *et al*., 2004; Gardas *et al*., 2007). 64
- 4.06 Radial distributions plots of cations and anions in  $[BMIM][PF_6]$ from the last 2 ns of MD simulation. 65
- 4.07 Radial distributions plots of cations and anions in [BMIM][BF4] from the last 2 ns of MD simulation. 66
- 4.08 Radial distributions plots of cations and anions in [BMIM][Cl] from the last 2 ns of MD simulation. 66
- 4.09 Radial distributions plots of cations and anions in [BMIM][TfO] from the last 2 ns of MD simulation. 67
- 4.10 Radial distributions of plots cations and anions in  $[BMIM][Tf_2N]$ from the last 2 ns of MD simulation. 67
- 4.11 Snapshot pictures of the final protein conformation of α-Chymotrypsin at each water percentages. All structures were aligned and superimposed to the one in water. Water percentages colored as red  $(5\%)$ , green  $(10\%)$ , blue  $(15\%)$ , magenta  $(20\%)$ , orange (50%) and teal (water). 69
- 4.12 RMSD plot of the α-Chymotrypsin protein structure (all heavy atoms) solvated by all RTILs with different water percentages. Values are averages over the last 2ns of three replicatesMDsimulations. 70
- 4.13 RMSF plots of α-Chymotrypsin main chain solvated in RTILs with different water percentages. RMSF values were averaged over the last 2 ns of three replicatesMDsimulations. 72
- Gardian et al., 2007.<br>
Madial distributions plots of cations and antions in [BMIM][PF<sub>6</sub>] 65<br>
Islam distributions plots of cations and antions in [BMIM][BF<sub>6</sub>] 66<br>
from the last 2 ns of MD simulation.<br>
4.07 Radial distrib 4.14 Flexibility of  $\alpha$ -Chymotrypsin structures in different RTILs at 15% of water. Theoretical b-factors were rendered using a spectrum of colors from dark blue (denotes lowest flexibility) to red (denotes highest flexibility). All structures were aligned to the aqueous system (a). Active site and hydrophobic pocket region were highlighted only once on panel (b) and (c), respectively. The protein segment between residues 4 and 15 was highlighted on panel (e). 73
	- 4.15 RMSD plots of the catalytic triad of α-Chymotrypsin in all RITLs and water percentage. The catalytic triad is composed by residues His57, Asp102 and Ser195. Values were average from the last 2 ns of three replicatesMDsimulations. 75
- 4.16 Surface representation of the hydrophobic pocket of α-Chymotrypsin (colored in blue). 76
- 4.17 RMSD plots of the hydrophobic pocket of α-Chymotrypsin in different RTILs and water percentage. RMSD values were averaged from the last 2 ns of three replicates MD simulations. 77
- 4.18 Cluster representation of water molecules colored in blue (a, c, e, g, and i) and anions colored in red (b, d, f, h and j) localized within 0.5 nm from the protein surface. The systems shown are the ones solvated with 15% water percentage. The protein surface is rendered with a yellow molecular surface and the active site region is colored in dark green and highlighted on panel (j).
- 4.19 Cluster distribution of [Cl] ions (rendered with a red surface) in the system solvated by [BMIM][Cl] and 15% of water percentage. The figure highlights the presence of ions located inside of the protein structure. The protein is rendered with a yellow molecular surface and a slab was applied to visualize the interior of the protein.
- 4.20 Average number of water molecules within 0.5 nm located from the protein's surface at different water percentages and RTILs. 80
- 4.18 Cluster representation of water molecules colored in blue (a, e, e, e, e, e, a, e) and i) and ations colored in red 0, 6.4. The and b) localized within  $15\%$  water percentage. The systems shown are the ones solved w 4.21 Water molecules within 0.5 nm from the  $\alpha$ -Chymotrypsin catalytic triad residues in  $[BMIM][Tf_2N]$  at 5% of water percentage. Key catalytic residues are labeled on the figure. The distance of the Ser 195 hydroxyl and a nearby water molecule is also indicated. Figure made from the final configuration of a 10 ns MD simulation trajectory.
	- 4.22 Snapshot pictures of the final protein conformation of thermolysin at each water percentages. All structures were aligned and superimposed to the one in water. Water percentages colored as red (5%), green (10%), blue (15%), magenta (20%), orange (50%) and teal (water). 84
	- 4.23 RMSD of the heavy atoms of thermolysin in RTILs at different water percentages. Values were averaged from the last 2 ns of triplicate MD simulations. 85

4.24 RMSF plots of thermolysin in RTILs at different water percentages. 86

4.25 Snapshot pictures of  $[Zn]^{2+}$  ions (ball representation) and the catalytic triad residues (stick representation) from the last configurations from 10 ns MD simulations. All configurations from water (teal),  $[BMIM][PF_6]$  (black),  $[BMIM][BF_4]$  (red), [BMIM][Cl] (green), [BMIM][TfO] (blue) and  $[BMIM][Tf_2N]$ were aligned to the one from the crystal structure (white). 87

78

79

82

- 4.26 Snapshot pictures of the final protein conformation of thermolysin in (a) water and (b)  $[BMIM][PF_6]$  (10% water) at different temperatures. All structures were aligned and superimposed to the one at 25 °C. Temperature indicated with different color: red (25  $°C$ ), green (60 °C), blue (70 °C), magenta (80 °C) and orange (90  $^{\circ}$ C). 88
- 4.27 Heavy atoms RMSD of thermolysin at different temperature. Values were averaged from the last 2 ns of triplicate MD simulations.

89

- 4.27 Heavy atoms RMSD of thermolysin at different temperature<br>
Values were severaged from the list 2 ns of triplicate MD<br>
simulations.<br>
4.28 Snapshot pictures of the final protein conformation of CALB at<br>
value can<br>
wart 4.28 Snapshot pictures of the final protein conformation of CALB at each water percentages. All structures were aligned and superimposed to the one in water. Water percentages colored as red (5%), green (10%), blue (15%), magenta (20%), orange (50%) and teal (water). 91
	- 4.29 Snapshot pictures of the final protein conformation of CRL at each water percentages. All structures were aligned and superimposed to the one in water. Water percentages colored as red (5%), green (10%), blue (15%), magenta (20%), orange (50%) and teal (water). 92
	- 4.30 RMSD plots of CALB (a) and CRL (b) heavy atoms in five RTILs at different water percentages, fitted against the respective initial configuration. Values are averaged over the last 2 ns of triplicate MD simulations. 93
	- 4.31 Atomic fluctuations of CALB (a) and CRL (b) main chains in five RTILs at different water concentration. Values are averaged from the last 2 ns of triplicate MD simulations. 96
	- 4.32 B-factor per residue of CALB (a) and CRL (b) main chain in aqueous and RTIL solutions with 15% of water content. 97
	- 4.33 Snapshot showing the local flexibility of CALB (a) and CRL (b) in aqueous, expressed by the b-factors. A spectrum of colors was used to represent the flexibility from most rigid (dark blue) to most flexible (red). 98
	- 4.34 Spatial distributions of water (light blue) and RTIL anions ( $[PF_6]$ <sup>-</sup> in black, [BF<sub>4</sub>] in red, [Cl] in green, [TfO] in dark blue and [Tf<sub>2</sub>N] in magenta) that were found around 0.5 nm from the surface of CALB from the last 2 ns of MD simulations. 100
	- 4.35 Spatial distributions of water (light blue) and RTIL anions ([PF6] in black, [BF<sub>4</sub>] in red, [Cl] in green, [TfO] in dark blue and [Tf<sub>2</sub>N] in magenta) that were found around 0.5 nm from the surface of CRL from the last 2 ns of MD simulations. 101
- 4.36 Average percentage of water molecules that were found located around 0.5 nm from the surface of CALB (a) and CRL (b) for different RTILs across the water percentages. Values are averaged from the last 2 ns of MD simulations. 102
- 4.37 Surface residues classification presented in percentages using pie chart representations. 104
- endo acid residues that were found exposed on the surface of  $\alpha$ -<br>
Chymotrypsin, CALB and CRL erystal structures.<br>
4.39 Parity plot showing consistency between the solvation free energy 107<br>
eductioned in work: as compare 4.38 Amino acid residues that were found exposed on the surface of α-Chymotrypsin, CALB and CRL crystal structures. 104
	- 4.39 Parity plot showing consistency between the solvation free energy calculated in water as compared to literature. 107
	- 4.40 Parity plot comparing the solvation free energy of amino acid side chain analogues in RTILs and in water. 109
	- 4.41 Diagram showing the order of solvation between water and different RTIL anions. 111

# **LIST OF APPENDICES**



- D.04 Simulation Parameters for Free-Energy Calculations by SD Simulations 163
- E.01 Average Root Mean Square Deviations of Enzymes in Aqueous from Triplicates MD Simulations 167
- E.02 Average Root Mean Square Deviations of Enzymes in RTILs at Different Water Percentages from Triplicates MD Simulations 168
- F.01 Secondary structure representation by colors throughout the last 2 ns of MD equilibration simulations of thermolysin in aqueous at different temperatures. Amino acid residues were count from bottom to top. 176
- Different water recoeninges from inputates *Sub* Simulations<br>
2 no of MD equilibration simulations of thermolysin in agrees at different inspectation by colors throughout this last<br>
and inferent inspectations with section F.02 Secondary structure representation by colors throughout the last 2 ns of MD equilibration simulations of thermolysin in [BMIM][PF6] (10% water) at different temperatures. Amino acid residues were count from bottom to top.

177

# **LIST OF ABBREVIATIONS**





#### **CHAPTER 1**

## **INTRODUCTION**

speciality food productions and modernic. A sub-proceding of bioccolonology sison and the highly<br>as industrial bioschnology, also known as "white biotechnology" is one of the highly<br>progressing fields in bioschnology appl Biotechnology can be generally defined as the application of living organisms for producing useful products. For many years, biotechnology has been applied in agricultural, food productions and medicine. A sub-specialty of biotechnology such as industrial biotechnology, also known as "white biotechnology" is one of the highly progressing fields in biotechnology applications. In an era where production of chemicals on industrial scale is in need of sustainable processes, chemical synthesis of organic compounds have benefited most from the use of natural catalysts such as enzymes. The process where organic compounds were transformed chemically using enzymes is called biocatalysis (Anthonsen, 2000). In organic synthesis, enzymes have shown a remarkable performance as a biocatalyst. Unlike the traditional chemical and metal-catalyzed reactions, biocatalysis is more environmental-friendly and is a sustainable process while producing excellent yields (Wohlgemuth, 2010). Even though this technology has been vastly applied by academicians and industrial companies worldwide, the future of biocatalysis field promises much more. The developments in enzyme engineering has allowed synthetic biocatalysts to emerge with the potential to be better than the natural ones (Coelho *et al.*, 2013; Narayan and Sherman, 2013). Besides enzyme modification such as immobilization, substantial efforts have been made to re-engineer the natural biocatalysts. Screening of enzyme variants, directed evolutions and rational designs were applied in order to produce mutants that can expand their functionality in biocatalysis (Zhang *et al.*, 2003; Kazlauskas, 2005; Wahab *et al.*, 2012).

On the other hand, advancements in peptide synthesis have encouraged the use of peptidomimetics in order to design smaller biocatalysts with similar functions as the natural enzymes (Fernandez *et al.*, 1995). One of the crucial parts in a biocatalytic reaction is the reaction media. A solvent could affect both the enzymes and substrates in a biocatalysis process. Thus, the selection of a solvent would crucially depend on its effects toward the enzyme and substrates that are involved in the targeted reaction. The use of non-aqueous systems especially organic solvents in biocatalysis has shown increased reaction rates and higher conversions or yields (Klibanov, 2001). Non-aqueous solvents can be characterized by their physical and physicochemical properties such as boiling point, volatility, polarity, hydrophobicity, and viscosity to name a few. In the new millennium era, a new class of non-aqueous solvents have emerged as an exciting media for biocatalysis, called the Ionic Liquids (IL)s. Like the classical molten salts such as sodium chloride, an IL is composed entirely of ions (Davis and Fox, 2003). To differentiate ILs from the classical molten salts, the ones which are "low melting" or exist in liquid state at a temperature of below 100 °C can be called Room Temperature Ionic Liquids (RTIL)s (Welton, 1999).

Enzymatic reactions carried out in the presence of RTILs have been reported to produce more yield (increased enzyme activity) as compared to conventional organic solvents (Eckstein *et al.*, 2002; Lozano *et al.*, 2003; Maruyama *et al.*, 2004; Noritomi *et al.*, 2009). A growing interests in RTILs have seen numerous researches conducted in them, involving many types of reactions, enzymes and co-solvents (Sheldon *et al.*, 2002; Yang and Pan, 2005). In particular, RTILs have shown a remarkable influence toward the productivity of hydrolases such as lipases and proteases in the various organic reactions. It is widely known that there are many factors affecting the performance of enzymes in biocatalysis reactions. One of the major influences is the stability of the protein conformation. Structural changes to the protein conformation, particularly at the active site, can affect enzyme's catalytic capability. Selectivity is also very important in order to get the better yield for the desired product. The flexibility of protein conformation played a major role in determining the selectivity of an enzyme that is used in a biocatalytic reaction (Broos, 2002). Controlling the enzyme flexibility is therefore an important characteristic of a good solvent. One of the major advantages of using organic solvents in biocatalysis is that they stabilize the enzyme conformation (Ogino and Ishikawa, 2001). Furthermore, in conventional organic solvents, the enzyme flexibility can be controlled by the water concentration in the system (Kurkal *et al.*, 2005).

toward the productivity of hyriolass such as higress and protass in the various<br>organic reactions. It is widely known that there are many factors affecting the<br>performance of envywes in biocalaylays reactions. One of the What makes organic solvents like RTILs fascinating is that they can be composed of cations and anions with different physicochemical properties. For example, [BMIM][BF4] consists of a hydrophobic cation in 1-butyl-3-methylimidazolium  $(\overline{[BMM]}^+)$  and a hydrophilic tetrafluoroborate  $(\overline{[BF_4]})$  anion. Theoretically, this RTIL can provide two distinctive characteristics toward the enzyme and substrates involved when used as the reaction media. Therefore, the physicochemical properties of RTIL cations and anions can have a huge influence on the solvation properties of the solute molecules. In a system which contains an enzyme, water and RTILs, the enzyme's structure and dynamics properties can be affected by the interactions between RTILs and water, particularly at the enzyme's surface. These interactions however, are extremely difficult to be observed experimentally. From many attempts, researchers have been trying to explain the mechanisms of RTILs interactions in chemical reactions (Zhao, 2010). Most of the published reports include the effect of RTILs on activity and stability of enzymes, but the interactions between RTILs, enzymes and water have been sparsely investigated. Only a small part of these researches were focusing on the structural and dynamics behavior of enzymes and RTILs in such system (Bourissou *et al.*, 2000; Raza *et al.*, 2001; Micaelo *et al.*, 2005; Logotheti *et al.*, 2009; Klähn *et al.*, 2011).

#### **1.1 Problem Identification**

Due to the complexity of RTILs, some may work well with a certain enzyme but not with others. These have been highlighted by several reports in recent years (Kaar *et al*., 2003; Park and Kazlauskas, 2003; Klähn *et al*., 2011). This phenomenon is related to the fact that the cation-anion combination can affect the performance of RTILs as the reaction media. Since a vast number of cation-anion combinations of RTILs have been introduced and studied in recent years, finding the best RTIL for a particular enzyme or a certain reaction is time, and resource-consuming. Many experimental works have been carried out to determine the effect of different cationanion combinations toward the performance of biocatalysts such as hydrolases (Irimescu and Kato, 2004; Paljevac *et al.*, 2006; Lee *et al.*, 2008; Herńandez-Ferńandez *et al.*, 2009; Zhao, 2010). However, the focus usually tends to go toward activity, reaction rate and yield but not onto the enzyme properties. Therefore, to relate the findings with structural and dynamics properties of enzymes such as stability, flexibility, surface interactions and solvation is a tricky task. To really understand how enzymes react with RTILs and how RTILs affect the enzyme performance as a whole, it is essential to look into the structural and dynamics properties in the presence of these solvents. Understanding of RTILs solvent effect in more detail could provide imperative support when the study focuses on the interactions between enzymes, water and cation/anion at enzyme's surface. This will ensure that before a particular RTIL is chosen to work with an enzyme, one will have an idea what criteria each must have in order to work well.

RTILL shave been introduced and statical in reconvergens finding the best RTILL for a neutron correlation and statical interest of a control content<br>membershave best and the observation of the determinant of the content c This can be predicted by using computational approach, such as molecular modeling and simulations, where the behavior of enzymes during solvation with RTILs can be predicted at molecular level. As one of the popular computer simulation methods, molecular dynamics (MD) has been proven as an excellent tool to distinguish the structural properties of biomolecules in aqueous and organic media (Lousa *et al.*, 2013). In MD, understanding of enzyme behavior in RTILs can be increased by mimicking the interactions between the enzyme and RTIL components in great atomic details. By combining reported experimental findings available and computer simulation studies, the relationship between enzyme activity and its molecular properties in RTILs can be further explained. It is believed that the effects of using different RTIL anions toward hydrolases' properties at molecular level can be revealed by the use of computer simulation techniques such as MD. From the analyses performed, a certain order can be established between anions, in relation toward the structural and dynamics properties of all hydrolases studied. The hypothesis is that the order is dependent on the physicochemical properties of the five anions and should coincide with experimental evidences. Meanwhile, thermodynamics characterizations from the free energy calculations can be used to predict the solvation properties of enzymes and substrates in different RTILs.

## **1.2 Objectives**

The main goal is to utilize molecular dynamics technique in order to investigate the structural and dynamics properties of different hydrolases such as α-Chymotrypsin, thermolysin, *Candida antarctica* Lipase B and *Candida rugosa* lipase when solvated in BMIM-based RTILs, composed of different anions such as  $[PF_6]$ ,  $[BF_4]$ ,  $[Cl]$ , [TfO] and [Tf<sub>2</sub>N]. Therefore, these objectives will be pursued:

- 1. To determine the effect of RTILs toward the structural stability and flexibility of hydrolases
- 2. To observe the effect of water concentration on enzyme properties in RTILs
- 3. To characterize enzyme:water:RTIL interactions on the protein surface and correlate with enzyme's structure and dynamics properties
- 4. To estimate the solvation free energy of small molecules in RTILs

1. To determine the effect of RTIs toward the structural subility and<br>
flexibility of hydrodases<br>
2. To obsarve the effect of water concentration on enzyme properties in RTILs<br>
3. Io characterize curgeme-water-RTIL intera The main focus of this project is on the effects when different RTIL anions are used. MD simulations was used to predict the behavior of several hydrolases in five [BMIM]-based RTILs. The five anions chosen for this project are consisted of four fluorine-based anions, commonly reported to increase enzyme activity. They were hexafluorophosphate ([PF<sub>6</sub>]<sup>-</sup>), tetrafluoroborate ([BF<sub>4</sub>]<sup>-</sup>), trifluoromethanesulfonate ([TfO]) and bis-trifluoromethylsulfonylimide ( $[Tf_2N]$ ). Chloride ([Cl]) anion was also selected due to its physical and physicochemical properties for comparison purposes. In the next chapter, the literatures related to the project will be discussed. The theoretical background and methodologies such as algorithms, parameters and analysis tools that were used during the project will be presented in the third chapter. In chapter four, the results from the simulations will be presented, correlated with experimental evidences and discussed. In order to verify that the models used can produce similar properties as determined experimentally, molecular modeling and simulations studies on selected RTILs were performed and reported. After validations, these models were used to investigate the effects of using different RTIL anions toward the structure and dynamics of an  $\alpha$ -Chymotrypsin at different hydration level. Due to the success of modeling the α-Chymotrypsin's behavior in RTILs, a similar approach was applied for thermolysin, which is currently underutilized in RTILs. The structural stability and flexibility of both proteases were compared. Lipases behavior in RTILs with different anions was compared between *Candida antarctica* Lipase B (CALB) and *Candida rugosa* Lipase (CRL). Lastly, the solvation thermodynamics of neutral amino acid side chain analogues in different RTIL anions were predicted and the results were discussed in relation to the solvation dynamics of enzymes and substrates in RTILs. In the last chapter, the summary of all findings will be provided. The structural stability and dynamics of all enzymes were summarized and a general trend on the effects of changing RTIL anions was elucidated. This was followed by the general conclusions that obtained by this project and recommendations for future works.

#### **REFERENCES**

- Abascal J. L. and Vega C. (2005) A general purpose model for the condensed phases of water: tip4p/2005. *Journal of Chemical Physics*, 123(23), 234505.
- Alder B. J. and Wainwright T. E. (1959) Studies in molecular dynamics. I. General method. *Journal of Chemical Physics*, 31(2), 459-466.
- Anderson J. L., Ding J., Welton T. and Armstrong D. W. (2002) Characterizing ionic liquids on the basis of multiple solvation interactions. *Journal of American Chemical Society*, 124(47), 14247-14254.
- Anthonsen T. (2000). Reactions catalyzed by enzymes. In A. J. J. Straathof & P. Adlercreutz (Eds.), *Applied Biocatalysis* (2nd ed.), Taylor & Francis, Amsterdam, pp. 18-59.
- Bennett C. H. (1976) Efficient estimation of free energy differences from Monte Carlo data. *Journal of Computational Physics*, 22(2), 245-268.
- Berendsen H. J. C., Postma J. P. M., Gunsteren W. F. v., A. DiNola and Haak J. R. (1984) Molecular dynamics with coupling to an external bath. *Journal of Chemical Physics*, 81(8), 3684-3690.
- Berg J. M., Tymoczko J. L. and Stryer. L. (2002). Section 9.1, Proteases: Facilitating a difficult reaction. *Biochemistry*  $(5<sup>th</sup>$  ed.), W H Freeman, New York, Retrieved from http://www.ncbi.nlm.nih.gov/books/NBK22526/.
- Bordusa F. (2002) Proteases in organic synthesis. *Chemical Reviews*, 102(12), 4817- 4868.
- Borodin O. (2009) Polarizable force field development and molecular dynamics simulations of ionic liquids. *The Journal of Physical Chemistry B*, 113(33), 11463-11478.
- Bourissou D., Guerret O., Gabbai F. P. and Bertrand G. (2000) Stable carbenes. *Chemical Reviews*, 100(1), 39-92.
- Boys S. F., Cook G. B., Reeves C. M. and Shavitt I. (1956) Automatic fundamental calculations of molecular structure. *Nature*, 178(4544), 1207-1209.
- Anderson J. L., Ding J., Wellon T. and Armstrong D. W. (2002) Characterizing ionic<br>
iquidis on the basis of multiple solvation interactions. Journal of American<br>
Chemical Society: 124(47), 14247-14254.<br>
Anthonsen T. (2000 Brennecke J. F., Fredlake C. P., Crosthwaite J. M., Hert D. G. and Aki S. N. V. K. (2004) Thermophysical properties of imidazolium-based ionic liquids. *Journal of Chemical and Engineering Data*, 19, 954-964.
	- Broos J. (2002) Impact of the enzyme flexibility on the enzyme enantio-selectivity in organic media towards specific and non-specific substrates. *Biocatalysis and Biotransformation*, 20(4), 291-295.
- Broos J., Visser A. J. W. G., Engbersen J. F. J., Verboom W., Hoek A. v. and Reinhoudt D. N. (1995) Flexibility of enzymes suspended in organic solvents probed by time-resolved fluorescence anisotropy. Evidence that enzyme activity and enantioselectivity are directly related to enzyme flexibility. *Journal of American Chemical Society*, 117(51), 12657-12663.
- Bussi G., Donadio D. and Parrinello M. (2007) Canonical sampling through velocity rescaling. *Journal of Chemical Physics*, 126(014101), 1-7.
- Carmichael A. J. and Seddon K. R. (2000) A polarity study of the 1-alkyl-3 methylimidazolium ambient temperature ionic liquids with the solvatochromic dye, Nile Red. *Journal of Physical and Organic Chemistry*, 13, 591-595.
- Carrera G. V. S. M., Branco L. C., Aires-de-Sousa J. and Afonso C. A. M. (2008) Exploration of quantitative structure-property relationships (QSPR) for the design of new guanidinium ionic liquids. *Tetrahedron Letters*, 64, 2216.
- Chaibakhsh N., Abdul Rahman M., Abd-Aziz S., Basri M., Salleh A. and Rahman R. (2009) Optimized lipase-catalyzed synthesis of adipate ester in a solventfree system. *Journal of Industrial Microbiology & Biotechnology*, 36(9), 1149-1155.
- rescaling. Journal of Chemical Physics, 126(014101), 1-7.<br>
Carmichal A. J. and Seddon K. R. (2000) A polarity study of the 1-silky1-3-<br>
methylimidazolium ambient temperature ionic liquids with the<br>
solventochromic dys. Ni ChemViews. (2013) Nobel Prize in Chemistry 2013, http://www.chemistryviews.org/details/ezine/5344441/Nobel Prize in Ch emistry 2013.html, NobelPrize.org, Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim
	- Chen J. and Brooks Iii C. L. (2008) Implicit modeling of nonpolar solvation for simulating protein folding and conformational transitions. *Physical Chemistry Chemical Physics,* 10(4), 471-481.
	- Coelho P. S., Brustad E. M., Kannan A. and Arnold F. H. (2013) Olefin cyclopropanation via carbene transfer catalyzed by engineered Cytochrome P450 enzymes. *Science*, 339(6117), 307-310.
	- Darden T., York D. and Pedersen L. (1993) Particle mesh Ewald: An *N*•log(*N*) method for Ewald sums in large systems. *Journal of Chemical Physics*, 98(12), 10089-10092.
	- Davis J. J. H. and Fox P. A. (2003) From curiosities to commodities: ionic liquids begin the transition. *Chemical Communications*, (11), 1209-1212.
	- de Andrade J., Böes E. S. and Stassen H. (2002) Computational study of room temperature molten salts composed by 1-alkyl-3-methylimidazolium cations force-field proposal and validation. *The Journal of Physical Chemistry B*, 106(51), 13344-13351.
	- de Ruiter A., Boresch S. and Oostenbrink C. (2013) Comparison of thermodynamic integration and Bennett acceptance ratio for calculating relative proteinligand binding free energies. *Journal of Computational Chemistry*, 34(12), 1024-1034.
- Earl D. J. and Deem M. W. (2008). Monte Carlo simulations. In A. Kukol (Ed.), *Molecular Modeling of Proteins* (Vol. 443). Humana Press, Totowa, New Jersey.
- Eckstein M., Sesing M., Kragl U. and Adlercreutz P. (2002) At low water activity  $\alpha$ chymotrypsin is more active in an ionic liquid than in non-ionic organic solvents. *Biotechnology Letters*, 24, 867-872.
- Eijsink V. G. H., Voltman O. R., Aukesma W., Vriend G. and Venema G. (1995)<br>
Structural determinants of the stability of thermolysia-like proteinases.<br>
Mature: Structural and Molecular Biology, 2(5), 374-379.<br>
Fersley J. Eijsink V. G. H., Veltman O. R., Aukema W., Vriend G. and Venema G. (1995) Structural determinants of the stability of thermolysin-like proteinases. *Nature: Structural and Molecular Biology*, 2(5), 374-379.
	- Emsley J. (1980) Very strong hydrogen bonding. *Chemical Society Reviews*, 9(1), 91-124.
	- Erbeldinger M., Mesiano A. J. and Russell A. J. (2000) Enzymatic catalysis of formation of Z-Aspartame in ionic liquid - an alternative to enzymatic catalysis in organic solvents. *Biotechnology Progress*, 16, 1129-1131.
	- Fernandez M. M., Margot A. O., Falender C. A., Blanch H. W. and Clark D. S. (1995) Enzymatic synthesis of peptides containing unnatural amino acids. *Enzyme and Microbial Technology*, 17(11), 964-971.
	- Fitzpatrick P. A. and Klibanov A. M. (1991) How can the solvent affect enzyme enantioselectivity. *Journal of American Chemical Society*, 113, 3166- 3171.
	- Fletcher R. and Powell M. J. D. (1963) A rapidly convergent descent method for minimization. *The Computer Journal*, 6(2), 163-168.
	- Freemantle M. (1998) Designer solvents. *Chemical & Engineering News Archive*, 76(13), 32-37.
	- G. Huddleston J. and D. Rogers R. (1998) Room temperature ionic liquids as novel media for 'clean' liquid-liquid extraction. *Chemical Communications*, (16), 1765-1766.
	- Gardas R. L. and Coutinho J. A. P. (2008) A group contribution method for viscosity estimation of ionic liquids. *Fluid Phase Equilibria*, 266, 195-201.
	- Gardas R. L. and Coutinho J. A. P. (2008) Applying a QSPR correlation to the prediction of surface tensions of ionic liquids. *Fluid Phase Equilibria*, 265, 57-65.
	- Gardas R. L., Freire M. G., Carvalho P. J., Marrucho I. M., Fonseca I. M. A., Ferreira A. G. M. and Coutinho J. A. P. (2007) High-pressure densities and derived thermodynamic properties of imidazolium-based ionic liquids. *Journal of Chemical and Engineering Data*, 52, 80-88.
	- Gorman L. A. S. and Dordick J. S. (1992) Organic solvents strip water off enzymes. *Biotechnology and Bioengineering*, 39(4), 392-397.
- Goto M., Moniruzzaman M. and Kamiya N. (2010) Activation and stabilization of enzymes in ionic liquids. *Organic Biomolecule Chemistry*, 8, 2887-2899.
- Gotor-Fernández V. and Vicente G. (2007). Use of lipases in organic synthesis. In J. Polaina & A. MacCabe (Eds.), *Industrial Enzymes*, Springer, Netherlands, pp. 301-315.
- Grochuski P, Li Y, Schotz P, D, Bouthiller F, Smith P, Harrison D, Rubin B,  $\mathbb{R}$  and Gxyler M. (1993) Insights into interficial activation from an open<br>stateture of Condita ruggosa lipses. Journal Bublogical Chemister Grochulski P., Li Y., Schrag J. D., Bouthillier F., Smith P., Harrison D., Rubin B. and Cygler M. (1993) Insights into interfacial activation from an open structure of *Candida rugosa* lipase. *Journal Biological Chemistry*, 268, 12843-12847
	- Gu Z. and Brennecke J. F. (2002) Volume expansivities and isothermal compressibilities of imidazolium and pyridinium-based ionic liquids. *Journal of Chemical & Engineering Data*, 47(2), 339-345.
	- Hanke C., Price S. and Lynden-Bell R. (2001) Intermolecular potentials for simulations of liquid imidazolium salts. *Molecular Physics*, 99(10), 801- 809.
	- Hartsough D. S. and Merz K. M. (1992) Protein flexibility in aqueous and nonaqueous solutions. *Journal of American Chemical Society*, 114, 10113- 10116.
	- Hernández-Fernández F. J., Ríos A. P. d. l., Tomás-Alonso F., Gómez D. and Víllora G. (2009) Stability of hydrolase enzymes in ionic liquids. *The Canadian Journal of Chemical Engineering*, 87(6), 910-914.
	- Hess B., Bekker H., Berendsen H. J. C. and Fraaije J. G. E. M. (1997) LINCS: A linear constraint solver for molecular simulations. *Journal of Computational Chemistry*, 18(12), 1463-1472.
	- Hess B., Kutzner C., Spoel D. v. d. and Lindahl E. (2008) GROMACS 4: Algorithms for highly efficient, load-balanced, and scalable molecular simulation. *Journal of Chemical Theory and Computation,* 4, 435-447.
	- Hloucha M. and Deiters U. K. (1998) Fast coding of the minimum image convention. *Molecular Simulation*, 20(4), 239-244.
	- Hockney R. W., Goel S. P. and Eastwood J. W. (1974) Quiet high-resolution computer models of a plasma. *Journal of Computational Physics*, 14(2), 148-158.
	- Holbrey J. D., Reichert W. M. and Rogers R. D. (2004) Crystal structures of imidazolium bis(trifluoromethanesulfonyl)imide 'ionic liquid' salts: the first organic salt with a cis-TFSI anion conformation. *Dalton Transactions*, 0(15), 2267-2271.
	- Holland D. R., Tronrud D. E., Pley H. W., Flaherty K. M., Stark W., Jansonius J. N., McKay D. B. and Matthews B. W. (1992) Structural comparison suggests that thermolysin and related neutral proteases undergo hinge-bending motion during catalysis. *Biochemistry*, 31(46), 11310-11316.
- Horn J. N., Romo T. D. and Grossfield A. (2013) Simulating the mechanism of antimicrobial lipopeptides with all-atom molecular dynamics. *Biochemistry*, 52(33), 5604-5610.
- Howells R. D. and McCown J. D. (1977) Trifluoromethanesulfonic acid and derivatives. *Chemical Reviews*, 77(1), 69-92.
- Huddleston J. G., Visser A. E., Reichert W. M., Willauer H. D., Broker G. A. and Rogers R. D. (2001) Characterization and comparison of hydrophilic and hydrophobic room temperature ionic liquids incorporating the imidazolium cation. *Green Chemistry*, 3(4), 156-164.
- Humphrey W., Dalke A. and Schulten K. (1996) VMD Visual Molecular Dynamics. *Journal of Molecular Graphics*, 14, 33-38.
- Hünenberger P. H., Mark A. E. and van Gunsteren W. F. (1995) Fluctuation and cross-correlation analysis of protein motions observed in nanosecond molecular dynamics simulations. *Journal of Molecular Biology*, 252(4), 492-503.
- Irimescu R. and Kato K. (2004) Investigation of ionic liquids as reaction media for enzymatic enantioselective acylation of amines. *Journal of Molecular Catalysis B: Enzymatic*, 30, 189-194.
- Itoh T., Akasaki E., Kudo K. and Shirakami S. (2001) Lipase-catalyzed enantioselective acylation in the ionic liquid solvent system: reaction of enzyme anchored to the solvent. *Chemistry Letters*, 30(3), 262-263.
- Jorgensen W. L., Chandrasekhar J., Madura J. D., Impey R. W. and Klein M. L. (1983) Comparison of simple potential functions for simulating liquid water. *Journal of Chemical Physics*, 79, 926-935.
- Huddetson J. G., Visser A. E., Reichert W. M., Willum El I. D., Droker G. A. and<br>
Rogers R. D. (2001) Chemeterization and comparison of hydrophilic and<br>
hydropholic contact menerature ionic liquids incorporating the imida Jorgensen W. L., Maxwell D. S. and Tirado-Rives J. (1996) Development and testing of the OPLS All-Atom force field on conformational energetics and properties of organic liquids. *Journal of American Chemical Society*, 118, 11225-11236.
	- Kaar J. L., Jesionowski A. M., Berberich J. A., Moulton R. and Russell A. J. (2003) Impact of ionic liquid physical properties on lipase activity and stability. *Journal of American Chemical Society*, 125, 4125-4131.
	- Kamal M. Z., Yedavalli P., Deshmukh M. V. and Rao N. M. (2013) Lipase in aqueous-polar organic solvents: Activity, structure, and stability. *Protein Science*, 22(7), 904-915.
	- Karplus M. and McCammon J. A. (2002) Molecular dynamics simulations of biomolecules. *Nature: Structural and Molecular Biology*, 9(9), 646-652.
- Katritzky A. R., Kuanar M., Stoyanova-Slavova I. B., Slavov S. H., Dobchev D. A., Karelson M. and Acree W. E. J. (2008) Quantitative structure-property relationship studies on Ostwald solubility and partition coefficients of organic solutes in ionic liquids. *Journal of Chemical and Engineering Data*, 53, 1085-1092.
- Kazlauskas R. J. (2005) Enhancing catalytic promiscuity for biocatalysis. *Current Opinion in Chemical Biology*, 9(2), 195-201.
- Opinion in Chomical Buology, 9(2), 195-201.<br>
Kielbasiiski P., Alhyrht M., Luozak J. and Mikolajcyk M. (2002) Enzymatic<br>
reactions in ionic liquids: liquical subproximation of necessine<br>
chiral hydroxynechiates and bydrox Kiełbasiński P., Albrycht M., Łuczak J. and Mikołajczyk M. (2002) Enzymatic reactions in ionic liquids: lipase-catalysed kinetic resolution of racemic, Pchiral hydroxymethanephosphinates and hydroxymethylphosphine oxides. *Tetrahedron: Asymmetry*, 13(7), 735-738.
	- Kim K.-W., Song B., Choi M.-Y. and Kim M.-J. (2001) Biocatalysis in ionic liquids: markedly enhanced enantioselectivity of lipase. *Organic Letters*, 3(10), 1507-1509.
	- Klähn M., Lim G. S. and Wu P. (2011) How ion properties determine the stability of a lipase enzyme in ionic liquids: A molecular dynamics study. *Physical Chemistry Chemical Physics*, 13, 18647-18660.
	- Klähn M., Lim G. S., Seduraman A. and Wu P. (2011) On the different roles of anions and cations in the solvation of enzymes in ionic liquids. *Physical Chemistry Chemical Physics*, 13, 1649-1662.
	- Klibanov A. M. (1989) Enzymatic catalysis in anhydrous organic solvents. *Trends in Biochemical Sciences*, 14(4), 141-144.
	- Klibanov A. M. (1997) Why are enzymes less active in organic solvents than in water? *Trends in Biotechnology*, 15(3), 97-101.
	- Klibanov A. M. (2001) Improving enzymes by using them in organic solvents. *Nature*, 409(6817), 241-246.
	- Köddermann T., Paschek D. and Ludwig R. (2007) Molecular dynamic simulations of ionic liquids: A reliable description of structure, thermodynamics and dynamics. *Chemical Physics Chemistry*, 8(17), 2464-2470.
	- Koshland D. E. (1959) Enzyme flexibility and enzyme action. *Journal of Cellular and Comparative Physiology*, 54(S1), 245-258.
	- Koshland D. E. (1963) The role of flexibility in enzyme action. *Cold Spring Harbor Symposia on Quantitative Biology*, 28, 473-482.
	- Kragl U., Eckstein M. and Kaftzik N. (2002) Enzyme catalysis in ionic liquids. *Current Opinion in Biotechnology*, 13, 565-571.
	- Kurkal V., Daniel R. M., Finney J. L., Tehei M., Dunn R. V. and Smith J. C. (2005) Enzyme activity and flexibility at very low hydration. *Biophysical Journal*, 89(2), 1282-1287.
- Laszlo J. A. and Compton D. L. (2001)  $\alpha$ -Chymotrypsin catalysis in imidazoliumbased ionic liquids. *Biotechnology and Bioengineering*, 75(2), 181-186.
- Lau R. M., Rantwijk F. v., Seddon K. R. and Sheldon R. A. (2000) Lipase-catalyzed reactions in ionic liquids. *Organic Letters*, 2(26), 4189-4191.
- Lee J., Suh S. W. and Shin S. (2000) Computational studies of essential dynamics of *pseudomonas cepacia* lipase. *Journal of Biomolecular Structure and Dynamics*, 18(2), 297-309.
- Lee S., Koo Y.-M. and Ha S. (2008) Influence of ionic liquids under controlled water activity and low halide content on lipase activity. *Korean Journal of Chemical Engineering*, 25(6), 1456-1462.
- Liu H., Sale K. L., Holmes B. M., Simmons B. A. and Singh S. (2010) Understanding the interactions of cellulose with ionic liquids: A molecular dynamics study. *The Journal of Physical Chemistry B*, 114(12), 4293- 4301.
- Logotheti G. E., Ramos J. and Economou L. G. (2009) Molecular modeling of imidazolium-based [Tf<sub>2</sub>N] ionic liquids: Microscopic structure, thermodynamic and dynamic properties, and segmental dynamics. *The Journal of Physical Chemistry B*, 113, 7211-7224.
- Lopes J. N. C. and Pádua A. A. H. (2004) Molecular force field for ionic liquids composed of triflate or bistriflylimide anions. *The Journal of Physical Chemistry B*, 108, 16893-16898.
- Lopes J. N. C. and Pádua A. A. H. (2006) Molecular force field for ionic liquids iii: imidazolium, pyridinium, and phosphonium cations; chloride, bromide, and dicyanamide anions. *The Journal of Physical Chemistry B*, 110, 19586-19592.
- Lopes J. N. C., Deschamps J. and Pádua A. A. H. (2004) Modeling ionic liquids using a systematic all-atom force field. *The Journal of Physical Chemistry B*, 108(6), 2038-2047.
- *pseudomouss cepacain* lipses. Journal of Biomolecular Structure and<br>
Dynamics 18(2), 297-309.<br>
Lec S. Koo Y.-M. and Ha S. (2008) Influence of ionic liquids under controlled water<br>
uctivity and luw halide content on lipse Lopes J. N. C., Pádua A. A. H. and Shimizu K. (2008) Molecular force field for ionic liquids iv: trialkylimidazolium and alkoxycarbonyl-imidazolium cations; alkylsulfonate and alkylsulfate anions. *The Journal of Physical Chemistry B*, 112, 5039-5046.
	- Lou W.-y., Zong M.-h. and Wu H. (2004) Enhanced activity, enantioselectivity and stability of papain in asymmetric hydrolysis of d,l-p-hydroxyphenylglycine methyl ester with ionic liquid. *Biocatalysis and Biotransformation*, 22(3), 171-176.
	- Lousa D., Baptista A. M. and Soares C. M. (2013) A molecular perspective on nonaqueous biocatalysis: contributions from simulation studies. *Physical Chemistry Chemical Physics*, 15(33), 13723-13736.
- Lozano P., De Diego T., Carrié D., Vaultier M. and Iborra J. L. (2003) Enzymatic ester synthesis in ionic liquids. *Journal of Molecular Catalysis B: Enzymatic*, 21(1–2), 9-13.
- Lozano P., Diego T. d., Gmouh S., Vaultier M. and Iborra J. L. (2004) Green enzymatic processes in ionic liquid. supercritical carbon dioxide. *Biotechnology Progress*, 20, 661-669.
- Lozano P., Diego T. d., Carrie D., Vaultier M. and Iborra J. L. (2001) Overstabilization of *Candida antarctica* lipase B by ionic liquids in ester synthesis. *Biotechnology Letters*, 23, 1529-1533.
- Lozano P., Diego T. d., Guegan J.-P., Vaultier M. and Iborra J. L. (2001) Stabilization of α-Chymotrypsin by ionic liquids in transesterification reactions. *Biotechnology and Bioengineering*, 75(5), 563-569.
- Lustig R. (1998) Microcanonical Monte Carlo simulation of thermodynamic properties. *Journal of Chemical Physics*, 109(20), 8816-8828.
- Lozamo P. Dicgo T. d., Carric D., Vaultier M. and Iborra J. L. (2001) Over-<br>stabilization of *Candida antarctica* lipase B by ionic liquids in ester<br>symbosis. *Biotechnology Letters*, 23, 1529-1533.<br>
Lozamo P., Diego T. d MacKerell A. D., Bashford D., Bellott, Dunbrack R. L., Evanseck J. D., Field M. J., Fischer S., Gao J., Guo H., Ha S., Joseph-McCarthy D., Kuchnir L., Kuczera K., Lau F. T. K., Mattos C., Michnick S., Ngo T., Nguyen D. T., Prodhom B., Reiher W. E., Roux B., Schlenkrich M., Smith J. C., Stote R., Straub J., Watanabe M., Wiórkiewicz-Kuczera J., Yin D. and Karplus M. (1998) All-atom empirical potential for molecular modeling and dynamics studies of proteins. *The Journal of Physical Chemistry B*, 102(18), 3586- 3616.
	- Maranas C. and Floudas C. (1994) Global minimum potential energy conformations of small molecules. *Journal of Global Optimization*, 4(2), 135-170.
	- Martínez L., Andrade R., Birgin E. G. and Martínez J. M. (2009) PACKMOL: A package for building initial configurations for molecular dynamics simulations. *Journal of Computational Chemistry*, 30(13), 2157-2164.
	- Maruyama T., Yamamura H., Kotani T., Kamiya N. and Goto M. (2004) Poly(ethylene glycol)-lipase complexes that are highly active and enantioselective in ionic liquids. *Organic & Biomolecular Chemistry*, 2(8), 1239-1244.
	- Mat Radzi S., Basri M., Bakar Salleh A., Ariff A., Mohammad R., Abdul Rahman M. B. and Abdul Rahman R. N. Z. R. (2005) High performance enzymatic synthesis of oleyl oleate using immobilised lipase from *Candida antartica*. *Electronic Journal of Biotechnology*, 8(3), 292-298.
	- Matthews B. W., Weaver L. H. and Kester W. R. (1974) The conformation of Thermolysin. *Journal of Biological Chemistry*, 249(24), 8030-8044.
	- McCammon J. A., Gelin B. R. and Karplus M. (1977) Dynamics of folded proteins. *Nature*, 267(5612), 585-590.
- McNaught A. D. and Wilkinson A. (1997). IUPAC. Compendium of chemical terminology, 2nd ed. (the "Gold Book"). Blackwell Scientific Publications, Oxford.
- Metropolis N., Rosenbluth A. W., Rosenbluth M. N., Teller A. H. and Teller E. (1953) Equation of state calculations by fast computing machines. *Journal of Chemical Physics*, 21(6), 1087-1092.
- Micaêlo N. M. and Soares C. M. (2007) Modeling hydration mechanisms of enzymes in nonpolar and polar organic solvents. *FEBS Journal*, 274, 2424-2436.
- Micaêlo N. M. and Soares C. M. (2008) Protein structure and dynamics in ionic liquids. Insights from molecular dynamics simulation studies. *The Journal of Physical Chemistry B*, 112(9), 2566-2572.
- Micaelo N. M., Baptista A. M. and Soares C. M. (2006) Parametrization of 1-butyl-3-methylimidazolium hexafluorophosphate/nitrate ionic liquid for the GROMOS force field. *The Journal of Physical Chemistry B*, 110, 14444- 14451.
- Micaelo N. M., Teixeira V. H., Baptista A. M. and Soares C. M. (2005) Water dependent properties of cutinase in nonaqueous solvents: a computational study of enantioselectivity. *Biophysical Journal*, 89(2), 999-1008.
- Millero F. J., Curry R. W. and Drost-Hansen W. (1969) Isothermal compressibility of water at various temperatures. *Journal of Chemical & Engineering Data*, 14(4), 422-425.
- Miyako E., Maruyama T., Kamiya N. and Goto M. (2003) Use of ionic liquids in a lipase-facilitated supported liquid membrane. *Biotechnology Letters*, 25(10), 805-808.
- Misaclo N. M. and Soarcs C. M. (2007) Modeling hydration mechanisms of enzymes<br>
in nonpolar and polar organic solvents. *FEBS Journal*, 274, 2424-2436.<br>
Micaleo N. M. and Soarcs C. M. (2008) Proteins simulation studies: Muramatsu M., Nagasawa Y. and Miyasaka H. (2011) Ultrafast solvation dynamics in room temperature ionic liquids observed by three-pulse photon echo peak shift measurements. *The Journal of Physical Chemistry A*, 115(16), 3886-3894.
	- Nara S. J., Harjani J. R., Salunkhe M. M., Mane A. T. and Wadgaonkar P. P. (2003) Lipase-catalysed polyester synthesis in 1-butyl-3-methylimidazolium hexafluorophosphate ionic liquid. *Tetrahedron Letters*, 44(7), 1371-1373.
	- Narayan A. R. H. and Sherman D. H. (2013) Re-engineering nature's catalysts. *Science*, 339(6117), 283-284.
	- Nilsson L. G. and Padró J. A. (1990) A time-saving algorithm for generalized Langevin-dynamics simulations with arbitrary memory kernels. *Molecular Physics*, 71(2), 355-367.
	- Norin M., Haeffner F., Hult K. and Edholm O. (1994) Molecular dynamics simulations of an enzyme surrounded by vacuum, water, or a hydrophobic solvent. *Biophysical Journal*, 67(2), 548-559.
- Noritomi H., Suzuki K., Kikuta M. and Kato S. (2009) Catalytic activity of α-Chymotrypsin in enzymatic peptide synthesis in ionic liquids. *Biochemical Engineering Journal*, 47, 27-30.
- Ogino H. and Ishikawa H. (2001) Enzymes which are stable in the presence of organic solvents. *Journal of Bioscience and Bioengineering*, 91(2), 109- 116.
- Olivier-Bourbigou H. and Magna L. (2002) Ionic liquids: perspectives for organic and catalytic reactions. *Journal of Molecular Catalysis A: Chemical*, 182– 183, 419-437.
- Olivier-Bourbigou H., Magna L. and Morvan D. (2010) Ionic liquids and catalysis: Recent progress from knowledge to applications. *Applied Catalysis A: General*, (373), 1-56.
- Paljevac M., Habulin M. and Knez Ž. (2006) Ionic liquids as (co)solvents for enzymatic reactions. *Chemical Industry and Chemical Engineering Quarterly*, 12(3), 181-186.
- Olivier-Bourhigon H. and Magna L. (2002) Ionic liquids: perspectives for organic and catalytic reactions. *Journal of Molecular Citalyts A:* Chemical, 182<br>
183.419437.<br>
Olivier-Bourhigon H, Magna L, and Morvan D (2010) Io Paluch A. S., Vitter C. A., Shah J. K. and Maginn E. J. (2012) A comparison of the solvation thermodynamics of amino acid analogues in water, 1-octanol and 1-n-alkyl-3-methylimidazolium bis(trifluoromethylsulfonyl)imide ionic liquids by molecular simulation. *Journal of Chemical Physics*, 137(18), 184504-184515.
	- Park S. and Kazlauskas R. J. (2001) Improved preparation and use of room temperature ionic liquids in lipase-catalyzed enantio- and regioselective acylations. *Journal of Organic Chemistry*, 66, 8395-8401.
	- Park S. and Kazlauskas R. J. (2003) Biocatalysis in ionic liquids advantages beyond green technology. *Current Opinion in Biotechnology*, 14, 432-437.
	- Parrinello M. and Rahman A. (1981) Polymorphic transitions in single crystals: A new molecular dynamics method. *Journal of Applied Physics*, 52(12), 7182-7190.
	- Pauliukaite R., Doherty A. P., Murnaghan K. D. and Brett C. M. A. (2011) Application of room temperature ionic liquids to the development of electrochemical lipase biosensing systems for water-insoluble analytes. *Journal of Electroanalytical Chemistry*, 656(1–2), 96-101.
	- Persson M. and Bornscheuer U. T. (2003) Increased stability of an esterase from Bacillus stearothermophilus in ionic liquids as compared to organic solvents. *Journal of Molecular Catalysis B: Enzymatic*, 22(1–2), 21-27.
	- Peter W. and Wilhelm K. (2000) Ionic liquids—New "solutions" for transition metal catalysis. *Angewandte Chemie International Edition*, 39(21), 3772-3789.
	- Peters G. H., van Aalten D. M., Edholm O., Toxvaerd S. and Bywater R. (1996) Dynamics of proteins in different solvent systems: analysis of essential motion in lipases. *Biophysical Journal*, 71(5), 2245-2255.
- Pham T. P. T., Cho C.-W. and Yun Y.-S. (2010) Environmental fate and toxicity of ionic liquids: A review. *Water Research*, 44(2), 352-372.
- Poole C. F. (2004) Chromatographic and spectroscopic methods for the determination of solvent properties of room temperature ionic liquids. *Journal of Chromatography A*, 1037, 49-82.
- Radzicka A. and Wolfenden R. (1988) Comparing the polarities of the amino acids: Side-chain distribution coefficients between the vapor phase, cyclohexane, 1-octanol, and neutral aqueous solution. *Biochemistry*, 27, 1664-1670.
- Rantwijk F. v., Secundo F. and Sheldon R. A. (2006) Structure and activity of *Candida antarctica* lipase B in ionic liquids. *Green Chemistry*, 8, 282-286.
- Raza S., Fransson L. and Hult K. (2001) Enantioselectivity in *Candida antarctica* lipase B: A molecular dynamics study. *Protein Science*, 10, 329-338.
- Rìos A. P. d. l., Herńandez-Ferńandez F. J., Martinez F. A., Rubio M. and Vìllora G. (2007) The effect of ionic liquid media on activity, selectivity and stability of *Candida antarctica* lipase B in transesterification reactions. *Biocatalysis and Biotransformation*, 25, 151-156.
- Rogers R. D. and Seddon K. R. (2003) Ionic liquids--Solvents of the future? *Science*, 302(5646), 792-793.
- Radzicka A. and Wolfenden R. (1988) Comparing the polaities of the anino accels.<br>
1. and wolfficients between the vapor phase, eyelchexans,<br>
1. ectand, and neutral agenous solution. *Buckennisty*, 27, 1664-1670.<br>
Ramtwijk Ru M. T., Dordick J. S., Reimer J. A. and Clark D. S. (1999) Optimizing the saltinduced activation of enzymes in organic solvents: Effects of lyophilization time and water content. *Biotechnology and Bioengineering*, 63(2), 233-241.
	- Ryckaert J. P. and Bellemans A. (1975) Molecular dynamics of liquid n-butane near its boiling point. *Chemical Physics Letters*, 30(1), 123-125.
	- Sambasivarao S. V. and Acevedo O. (2009) Development of OPLS-AA force field parameters for 68 unique ionic liquids. *Journal of Chemical Theory and Computation*, 5(4), 1038-1050.
	- Sandoval M., Cortes A., Civera C., Trevino J., Ferreras E., Vaultier M., Berenguer J., Lozano P. and Hernaiz M. J. (2012) Efficient and selective enzymatic synthesis of N-acetyl-lactosamine in ionic liquid: a rational explanation. *RSC Advances*, 2(15), 6306-6314.
	- Schlick T. (2013) The 2013 Nobel Prize in Chemistry celebrates computations in chemistry and biology. *SIAM News*, 46(10), 1-4.
	- Schőfer S. H., Kaftzik N., Wasserscheid P. and U.Kragl. (2001) Enzyme catalysis in ionic liquids: lipase catalyzed kinetic resolution of 1-phenylethanol with vinyl acetate. *Chemical Communications*, 425-426.

Schrodinger, LLC. (2010). The PyMOL Molecular Graphics System, version 1.3r1.

- Schuler L. D., Daura X. and van Gunsteren W. F. (2001) An improved GROMOS96 force field for aliphatic hydrocarbons in the condensed phase. *Journal of Computational Chemistry*, 22(11), 1205-1218.
- Seddon K. R. (2003) Ionic liquids: A taste of the future. *Nature: Materials*, 2(6), 363-365.
- Seddon K. R., Stark A. and Torres M.-J. (2000) Influence of chloride, water, and organic solvents on the physical properties of ionic liquids. *Pure Applied Chemistry*, 72(12), 2275-2287.
- Shah J. K. and Maginn E. J. (2005) Monte Carlo simulations of gas solubility in the ionic liquid 1-n-butyl-3-methylimidazolium hexafluorophosphate. *The Journal of Physical Chemistry B*, 109(20), 10395-10405.
- Sheldon R. A., Lau R. M., Sorgedrager M. J., van Rantwijk F. and Seddon K. R. (2002) Biocatalysis in ionic liquids. *Green Chemistry*, 4(2), 147-151.
- Seddon K. R., Statk A. and Tores M.-1. (2000) Influence of coloride, water, and Consinical Cheemistry, 72(12), 2275-2287.<br>
Shuh J. K. and Maginn T. J. (2006) Monte Carlo simulations of gas solutions of the limit is limit Shimizu K., Almantariotis D., Gomes M. F. C., Pádua A. l. A. H. and Canongia Lopes J. N. (2010) Molecular force field for ionic liquids v: hydroxyethylimidazolium, dimethoxy-2- methylimidazolium, and fluoroalkylimidazolium cations and bis(fluorosulfonyl)amide, perfluoroalkanesulfonylamide, and fluoroalkylfluorophosphate anions. *The Journal of Physical Chemistry B*, 114(10), 3592-3600.
	- Shirts M. R. and Pande V. S. (2005) Solvation free energies of amino acid side chain analogs for common molecular mechanics water models. *Journal of Chemical Physics*, 122(13), 134508-134513.
	- Shirts M. R., Pitera J. W., Swope W. C. and Pande V. S. (2003) Extremely precise free energy calculations of amino acid side chain analogs: Comparison of common molecular mechanics force fields for proteins. *Journal of Chemical Physics*, 119(11), 5740-5761.
	- Simon G. M. and Cravatt B. F. (2010) Activity-based proteomics of enzyme superfamilies: Serine hydrolases as a case study. *Journal of Biological Chemistry*, 285(15), 11051-11055.
	- Singh T. and Kumar A. (2007) Aggregation behavior of ionic liquids in aqueous solutions: Effect of alkyl chain length, cations, and anions. *The Journal of Physical Chemistry B*, 111(27), 7843-7851.
	- Starzak M. (2010). Maxwell–Boltzmann distributions energy and entropy. Springer, New York, pp. 197-216.
	- Straatsma T. P. and Berendsen H. J. C. (1988) Free energy of ionic hydration: Analysis of a thermodynamic integration technique to evaluate free energy differences by molecular dynamics simulations. *Journal of Chemical Physics*, 89(9), 5876-5886.
	- Sun P. and Armstrong D. W. (2010) Ionic Liquids in analytical chemistry. *Analytica Chimica Acta*, 661, 1.
- Swatloski R. P., Spear S. K., Holbrey J. D. and Rogers R. D. (2002) Dissolution of cellose with ionic liquids. *Journal of American Chemical Society*, 124(18), 4974-4975.
- Swope W. C., Andersen H. C., Berens P. H. and Wilson K. R. (1982) A computer simulation method for the calculation of equilibrium constants for the formation of physical clusters of molecules: Applications to small water clusters. *Journal of Chemical Physics*, 76(1), 637-649.
- Tajima M., Urabe I., Yutani K. and Okada H. (1976) Role of calcium ions in the thermostability of Thermolysin and *Bacillus subtilis var. amylosacchariticus* neutral protease. *European Journal of Biochemistry*, 64(1), 243-247.
- Tejo B. A., Salleh A. B. and Pleiss J. (2004) Structure and dynamics of *Candida rugosa* lipase: the role of organic solvent. *Journal of Molecular Modeling*, 10, 358–366.
- Terranova Z. L. and Corcelli S. A. (2013) On the mechanism of solvation dynamics in imidazolium-based ionic liquids. *The Journal of Physical Chemistry B*, 117 (49), 15659–15666.
- Tokuda H., Hayamizu K., Ishii K., Susan M. A. B. H. and Watanabe M. (2005) Physicochemical properties and structures of room temperature ionic liquids. 2. Variation of alkyl chain length in imidazolium cation. *The Journal of Physical Chemistry B,* 109(13), 6103-6110.
- Tokuda H., Tsuzuki S., Susan M. A. B. H., Hayamizu K. and Watanabe M. (2006) How ionic are room-temperature ionic liquids? An indicator of the physicochemical properties. *The Journal of Physical Chemistry B*, 110, 19593-19600.
- clusters. Journal of Chemical Physics, 76(1), 637-649.<br>
Tajima M., Unabe I, Yutani K. and Okada H. (1976) Role of calcium ions in the<br>
thermostobitity of Thermolysin and Bacillus subilis vary<br>
amyloanceMeritiests nottral Tomé L. I. N., Jorge M., Gomes J. R. B. and Coutinho J. A. P. (2012) Molecular dynamics simulation studies of the interactions between ionic liquids and amino acids in aqueous solution. *The Journal of Physical Chemistry B*, 116(6), 1831-1842.
	- Tran C. D., De Paoli Lacerda S. H. and Oliveira D. (2003) Absorption of water by room-temperature ionic liquids: Effect of anions on concentration and state of water. *Applied Spectroscopy*, 57(2), 152-157.
	- Trodler P. and Pleiss J. (2008) Modeling structure and flexibility of *Candida antarctica* lipase B in organic solvents*. BMC Structural Biology*, 8, 9-18.
	- Turner M. B., Spear S. K., Huddleston J. G., Holbrey J. D. and Rogers R. D. (2003) Ionic liquid salt-induced inactivation and unfolding of cellulase from *Trichoderma reesei*. Green Chemistry, 5, 443-447.
	- Uppenberg J., Hansen M. T., Patkar S. and Jones T. A. (1994) The sequence, crystal structure determination and refinement of two crystal forms of lipase B from *Candida antarctica*. *Structure*, 2(4), 293-308.
- Uppenberg J., Ohrner N., Norin M., Hult K., Kleywegt G. J., Patkar S., Waagen V., Thorleif A. and Jones T. A. (1995) Crystallographic and molecularmodeling studies of lipase B from *Candida antarctica* reveal a stereospecificity pocket for secondary alcohols. *Biochemistry*, 34, 16838- 16855.
- Vakos H., Kaplan H., Black B., Dawson B. and Hefford M. (2000) Use of the pH memory effect in lyophilized proteins to achieve preferential methylation of α-amino groups. *Journal of Protein Chemistry*, 19(3), 231-237.
- van Gunsteren W. F. and Berendsen H. J. C. (1990) Computer simulation of molecular dynamics: Methodology, applications, and perspectives in chemistry. *Angewandte Chemie International Edition*, 29(9), 992-1023.
- van Gunsteren W. F., Berendsen H. J. C. and Rullmann J. A. C. (1981) Stochastic dynamics for molecules with constraints. *Molecular Physics*, 44(1), 69-95.
- Verlet L. (1967) Computer 'experiments' on classical fluids. I. Thermodynamical properties of Lennard-Jones molecules. *Physical Reviews*, 159, 98-103.
- Vidya P. and Chadha A. (2010) *Pseudomonas cepacia* lipase catalyzed esterification and transesterification of 3-(furan-2-yl) propanoic acid/ethyl ester: A comparison in ionic liquids vs hexane. *Journal of Molecular Catalysis B: Enzymatic*, 65(1–4), 68-72.
- Villa A. and Mark A. E. (2002) Calculation of the free energy of solvation for neutral analogs of amino acid side chains. *Journal of Computational Chemistry*, 23(5), 548-553.
- menoy effect in lyophilized proteins to achieve preiscential methodom<br>
of training upmy, Jonard of Protein Chemistry, 19(3), 231-237<br>
van Gunstert W. F. and Berendsen H. J. C. (1990) Computer simulation of<br>
molecular dyna Wahab R. A., Basri M., Rahman M. B. A., Rahman R. N. Z. R. A., Salleh A. B. and Leow T. C. (2012) Combination of oxyanion Gln114 mutation and medium engineering to influence the enantioselectivity of thermophilic lipase from *Geobacillus zalihae*. *International Journal of Molecular Sciences*, 13(9), 11666-11680.
	- Wang D., Bode W. and Huber R. (1985) Bovine chymotrypsinogen A X-ray crystal structure analysis and refinement of a new crystal form at 1.8 Å resolution. *Journal of Molecular Biology*, 185, 595-624.
	- Wang J., Wolf R. M., Caldwell J. W., Kollman P. A. and Case D. A. (2004) Development and testing of a general amber force field. *Journal of Computational Chemistry*, 25(9), 1157-1174.
	- Welton T. (1999) Room-temperature ionic liquids. Solvents for synthesis and catalysis. *Chemical Reviews*, 99, 2071-2083.
	- Wilkes J. S. (2002) A short history of ionic liquids-from molten salts to neoteric solvents. *Green Chemistry*, 4(2), 73-80.
- Wilkes J. S., Levisky J. A., Wilson R. A. and Hussey C. L. (1982) Dialkylimidazolium chloroaluminate melts: a new class of roomtemperature ionic liquids for electrochemistry, spectroscopy and synthesis. *Inorganic Chemistry*, 21(3), 1263-1264.
- Wohlgemuth R. (2010) Biocatalysis: key to sustainable industrial chemistry. *Current Opinion in Biotechnology*, 21(6), 713-724.
- Xuebing X., Bena-Marie L. and Zheng G. (2010) Effect of room temperature ionic liquid structure on the enzymatic acylation of flavonoids. *Process Biochemistry*, 45, 1375-1382.
- Yang L., Dordick J. S. and Garde S. (2004) Hydration of enzyme in nonaqueous media is consistent with solvent dependence of its activity. *Biophysical Journal*, 87(2), 812-821.
- Yang Z. and Pan W. (2005) Ionic Liquids: Green solvents for nonaqueous biocatalysis. *Enzyme and Microbial Technology*, 37, 19-28.
- Yang Z., Yue Y.-J., Huang W.-C., Zhuang X.-M., Chen Z.-T. and Xing M. (2009) Importance of the ionic nature of ionic liquids in affecting enzyme performance. *Journal of Biochemistry*, 145(3), 355-364.
- Zaks A. and Klibanov A. M. (1985) Enzyme-catalyzed processes in organic solvents. *Proceedings of the National Academy of Sciences*, 82(10), 3192-3196.
- Zaks A. and Klibanov A. M. (1988) The effect of water on enzyme action in organic media. *Journal of Biological Chemistry*, 263(17), 8017-8021.
- Xucbing X, Bona-Marie L and Zheng G, (2010) Effect of room temperature conical studients and studients of the energy and and studient of the energy of the monagrees  $\frac{1}{2}$  Yang I., Dordick J. S. and Garde S, (2004) Hyd Zhang N., Suen W. C., Windsor W., Xiao L., Madison V. and Zaks A. (2003) Improving tolerance of *Candida antarctica* lipase B towards irreversible thermal inactivation through directed evolution. *Protein Engineering*, 16(8), 599-605.
	- Zhang S., Sun N., He X., Lu X. and Zhang X. (2006) Physical properties of ionic liquids: Database and evaluation. *Journal of Physical Chemistry Reference Data*, 35(4), 1475-1517.
	- Zhao H. (2010) Methods for stabilizing and activating enzymes in ionic liquids-a review. *Journal of Chemical Technology & Biotechnology*, 85(7), 891- 907.
	- Zhao H., Olubajo O., Song Z., Sims A. L., Person T. E., Lawal R. A., Holley and LaDena A. (2006) Effect of kosmotropicity of ionic liquids on the enzyme stability in aqueous solutions. *Biooorganic Chemistry*, 34, 15-25.
	- Zhou T., Chen L., Ye Y., Chen L., Qi Z., Freund H. and Sundmacher K. (2012) An overview of mutual solubility of ionic liquids and water predicted by COSMO-RS. *Industrial & Engineering Chemistry Research*, 51(17), 6256- 6264.