

## **UNIVERSITI PUTRA MALAYSIA**

## INFLUENCE OF MEDIA, NITROGEN, INDIGENOUS MICROORGANISM AND WATER STRESS ON BIOACTIVE COMPOUNDS AND BIOMASS YIELD OF Andrographis palliculata (Burm.f.) WALL. EX NEES

# SHARA SALIH ALI

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By

SHARA SALIH ALI

Thesis Submitted to School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Master of Science

December 2014

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Master of Science

## INFLUENCE OF MEDIA, NITROGEN, INDIGENOUS MICROORGANISM AND WATER STRESS ON BIOACTIVE COMPOUNDS AND BIOMASS YIELD OF Andrographis paniculata (Burm.f.) WALL. EX NEES

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### SHARA SALIH ALI

December 2014

#### Chairman: Siti Zaharah Sakimin, PhD

#### **Faculty: Agriculture**

The current interest in reducing the herbal product importation by cultivating medicinal plants as Hempedu bumi, may have great impact on increasing the production of herbs in the local herbal industry. Growing plants under soilless media enable year round production of Hempedu bumi biomass. Although, some of soilless media (SM) components are cost effective and finding an alternative which is cheap and available locally in combination with indigenous microorganism might reduce applied rate of N chemical fertilizer. Besides the fact that organic and microbial amend d technique can enhance the growth of healthy herb, but also there is a need to increase the secondary metabolites in the pharmaceutical herb industry and this could be achieved by undergoing the plant through water stressed condition.

A preliminary study was carried out to determine the physical, chemical and microbiological properties of the soilless media that has used in the study which where; (coconut coir dust (CCD), empty fruit bunch compost (EFBC) and peat. Results of these study showed that the properties of some SM such as (EFBC) were suitable for the plant growth and development of *Andrographis paniculata*. In the first glass house experiment, five different combinations of SM were used as growing media which includes; C1 as control = CCD only (1: -), C2 = EFBC + CCD (7:3), C3 = EFBC + CCD (3:7), C4 = CCD + Peat (7:3), C5 = CCD + Peat (3:7) in combination with four nitrogen rates (NR) (0, 30, 60, and 90 kg ha<sup>-1</sup>) along with and without indigenous microorganism (IMO) application. The growth performance, physiology, biomass production of root and shoot, macronutrient content of leaf tissue and microbial populations in rhizosphere were measured at 30 and 60 day after planting (DAP). At 30 and 60 DAP with application of IMO the soilless media that gave highest growth performance parameter was under C3 media with fertilization of 60 kg N ha<sup>-1</sup> compared to control media C1 and other media C2, C4 and C5, plant

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height (25, 40.8 cm, respectively), number of leaves (121, 132 leaves plant<sup>-1</sup>) and total leaf area (122.8, 156.7 cm<sup>2</sup>), dry matter biomass (dry weight of shoot (3.1, 5.1 g), root (0.5, 1.1 g) and total dry biomass (3.6, 6.2 g)) physiological response (photosynthetic rate (9, 10  $\mu$ mol m<sup>-2</sup> s<sup>-2</sup>), stomatal conductance (410.4, 469.4 mmol m<sup>-2</sup> s<sup>-1</sup>) and transpiration rate (3.9, 4.2 mmol H<sub>2</sub>O m<sup>-2</sup> s<sup>-1</sup>)) and a decline in all parameter noticed at 90 kg N ha<sup>-1</sup>. With 90 kg N ha<sup>-1</sup> the parameters declined under C3 media in which plant height (19, 37.5 cm, respectively), number of leaves (74.8, 105.8 leaves plant<sup>-1</sup>) and total leaf area (88, 93.2 cm<sup>2</sup>), dry matter biomass (dry weight of shoot (1.7, 3.6 g), root (0.3, 0.9 g) and total dry biomass (2, 4.5 g)) physiological response (photosynthetic rate (8.5, 9  $\mu$ mol m<sup>-2</sup> s<sup>-2</sup>), stomatal conductance (234.8, 363 mmol m<sup>-2</sup> s<sup>-1</sup>) and transpiration rate (2.3, 2.6 mmol H<sub>2</sub>O m<sup>-2</sup> s<sup>-1</sup>)).

While without IMO application, the above mentioned parameters increased with increasing rate of N fertilizer to 90 kg N ha<sup>-1</sup> at both planting periods. The combined application of IMO and N fertilizer created positive impact on most of the plant character and total biomass yield of Hempedu bumi grown under C3 media compared to C1 (control). Despite of the fact that EFBC is slow in releasing nutrient but additional of IMO could positively enhance nutrient release from the compost and resulting in highest biomass production of the plant. In second experiment, two soilless media (C1 and C3) from previous studies were chosen base on their effects on the performance of plant growth, physiological parameter and biomass production to further determine their effect on bioactive compound of Andrographis paniculata under WS condition. Media C1 as control and C3 supplied with or without fertilizer. Plant with fertilizer was supplied with 60 kg N ha<sup>-1</sup>, 15 kg P ha<sup>-1</sup>, 20 kg K ha<sup>-1</sup> and no fertilizer at all was given to the plant without fertilizer. The plants was subjected into three WS levels which include; well watered (WW), moderate water stressed (MS) and severe stressed (SS). The plant received equal amounts of water twice a day for plant establishment and WS treatments were introduced 60 DAP for a period of 21 days. During WS treatments, the leaf relative water content (LRWC) and stomatal conductance  $(g_s)$  were recorded at 5 days interval and the leaf increments were measured at 3 days interval. Data for biomass production and bioactive compound was determined after 21 days of WS treatment. Results showed that the highest plant biomass production (dry weight of shoot (11.5 and root 1.4 g) was observed from plants grown in C3 media under WW with application of fertilizer. Averaged overall WS treatment with fertilizer application the mean number of leaves and total leaf area (TLA) of plant grown under media C3 exhibited 67% and 31% higher than that of plants grown in C1. While without fertilizer application the number of leaves and total leaf area (TLA) of plant grown in C3 media was 69% and 33% higher than that of plants grown under control media C1. The biochemical compound of the plant was significantly affected by WS condition in which the highest content of andrographolide (0.06  $\mu$ g mg<sup>-1</sup> dry leaf ), proline (3.4  $\mu$ mole g<sup>-1</sup> FW), total phenolic (282.4 (  $\mu g m g^{-1}$  extract) and flavonoid (381.8  $\mu g m g^{-1}$  extract) content were observed in SS plants grown under media C3. When averaged across different WS condition, the plant water status and chlorophyll content of plants grown under media C3 were adversely affected by stress condition when the result showed 16% and 57% higher compared to control either with or without fertilizer application, respectively In conclusion, results of this study suggest that the best media proportion was C3 media which consisted of 7: 3 of CCD: EFBC which gave best growth performance as well as biomass production of *Andrographis paniculata*. Application of IMO as an amendment to soilless media resulted in reducing the used amount of chemical N fertilizer in which the best biomass production was determined when the plant applied with 60 kg N ha<sup>-1</sup> and a decline in the biomass was observed with increasing N rate to 90 kg N ha<sup>-1</sup>. The biochemical content and the secondary metabolite of the plant increased with subjecting the plant to water stress condition. Overall, improvement of SM amended with IMO and maximizing active compound of the herb can be implemented to increase herbal industry in Malaysia.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

## PENGARUH MEDIA, NITROGEN, MIKROORGANISMA ASLI DAN TEKANAN AIR KE ATAS SEBATIAN BIOAKTIF DAN HASIL BIOMASS DALAM Andrographis paniculata (Burm.f.) WALL. EX NEES

Oleh

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#### **Disember 2014**

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Tarikan utama dalam usaha mengurangkan import produk herba ialah melalui usaha penanaman tanaman herba seperti Hempedu bumi, usaha ini akan memberi kesan kepada peningkatan produktiviti herba dalam pasaran tempatan. Proses penanaman dengan menggunakan kultur media tanpa tanah mampu menyediakan penghasilan Hempedu bumi sepanjang tahun. Walaupun sesetengah komponen media tanpa tanah memerlukan kos yang tinggi tetapi terdapat alternatif dalam mengurangkan kos iaitu dengan melibatkan penggunaan produk tempatan dengan kombinasi bersama mikroorganisma bermanfaat yang boleh mengurangkan kadar penggunakan baja kimia Selain daripada penerapan teknik organik dan penggunaan mikrob untuk meningkatkan pertumbuhan herba, terdapat juga teknik lain yang boleh digunakan untuk meningkatkan penghasilan metabolik sekunder dalam industri farmaseutikal herba iaitu melalui pendedahan tumbuhan kepada keadaan ketegangan air. Kajian preliminari telah dijalankan untuk mengenalpasti sifat-sifat fizikal, kimia dan mikrobiologi dalam media tanpa tanah dengan melibatkan sabut kelapa (CCD), kompos tandan buah kosong (EFBC) dan tanah gambut. Hasil kajian menunjukkan beberapa sifat SM seperti EFBC sesuai untuk pertumbuhan dan perkembangan Andrographis paniculata. Dalam eksperimen rumah kaca yang pertama, lima kombinasi SM telah digunakan sebagai media pertumbuhan iaitu; C1 sebagai kawalan = CCD sahaja (1:-), C2 = EFBC + CCD (7:3), C3 = EFBC + CCD (3:7), C4= CCD + Gambut (7:3), C5 = CCD + Gambut (3:7) dalam kombinasi kadar N yang berbeza (0,30, 60,90 kg ha<sup>-1</sup>) bersama dengan penggunaan IMO dan tanpa IMO. Kadar pertumbuhan, fisiologi, penghasilan biomass pucuk dan akar, kandungan micronutrient dan populasi rhizosfera diukur 30 dan 60 hari selepas menanam (DAP).

Pada 30 dan 60 DAP dengan penggunaan IMO, media yang menunjukkan kadar pertumbuhan yang tertinggi ialah media C3 dengan pembajaan sebanyak 60 kg ha<sup>-1</sup> berbanding dengan media kawalan C1, C2, C4 dan C5, peningkatan yang

ditunjukkan ialah ketinggian pertumbuhan (24, 40.8 cm), jumlah daun (121, 132 daun per pokok) dan jumlah luas daun (122.8, 156.7 cm<sup>2</sup>), berat kering biomass ( berat kering pucuk (3.1, 5.1 g), akar (0.5, 1.1 g) dan jumlah bahan kering (3.6, 6.2 g)), tindak balas fisiologi ((kadar fotosintesis (9, 10  $\mu$ mol m<sup>-2</sup> s<sup>-2</sup>), konduksi stomata  $(410.4, 469.4 \text{ mmol m}^{-2} \text{ s}^{-1})$  dan kadar transpirasi  $(3.9, 4.2 \text{ mmol } \text{H}_2\text{O} \text{ m}^{-2} \text{ s}^{-1}))$  dan semua parameter mengalami penurunan pada kadar baja 90 kg N ha<sup>-1</sup>. Dengan penggunaan baja 90 kg N ha<sup>-1</sup> dalam media C3 menunjukkan penurunan bagi ketinggian pokok (19, 37.5 cm), bilangan daun (74.8, 105.8 daun pokok<sup>-1</sup>) dan luas daun (88, 93.2 cm<sup>2</sup>), biomass bahan kering (berat kering pucuk (1.7, 3.6 g), akar (0.3, 0.9 g) dan jumlah berat biomass (2, 4.5 g)), manakala bagi respon fisiologi seperti (kadar fotosintesis (8.5, 9  $\mu$ mol m<sup>-2</sup> s<sup>-2</sup>), konduksi stomata (234.8, 363 mmol m<sup>-2</sup> s<sup>-1</sup>) dan kadar transpirasi (2.3, 2.6 mmol  $H_2O \text{ m}^{-2} \text{ s}^{-1}$ )). Walaupun tanpa kehadiran IMO, semua parameter yang dinyatakan diatas meningkat dengan penggunaan baja N sebanyak 90 kg N ha<sup>-1</sup> sepanjang dua musim penanaman. Gabungan aplikasi IMO dan baja N memberi kesan positif kepada kebanyakan pertumbuhan pokok dan jumlah penghasilan biomass Hempedu bumi yang ditanam dalam media C3 berbanding dengan media C1 (kawalan). Walaupun terdapat fakta yang menyatakan bahawa EFBC lambat melepaskan nutrien kepada tumbuhan tetapi dengan penambahan IMO ia menunjukkan kesan positif dalam meningkatkan pembebasan nutrient dari baja dan penghasilan biomass yang tinggi bagi setiap tanaman.

Dalam eksperimen kedua, dua media telah dipilih (C1 dan C3) daripada eksperimen sebelum ini berdasarkan peningkatan tindak balas dalam pertumbuhan pokok, fisiologi dan penghasilan biomass yang bertujuan untuk menentukan kesan media terhadap kandungan komposisi bioaktif dalam Andrographis paniculata dalm keadaan WS. Media C1 sebagai kawalan and C3 digunakan dengan aplikasi baja dan tanpa pembajaan. Tumbuhan yang diberikan baja ialah dengan kadar sebanyak 60 kg N ha<sup>-1</sup>, 15 kg P ha<sup>-1</sup>, 20 kg K ha<sup>-1</sup> dan tiada baja diberikan kepada pokok dalam kategori tanpa pembajaan. Tumbuhan diletakkan dalam 3 keadaan WS iaitu; disiram (WW), tegangan air sederhana (MS) dan tegangan air yang serius (SS). Sepanjang tempoh pengenalan rawatan, tanaman menerima jumlah air yang sama banyak iaitu dua kali sehari dan rawatan WS diberikan selepas 60 hari dalam tempoh 21 hari. Sepanjang rawatan WS, kandungan relatif air dalam daun dan konduksi stomata  $(g_s)$ dicatatkan selang 5 hari manakala pertambahan saiz daun diukur selang 3 hari. Data untuk penghasilan biomas dan komposisi bioaktif ditentukan selepas 21 hari rawatan. Keputusan menunjukkan penghasilan biomass yang tertinggi (berat kering pucuk (11.5 dan akar 1.4 g) oleh tumbuhan yang ditanam dalam media C3 dibawah WS dengan kombinasi aplikasi pembajaan. Purata keseluruhan rawatan WS dengan aplikasi baja menunjukkan min bilangan daun dan jumlah luas daun (TLA) bagi tumbuhan yang ditanam dalam media C3 menunjukkan peningkatan 67% dan 31% lebih tinggi daripada tumbuhan yang ditanam dalam media C1. Manakala rawatan tanpa pembajaan menunjukkan bilangan daun dan jumlah luas daun bagi tumbuhan yang ditanam dalam media C3 menunjukkan peningkatan 69% dan 33% lebih tinggi daripada tumbuhan yang ditanam dalam media C1. Sebatian kimia dalam tumbuhan turut menunjukkan kesan yang ketara dalam keadaan dimana jumlah tertinggi andrographolide (0.06  $\mu$ g mg<sup>-1</sup> daun kering ), proline (3.4  $\mu$ mole g<sup>-1</sup> FW), jumlah phenolic (282.4 (  $\mu$ g mg<sup>-1</sup> extrak) dan kandungan flavonoid (381.8  $\mu$ g mg<sup>-1</sup> extrak) direkodkan bagi tumbuhan yang ditanam dalam media C3. Dalam purata keadaan WS yang berbeza, status kandungan air pokok dan klorofil daun bagi pokok yang ditanam dalam media C3 menunjukkan kesan tekanan yang berbeza seperti yang dicatatkan iaitu 16% dan 57% lebih tinggi daripada media kawalan sama ada dengan atau tanpa pembajaan.

Kesimpulannya , hasil kajian ini mencadangkan bahawa media C3 yang terdiri daripada 7: 3 daripada CCD: EFBC telah memberikan peningkatan pertumbuhan yang terbaik dan penghasilan biomass tertinggi bagi *Andrographis paniculata*. Penggunaan sebagai perapi media tanpa tanah menunjukkan pengurangan dalam kadar penggunaan baja N dimana penghasilan jumlah biomas yang tertinggi dicatatkan oleh tumbuhan yang menggunakan 60 kg N ha<sup>-1</sup> dan penghasilan biomas meunjukkan penurunan apabila kadar baja N dinaikkan kepada 90 kg N ha<sup>-1</sup>. Kandungan sebatian kimia dan metabolit sekunder bagi tumbuhan ini meningkatan dengan kenaikan kadar tegangan air yang dikenakan.secara keseluruhannya, penggunaan SM dengan kombinasi IMO boleh ditingkatkan penggunaannya unuk penghasilan komposisi bahan aktif yang maksimum bagi meningkatkan industri herba di Malaysia.

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#### 7 GENERAL CONCLUSION

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## LIST OF SYMBOLS AND ABBREVATIONS

- × Multiply/interaction between
- + Plus
- Minus
- > Greater than
- < Less than
- $\geq$  Greater than or equal to
- $\leq$  Less than or equal to
- ± Plus minus
- / Divide
- = Equal to
- ° Degree
- °C Degree celcious
- % Per cent
- $\lambda$  Lambda
- μg Microgram(s)
- μL Microlitre(s)
- μM Micromolar(s)
- μm Micrometre(s)
- µmol Micromole(s)
- ABA Abscisic acid
- ADP Adenosine diphosphate
- AlCl<sub>3</sub> Aluminum Trichloride
- AM Arbuscular mycorrhizal
- ANOVA Analysis of variance
- AU Absorbance units

	С	Carbon
	Ca	Calcium
	CaCl2	Calcium chloride
	CCD	Coconut coir dust
	CEC	Cation exchange capacity
	Cfu	Colony forming unit
	$CO_2$	Carbon dioxide / respiration
	conc.	Concentration
	CIRP	Christmas Island rock phosphate
	d	Day(s)
	DAP	Days after planting
	dH <sub>2</sub> O	Distilled water
	DPPH	2, 2-diphenyl-1-picryl-hydrazyl
	Dr.	Doctor
	EC	Electrical conductivity
	EFBC	Empty fruit bunch compost
	EM	Effective microorganism
	FFTC	Food and fertilizer technology center
	g	Gram(s)
	GA	Gibberellic acid
	GA	Gallic Acid
	h	Hour(s)
	Н	Hydrogen
	ha	Hectare(s)
	HCl	Hydrochloric acid
	$H_2O$	Water

$H_2O_2$	Hydrogen peroxide
$H_2SO_4$	Sulfuric acid
IMO	Indigenous microorganism
INM	Integrated nutrient management
Κ	Potasium
kg	Kilogram(s)
KOH	Potassium hydroxide
KNF	Korean Natural Farming
L	Liter(s)
LI	Leaf increment
Log	Logarithm
m	Metre(s)
М	Molar
mAU	Milliabsorbance unit
MeOH	Methanol
mg	Milligram(s)
Mg	Magnesium
min	Minute(s)
mL	Millilitre(s)
mm	Millimetre(s)
mM	Millimolar(s)
mmol	Millimole(s)
MPOB	Malaysian palm oil board
Ν	Nitrogen
NaCl	Sodium chloride
NaF	Sodium fluoride

C

NaHSO <sub>3</sub>	Sodium hydrogen sulphite		
NaOH	Sodium hydroxide		
NaOCl	Sodium hypochlorite		
NO <sub>3</sub>	Nitrate		
NR	Nitrogen rate		
NS	Not significant		
Pa	Pascals		
$O_2$	Oxygen		
OPT	Oil palm trunk		
Р	Probability		
Р	Peat		
Р	Phosphorus		
рН	Symbol denoting hydrogen ion in a solution		
PGPR	Plant growth promoting rhizobacteria		
POME	Palm oil mill effluent		
Prof.	Professor		
PSB	Phosphorus solubilizing bacteria		
RD	Root diameter		
RDW	Root dry weight		
RH	Relative humidity		
RL	Root length		
RLWC	Relative leaf water content		
RM	Malaysian ringgit		
RSA	Root surface area		
RV	Root volume		
R: S	Root to shoot ratio		

0

RU	Rutin
8	Second(s)
SAS	Statistical Analysis System
SDW	Shoot dry weight
S.E.	Standard error
sp.	Species
SM	Soilless media
Std	Standard
UFLC	Ultra-fast liquid chromatography
UK	United Kingdom
USA	United States of America
UV	Ultra-violet
Vol.	Volume(s)
v/v	Volume by volume
viz.	Videlicet (namely)
WS	Water stress
w/w	Weight by weight

#### CHAPTER 1

#### **INTRODUCTION**

In Malaysia, Andrographis paniculata is known as 'Hempedu bumi', which means 'bile of the earth'. Commonly it is known as 'king of bitter' since it is one of the bitterest herbs that is regularly used in traditional medicine. Before antibiotics were invented Indian people used this plant for natural ailment remedy (Kumar, Dora, Singh, & Tripathi, 2012). The pharmaceutical value of this plant is in the aerial part and the root which have a very bitter taste due to the existence of active ingredient viz. andrographolide, 14-deoxy-11, 12 didehydroandrographolide and neoandrographolide. However, maximum amount of these compounds contained in mature leaves of the plant (Suriyo et al., 2014). Whole plant parts traditionally used for the treatment of hepatitis, cough, fever, mouth ulcers, sores, venomous snake bites, common cold, and urinary tract infections. It is also discovered that the plant has anticancer and immune stimulatory (Bhattacharya, Puri, Jamwal, & Sharma, 2012). Besides applications of aerial part of the plant for liver disease, dysentery and common colds, it also applied in the therapy of hypertension and diabetes mellitus (Agarwal, Sulaiman, & Mohamed, 2005; Wiart, 2002; Zhang & Tan, 1996). Study have found that the methanolic extract of Andrographis paniculata have high potential as an antiviral activity on dengue fever (Tang, Ling, Koh, Chye, & Voon, 2012).

In Malaysia the production of traditional and herbal medicine product is expands as a result of increased dependent on the import of raw materials for the local herbal industry which is an economic scarcity. Charles et al. (1993) reported that in 2009 the trading of herbal based products in Malaysia reached RM777 billion which is expected to triple by 2020. Among the valuable herbal plants in high demands in Peninsular Malaysia is 'Hempedu bumi' which has enormous pharmaceutical utilization. Therefore, it would be justifiable to increase or enhance the production of safety products in the local herbal industry by using new cultivation technology of organic based soilless media. Soilless media is an artificial method of providing plants with support and a reservoir for nutrients and water. There are very limited types of organic soilless media that can be used as potting media for plant production such as peat, coconut coir dust (CCD) and inorganic substrate such as rockwool and perlite. Currently the environmental and ecological concerns against using peat as a growth substrate is increasing due to its contribution in increasing global warming gases (Kip et al., 2010). Moreover, the rising costs for peat as a growing media in horticulture have led to search for high quality and cost effective alternative for peat (Saffigna, Powlson, Brookes, & Thomas, 1989). The price of peat is increasing because they have to be imported (Dash & Mohanty, 2001). However the price of CCD is reasonable in comparison with peat but still there is a need to find out other substrates that are more reasonable. Oil palm is the most essential product in Malaysia and empty fruit bunch (EFB) is a raw by-product of palm oil that can be utilized as a growing media to be used in agriculture industries. Recycling and managing these residue of oil palm is targeted to reduce the major disposal problem and recover the energy (Wiart, 2002). Empty fruit bunch compost (EFBC) is one of the by-products that generated from palm oil which is rich in organic matter and nutrient content (Caliman, Hardianto, & Ng, 2001). The application of EFBC as media enrichment increased the nutrient uptake and yield of oil palm in palm oil plantation (Zaharah & Lim, 2000). Hence, as an alternative, empty fruit bunch compost (EFBC) probably can be fully utilized as a growing media.

The characteristic of growing media is one of the important factors that affect the growth performance of the plant because plant requires sufficient support, nutrients and moisture from the media in which they are grown. Use of this organic substrate as potting media could reduce the decline in fertilizer availability, reduce the rate of excessive and continues usage of these fertilizers which contaminate the environment and disturbs the quality of soil and increase economic return to the farmer (Agarwal et al., 2005). However soilless media may have good water holding and aeration properties, but can be limited on their capability to hold nutrients. Fertility management of this media is extremely important so that, the nutrient levels should be optimized in the growing media prior to planting in order to gain maximum crop growth. Balance among the essential plant nutrients, especially potassium, calcium, and magnesium is important. The application of inorganic and organic source of nutrient to increase the biomass production of plant is called integrated nutrient management (INM), a technique which modifies the plant nutrient supply to an optimum level. The highest growth parameters, nutrient uptake, yield and andrographolide content of Andrographis paniculata were observed when cultivated under INM system (Abdullah & Sulaiman, 2013).

The nutrient release of EFBC is slow during high nutrients appeal at peak time and in order to release nutrients from EFBC, microbial assisted biodegradation should be increased. Application of microbial component such as indigenous microorganism (IMO) could increase the nutrient emission from the compost (Abdul Mutalib, 2009). The IMO is a beneficial microorganism with the ability of releasing nutrient from the media. Due to the availability of filamentous fungi, yeasts and bacteria, IMO have many important functions which are fermentation, decomposition and N fixation (Jensen, Guilaran, Jaranilla, & Garingalao, 2006). In addition, effect on the microbial biomass and growing media health when combination of organic and chemical fertilizer is applied has been reported earlier (Dutta, Pal, Chakeraborty, & Chakrabarti, 2003).

As mentioned earlier due to the availability of many active compounds in 'Hempedu bumi' the plant is important as a pharmaceutical herb. Increase in the secondary metabolite or active compounds of the plant reached the demand of safety plant production for human health as well as environment. The productivity of the secondary metabolite in the plant can be enhanced by subjecting the plant through abiotic stress condition. Although, drought reduce the growth performance of the plant (Flexas, Bota, Loreto, Cornic, & Sharkey, 2004) but with application of appropriate fertilizer and microbial consortium the adverse effect of water stress on growth of plant could be alleviated. In addition, drought is one of the most essential abiotic stress factors in order to enhance the biochemical components of the plants (Zobayed, Afreen, & Kozai, 2007). It has been reported that water stress or drought stress increased the production of active compound in many medicinal plant such as in *Artemisia annua* (Charles et al., 1993), *Hypericum perforatum* (Zobayed et al., 2007). In addition, adjusting plants to drought stressed condition will increase water used efficiency and water availability for human supply. Researcher have reported that the agricultural irrigation used over 70% of the world supplies of clean water and most of the water is especially used in the protected environments (Ali, Ismail, Mohd Saud, & Manan, 2004). By incorporating with IMO, the amendment of EFBC in soilless media is hoping to provide a sustainable growing media which can be used to increase the growth performance of 'Hempedu bumi'.

The current interest in reducing the herbal product importation by cultivating medicinal plants as 'Hempedu bumi', may have great impact on increasing the production of herbs in the local herbal industry. Growing plants under soilless media enable year round production of Hempedu bumi biomass. Although, some of soilless media (SM) components are cost effective and finding an alternative which is cheap and available locally in combination with indigenous microorganism might reduce applied rate of N chemical fertilizer. Besides the fact that organic and microbial amend d technique can enhance the growth of healthy herb, but also there is a need to increase the secondary metabolites in the pharmaceutical herb industry and this could be achieved by undergoing the plant through water stressed condition. Traditionally choices of soilless media that are available in the market are limited mainly to peat (Ramahsamy, Bakar, Abdullah, & Ishak, 2012a) and coconut (Mazuela, Urrestarazu, & Bastias, 2012). Although, EFBC have been used in agriculture but very little work has been reported on using EFBC as an alternative soilless media which increase recycling residue of oil palm and leads to use locally available residue. It has been reported that the empty fruit bunch compost (EFBC) increased nutrient uptake and yield of palm oil plantation when used as cultivation media (Zaharah & Lim, 2000). Abdul Mutalib (2009) reported that EFBC has great potential to be used in organic cabbage production replacing inorganic fertilizer. The choices of combination of soilless media have great effect on the plant growth as in previous study reported by Wira, Razi, and Jamil (2011a) which claimed that combination of EFC and CCD soilless media in 3:7, v/v ratio gave highest shoot biomass of rock melon while, combination of CCD and peat in 7:3, v/v ratio gave lowest shoot biomass compared to 100% CCD as control media.

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However, studies on the application of IMO with soilless media to decrease the applied rate of N fertilizer and their effect on the biomass of Hempedu bumi have not been done. To add to that, study on the effect of water stress on the secondary metabolite of Hempedu bumi has not been done yet. In view on the potential benefit of EFBC as growing media it was therefore hypothesized that the cultivation of 'Hempedu bumi' under different combination of soilless media (EFBC, CCD and peat) with application of indigenous microorganism (IMO) and different nitrogen rates enhance the growth performance, as well as the secondary metabolite of the plant might improves by subjecting the plant to water stress condition. Therefore, the aims of this research work were:

1. To determine the physical, chemical and microbiological properties of soilless media before planting.

2. To determine the best soilless media, optimum rate of N fertilizer and effect of IMO application on the growth and biomass production of *Andrographis paniculata* grown under different combination of soilless.

3. To evaluate the effect of water stress on the biomass and bioactive compound of *Andrographis paniculata* grown under selected soilless media and optimum level of N fertilizer.



#### REFERENCES

- Abad, M., Fornes, F., Carrión, C., Noguera, V., Noguera, P., Maquieira, Á., & Puchades, R. (2005). Physical properties of various coconut coir dusts compared to peat. *Horticulture Science* 40(7), 2138-2144.
- Abdalla, J. A. (2005). Genetic Variation and Anticancer Activity of Adrographis Paniculata Germplasm from Malaysia (Master Thesis). Universiti Putra Malaysia.
- Abdalla, M., & El-Khoshiban, N. (2007). The influence of water stress on growth, relative water content, photosynthetic pigments, some metabolic and hormonal contents of two Triticium aestivum cultivars. *Journal of Applied Sciences Research*, *3*(12), 2062-2074.
- Abdul Aziz, H. (2007). Reactive Extraction Of Sugars From Oil Palm Empty Fruit Bunch Hydrolysate Using Naphthalene-2-Boronic Acid (Master Thesis). Universiti Sains Malaysia.
- Abdul Mutalib, F. a. M., Fatkhiah and H., Siti Aishah. (2009). Effects of Empty Fruit Bunch (EFB) Compost and Indigenous Microbes on Growth Performance of Cabbage (*Brassica oleracea* var. Capitata). In: 20th Malaysian Society of Plant Physiology Conference (MSPPC 2009), 24-26 July 2009, Avillion Admiral Cove, Port Dickson, Negeri Sembilan.Abdullah, N., & Sulaiman, F. (2013). *The Oil Palm Wastes in Malaysia, Biomass Now Sustainable Growth and Use. 2013*.
- Adam, F., & Barakbah, S. (1990). Response to water stress in banana, peanut and rice a comparative study. *Transaction of the Malaysian Society of Plant Physiology*, 1, 99-104.
- Agarwal, R., Sulaiman, S. A., & Mohamed, M. (2005). Open label clinical trial to study adverse effects and tolerance to dry powder of the aerial part of Andrographis paniculata in patients type 2 with diabetes mellitus. *The Malaysian Journal of Medical Sciences: MJMS*, 12(1), 13.
- Ahmad, A., Ismail, M. R., Yusop, M. K., Mahmood, M., & Mohd, S. (2004). Physical and Chemical properties of coconut coir dust and oil palm empty fruit bunch and the growth of hybrid heat tolerant cauliflower plant. *Pertanika Journal of Tropical Agricultural Science*, 27(2), 121-133.
- Aisueni, N., & Omoti, U. (2001). The role of compost in sustainable oil palm production. Paper presented at the Cutting-edge technologies for sustained competitiveness: Proceedings of the 2001 PIPOC International Palm Oil Congress, Agriculture Conference, Kuala Lumpur, Malaysia, 20-22 August 2001.
- Ajaya Kumar, R., Sridevi, K., Vijaya Kumar, N., Nanduri, S., & Rajagopal, S. (2004). Anticancer and immunostimulatory compounds from Andrographis paniculata. Journal of Ethnopharmacology, 92(2), 291-295.

- Akbar, S. (2011). Andrographis paniculata: a review of pharmacological activities and clinical effects. Alternative Medicine Review, 16(1), 66-77.
- Akowuah, G., Zhari, I., Norhayati, I., & Mariam, A. (2006). HPLC and HPTLC densitometric determination of andrographolides and antioxidant potential of *Andrographis paniculata*. Journal of Food Composition and Analysis, 19(2), 118-126.
- Al Rawahy, M., Al Raisy, F., & Al Makhmari, S. (2009). Evaluation of cucumber in different culture media under soilless growing technique (open system) in non cooled screenhouse conditions. *Acta Horticulturae*, 807, 481-484.
- Alexieva, V., Sergiev, I., Mapelli, S., & Karanov, E. (2001). The effect of drought and ultraviolet radiation on growth and stress markers in pea and wheat. *Plant, Cell & Environment, 24*(12), 1337-1344.
- Ali, H. I., Ismail, M. R., Mohd Saud, H., & Manan, M. M. (2004). Effect of Partial Rootzone Drying (PRD) on Growth, Water Use Efficiency (WUE) and Yield of Tomatoes Grown in Soilless Culture. *Pertanika Journal of Tropical Agricultural Science*, 27(2), 143-149.
- Allam MZ, H. S., 2007. (2007). Accumulation of Protein, Chlorophyll and Relative Leaf Water Content in Barley (*Hordeum Vulgare L.*) In Relation to Sowing Time and Nitrogen Fertilizer. *Research Journal of Agricultural and Biological Science*, 3(3), 4.
- Allen, R. D., Webb, R. P., & Schake, S. A. (1997). Use of transgenic plants to study antioxidant defenses. *Free Radical Biology and Medicine*, 23(3), 473-479.
- Amar Bahadura, D. P. S., Pratibha Singhb, J.S. Singhb, U. P. Singh. (2012). Phenolic Constituents of Centella asiatica and Andrographis paniculata. Archives of Applied Science Research, 4(6), 2424-2426.
- Amujoyegbe, B., Opabode, J., & Olayinka, A. (2007). Effect of organic and inorganic fertilizer on yield and chlorophyll content of maize (Zea mays L.) and sorghum Sorghum bicolour (L.) Moench). African Journal of Biotechnology, 6(16).
- Angadi, S., & Entz, M. (2002). Water relations of standard height and dwarf sunflower cultivars. *Crop Science*, 42(1), 152-159.
- Angle, J. S., Gagliardi, J. V., McIntosh, M., & Levin, M. (1996). Enumeration and expression of bacterial counts in the rhizosphere. *Soil and Biochemistry*. *Marcel Dekker, Inc. NY*, 233-251.
- Aref, I., El Atta, H., Khan, P., Iqbal, M., El Obeid, M., & Ahmed, A. (2013). Effect of Water Stress on Relative Water and Chlorophyll Contents of Juniperus Procera Hochst. Ex Endlicher in Saudi Arabia. *Life Science Journal*, 10(4).
- Arpana, J., & Bagyaraj, D. (2007). Response of Kalmegh to an arbuscular mycorrhizal fungus and a plant growth promoting rhizomicroorganism at two

levels of phosphorus fertilizer. *American-Eurasian Journal of Agricultural & Environmental Science*, 2, 33-38.

- Asada, K. (1999). The water-water cycle in chloroplasts: scavenging of active oxygens and dissipation of excess photons. *Annual review of plant biology*, 50(1), 601-639.
- Asami, D. K., Hong, Y.-J., Barrett, D. M., & Mitchell, A. E. (2003). Comparison of the total phenolic and ascorbic acid content of freeze-dried and air-dried marionberry, strawberry, and corn grown using conventional, organic, and sustainable agricultural practices. *Journal of Agricultural and Food Chemistry*, 51(5), 1237-1241.
- Ashok, K., Amit, A., Sujatha, M., Murali, B., & Anand, M. (2002). Effect of aging on andrographolide content in *Andrographis paniculata*. *Journal of Natural Remedies*, 2(2), 179-181.
- Ashraf, M., Azmi, A., Khan, A., & Ala, S. (1994). Effect of water stress on total phenols, peroxidase activity and chlorophyll content in wheat [*Triticum aestivum* L.]. Acta Physiologiae Plantarum, 16(3).
- Atiyeh, R. M., Edwards, C. A., Subler, S., & Metzger, J. D. (2000). Earthwormprocessed organic wastes as components of horticultural potting media for growing marigold and vegetable seedlings. *Compost Science & Utilization*, 8(3), 215-223.
- Augé, R. M. (2001). Water relations, drought and vesicular-arbuscular mycorrhizal symbiosis. *Mycorrhiza*, 11(1), 3-42.
- Awang, Y., Shaharom, A. S., Mohamad, R. B., & Selamat, A. (2009). Chemical and Physical Characteristics of Cocopeat-Based Media Mixtures and Their Effects on the Growth and Development of Celosia cristata. American Journal of Agricultural and Biological Sciences, 4(1), 63.
- Aziz, R. (2003). Turning Malaysia Into A Global Herbal Producer: A Personal Perspective. Kuala Lumpur, Malaysia: University Technology Malaysia, Professional Inaugural Lecture Series.
- Bachman, G., & Metzger, J. (2008). Growth of bedding plants in commercial potting substrate amended with vermicompost. *Bioresource Technology*, 99(8), 3155-3161.
- Bacon, M. A., Wilkinson, S., & Davies, W. J. (1998). pH-regulated leaf cell expansion in droughted plants is abscisic acid dependent. *Plant Physiology*, *118*(4), 1507-1515.
- Bahari, Z., Pallardy, S., & Parker, W. (1985). Photosynthesis, water relations, and drought adaptation in six woody species of oak-hickory forests in central Missouri. *Forest Science*, *31*(3), 557-569.

- Balazová, A., Bilka, F., Blanáriková, V., & Psenák, M. (2002). Effect of a fungal elicitor on levels of sanguinarine and polyphenoloxidase activity in a suspension culture of *Papaver somniferum* L. Ceska a Slovenska farmacie: casopis Ceske farmaceuticke spolecnosti a Slovenske farmaceuticke spolecnosti, 51(4), 182-185.
- Baloğlu, M. C., Kavas, M., Aydin, G., Öktem, H. A., & Yücel, A. M. (2012). Antioxidative and physiological responses of two sunflower (*Helianthus annuus*) cultivars under PEG-mediated drought stress. *Turkish Journal of Botany*, 36(6).
- Banerjee, A., Datta, J., & Mondal, N. (2012). Biochemical changes in leaves of mustard under the influence of different fertilizers and cycocel. *Journal of Agricultural Science and Technlogy*, 8(4), 1397-1411.
- Barea, J., Gryndler, M., Lemanceau, P., Schiiepp, H., & Azcon, R. (2002). The rhizosphere of mycorrhizal plants. *Mycorrihizal Technology in Agriculture: From Genes to Bioproducts*, 1.
- Barry, P., Evershed, R. P., Young, A., Prescott, M. C., & Britton, G. (1992). Characterization of carotenoid acylesters produced in drought-stressed barley seedlings. *Phytochemistry*, *31*(9), 3163-3168.
- Bates, L., Waldren, R., & Teare, I. (1973). Rapid determination of free proline for water-stress studies. *Plant and Soil*, 39(1), 205-207.
- Begg, J., Turner, N., & Kramer, P. (1980). Morphological adaptations of leaves to water stress. Adaptation of Plants to Water and High Temperature Stress., 33-42.
- Begg, J. E., & Turner, N. C. (1976). Crop water deficits. *Advances in agronomy*, 28(161217.1976).
- Bhan, M., Dhar, A., Khan, S., Lattoo, S., Gupta, K., & Choudhary, D. (2006). Screening and optimization of *Andrographis paniculata* (Burm. f.) Nees for total andrographolide content, yield and its components. *Scientia Horticulturae*, 107(4), 386-391.
- Bhattacharya, S., Puri, S., Jamwal, A., & Sharma, S. (2012). Studies on Seed Germination and Seedling Growth in Kalmegh (*Andrographis paniculata* Wall. Ex Nees) under Abiotic Stress Conditions. *International Journal of Science, Environment and Technology, 1*(3), 7.
- Bímová, P., & Pokluda, R. (2009). Impact of organic fertilizers on total antioxidant capacity in head cabbage. *Horticultural Science*, *36*, 21-25.
- Blackman, P., & Davies, W. (1985). Root to shoot communication in maize plants of the effects of soil drying. *Journal of Experimental Botany*, *36*(1), 39-48.

- Blum, A., & Ebercon, A. (1976). Genotypic responses in sorghum to drought stress.
   III. Free proline accumulation and drought resistance. *Crop Science*, 16(3), 428-431.
- Bokhtiar, S., & Sakurai, K. (2005). Effects of organic manure and chemical fertilizer on soil fertility and productivity of plant and ratoon crops of sugarcane. *Archives of Agronomy and Soil science*, *51*(3), 325-334.
- Boutraa, T., & Sanders, F. (2001). Effects of interactions of moisture regime and nutrient addition on nodulation and carbon partitioning in two cultivars of bean (*Phaseolus vulgaris* L.). Journal of Agronomy and Crop Science, 186(4), 229-237.
- Boyer, J. (1976). Water deficits and photosynthesis. *Water deficits and plant growth*, 4, 153-190.
- Brouwer, R. (1963). Some aspects of the equilibrium between overground and underground plant parts. Jaarboek van het Instituut voor Biologisch en Scheikundig onderzoek aan Landbouwgewassen, 1963, 31-39.
- Caliman, J., Hardianto, J., & Ng, M. (2001). Strategy for fertilizer management during low commodity price. Paper presented at the Cutting-edge technologies for sustained competitiveness: Proceedings of the 2001 PIPOC International Palm Oil Congress, Agriculture Conference, Kuala Lumpur, Malaysia, 20-22 August 2001.
- Carter Jr, J., & Patterson, R. (1985). Use of relative water content as a selection tool for drought tolerance in soybean. Paper presented at the Fide Agron abstract 77th Annual Meeting.
- Cechin, I., & de Fátima Fumis, T. (2004). Effect of nitrogen supply on growth and photosynthesis of sunflower plants grown in the greenhouse. *Plant Science*, *166*(5), 1379-1385.
- Chao, W.-W., & Lin, B.-F. (2010). Review Isolation and identification of bioactive compounds in *Andrographis paniculata* (Chuanxinlian). *growth*, 10, 44.
- Charles, D. J., Simon, J. E., Shock, C. C., Feibert, E. B., Smith, R. M., Janick, J., & Simon, J. (1993). *Effect of water stress and post-harvest handling on artemisinin content in the leaves of Artemisia annua L.* Paper presented at the Proceedings of the second national symposium: New crops, Exploration, Research and Commercialization.
- Chen, Y., Guo, Q., Liu, L., Liao, L., & Zhu, Z. (2011). Influence of fertilization and drought stress on the growth and production of secondary metabolites in *Prunella vulgaris* L. *Journal of Medicinal Plant Research*, 5(9), 1749.
- Chiramel, T., Bagyaraj, D., & Patil, C. (2006). Response of Andrographis paniculata to different arbuscular mycorrhizal fungi. Journal of Agricicultural Technology, 2(2), 221-228.

- Cho, H. K., & Koyama, A. (1997). Korean natural farming.
- Chong, T. M., Abdullah, M. A., Lai, O. M., Nor'Aini, F. M., & Lajis, N. H. (2005). Effective elicitation factors in *Morinda elliptica* cell suspension culture. *Process Biochemistry*, 40(11), 3397-3405.
- Coombs, J., Hall, D. O., Long, S., & Scurlock, J. (1985). *Techniques in bioproductivity and photosynthesis*: Pergamon Press.
- Cornic, G. (2000). Drought stress inhibits photosynthesis by decreasing stomatal aperture–not by affecting ATP synthesis. *Trends in plant science*, 5(5), 187-188.
- Cruz, J., Mosquim, P., Pelacani, C., Araújo, W., & DaMatta, F. (2003). Photosynthesis impairment in cassava leaves in response to nitrogen deficiency. *Plant and Soil*, 257(2), 417-423.
- Daneshmand, A., Shiranirad, A. H., Nourmohammadi, G., Zareei, G., & Daneshian, J. (2008). Effect of water deficit and different nitrogen rates on yield, yield components and physiological traits of two rapeseed (*Brassica napus* L.) cultivars. *Journal of Agricultural Sciences and Natural Resources*.
- Darlington, W. (2005). Compost-A Guide for evaluating and using compost materials as soil amendments.
- Dash, S., & Mohanty, N. (2001). Evaluation of assays for the analysis of thermotolerance and recovery potentials of seedlings of wheat (*Triticum aestivum* L.) cultivars. *Journal of Plant Physiology*, 158(9), 1153-1165.
- Davies, W. J., Bacon, M. A., Thompson, D. S., Sobeih, W., & Rodríguez, L. G. (2000). Regulation of leaf and fruit growth in plants growing in drying soil: exploitation of the plants' chemical signalling system and hydraulic architecture to increase the efficiency of water use in agriculture. *Journal of Experimental Botany*, 51(350), 1617-1626.
- Davies, W. J., & Zhang, J. (1991). Root signals and the regulation of growth and development of plants in drying soil. *Annual review of Plant Biology*, 42(1), 55-76.
- De Boodt, M., & Verdonck, O. (1971). The physical properties of the substrates in horticulture. *Acta Horticulture*, 26, 37-44.
- De Brito, A., Gagne, S., & Antoun, H. (1995). Effect of compost on rhizosphere microflora of the tomato and on the incidence of plant growth-promoting rhizobacteria. *Applied and Environmental Microbiology*, *61*(1), 194-199.
- De Souza, C. R., Maroco, J. P., dos Santos, T. P., Rodrigues, M. L., Lopes, C. M., Pereira, J. S., & Chaves, M. M. (2003). Partial rootzone drying: regulation of stomatal aperture and carbon assimilation in field-grown grapevines (*Vitis vinifera cv. Moscatel*). *Functional Plant Biology*, 30(6), 653-662.

- Demmig-Adams, B., & Adams Iii, W. (1992). Photoprotection and other responses of plants to high light stress. *Annual review of plant biology*, 43(1), 599-626.
- Dhanda, S., & Sethi, G. (2002). Tolerance to drought stress among selected Indian wheat cultivars. *The Journal of Agricultural Science*, 139(03), 319-326.
- Dietz, K.-J., & Harris, G. (1996). Photosynthesis under nutrient deficiency. *Handbook of Photosynthesis*.
- Dos Santos, T. P., Lopes, C. M., Rodrigues, M. L., de Souza, C. R., Maroco, J. P., Pereira, J. S., Chaves, M. M. (2003). Partial rootzone drying: effects on growth and fruit quality of field-grown grapevines (*Vitis vinifera*). *Functional Plant Biology*, 30(6), 663-671.
- Douds Jr, D., Galvez, L., Franke-Snyder, M., Reider, C., & Drinkwater, L. (1997). Effect of compost addition and crop rotation point upon VAM fungi. *Agriculture, Ecosystems & Environment, 65*(3), 257-266.
- Dubey, R., & Pessarakli, M. (2002). Physiological mechanisms of nitrogen absorption and assimilation in plants under stressful conditions. *Handbook of Plant and Crop Physiology. 2nd ed. Marcel Dekker, NY*, 637-655.
- Dutta, S., Pal, R., Chakeraborty, A., & Chakrabarti, K. (2003). Influence of integrated plant nutrient phosphorus and sugarcane and sugar yields. *Field Crop Research*, 77, 43-49.
- Efeoğlu, B., Ekmekci, Y., & Cicek, N. (2009). Physiological responses of three maize cultivars to drought stress and recovery. *South African Journal of Botany*, 75(1), 34-42.
- Efthimiadou, A., Bilalis, D., Karkanis, A., Froud-Williams, B., & Eleftherochorinos, I. (2009). Effects of cultural system (organic and conventional) on growth, photosynthesis and yield components of sweet corn (*Zea mays L.*) under semi-arid environment. *Notulae Botanicae Horticulture Agrobotanici Cluj-Napoca*, 37(2), 104-111.
- Esen, A. H. S., Özgür, R., Uzilday, B., Tanyolaç, Z. Ö., & Dinc, A. (2012). The response of the xerophytic plant *Gypsophila aucheri* to salt and drought stresses: the role of the antioxidant defence system. *Turkish Journal of Botany*, 36(6).
- Eskling, M., Arvidsson, P. O., & Åkerlund, H. E. (1997). The xanthophyll cycle, its regulation and components. *Physiologia Plantarum*, *100*(4), 806-816.
- Farahzety, A., & Aishah, H. S. (2013). Effects of organic fertilizers on performance of cauliflower (*Brassica oleracea* var. botrytis) grown under protected structure. *Journal of Tropical Agriculture and Food Scince Trop*, 41(1), 15-25.

- Farooq, M., Wahid, A., Kobayashi, N., Fujita, D., & Basra, S. (2009). Plant drought stress: effects, mechanisms and management *Sustainable Agriculture* (pp. 153-188): Springer.
- Farquhar, G. D., Ehleringer, J. R., & Hubick, K. T. (1989). Carbon isotope discrimination and photosynthesis. *Annual Review of Plant Biology*, 40(1), 503-537.
- Farquhar, G. D., & Sharkey, T. D. (1982). Stomatal conductance and photosynthesis. *Annual Review of Plant Physiology*, 33(1), 317-345.
- Flexas, J., Bota, J., Loreto, F., Cornic, G., & Sharkey, T. (2004). Diffusive and metabolic limitations to photosynthesis under drought and salinity in C3 plants. *Plant Biology*, 6(3), 269-279.
- Fotovat, R., Valizadeh, M., & Toorchi, M. (2007). Association between water-use efficiency components and total chlorophyll content (SPAD) in wheat (*Triticum aestivum* L.) under well-watered and drought stress conditions. *Journal of Food Agriculture and Environment*, 5(3/4), 225.
- Gardner, F. P., Pearce, R. B., & Mitchell, R. L. (2003). *Physiology of crop plants* (Vol. 8).
- Garside, A., Lawn, R., & Byth, D. (1992). Irrigation management of soybean (*Glycine max* (L.) Merrill) in a semi-arid tropical environment. I. Effect of irrigation frequency on growth, development and yield. *Crop and Pasture Science*, 43(5), 1003-1017.
- Ghosh, K. B., Datta, K. A., Mandal, A., Dubey, K. P., & Halder, S. (2012). An Overview On Andrographis paniculata (Burm. F.) Nees. International Journal of Research in Ayurveda & Pharmacy, 3(5).
- Girija, C., Smith, B., & Swamy, P. (2002). Interactive effects of sodium chloride and calcium chloride on the accumulation of proline and glycinebetaine in peanut (*Arachis hypogaea* L.). *Environmental and experimental botany*, 47(1), 1-10.
- Gopal, M. S. S. a. P. D. V. R. S. (2013). Studies on Indigenous Microorganisms (IMOs) increasing Growth of Leaves Germination, Chlorophyll content and Differentiation between IMOs and Chemical Fertilizers in various crop plants. *International Journal of Emerging Technologies in Computational and Applied Sciences (IJETCAS)*, 6.
- Gowing, D., Davies, W., & Jones, H. (1990). A positive root-sourced signal as an indicator of soil drying in apple, Malus x domestica Borkh. *Journal of Experimental Botany*, 41(12), 1535-1540.
- Griffiths, R. I., Whiteley, A. S., O'Donnell, A. G., & Bailey, M. J. (2003). Physiological and community responses of established grassland bacterial populations to water stress. *Applied and Environmental Microbiology*, 69(12), 6961-6968.

- Grillas, S., Lucas, M., Bardopolou, E., Sarafopoulos, S., & Voulgari, M. (2001). Perlite based soilless culture systems: Current commercial applications and prospects. Acta Horticulturae, 105-114.
- Gruda, N. (2012). Do soilless culture systems have an influence on product quality of vegetables? *Journal of Applied Botany and Food Quality*, 82(2), 141-147.
- Gullo, M., & Salleo, S. (1988). Different strategies of drought resistance in three Mediterranean sclerophyllous trees growing in the same environmental conditions. *New Phytologist*, 108(3), 267-276.
- Hale, B. K., Herms, D. A., Hansen, R. C., Clausen, T. P., & Arnold, D. (2005). Effects of drought stress and nutrient availability on dry matter allocation, phenolic glycosides, and rapid induced resistance of poplar to two lymantriid defoliators. *Journal of Chemical Ecology*, 31(11), 2601-2620.
- Hare, P., Cress, W., & Van Staden, J. (1998). Dissecting the roles of osmolyte accumulation during stress. *Plant, Cell & Environment, 21*(6), 535-553.
- Harris, D. G. (1973). Photosynthesis, diffusion resistance and relative plant water content of cotton as influenced by induced water stress. *Crop Science*, 13(5), 570-572.
- Harris, R. W. (1992). Root-shoot ratios. Journal of Arboriculture, 18(1), 39-42.
- Hashemimajd, K., & Jamaati-e-Somarin, S. (2011). Investigating the effect of iron and zinc enriched vermicompost on growth and nutritional status of peach trees. *Scientific Research and Essays*, 6(23), 5004-5007.
- Hayatu, M., & Mukhtar, F. (2010). Physiological responses of some drought resistant cowpea genotypes (Vigna unguiculata (L.) Walp) to water stress. Bayero Journal of Pure and Applied Sciences, 3(2), 69-75.
- Haynes, R. (1980). Competitive aspects of the grass-legume association. Advances in agronomy, 33, 227-261.
- Heinstein, P. F. (1985). Future approaches to the formation of secondary natural products in plant cell suspension cultures. *Journal of Natural Products*, 48(1), 1-9.
- Herbinger, K., Tausz, M., Wonisch, A., Soja, G., Sorger, A., & Grill, D. (2002). Complex interactive effects of drought and ozone stress on the antioxidant defence systems of two wheat cultivars. *Plant Physiology and Biochemistry*, 40(6), 691-696.
- Holaday, A. S., Martindale, W., Alred, R., Brooks, A. L., & Leegood, R. C. (1992). Changes in activities of enzymes of carbon metabolism in leaves during exposure of plants to low temperature. *Plant physiology*, 98(3), 1105-1114.
- Holford, I. (1997). Soil phosphorus: its measurement, and its uptake by plants. *Australian Journal of Soil Research*, 35(2), 227-240.

- Holman, J., Bugbee, B., & Chard, J. (2005). A Comparison of Coconut Coir and Sphagnum Peat as Soil-less Media Components for Plant Growth. Utah State University. Department of Plants, Soils and Biometeorology.
- Hsiao, T., Fereres, E., Acevedo, E., & Henderson, D. (1976). Water stress and dynamics of growth and yield of crop plants *Water and plant life* (pp. 281-305): Springer.
- Hsiao, T. C. (1973). Plant responses to water stress. Annual review of plant physiology, 24(1), 519-570.
- Hura, T., Hura, K., & Grzesiak, S. (2009). Possible contribution of cell-wall-bound ferulic acid in drought resistance and recovery in triticale seedlings. *Journal* of Plant Physiology, 166(16), 1720-1733.
- Iberahim, I. (2001). Improvement of Soilless Media by Incorporation of Trichoderma Inoculant for the Production of Cauliflower. Universiti Putra Malaysia.
- Ibrahim, M. H., & Jaafar, H. Z. (2011). Photosynthetic capacity, photochemical efficiency and chlorophyll content of three varieties of Labisia pumila Benth. exposed to open field and greenhouse growing conditions. *Acta Physiologiae Plantarum*, 33(6), 2179-2185.
- Ibrahim, M. H., Jaafar, H. Z., Karimi, E., & Ghasemzadeh, A. (2013). Impact of Organic and Inorganic Fertilizers Application on the Phytochemical and Antioxidant Activity of Kacip Fatimah (*Labisia pumila* Benth). *Molecules*, 18(9), 10973-10988.
- Ismail, M. R. (1995). Growth, physiological processes and yield of tomatoes grown in different root zone volumes using sand culture. *Pertanika Journal of Tropical Agricultural Science*, 18(2), 141-147.
- Ismail, M. R., Abdul Aziz, M., & Hasim, T. (1994). Growth, Water Relations and Physiological Changes of Young Durian (Durio zibenthinus Murr) as Influenced by Water Availability. Pertanika Journal of Tropical Agricultural Science, 17(3), 149-156.
- Ismail, M. R., Saud, H. M., Othman, R., Habib, S., Kausar, H., & Naher, L. (2013). Effect of oil palm frond compost amended coconut coir dust soilless growing media on growth and yield of cauliflower. *International Journal of Agriculture and Biology*, *15*(4), 731-736.
- Iturbe-Ormaetxe, I., Escuredo, P. R., Arrese-Igor, C., & Becana, M. (1998). Oxidative damage in pea plants exposed to water deficit or paraquat. *Plant Physiology*, *116*(1), 173-181.
- Jagtap, V., Bhargava, S., Streb, P., & Feierabend, J. (1998). Comparative effect of water, heat and light stresses on photosynthetic reactions in *Sorghum bicolor* (L.) Moench. *Journal of Experimental Botany*, 49(327), 1715-1721.

- Jaleel, A., Manivannan, P., Sankar, B., Kishorekumar, A., Gopi, R., Somasundaram, R., & Panneerselvam, R. (2007). Pseudomonas fluorescens enhances biomass yield and ajmalicine production in *Catharanthus roseus* under water deficit stress. *Colloids and Surfaces B: Biointerfaces*, 60(1), 7-11.
- Jaleel, C. A., Manivannan, P., Wahid, A., Farooq, M., Al-Juburi, H. J., Somasundaram, R., & Panneerselvam, R. (2009). Drought stress in plants: a review on morphological characteristics and pigments composition. *International Journal of Agriculture and Biology*, 11(1), 100-105.
- Jaleel, C. A., Sankar, B., Murali, P., Gomathinayagam, M., Lakshmanan, G., & Panneerselvam, R. (2008). Water deficit stress effects on reactive oxygen metabolism in *Catharanthus roseus*; impacts on ajmalicine accumulation. *Colloids and Surfaces B: Biointerfaces*, 62(1), 105-111.
- Jensen, H., Guilaran, L., Jaranilla, R., & Garingalao, G. (2006). Nature Farming Manual. *Retrieved on January*, 27, 2010.
- Jiang, M., & Zhang, J. (2002). Water stress induced abscisic acid accumulation triggers the increased generation of reactive oxygen species and up regulates the activities of antioxidant enzymes in maize leaves. *Journal of Experimental Botany*, 53(379), 2401-2410.
- Johnson, R., Nguyen, H., & Croy, L. (1984). Osmotic adjustment and solute accumulation in two wheat genotypes differing in drought resistance. *Crop Science*, 24(5), 957-962.
- Jones, M. B., & Lazenby, A. (1988). *The grass crop. The physiological basis of production*: Chapman and Hall.
- Joseph Nda Ainika, E. B. A., Christina Omoleye Olonitola, Patrick Chiweta Okutu, Emmanuel Yusuf Dodo. (2012). Effect of Organic and Inorganic Fertilizer on Growth and Yield of *Amaranthus caudatus* L. in Northern Guinea Savanna of Nigeria. *World Journal of Engineering and Pure and Applied Science*, 2(2), 26.
- Kala, D., Rosenani, A., Fauziah, C., & Thohirah, L. (2009). Composting oil palm wastes and sewage sludge for use in potting media of ornamental plants. *Malaysian Journal of Soil Science*, 13, 77-91.
- Kang, S., Liang, Z., Hu, W., & Zhang, J. (1998). Water use efficiency of controlled alternate irrigation on root-divided maize plants. *Agricultural Water Management*, 38(1), 69-76.
- Kao, C. H. (1981). Senescence of rice leaves VI. Comparative study of the metabolic changes of senescing turgid and water-stressed excised leaves. *Plant and cell Physiology*, 22(4), 683-688.
- Karkanis, A., Bilalis, D., & Efthimiadou, A. (2007). The effect of green manure and irrigation on morphological and physiological characteristics of Virginia

(flue-cured) organic tobacco (*Nicotiana tabacum*). International Journal of Agricultural Research, 2(11), 910-919.

- Karthick Raja, N. (2012). Effect of Compost Derived From Decomposed Fruit Wastes by Effective Microorganism (EM) Technology on Plant Growth Parameters of Vigna mungo. Journal of Bioremediation and Biodegradation.
- Kaschuk, G., Kuyper, T. W., Leffelaar, P. A., Hungria, M., & Giller, K. E. (2009). Are the rates of photosynthesis stimulated by the carbon sink strength of rhizobial and arbuscular mycorrhizal symbioses? *Soil Biology and Biochemistry*, 41(6), 1233-1244.
- Kavitha, B., Jothimani, P., & Rajannan, G. (2013). Empty fruit bunch- a potential organic manure for agriculture. *International Journal of Science*, *Environment*, 2(5), 930-937.
- Kennedy, A., & Smith, K. (1995). Soil microbial diversity and the sustainability of agricultural soils. *Plant and soil*, 170(1), 75-86.
- Kent, A. D., & Triplett, E. W. (2002). Microbial communities and their interactions in soil and rhizosphere ecosystems. *Annual Reviews in Microbiology*, 56(1), 211-236.
- Khaddar, V., & Yadav, S. (2006). Effect of integrated nutrient management practices on soil microbial population in a soybean-wheat cropping sequence. *Journal* of Environmental Research And Development 1(2).
- Khaliq, A., Abbasi, M. K., & Hussain, T. (2006). Effects of integrated use of organic and inorganic nutrient sources with effective microorganisms (EM) on seed cotton yield in Pakistan. *Bioresource Technology*, 97(8), 967-972.
- Kharadi, R., Upadhyaya, S., Upadhyay, A., & Sagar, N. P. (2011). Differential responses of plumbagin content in *Plumbago zeylanica* L.(Chitrak) under controlled water stress treatments. *Journal of Stress Physiology & Biochemistry*, 7, 114-121.
- Khonje, D., Varsa, E., & Klubek, B. (1989). The acidulation effects of nitrogenous fertilizers on selected chemical and microbiological properties of soil. *Communications in Soil Science & Plant Analysis*, 20(13-14), 1377-1395.
- Kip, N., van Winden, J. F., Pan, Y., Bodrossy, L., Reichart, G.-J., Smolders, A. J., den Camp, H. J. O. (2010). Global prevalence of methane oxidation by symbiotic bacteria in peat-moss ecosystems. *Nature Geoscience*, 3(9), 617-621.
- Kirakosyan, A., Kaufman, P., Warber, S., Zick, S., Aaronson, K., Bolling, S., & Chul Chang, S. (2004). Applied environmental stresses to enhance the levels of polyphenolics in leaves of hawthorn plants. *Physiologia Plantarum*, 121(2), 182-186.

- Kirda, C., Cetin, M., Dasgan, Y., Topcu, S., Kaman, H., Ekici, B., . . . Ozguven, A. (2004). Yield response of greenhouse grown tomato to partial root drying and conventional deficit irrigation. *Agricultural Water Management*, 69(3), 191-201.
- Kogbe, J., & Adediran, J. (2004). Influence of nitrogen, phosphorus and potassium application on the yield of maize in the savanna zone of Nigeria. *African Journal of Biotechnology*, 2(10), 345-349.
- Kozlowski, T., & Pallardy, S. (2002). Acclimation and adaptive responses of woody plants to environmental stresses. *The Botanical Review*, 68(2), 270-334.
- Kramer, P. J. (1986). The role of physiology in forestry. *Tree Physiology*, 2(1-2-3), 1-16.
- Kulshreshtha, S., Mishra, D., & Gupta, R. (1987). Changes in contents of chlorophyll, proteins and lipids in whole chloroplasts and chloroplast membrane fractions at different leaf water potentials in drought resistant and sensitive genotypes of wheat. *Photosynthetica (Czechoslovakia)*.
- Kumar, A., Dora, J., Singh, A., & Tripathi, R. (2012). A review of king of bitter (Kalmegh) International Journal of Research in Pharmacy an Chemistry 2(1), 116-124.
- Kumar, S., & Kumar, A. (2013). Spatial and harvesting influence on growth, yield, quality and economic potential of Kalmegh (Andrographis paniculata Wall Ex. Nees). Journal of Agriculture and Rural Development in the Tropics and Subtropics 114(1), 69-76.
- Kyparissis, A., Petropoulou, Y., & Manetas, Y. (1995). Summer survival of leaves in a soft-leaved shrub (*Phlomis fruticosa* L., Labiatae) under Mediterranean field conditions: avoidance of photoinhibitory damage through decreased chlorophyll contents. *Journal of Experimental Botany*, 46(12), 1825-1831.
- Lawn, R. (1982). Response of four grain legumes to water stress in south-eastern Queensland. I. Physiological response mechanisms. *Crop and Pasture Science*, 33(3), 481-496.
- Leonard, D., & Corps, P. (1986). Soils, Crops, and Fertilizer Use: A Field Manual for Development Workers: Peace Corps, Information Collection & Exchange.
- Li, L., & Van Staden, J. (1998). Effects of plant growth regulators on the antioxidant system in callus of two maize cultivars subjected to water stress. *Plant Growth Regulation*, 24(1), 55-66.
- Liang, B., & MacKenzie, A. (1995). Effect of fertilization on organic and microbial biomass nitrogen using15N under corn (*Zea mays* L.) in two Quebec soils. *Fertilizer research*, 44(2), 143-149.

- Liew Voon Kheong, Z. A. R., Mohamed Hanafi Musa, Aminudin Hussein. (2010). Nutrient absorption by oil palm primary roots as affected by empty fruit bunch application. *Journal of Oil Palm Research*, 22, 711-720.
- Lima, J., Mosquim, P., & Da Matta, F. (1999). Leaf gas exchange and chlorophyll fluorescence parameters in *Phaseolus vulgaris* as affected by nitrogen and phosphorus deficiency. *Photosynthetica*, *37*(1), 113-121.
- Liu, F., & Stützel, H. (2004). Biomass partitioning, specific leaf area, and water use efficiency of vegetable amaranth in response to drought stress. *Scientia Horticulturae*, *102*(1), 15-27.
- Liu, H., Wang, X., Wang, D., Zou, Z., & Liang, Z. (2011). Effect of drought stress on growth and accumulation of active constituents in *Salvia miltiorrhiza* Bunge. *Industrial crops and products*, 33(1), 84-88.
- Loewenstein, N. J., & Pallardy, S. G. (1998). Drought tolerance, xylem sap abscisic acid and stomatal conductance during soil drying: a comparison of young plants of four temperate deciduous angiosperms. *Tree physiology*, *18*(7), 421-430.
- Lugojan, C., & Ciulca, S. (2011). Evaluation of relative water content in winter wheat. *Journal of Horticulture, Forestry and Biotechnology*, 15(2), 173-177.
- Lugtenberg, B., & Kamilova, F. (2009). Plant-growth-promoting rhizobacteria. Annual review of microbiology, 63, 541-556.
- Luvaha, E., Netondo, G., & Ouma, G. (2008). Effect of water deficit on the physiological and morphological characteristics of mango (*Mangifera indica*) rootstock seedlings. *American Journal of Plant Physiology*, 3(1), 1-15.
- Lyngwi, N. A., Koijam, K., Sharma, D., & Joshi, S. (2013). Cultivable bacterial diversity along the altitudinal zonation and vegetation range of tropical Eastern Himalaya. *Revista de Biologia Tropical* 61(1), 467-490.
- Mafakheri, A., Siosemardeh, A., Bahramnejad, B., Struik, P., & Sohrabi, Y. (2010). Effect of drought stress on yield, proline and chlorophyll contents in three chickpea cultivars. *Australian Journal of Crop Science*, 4(8), 580-585.
- Maggio, A., Miyazaki, S., Veronese, P., Fujita, T., Ibeas, J. I., Damsz, B., Bressan, R. A. (2002). Does proline accumulation play an active role in stress induced growth reduction? *The Plant Journal*, *31*(6), 699-712.
- Majumdar, S., Ghosh, S., Glick, B. R., & Dumbroff, E. B. (1991). Activities of chlorophyllase, phosphoenolpyruvate carboxylase and ribulose1, 5 bisphosphate carboxylase in the primary leaves of soybean during senescence and drought. *Physiologia Plantarum*, 81(4), 473-480.
- Manivannan, P., Jaleel, C. A., Sankar, B., Kishorekumar, A., Somasundaram, R., Lakshmanan, G. A., & Panneerselvam, R. (2007). Growth, Biochemical

modifications and proline metabolism in *Helianthus annuus* L. as induced by drought stress. *Colloids and Surfaces B: Biointerfaces, 59*(2), 141-149.

- Marcelis, L., Heuvelink, E., & Goudriaan, J. (1998). Modelling biomass production and yield of horticultural crops: a review. *Scientia Horticulturae*, 74(1), 83-111.
- Marschner, H., & Rimmington, G. (1988). Mineral nutrition of higher plants. *Plant, Cell and Environment, 11*, 147-148.
- Marti, H. R., & Mills, H. A. (1991). Nutrient uptake and yield of sweet pepper as affected by stage of development and N form. *Journal of Plant Nutrition*, 14(11), 1165-1175.
- Matcha, S. K. (2007). Effects of Nitrogen Deficiency on Plant Growth, Leaf Photosynthesis, and Hyperspectral Reflectance Properties in Castor (Ricinus Communis L.). Mississippi State University.
- Matin, M., Brown, J. H., & Ferguson, H. (1989). Leaf water potential, relative water content, and diffusive resistance as screening techniques for drought resistance in barley. *Agronomy Journal*, *81*(1), 100-105.
- Mazuela, P., Urrestarazu, M., & Bastias, E. (2012). Vegetable waste compost used as substrate in soilless culture. *Crop Production Technologies. Rijeka, Croazia, In Tech Europe*, 180-198.
- Mediavilla, S., & Escudero, A. (2004). Stomatal responses to drought of mature trees and seedlings of two co-occurring Mediterranean oaks. *Forest Ecology and Management*, 187(2), 281-294.
- Meerow, A. W. (1994). Growth of two subtropical ornamentals using coir (coconut mesocarp pith) as a peat substitute. *Horticultural Science*, 29(12), 1484-1486.
- Merah, O. (2001). Potential importance of water status traits for durum wheat improvement under Mediterranean conditions. *The Journal of Agricultural Science*, 137(02), 139-145.
- Mia, M., Shamsuddin, Z., Zakaria, W., & Marziah, M. (2000). Growth and physiological attributes of hydroponically-grown bananas inoculated with plant growth promoting rhizobacteria. Transac. Malaysian Soc. *Plant Physiology*, *9*, 324-327.
- Miller, J. H., & Jones, N. (1995). Organic and compost-based growing media for tree seedling nurseries: World Bank Publications.
- Mills and Jones, J. J. B. (1996). *Plant Analysis* (Vol. 2). Ghana: Macromicro Publishing, Athens.
- Mishra, S., & Jain, A. (2014). Effect of INM on Vegetative Growth, Flowering and Fruiting of Andrographis paniculata. Universal Journal of Agricultural Research 2(3), 93-96.

- Mishra, S., Tiwari, S., Kakkar, A., & Pandey, A. (2010). Chemoprofiling of *Andrographis paniculata* (kalmegh) for its andrographolide content in Madhya Pradesh, India. *International Journal of Pharma and Bio Sciences*
- Mokhtari, S. (2010). Use of Organic Enrichment as Additives in Coconut Coir Dust Soilles Culture for Growth and Yield of Tomatoes (Lycopersicon Esculentum Mill. cv. Beril). Universiti Putra Malaysia.
- Molitor, H.-D., & Brückner, U. (1997). Waste paper-a substitute for peat in horticulture. Acta Horticulturae, 450, 47-56.
- Moreno, J., Garcia, C., Landi, L., Falchini, L., Pietramellara, G., & Nannipieri, P. (2001). The ecological dose value (ED 50) for assessing Cd toxicity on ATP content and dehydrogenase and urease activities of soil. *Soil Biology and Biochemistry*, 33(4), 483-489.
- MPOB. (2013). Malaysia Palm Oil Board, from http://www.mpob.gov.my
- Munns, R. (2002). Comparative physiology of salt and water stress. *Plant, Cell & Environment, 25*(2), 239-250.
- Nacif de Abreu, I., & Mazzafera, P. (2005a). Effect of water and temperature stress on the content of active constituents of *Hypericum brasiliense* Choisy. *Plant Physiology and Biochemistry*, 43(3), 241-248.
- Nacif de Abreu, I., & Mazzafera, P. (2005b). Effect of water and temperature stress on the content of active constituents of *Hypericum brasiliense* Choisy. *Plant Physiology and Biochemistry*, 43(3), 241-248.
- Nageswara Rao, R., Talwar, H., & Wright, G. (2001). Rapid assessment of specific leaf area and leaf nitrogen in peanut (*Arachis hypogaea* L.) using a chlorophyll meter. *Journal of Agronomy and Crop Science*, 186(3), 175-182.
- Naher, U. A., Othman, R., & Panhwar, Q. A. (2013). Culturable total and beneficial microbial occurrences in long-term nutrient deficit wetland rice soil. *Australian Journal of Crop Science*, 7(12), 1848.
- Namdeo, A., Patil, S., & Fulzele, D. P. (2002). Influence of fungal elicitors on production of ajmalicine by cell cultures of *Catharanthus roseus*. *Biotechnology progress*, 18(1), 159-162.
- Navari-Izzo, F., & Rascio, N. (1999). Plant response to water-deficit conditions. *Handbook of plant and crop stress*, 2.
- Neil C, T. (1979). Drought resistance and adaptation to water deficits in crop plants. *Stress Physiology in Crop Plants. Eds. H. Mussel and RC Staples. Wiley, New York*, 344-372.
- Neil C, T. (1986). Crop water deficits: a decade of progress. *Advances in agronomy*, *39*.

- Neil NC, T. (1997). Further progress in crop water relations. *Advances in agronomy*, 58.
- Niranjan, A., Tewari, S., & Lehri, A. (2010). Biological activities of Kalmegh (*Andrographis paniculata* Nees) and its active principles-a review. *Indian Journal of Natural Products and Resources* 1(2), 125-135.
- Noguera, P., Abad, M., Noguera, V., Puchades, R., & Maquieira, A. (1998). Coconut coir waste, a new and viable ecologically-friendly peat substitute. Paper presented at the XXV International Horticultural Congress, Part 7: Quality of Horticultural Products 517.
- Noorhanin, D., Puteri Edaroyati, M.W. \*and Siti Aishah, H. (2013). Response of Different Media Ratio on Growth and Biomass Production of Andrographis paniculata (Hempedu Bumi) Grown under Soilless Culture System. Transaction of the Malaysian Society of Plant Physiology, 21.
- Norman, J. C. (2004). Tropical Horticulture. *National Science and Technology Press Ghana*, 23-32.
- Nyachiro, J., Briggs, K., Hoddinott, J., & Johnson-Flanagan, A. (2001). Chlorophyll content, chlorophyll fluorescence and water deficit in spring wheat. *Cereal Research Communications*, 29(1-2), 135-142.
- Ommen, O., Donnelly, A., Vanhoutvin, S., Van Oijen, M., & Manderscheid, R. (1999). Chlorophyll content of spring wheat flag leaves grown under elevated CO<sub>2</sub> concentrations and other environmental stresses within the 'ESPACE-wheat' project. *European Journal of Agronomy*, 10(3), 197-203.
- Orhan, E., Esitken, A., Ercisli, S., Turan, M., & Sahin, F. (2006). Effects of plant growth promoting rhizobacteria (PGPR) on yield, growth and nutrient contents in organically growing raspberry. *Scientia Horticulturae*, 111(1), 38-43.
- Osuagwu, G., & Edeoga, H. (2012). Effect of inorganic fertilizer application on the flavonoid, phenol and steroid content of the leaves of *Ocimum gratissimum* (L) and Gongronema latifolium (Benth). *International Journal of Medicinal and Aromatic Plants*, 2(2), 254-262.
- Otake, T., Mori, H., Morimoto, M., Ueba, N., Sutardjo, S., Kusumoto, I. T., Namba, T. (1995). Screening of Indonesian plant extracts for anti human immunodeficiency virus type 1 (HIV1) activity. *Phytotherapy Research*, *9*(1), 6-10.
- Parashar, R., Upadhyay, A., Singh, J., Diwedi, S. K., & Khan, N. A. (2011). Morphophysiological evaluation of *Andrographis paniculata* at different growth stages. *World Journal of Agricultural Science*, 7(2), 124-127.

- Parchmann, S., Gundlach, H., & Mueller, M. J. (1997). Induction of 12-oxophytodienoic acid in wounded plants and elicited plant cell cultures. *Plant Physiology*, 115(3), 1057-1064.
- Parvin, S. (2007). Response of kalmegh (Andrographis paniculata) to nitrogen and cowdung (Master Thesis). Master, The Bangabandhu Sheikh Mujibur Rahman Agricultural University
- Patterson, T., Guy, R., & Dang, Q. (1997). Whole-plant nitrogen-and water-relations traits, and their associated trade-offs, in adjacent muskeg and upland boreal spruce species. *Oecologia*, *110*(2), 160-168.
- Pearcy, R., & Ehleringer, J. (1984). Comparative ecophysiology of C3 and C4 plants. *Plant, Cell & Environment, 7*(1), 1-13.
- Peet, M. M., Rippy, J. M., Nelson, P., & Catignani, G. L. (2004). Organic production of greenhouse tomatoes utilizing the bag system and soluble organic fertilizers. Paper presented at the VII International Symposium on Protected Cultivation in Mild Winter Climates: Production, Pest Management and Global Competition 659.
- Pérez-López, A. J., del Amor, F. M., Serrano-Martínez, A., Fortea, M. I., & Núñez-Delicado, E. (2007). Influence of agricultural practices on the quality of sweet pepper fruits as affected by the maturity stage. *Journal of the Science* of Food and Agriculture, 87(11), 2075-2080.
- Perrenoud, S. (1983). Fertilising for high yield potato. *IPI Bulletin*(8).
- Pessarakli, M. (2002). Handbook of plant and crop stress: CRC Press.
- Phurailatpam A., B. A. a. M. S. (2013). An assessment of andrographolide production in Andrographis paniculata grown in different agroclimatic locations. African Journal of Agricultural Research, 8(48), 9.
- Poorter, L., & Markesteijn, L. (2008). Seedling traits determine drought tolerance of tropical tree species. *Biotropica*, 40(3), 321-331.
- Pospisilova, J., Vagner, M., Malbeck, J., Travnickova, A., & Batkova, P. (2005). Interactions between abscisic acid and cytokinins during water stress and subsequent rehydration. *Biologia Plantarum*, 49(4), 533-540.
- Prabhu, S., & Thomas, G. V. (2002). Biological conversion of coir pith into a valueadded organic resource and its application in agri-horticulture: Current status, prospects and perspective. *Journal of Plantation Crops*, *30*(1), 1-17.
- Prasithikhet, J., Mongkolporn, P., Sritanan, V., and Sonmuang, P. . (1993). Use of organic and inorganic fertilizers in farmers rice fields in the Northeast. *Soil Management Abstracts* 5(2), 1458.

- Prathanturarug, S., Soonthornchareonnon, N., Chuakul, W., & Saralamp, P. (2007). Variation in growth and diterpene lactones among field-cultivated Andrographis paniculata. Journal of Natural Medicines, 61(2), 159-163.
- Ghana. Master, Kwame Nkrumah University of Science and Technology Kumasi, Ghana.
- Rahbarian, P., Afsharmanesh, G., & Shirzadi, M. (2010). Effects of drought stress and manure on relative water content and cell membrane stability in dragonhead (*Dracocephalum moldavica*). *Physiological Science of Herbs*, 2(1), 13-19.
- Rahimi, R., Madah Hosseini, S., Pooryoosef, M., & Fateh, I. (2010). Variation of leaf water potential, relative water content and SPAD under gradual drought stress and stress recovery in two medicinal species of Plantago ovata and *P. psyllium. Journal of Plant Ecophysiology*, 2(2), 53-60.
- Ramahsamy, K. D., Bakar, R. A., Abdullah, T. L., & Ishak, C. F. (2012a). Oil Palm Waste-Sewage Sludge Compost As A Peat Substitute In A Soilless Potting Media for Chrysanthemum. *Global Journal of Science Frontier Research-D: Agriculture and Veterinary*, 12(2).
- Ramahsamy, K. D., Bakar, R. A., Abdullah, T. L., & Ishak, C. F. (2012b). Oil Palm Waste-Sewage Sludge Compost As A Peat Substitute In A Soilless Potting Media for Chrysanthemum. *Global Journal of Science Frontier Research*, 12(2-D).
- Raviv, M., & Blom, T. J. (2001). The effect of water availability and quality on photosynthesis and productivity of soilless-grown cut roses. *Scientia Horticulturae*, 88(4), 257-276.
- Rechcigl, J. E. (1995). Soil amendments and environmental quality (Vol. 2): CRC Press.
- Reddy, A. R., Chaitanya, K. V., & Vivekanandan, M. (2004). Drought-induced responses of photosynthesis and antioxidant metabolism in higher plants. *Journal of Plant Physiology*, *161*(11), 1189-1202.
- Riaz, A., Arshad, M., Younis, A., Raza, A., & Hameed, M. (2008). Effects of different growing media on growth and flowering of *Zinnia elegans* cv. blue point. *Pakistan Journal of Botany*, 40(4), 1579-1585.
- Rita N. Kumar, S. C. a. N. K. J. I. (2011). Methods to break seed dormancy of Andrographis paniculata (Burm. f. Nees): an important medicinal herb of tropical Asia. Asian Journal of Experimental Biological Sciences, 2(1), 143-146.
- Ritchie, S. W., Nguyen, H. T., & Holaday, A. S. (1990). Leaf water content and gasexchange parameters of two wheat genotypes differing in drought resistance. *Crop Science*, 30(1), 105-111.

- Roe, N. E. (1998). Compost utilization for vegetable and fruit crops. *Horticutural Science*, *33*(6), 934-937.
- Rosales-Serna, R., Kohashi-Shibata, J., Acosta-Gallegos, J. A., Trejo-López, C., Ortiz-Cereceres, J. n., & Kelly, J. D. (2004). Biomass distribution, maturity acceleration and yield in drought-stressed common bean cultivars. *Field Crops Research*, 85(2), 203-211.
- Rouhi, V., Samson, R., Lemeur, R., & Damme, P. V. (2007). Photosynthetic gas exchange characteristics in three different almond species during drought stress and subsequent recovery. *Environmental and Experimental Botany*, 59(2), 117-129.
- Routley, D. (1966). Proline accumulation in wilted ladino clover leaves. *Crop Science*, *6*(4), 358-361.
- Ruiz Lozano, J., & Azcón, R. (1995). Hyphal contribution to water uptake in mycorrhizal plants as affected by the fungal species and water status. *Physiologia Plantarum*, 95(3), 472-478.
- Ruiz Lozano, J. M., Gómez, M., & Azcón, R. (1995). Influence of different *Glomus* species on the time-course of physiological plant responses of lettuce to progressive drought stress periods. *Plant Science*, 110(1), 37-44.
- Safarnejad, A. (2004). Characterization of somaclones of Medicago sativa L. for drought tolerance. *Journal of Agricicultural Science*, 6, 121-127.
- Saffigna, P., Powlson, D., Brookes, P., & Thomas, G. (1989). Influence of sorghum residues and tillage on soil organic matter and soil microbial biomass in an Australian Vertisol. *Soil Biology and Biochemistry*, 21(6), 759-765.
- Safir, G., Boyer, J., & Gerdemann, J. (1971). Mycorrhizal enhancement of water transport in soybean. *Science*, *172*(3983), 581-583.
- Safir, G., Boyer, J., & Gerdemann, J. (1972). Nutrient status and mycorrhizal enhancement of water transport in soybean. *Plant physiology*, 49(5), 700-703.
- Sagoe, C. I., Ando, T., Kouno, K., & Nagaoka, T. (1998). Relative importance of protons and solution calcium concentration in phosphate rock dissolution by organic acids. *Soil science and plant nutrition*, 44(4), 617-625.
- Sailo, G. L., & Bagyaraj, D. J. (2005). Influence of different AM-fungi on the growth, nutrition and forskolin content of Coleus forskohlii. *Mycological Research*, 109(07), 795-798.
- Sairam, R., Deshmukh, P., & Shukla, D. (1997). Tolerance of drought and temperature stress in relation to increased antioxidant enzyme activity in wheat. *Journal of Agronomy and Crop Science*, *178*(3), 171-178.
- Sairam, R. K., Rao, K. V., & Srivastava, G. (2002). Differential response of wheat genotypes to long term salinity stress in relation to oxidative stress,

antioxidant activity and osmolyte concentration. *Plant Science*, *163*(5), 1037-1046.

- Saliendra, N. Z., Sperry, J. S., & Comstock, J. P. (1995). Influence of leaf water status on stomatal response to humidity, hydraulic conductance, and soil drought in *Betula occidentalis*. *Planta*, 196(2), 357-366.
- Sánchez, F. J., Manzanares, M. a., de Andres, E. F., Tenorio, J. L., & Ayerbe, L. (1998). Turgor maintenance, osmotic adjustment and soluble sugar and proline accumulation in 49 pea cultivars in response to water stress. *Field Crops Research*, 59(3), 225-235.
- Santos, K. M., Fisher, P. R., & Argo, W. R. (2008). A survey of water and fertilizer management during cutting propagation. *HortTechnology*, 18(4), 597-604.
- Saraswathy, S., Manavalan, R., Vadivel, E., Manian, K., & Subramanian, S. (2004). Studies on seed germination in kalmegh (Andrographis paniculata Nees.). South Indian Horticulture, 52(1/6), 286.
- Saravanan, R., Khristi, S., Gajbhiye, N., & Maiti, S. (2009). Effect of plant population and soil moisture stress on herbage yield and andrographolide content in Andrographis paniculata. Indian Journal of Horticulture, 66(1), 120-125.
- Sarkar, S., Y. K., M. E., S. U., & Nukaya, a. A. (2008). Possibility of high soluble solid content tomato production under water stress conditions controlled by matric potential. *Journal of the Japanese Society for Horticultural Science*, 77(3), 251-258.
- Sauter, A., Davies, W. J., & Hartung, W. (2001). The long distance abscisic acid signal in the droughted plant: the fate of the hormone on its way from root to shoot. *Journal of Experimental Botany*, *52*(363), 1991-1997.
- Schlemmer, M. R., Francis, D. D., Shanahan, J., & Schepers, J. S. (2005). Remotely measuring chlorophyll content in corn leaves with differing nitrogen levels and relative water content. *Agronomy Journal*, 97(1), 106-112.
- Schonfeld, M. A., Johnson, R. C., Carver, B. F., & Mornhinweg, D. W. (1988). Water relations in winter wheat as drought resistance indicators. *Crop Science*, 28(3), 526-531.
- Serraj, R., & Sinclair, T. (2002). Osmolyte accumulation: can it really help increase crop yield under drought conditions? *Plant, Cell & Environment, 25*(2), 333-341.
- Shao, H.-B., Chu, L.-Y., Jaleel, C. A., & Zhao, C.-X. (2008). Water-deficit stressinduced anatomical changes in higher plants. *Comptes Rrendus Biologies*, 331(3), 215-225.
- Sharma, M. K., & Kumawat, D. (2012). Nitrogen fixation in JS-7322 cultivar of soybean. *International journal of phytothearpy research*, 2(1), 19-26.

- Shen, J., Li, R., Zhang, F., Fan, J., Tang, C., & Rengel, Z. (2004). Crop yields, soil fertility and phosphorus fractions in response to long-term fertilization under the rice monoculture system on a calcareous soil. *Field Crops Research*, 86(2), 225-238.
- Sheteawi, S., & Tawfik, K. (2007). Interaction effect of some biofertilizers and irrigation water regime on mung bean (*Vigna radiata*) growth and yield. *Journal of Applied Science and Technology*, 3(3), 251-262.
- Shimber, T. (1994). Influence of Shading, Mulching and Watering Frequency on Seedling Growth of Arabica Coffee.
- Shimshi, D., Mayoral, M. L., & Atsmon, D. (1982). Responses to water stress in wheat and related wild species. *Crop Science*, 22(1), 123-128.
- Simmons, G., & Pope, P. (1987). Influence of soil compaction and vesiculararbuscular mycorrhizae on root growth of yellow poplar and sweet gum seedlings. *Canadian Journal of Forest Research*, 17(8), 970-975.
- Sinclair, T., & Ludlow, M. (1985). Who taught plants thermodynamics? The unfulfilled potential of plant water potential. *Functional Plant Biology*, 12(3), 213-217.
- Singh, J., & Trehan, S. (1997). Balanced fertilization to increase the yield of potato. Paper presented at the Proceeding of the IPI-PRI-PAU workshop on "Balanced Fertilization in Punjab Agriculture" held at PAU, Ludhiana, India.
- Smetanska, I. (2008). Production of secondary metabolites using plant cell cultures *Food Biotechnology* (pp. 187-228): Springer.
- Smirnoff, N. (1993). Tansley Review No. 52. The role of active oxygen in the response of plants to water deficit and desiccation. *New Phytologist*, 27-58.
- Smirnoff, N. (1995). Antioxidant systems and plant response to the environment. Environment and plant metabolism: Flexibility and Acclimation, 42, 1981.1255-1258.
- Smith, M. W., Wazir, F. K., & Akers, S. W. (1989). The influence of soil aeration on growth and elemental absorption of greenhouse grown seedling pecan trees 1. *Communications in Soil Science & Plant Analysis*, 20(3-4), 335-344.

Smith, S. E., & Read, D. J. (1996). Mycorrhizal symbiosis: Academic press.

- Sobrado, M., & Turner, N. C. (1986). Photosynthesis, dry matter accumulation and distribution in the wild sunflower *Helianthus petiolaris* and the cultivated sunflower Helianthus annuus as influenced by water deficits. *Oecologia*, 69(2), 181-187.
- Srirangan, K., Pyne, M. E., & Perry Chou, C. (2011). Biochemical and genetic engineering strategies to enhance hydrogen production in photosynthetic algae and cyanobacteria. *Bioresource technology*, *102*(18), 8589-8604.

- Stankovic, M. S. (2011). Total phenolic content, flavonoid concentration and antioxidant activity of *Marrubium peregrinum* L. extracts. *Kragujevac Journal of Science*, 33(2011), 63-72.
- Stanojević, L., Stanković, M., Nikolić, V., Nikolić, L., Ristić, D., Čanadanovic-Brunet, J., & Tumbas, V. (2009). Antioxidant activity and total phenolic and flavonoid contents of *Hieracium pilosella* L. extracts. *Sensors*, 9(7), 5702-5714.
- Stoll, M., Loveys, B., & Dry, P. (2000). Hormonal changes induced by partial rootzone drying of irrigated grapevine. *Journal of Experimental Botany*, 51(350), 1627-1634.
- Sturz, A., Christie, B., & Nowak, J. (2000). Bacterial endophytes: potential role in developing sustainable systems of crop production. *Critical Reviews in Plant Sciences*, 19(1), 1-30.
- Suhaimi, M., & Ong, H. (2001). Composting empty fruit bunches of oil palm. *Extension Bulletin-Food & Fertilizer Technology Center*(505), 1-8.
- Sumathi, T., Janardhan, A., Srilakhmi, A., Gopal, D. S., & Narasimha, G. (2012). Impact of indigenous microorganisms on soil microbial and enzyme activities. Archives of Applied Science Research, 4, 1065-1073.
- Sundara, B., Natarajan, V., & Hari, K. (2002). Influence of phosphorus solubilizing bacteria on the changes in soil available phosphorus and sugarcane and sugar yields. *Field crops research*, 77(1), 43-49.
- Suriyo, T., Pholphana, N., Rangkadilok, N., Thiantanawat, A., Watcharasit, P., & Satayavivad, J. (2014). Andrographis paniculata Extracts and Major Constituent Diterpenoids Inhibit Growth of Intrahepatic Cholangiocarcinoma Cells by Inducing Cell Cycle Arrest and Apoptosis. Planta Medica (EFirst).
- Syunarti, S. (2008). The effect of inorganic fertilizer and biofertilizer on the growth of patchouli plant (Pogostemon Cablin) (Master Thesis). Universiti Malaysia Pahang.
- Talei, D., Valdiani, A., Abdullah, M. P., & Hassan, S. A. (2012). A rapid and effective method for dormancy breakage and germination of King of Bitters (*Andrographis paniculata* Nees.) seeds. *Maydica*, 57(2), 98-105.
- Tan, C., & Buttery, B. (1982). The effect of soil moisture stress to various fractions of the root system on transpiration, photosynthesis, and internal water relations of peach seedlings. *Journal American Society for Horticultural Science*.
- Tang, L. I., Ling, A. P., Koh, R. Y., Chye, S. M., & Voon, K. G. (2012). Screening of anti-dengue activity in methanolic extracts of medicinal plants. *BMC Complementary and Alternative Medicine*, 12(1), 3.

- Tanguilig, V., Yambao, E., O'toole, J., & De Datta, S. (1987). Water stress effects on leaf elongation, leaf water potential, transpiration, and nutrient uptake of rice, maize, and soybean. *Plant and Soil*, 103(2), 155-168.
- Tardieu, F., Zhang, J., Katerji, N., Bethenod, O., Palmer, S., & Davies, W. (1992). Xylem ABA controls the stomatal conductance of field grown maize subjected to soil compaction or soil drying. *Plant, Cell & Environment, 15*(2), 193-197.
- Tesfaye, S., Razi, I. M., & Maziah, M. (2008). Effects of deficit irrigation and partial rootzone drying on growth, dry matter partitioning and water use efficiency in young coffee (*Coffea arabica* L.) plants. *Journal of Food, Agriculture and Environment*, 6(384), 312-317.
- Tipakorn, N. (2002). Effects of Andrographis paniculata (Burm. F.) Nees on Performance, Mortality and Coccidiosis in Broiler Chickens [disertasi]. Göttingen, Germany: Doctor of Agricultural Sciences of the Faculty of Agricultural Sciences. Georg August-University.
- Tiwari, N., Purohit, M., Sharma, G., & Nautiyal, A. (2013). Changes in Morpho-Physiology of Jatropha curcas grown under different water regimes. *Nature and Science of Sleep*, *11*(9), 76-83.
- Tripepi, A. D. L. a. R. R. (1993). *Plant Science Division*. Moscow: University of Idaho.
- Tucker, M. R. (1999). Essential plant nutrients: their presence in North Carolina soils and role in plant nutrition: Department of Agriculture and Consumer Services, Agronomic Division.
- Turner, N. (1990). Plant water relations and irrigation management. Agricultural water management, 17(1), 59-73.
- Uwumarongie Ilori, E., Sulaiman-Ilobu, B., Ederion, O., Imogie, A., Imoisi, B., Garuba, N., & Ugbah, M. (2012). Vegetative Growth Performance of Oil Palm (*Elaeis guineensis*) Seedlings in Response to Inorganic and organic fertilizers. *Greener Journal of Agricultural Sciences* 2(2276-7770), 026-030.
- Vakili, M., & Haque, A. A. (2012). Assessment of Palm Oil Bio-waste with Cowdung for Compost Production. Paper presented at the Energy, biomass and biological residues. International Conference of Agricultural Engineering-CIGR-AgEng 2012: Agriculture and Engineering for a Healthier Life, Valencia, Spain, 8-12 July 2012.
- Van Rensburg, L., & Krüger, G. (1994). Evaluation of Components of Oxidative Stress Metabolism for Use in Selection of Drought Tolerant Cultivars of *Nicotiana tabacum* L. *Journal of plant physiology*, 143(6), 730-737.

- Vasanthakrishna, M., Bagyaraj, J. D., & Nirmalnath, J. P. (1995). Selection of efficient VA mycorrhizal fungi for Casuarina equisetifolia—second screening. *New Forests*, 9(2), 157-162.
- Verbruggen, N., & Hermans, C. (2008). Proline accumulation in plants: a review. *Amino acids*, 35(4), 753-759.
- Vessey, J. K. (2003). Plant growth promoting rhizobacteria as biofertilizers. *Plant and soil*, 255(2), 571-586.
- Vivas, A., Marulanda, A., Ruiz-Lozano, J. M., Barea, J. M., & Azcón, R. (2003). Influence of a Bacillus sp. on physiological activities of two arbuscular mycorrhizal fungi and on plant responses to PEG-induced drought stress. Mycorrhiza, 13(5), 249-256.
- Wahid, A., Perveen, M., Gelani, S., & Basra, S. (2007). Pretreatment of seed with H<sub>2</sub>O<sub>2</sub> improves salt tolerance of wheat seedlings by alleviation of oxidative damage and expression of stress proteins. *Journal of plant physiology*, 164(3), 283-294.
- Wahid, A. N. A., Noordin, L., & Hoe, P. C. K. (2011). Bio fertilizer Application in a Fertigation System.
- Wahid, A. N. A., Noordin, L., Hoe, P. C. K., & Razak, A. (2012). Development of Multifunctional Biofertilizer Formulation from Indigenous Microorganisms and Evaluation of Their N<sub>2</sub>-Fixing Capabilities on Chinese Cabbage Using 15 N Tracer Technique Pertanika Journal of Tropical and Agricultural Science.
- Wahid, A. N. A. N., Latiffah Hoe, Phua Choo Kwai Razak, Abdul. (2012). Development of Multifunctional Biofertilizer Formulation from Indigenous Microorganisms and Evaluation of Their N<sub>2</sub> -Fixing Capabilities on Chinese Cabbage Using 15 N Tracer Technique *Pertanika Journal of Tropical and Agricultural Science*, 35 (3), 673 - 679.
- Wakrim, R., Wahbi, S., Tahi, H., Aganchich, B., & Serraj, R. (2005). Comparative effects of partial root drying (PRD) and regulated deficit irrigation (RDI) on water relations and water use efficiency in common bean (*Phaseolus vulgaris* L.). Agriculture, Ecosystems & Environment, 106(2), 275-287.
- Wang, C., Wu, J., & Mei, X. (2001). Enhancement of taxol production and excretion in Taxus chinensis cell culture by fungal elicitation and medium renewal. *Applied Microbiology and Biotechnology*, 55(4), 404-410.
- Weibel, F., Bickel, R., Leuthold, S., & Alföldi, T. (1998). Are organically grown apples tastier and healthier? A comparative field study using conventional and alternative methods to measure fruit quality. Paper presented at the XXV International Horticultural Congress, Part 7: Quality of Horticultural Products 517.

Wiart, C. (2002). Medicinal plants of Southeast Asia: Prentice Hall.

- Wira, A., Mohd, R., & Abd, J. (2011). Composts as additives in coconut coir dust culture for growing rockmelon (*Cucumis melo L.*). Journal of Tropical Agriculture and Food Science, 39(2), 229-237.
- Wira, A., Razi, I. M., & Jamil, Z. A. (2011a). Composts as additives in coconut coir dust culture for growing rockmelon (*Cucumis melo* 1.). Journal of Tropical Agriulture. and Food Science, 39(2), 229-237.
- Wira, A., Razi, I. M., & Jamil, Z. A. (2011b). Composts as additives in coconut coir dust culture for growing rockmelon (Cucumis melo 1.). *Journal of Tropical Agriculture and Food Sciene*, 39(2), 229-237.
- Wongkittipong, R., Prat, L., Damronglerd, S., & Gourdon, C. (2004). Solid–liquid extraction of andrographolide from plants experimental study, kinetic reaction and model. *Separation and Purification Technology*, 40(2), 147-154.
- Wright, G., Rao, R., & Farquhar, G. (1994). Water-use efficiency and carbon isotope discrimination in peanut under water deficit conditions. *Crop Science*, 34(1), 92-97.
- Wu, F., Bao, W., Li, F., & Wu, N. (2008). Effects of drought stress and N supply on the growth, biomass partitioning and water-use efficiency of Sophora davidii seedlings. Environmental and Experimental Botany, 63(1), 248-255.
- Wu, X. H., Tang, Z.H. and Zn, Y.G. (2006). Effect of water stress on free amino acid content in *Rosmarinus officinalis L. Jornal of North East Forcs*. Univiversity, 34, 57-58.
- Wullschleger, S. D., Yin, T., DiFazio, S., Tschaplinski, T., Gunter, L., Davis, M., & Tuskan, G. (2005). Phenotypic variation in growth and biomass distribution for two advanced-generation pedigrees of hybrid poplar. *Canadian Journal of Forest Research*, 35(8), 1779-1789.
- Yancey, P. H., Clark, M. E., Hand, S. C., Bowlus, R. D., & Somero, G. N. (1982). Living with water stress: evolution of osmolyte systems. *Science*, 217(4566), 1214-1222.
- Yang, Y., He, F., Yu, L., Chen, X., Lei, J., & Ji, J. (2007). Influence of drought on oxidative stress and flavonoid production in cell suspension culture of *Glycyrrhiza inflata* Batal. *Zeitschrift fur Naturforschung C-Journal of Biosciences*, 62(5-6), 410-416.
- Yuan, Y., Liu, Y., Wu, C., Chen, S., Wang, Z., Yang, Z., Huang, L. (2012). Water deficit affected flavonoid accumulation by regulating hormone metabolism in *Scutellaria baicalensis* Georgi roots. *PLOS one*, 7(10), e42946.
- Zaharah, A., & Lim, K. (2000). Oil palm empty fruit bunch as a source of nutrients and soil ameliorant in oil palm plantation, Malaysia J. *Soil Science*, *4*, 51-66.
- Zarrouk, O., Gogorcena, Y., Gómez-Aparisi, J., Betrán, J., & Moreno, M. (2005). Influence of almond× peach hybrids rootstocks on flower and leaf mineral

concentration, yield and vigour of two peach cultivars. *Scientia Horticulturae*, 106(4), 502-514.

- Zhang, C., & Tan, B. (1996). Hypotensive activity of aqueous extract of Andrographis paniculata in rats. Clinical and Experimental Pharmacology and Physiology, 23(8), 675-678.
- Zhang, J., & Davies, W. (1989). Abscisic acid produced in dehydrating roots may enable the plant to measure the water status of the soil. *Plant, Cell & Environment, 12*(1), 73-81.
- Zhang, M., Duan, L., Zhai, Z., Li, J., Tian, X., Wang, B., Li, Z. (2004). Effects of plant growth regulators on water deficit-induced yield loss in soybean. Paper presented at the Proceedings of the 4th International Crop Science Congress, Brisbane, Australia.
- Zhao, D., Reddy, K. R., Kakani, V. G., & Reddy, V. (2005). Nitrogen deficiency effects on plant growth, leaf photosynthesis, and hyperspectral reflectance properties of sorghum. *European Journal of Agronomy*, 22(4), 391-403.
- Zhao, J., Zhu, W.-H., & Hu, Q. (2001). Selection of fungal elicitors to increase indole alkaloid accumulation in *Catharanthus roseus* suspension cell culture. *Enzyme and Microbial Technology*, 28(7), 666-672.
- Zhu, Z., Liang, Z., Han, R., & Wang, X. (2009a). Impact of fertilization on drought response in the medicinal herb *Bupleurum chinense* DC.: Growth and saikosaponin production. *Industrial Crops and Products*, 29(2), 629-633.
- Zhu, Z., Liang, Z., Han, R., & Wang, X. (2009b). Impact of fertilization on drought response in the medicinal herb *Bupleurum chinense* DC.: Growth and saikosaponin production. *Industrial Crops and Products*, 29(2), 629-633.
- Zobayed, S., Afreen, F., & Kozai, T. (2007). Phytochemical and physiological changes in the leaves of St. John's wort plants under a water stress condition. *Environmental and Experimental Botany*, 59(2), 109-116.
- Zulkarami, B., Ashrafuzzaman, M., & Razi, I. M. (2010). Morpho-physiological growth, yield and fruit quality of rock melon as affected by growing media and electrical conductivity. *Journal of Food Agriculture & Environment*, 8, 249-252.
- Zulkarami, B., Tajul, M., Fariz, A., Husni, M., Norazrin, A., Radziah, O., Rafii, M. (2012). Effects of bacteria and arbuscular mycorhizae inoculation at different electrical conductivity level on growth and yield of rockmelon ('*Cucumis melo*') under soilless culture. *Australian Journal of Crop Science*, 6(11), 1494.
- Zydlik, P., & Zydlik, Z. (2008). Impact of biological effective microorganisms [EM] preparations on some physico-chemical properties of soil and the vegetative

growth of apple-tree rootstocks. Nauka Przyroda Technologie. Uniwersytet Przyrodniczy w Poznaniu, 2(1).