



UNIVERSITI PUTRA MALAYSIA

***GanoCare™ FERTILIZATION EFFECTS ON GROWTH AND
SUPPRESSION
OF BASAL STEM ROT DISEASE OF OIL PALM SEEDLINGS***

NUR AKMAL BINTI REBITANIM

ITA 2016 7



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OF BASAL STEM ROT DISEASE OF OIL PALM SEEDLINGS**

By

NUR AKMAL BINTI REBITANIM

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Master of Science**

February 2016



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DEDICATION

This thesis is dedicated to Ayah, Mama, Ika, Aci & Azman.



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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Master of Science

GanoCare™ FERTILIZATION EFFECTS ON GROWTH AND SUPPRESSION OF BASAL STEM ROT DISEASE OF OIL PALM SEEDLINGS

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February 2016

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Institute : Tropical Agriculture

The oil palm industry is on alert for the basal stem rot (BSR) disease. The BSR which is caused by *Ganoderma boninense*, is the major disease of oil palm and is a serious threat to the palm oil production in Malaysia. One of the ideal solutions to manage BSR disease is through supplementation of nutrients. With no definitive cure at the present, thus this study introduced a new fertilizer technology called GanoCare™, as an effort to suppress BSR incidence in oil palm. The GanoCare™ fertilizer was produced from powdered oil palm empty fruit bunches (EFB) incorporated with beneficial elements. The optimum levels of GanoCare™ from two different application frequencies for the growth of oil palm was determined in a six-month nursery trial, and subsequently extended in the field trial for 21 months. All growth and physiological parameters evaluated were positively correlated with GanoCare™ levels up to a certain level. Two rates at 10 g/palm for every month application and 30 g/palm for every three months application were identified as optimum levels for the growth of oil palm seedlings. In the field, the optimum levels were at 100 g/palm for every month application and 300 g/palm for every three months application. The effect of optimized rate of GanoCare™ 1 on growth, physiological and BSR disease suppression were assessed in the nursery stage using sitting (bait) technique, in which the treatments added were CT (control), T1 (pre-treatment with 10 g/month), T2 (pre-treatment with 30 g/three months), T3 (pre-treatment and continuous treatment with 10 g/month) and T4 (pre-treatment and continuous treatment with 30 g/three months). A follow up in the field stage was carried out for 21 months to test on severity of *Ganoderma* using baiting technique under natural condition. The treatments were similar as in the nursery stage, however with some alterations to suit the condition of the experiment. In nursery, the application of GanoCare™ 1 significantly enhanced the height, girth, frond count, total leaf area, chlorophyll content, photosynthesis rate, fresh and dry weight of oil palm seedlings when compared to the control. Despite some modifications done to the treatments, the results from the field trial validate the pathological results from the nursery, in which both experiments similarly showed that application of GanoCare™ was able to suppress the incidence of BSR disease compared to the control seedlings. For both studies in nursery and field, the disease reduction was higher for seedlings given pre-

treatment and continuous treatment (T3 and T4) compared to the seedlings that were pre-treated only (T1 and T2), suggesting that continuous application is required to provide maximum protection from *Ganoderma* disease, and that pre-treatment alone was not sufficient to allow the development of a strong defense system in the seedlings. Combination of pre-treatment and continuous treatment with 30 g of GanoCare™ at three-month intervals (T4) was the most effective treatment in both nursery and field by reducing the BSR disease by 77.78 and 82.36%, respectively.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**KESAN PEMBAJAAN GanoCare™ KE ATAS PERTUMBUHAN DAN
PENYEKATAN PENYAKIT REPUT PANGKAL BATANG ANAK KELAPA
SAWIT**

Oleh

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Industri kelapa sawit kini sedang dikejutkan dengan penyakit reput pangkal batang (BSR). Penyakit BSR yang disebabkan oleh *Ganoderma boninense*, adalah penyakit utama kelapa sawit dan merupakan satu ancaman serius kepada pengeluaran minyak sawit di Malaysia. Salah satu penyelesaian yang berpotensi untuk mengawal penyakit BSR adalah melalui rawatan nutrien. Disebabkan tiada langkah berkesan pada masa ini, maka kajian ini memperkenalkan teknologi baja baru yang dikenali sebagai GanoCare™, sebagai usaha untuk mengurangkan kejadian BSR kelapa sawit. Baja GanoCare™ dihasilkan daripada serbuk tandan buah kosong sawit (EFB) yang digabungkan bersama unsur-unsur yang berfaedah. Tahap optimum GanoCare™ dari dua frekuensi aplikasi yang berlainan bagi menilai pertumbuhan kelapa sawit telah dijalankan di tapak semaian selama 6 bulan, yang kemudiannya dilanjutkan ke kajian lapangan selama 21 bulan. Kesemua penilaian ke atas parameter pertumbuhan dan fisiologi menunjukkan perhubungan positif dengan tahap aplikasi GanoCare™ sehingga ke suatu tahap. Dua kadar iaitu 10 g/pokok bagi aplikasi setiap bulan dan 30 g/pokok bagi aplikasi setiap tiga bulan telah dikenal pasti sebagai tahap optimum untuk pertumbuhan anak pokok kelapa sawit. Untuk kajian lapangan, tahap optimum ialah 100 g/pokok bagi aplikasi setiap bulan dan 300 g/pokok bagi aplikasi setiap tiga bulan. Kesan kadar optimum GanoCare™ 1 pada pertumbuhan, fisiologi dan penyekatan penyakit BSR dinilai di peringkat tapak semaian menggunakan teknik duduk (umpan), di mana rawatan yang diberikan adalah CT (kawalan), T1 (pra-rawatan dengan 10 g/bulan), T2 (pra-rawatan dengan 30 g/tiga bulan), T3 (pra-rawatan dan rawatan berterusan dengan 10 g/bulan) dan T4 (pra-rawatan dan rawatan berterusan dengan 30 g/tiga bulan). Susulan seterusnya dijalankan di peringkat lapangan selama 21 bulan untuk menguji keterukan serangan *Ganoderma* dengan menggunakan teknik umpan di bawah keadaan semula jadi. Rawatan adalah sama seperti di peringkat tapak semaian, bagaimanapun dengan beberapa pengubahsuaian untuk disesuaikan dengan keadaan eksperimen. Di tapak semaian, aplikasi GanoCare™ 1 dengan berkesannya meningkatkan tinggi, lilitan, kiraan daun, jumlah permukaan daun, kandungan klorofil, kadar fotosintesis, berat basah dan kering anak pokok kelapa sawit jika dibandingkan

dengan pokok kawalan. Keputusan daripada percubaan lapangan mengesahkan keputusan patologi dari tapak semaian, di mana kedua-dua eksperimen menunjukkan bahawa penggunaan GanoCare™ dapat mengurangkan kejadian penyakit BSR berbanding dengan benih kawalan. Untuk kedua-dua kajian di tapak semaian dan lapangan, pengurangan penyakit adalah lebih tinggi untuk benih yang telah diberikan pra-rawatan dan rawatan berterusan (T3 dan T4) berbanding dengan benih yang diberi pra-rawatan sahaja (T1 dan T2), dan ini menunjukkan bahawa aplikasi berterusan diperlukan bagi memberi perlindungan maksimum daripada penyakit *Ganoderma*, dan pra-rawatan sahaja tidak mencukupi untuk membolehkan pembangunan sistem pertahanan yang kuat dalam benih. Kombinasi pra-rawatan dan rawatan berterusan dengan 30 g GanoCare™ setiap tiga bulan (T4) adalah rawatan yang paling berkesan untuk kedua-dua tapak semaian dan lapangan dengan masing-masing dapat mengurangkan penyakit BSR sebanyak 77.78 dan 82.36%.



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LIST OF ABBREVIATIONS

%	Percent
\$	Dollars
°C	Degree celcius
AMF	Arbuscular mycorrhiza fungi
ANOVA	Analysis of variance
AUDPC	Area under the disease progress curve
B	Boron
BSR	Basal stem rot
C	Carbon
Ca	Calcium
CaNO ₃	Calcium nitrate
Cl	Chlorine
cm	Centimeter
Co	Cobalt
CO ₂	Carbon dioxide
CRD	Completely Randomized Design
Cu	Copper
DI	Disease incidence
DR	Disease reduction
DS	Disease severity
DSIB	Disease severity of bole index
DSIF	Disease severity of foliar index
DSIR	Disease severity of root index
EB	Endophytic bacteria
EFB	Empty fruit bunch
Fe	Iron
FELCRA	Federal Land Consolidation and Rehabilitation Authority
FELDA	Felda Land Development Authority
FFB	Fresh fruit bunch
FPSSB	FELCRA Plantation Services Sdn. Bhd.
g	Gram
GSM	<i>Ganoderma</i> selective medium
H	Hydrogen
H ₂ O ₂	Hydrogen peroxide
ha	Hectare
ISR	Induced systemic resistance
K	Potassium
K ₂ O	Potassium oxide
kg	Kilogram
LSD	Least significant difference
m	Meter
MARDI	Malaysian Agricultural Research and Development Institute
MEA	Malt extract agar
Mg	Magnesium
mg/L	Milligram per liter
MgO	Magnesium oxide
mL	Milliliter
mm	Millimeter

Mn	Manganese
Mo	Molybdenum
MPOB	Malaysian Palm Oil Board
N	Nitrogen
Nm	Nanometer
O	Oxygen
P	Phosphorus
P ₂ O ₅	Phosphorus pentoxide
PAL	Phenylalanine ammonia-lyase
PDA	Potato dextrose agar
pH	Hydrogen potential
POD	Peroxidase
ppm	Part per million
PPO	Polyphenol oxidase
RCBD	Randomized complete block design
RNA	Ribonucleic acid
RWB	Rubber wood block
S	Sulphur
SFS	Severity of foliar symptoms
Si	Silicon
SiO ₂	Silicon dioxide
TE	Trace element
TEM	Transmission electron microscopy
™	Trademark
UPM	Universiti Putra Malaysia
v/v	Volume to volume
Zn	Zinc

CHAPTER 1

INTRODUCTION

Oil palm (*Elaeis guineensis* Jacq.) was cultivated as early as 3000 B.C in Egypt (Friedel, 1987). It is a perennial crop that exists in the equatorial tropics of Africa, America and South-East Asia (Hartley, 1988). From growing wild, oil palm was developed into an important agriculture crop and has been planted in all tropical parts of the world, especially in South East Asia countries, such as Malaysia, Thailand and Indonesia (Shuit et al., 2009). After Indonesia, Malaysia is the second major producer and exporter in oil palm production, with both countries accounted for nearly 90% of the total world crude oil production (Carter et al., 2007; Sharma, 2007). In developing countries, such as Malaysia and Indonesia, oil palm industry is one of the important tools to alleviate poverty. Oil palm is said as the “golden crop of Malaysia” since it plays important key role in the Malaysian economy and generates profitable export earnings for this country (Bivi et al., 2010).

Oil palm is cultivated mostly for its oil in which the global demand for palm oil is tremendously increase for the last few decades. During the Industrial Revolution in Europe, oil palm was largely cultivated due to massive demands in manufacturing of lubricants and soap (Hardon, 1995). Presently, palm oil is commonly used as edible vegetable oil for human consumption and also a significant source of biodiesel and biogas. The billion dollar oil palm industry brings huge profitability more than any other vegetable oil, such as corn oil, linseed oil, sunflower oil, soybean oil and canola oil (Carrere, 2001). In 2010, about 47.9% of the world palm oil production was contributed by Malaysia, with export revenue earnings from oil palm products of more than RM 60 billion (Simeh and Fairuz, 2009; Abas et al., 2011). In 2011, oil palm produced 31.3% of global oils and fats, with only 5.3% of global land used for cultivation, showing that this crop brings high productivity with little land requirement (Mielke, 2012).

Nonetheless, oil palm is vulnerable to various pests and diseases, among which is the basal stem rot (BSR) disease. The BSR disease is caused by white rot fungus called *Ganoderma boninense*, which this pathogen has brought a massive damage to the oil palm plantation in Pacific Islands and Papua New Guinea, and in Southeast Asia, particularly in Indonesia and Malaysia (Ariffin, 1990; Flood and Hasan, 2004; Pilotti, 2005). On coastal soils of Malaysia, palm losses raised as high as 85% by the time palms were replanted at 25 years. About 31 to 67% of disease incidence has caused up to 46% yield reduction in Malaysia (Singh, 1991). It was estimated that only 1% disease incidence can bring a loss of up to 38 million US\$ annually to Indonesian industry (Flood and Hasan, 2004). In Malaysia, the BSR incidence reported was 3.71% and total area impacted was 59,148 hectares, with the most severe areas affected particularly in Perak, Johor and Sabah (Idris et al., 2011).

Among the early signs of BSR disease are leaves yellowing and spear rotting. The fronds are usually folded like an arrow, thus restricting the oil palms' ability to photosynthesize (Idris et al., 2000). The foliar symptoms of BSR will only emerged after substantial portion of the bole is infected. The internal xylem vessels which are vital to the oil palm's transport systems usually deteriorate, thus inhibiting the access of

water and plant nourishment. Originally, BSR disease was reported to only attacks matured palms over 25 years old, however the disease symptoms have been detected to infect younger palms (Turner, 1981; Khairudin, 1993; Ariffin et al., 2000; Susanto et al., 2005). Unfortunately, to date, no definitive solution is available to cure this disease.

A variety of methods are currently being practiced as an integrated approach to manage BSR disease. Development of oil palm progenies resistant to *Ganoderma* may offer the most effective method and long term solution for controlling BSR disease. Even though breeding programs for lightly resistant progenies to *Ganoderma* are actively being undertaken by few research organizations, including Malaysian Palm Oil Board (MPOB), however tolerant oil palm progenies are yet unavailable due to the genetic diversity of the fungus. Utilization of fungicides is an important component in the BSR management. Fungicides, such as carbendazim, cycloheximide and drazoxolone are reported to be strongly inhibitory to the fungus (Ramasamy, 1972; Jollands, 1983; Khairudin, 1990). However, application of fungicides only assist to slow the infection of *Ganoderma* and arise of public alertness on the effect of fungicides leads to detrimental effects of a non-target organisms and environmental issues, such as ground water contamination (Sathya et al., 2010; Sundram, 2012).

This eventually resulted to a more environmental friendly practice, which is the use of antagonistic microbes as biological control agents (Sundram, 2005). Several promising antagonists, such as *Trichoderma* spp. (Ilias, 2000; Sundram, 2005), *Penicillium* spp. (Dharmaputra et al., 1989), *Aspergillus* (Shukla and Uniyal, 1989), *Pseudomonas* spp. and *Bacillus* spp. (Susanto et al., 2005) have demonstrated potential efficacy against *G. boninense*. Incidence of BSR disease on four months old oil palm seedlings was reduced when treated with *Trichoderma* spp. as aggregates on paddy grain (Zakaria et al., 2010). In addition, treatment with conidial suspension of *Trichoderma harzianum* (isolate FA 1132) suppressed the incidence of BSR in oil palm (Nur Ain Izzati and Abdullah, 2008). Even so, biological approach itself cannot stop the emergence of *G. boninense* in oil palm.

Owing to these limitations, developing another alternative which is cost effective and more environmentally safe by improvement of the oil palm defense system, through addition of required mineral nutrients becomes an imperative solution. Nutrient supplementation to the soil as fertilizer is known to affect the susceptibility of plants towards some diseases caused by fungal pathogens (Veresoglou et al., 2013). In most cases, a balanced of mineral nutrient added to the soil in the form of fertilizer improves the plants' ability to disease resistance (Usherwood, 1980; Fageria, 2009). This approach may not necessarily be a cure for the BSR disease but it could arrest the spread of disease in the oil palm. A reduction in BSR incidence was observed when oil palm seedlings were supplemented with Cu, Ca and combination of Cu and Ca compared to the control seedlings (Nur Sabrina et al., 2012). This ideal solution to delay BSR disease development in oil palm is through manipulation of lignin by plant nutrient management as it assists in enhancing the defense mechanism in plants (Paterson et al., 2009).

Therefore, the present study was undertaken to investigate the effect of GanoCare™ in suppressing BSR disease in oil palm. The GanoCare™ (consisted of GanoCare™ 1 and GanoCare™ 2) is a new fertilizer technology, formulated to prevent BSR disease in oil palm. The fertilizer was derived by combining powdered empty fruit bunches (EFB) with beneficial element. The effect of GanoCare™ 1 on vegetative growth,

physiological parameters and suppression of BSR disease of oil palm seedlings was investigated through the nursery trials using sitting technique. Most research studies have been limited in the nursery without validation assessment from the field trial. Thus, in this study, the scope was broadened by carrying out the GanoCare™ 1 treatments in the field stage based on seedling baiting technique.

Thus, the study was carried out with these objectives:

1. To determine the optimum levels of GanoCare™ from two different application frequencies for both nursery and field trials.
2. To investigate the effect of optimized rates of GanoCare™ 1 on the performance of oil palm seedlings in the nursery trial.
3. To investigate the effect of GanoCare™ 1 on BSR disease suppression of oil palm seedlings in the nursery trial using sitting (bait) technique.
4. To evaluate the effect of GanoCare™ 1 on BSR disease suppression of oil palm seedlings in the field trial using baiting technique.

REFERENCES

- Abas, R., Kamarudin, M. F., Borhan, A. A. N. and Simeh, M. A. (2011). A study on the Malaysian oil palm biomass sector- supply and perception of palm oil millers. *Oil Palm Industry Economic Journal*, 11(1): 28-41.
- Abdjad, A. N., Ika, D., Suryo, W. and Juang Gema, K. (2011). Selection and characterization of endophytic bacteria as biocontrol agents of tomato bacterial wilt disease. *Journal of Biosciences*, 18(2): 66-70.
- Abdullah, F., Ilias, G. N. M., Nelson, M., Nur Ain Izzati, M. Z. and Umi Kalsom, Y. (2003). Disease assessment and the efficacy of *Trichoderma* as a biocontrol agent of basal stem rot of oil palms. *Research Bulletin Science Putra*, 11: 31-33.
- Abdullah, F., Ilias, G. N. M., Vijaya, S. K. and Leong, T. T. (1999). Diversity of *Trichoderma* and its *in vivo* efficacy against *Ganoderma boninense*. In: Sidek, Z., Bong, S. K., Ong, C. A. and Husan, A. K. (Eds.), *Sustainable Crop Protection Practices in the Next Millenium, MCB-MAPPS Plant Protection Conference 1999*, 2-3 November 1999. Kota Kinabalu, Sabah.
- Abdullah, F., Mohd Hefni, R. and Noorhaida, S. (2005). Bacteria from oil palm agricultural system and their interactions with *Ganoderma boninense* and *Trichoderma harzianum*. *Pertanika Journal of Tropical Agricultural Science*, 28(2): 95-102.
- Agarie, S., Agata, W., Uchida, H., Kubota, F. and Kaufman, P. B. (1996). Function of silica bodies in the epidermal system of rice (*Oryza sativa* L.): testing the window hypothesis. *Journal of Experimental Botany*, 47(299): 655-660.
- Agrios, G. N. (2005). *Plant Pathology 5th Edition*. San Diego: Elsevier-Academic Press.
- Akbar, U., Kusnadi, M. and Ollagnier, M. (1971). Influence of the type of planting materials and of mineral nutrients on oil palm stem rot due to *Ganoderma*. *Oleagineux*, 26: 527-534.
- Andrews, J. H. (1992). Biological control of the phyllosphere. *Annual Review of Phytopathology*, 30: 603-605.
- AOAC (1975). *Official Methods of Analysis 12th Edition*. Washington, DC: Association of Official Analytical Chemists.
- Ariffin, D. (1990). Progress on *Ganoderma* research at PORIM. In: *Proceedings of Ganoderma Workshop*, 11 September 1990. PORIM, Bangi.
- Ariffin, D. (2000). Major diseases of oil palm. In: Yusof, B., Jalani, B. S. and Chan, K. W. (Eds.), *Advances in Oil Palm Research (pp. 596-622)*. Malaysian Palm Oil Board.

- Ariffin, D. and Idris, A. S. (1991). A selective medium for the isolation of *Ganoderma* from diseased tissues. In: Yusof et al. (Eds.), *Proceedings of the International Palm Oil Conference, Progress, Prospects and Challenges towards the 21st Century*, 9-14 September 2001. Palm Oil Research Institute of Malaysia, Malaysia.
- Ariffin, D. and Idris, A. S. (1992). The *Ganoderma*-selective medium (GSM). PORIM Information Series, ISSN 0128-5726.
- Ariffin, D., Idris, A. S. and Marzuki, A. (1995). *Proceedings of the 1995 PORIM National Oil Palm Conference "Technologies in Plantation- the way forward"*, 11-12 July 1995. PORIM, Bangi.
- Ariffin, D., Idris, A. S. and Singh, G. (2000). Status of *Ganoderma* in oil palm. In: Flood, J., Bridge, P. D. and Holderness, M. (Eds.), *Ganoderma Disease of Perennial Crops* (pp. 49-68). UK: CABI Publishing.
- Arnold, A. E., Meffa, L. C., Kyollo, D., Rojas, E. I., Maynard, Z., Robbins, N. and Herre, E. A. (2003). Fungal endophytes limit pathogen damage in a tropical tree. *Proceedings of the National Academic Sciences*, 23 December 2003. USA.
- Azahar, T. M., Jawahir, C. M., Mazliham and Patrice, B. (2011). Temporal analysis of basal stem rot disease in oil palm plantations. *International Journal of Engineering and Technology*, 11(3): 96-101.
- Benhamou, N. and Nicole, M. (1999). Cell biology of plant immunization against microbial infection: the potential of induced resistance in controlling plant diseases. *Plant Physiology and Biochemistry*, 37: 703-719.
- Biljana, G. and Jugoslav, Z. (2011). *Trichoderma harzianum* as a biocontrol agent against *Alternaria alternata* on tobacco. *Applied Technologies and Innovations*, 7(2): 67-76.
- Bivi, M. R., Farhana, M. S. N., Khairulmazmi, A. and Idris, A. (2010). Control of *Ganoderma boninense*: a causal agent of basal stem rot disease in oil palm with endophyte bacteria *in vitro*. *International Journal of Agriculture and Biology*, 12: 833-839.
- Breton, F., Hasan, Y., Hariadi, S., Lubis, Z. and De Franqueville, H. (2006). Characterization of parameters for the development of an early screening test for basal stem rot tolerance in oil palm progenies. *Journal of Oil Palm Research*, 24-36.
- Broadley, M., Brown, P., Cakmak, I., Rengel, Z. and Zhao, F. (2012). Functions of nutrients: micronutrients. In: Marschner, P. (Ed.), *Marschner's Mineral Nutrition of Higher Plants* (pp. 191-248). USA: Academic Press.
- Broadley, M., White, P. J., Hammond, J. P., Zelko, I. and Lux, A. (2007). Zinc in plants. *New Phytologist*, 173: 677-702.

- Cai, K., Gao, D., Luo, S., Zeng, R., Yang, J. and Zhu, X. (2008). Physiological and cytological mechanisms of silicon-induced resistance in rice against blast disease. *Physiologia Plantarum*, 134: 324-333.
- Caiyun, L., Jian, M. A., Xin, C., Xudong, Z. and Yinshi, B. (2010). Effect of nitrogen fertilizer and maize straw incorporation on NH₄ and NO₃ accumulation in black soil of Northeast China among three consecutive cropping cycles. *Journal of Soil Science Plant Nutrition*, 10(4): 443-453.
- Campbell, C. L. and Madden, L. V. (1990). *Introduction to Plant Disease Epidemiology* (pp. 113-121). USA: John Wiley and Sons.
- Carrere, R. (2001). *The Bitter Fruit of Oil Palm: Dispossession and Deforestation. World Rainforest Movement*. Novib and the Swedish Society for Nature Conversation.
- Carter, C., Finley, W., Fry, J., Jackson, D. and Willis, L. (2007). Palm oil markets and future supply. *European Journal of Lipid Science and Technology*, 109: 307-314.
- Chérif, M., Asselin, A. and Bélanger, R. (1994). Defense responses induced by soluble silicon in cucumber roots infected by *Pythium* spp. *Phytopathology*, 84: 236-242.
- Chong, K. P. (2012). An evaluation of *Ganoderma* fungal colonisation using ergosterol analysis and quantification. *The Planter*, 88: 311-319.
- Chung, G. F. (2011). Management of *Ganoderma* diseases in oil palm plantations. *The Planter*, 87(1022): 325-339.
- Clemens, S. (2006). Toxic metal accumulation, responses to exposure and mechanisms of tolerance in plants. *Biochimie*, 88: 1707-1719.
- Corden, M. E. (1965). Influence of calcium nutrition on fusarium wilt of tomato and polygalacturonase activity. *Phytopathology*, 55: 222-224.
- Corley, R. H. V. and Mok, C. K. (1972). Effects of nitrogen, phosphorus, potassium and magnesium on growth of the oil palm. *Experimental Agriculture*, 8: 347-353.
- Corley, R. H. V. and Tinker, P. B. (2003). *The Oil Palm* (pp. 284). Oxford, UK: Blackwell Science.
- Correia, M. J., Pereira, J. S., Chaves, M. M., Rodrigues, M. L. and Pacheco, C. A. (1995). ABA xylem concentration determines maximum daily leaf conductance of field-grown *Vitis vinifera* L. plants. *Plant Cell Environment*, 18: 511-52.

- Darmono, T. W. (1998). *Ganoderma in Oil Palm in Indonesia: Current Status and Prospective Use of Antibodies for the Detection of Infection*. UK: Taylor & Francis Ltd.
- Dharmaputra, O. S., Tjitrosomo, H. S. and Abadi, A. I. (1989). Antagonistic effect of four fungal isolates to *Ganoderma boninense*, the causal agent of basal stem rot of oil palm. *Biotropia*, 3: 41-49.
- Dik, A. J. and Albajes, R. (1999). Integrated pest and disease management in greenhouse crops. In: Albajes, R., Lodovica Gullino, M., J van Lenteren, J. C. and Elad, Y. (Eds.), *Principles of Epidemiology, Population Biology, Damage Relationships and Integrated Control of Diseases and Pests*. Hiedelber: Springer.
- Duffy, B. (2007). Zinc and plant disease. In: Datnoff, L. E. (Ed.), *Mineral Nutrition and Plant Disease* (pp. 155-176). St Paul: The American Phytopathological Society.
- Durand, T. G., Asmady, H., Flori, A., Jacquemard, J. G., Hayun, Z., Breton, F. and Franqueville, H. (2005). Possible sources of genetic resistant in oil palm (*Elaeis guineensis* Jacq.) to basal stem rot caused by *Ganoderma boninense*-prospects for future breeding. *Mycopathologia*, 159: 93-100.
- Fageria, N. K. (2009). *The Use of Nutrients in Crop Plants* (pp. 436-460). Boca Raton, FL: CRC Press.
- Fageria, N. K., Baligar, V. C. and Jones, C. A. (2011). *Growth and Mineral Nutrition of Field Crops* (pp. 195-218). Boca Raton, FL: CRC Press.
- Fawe, A., Abou-Zaid, M., Menzies, J. G. and Be langer, R. R. (1998). Silicon-mediated accumulation of flavonoid phytoalexins in cucumber. *Phytopathology*, 88: 396-401.
- Fekadu, A. and Tesfaye, A. (2013). Antifungal activity of secondary metabolites of *Pseudomonas fluorescens* isolates as a biocontrol agent of chocolate spot disease of faba bean in Ethiopia. *African Journal of Microbiology Research*, 7(47): 5364-5373.
- Fitzherbert, E. B., Struebig, M. J., Morel, A., Danielsen, F., Brühl, C. A., Donald, P. F. and Phalan, B. (2008). How will oil palm expansion affect biodiversity? *Trends in Ecology and Evolution*, 23(10): 538-545.
- Flood, J. and Hasan, Y. (2004). Basal stem rot- taxonomy, biology, epidemiology, economic status and control in Southeast Asia and Pacific Islands. Paper 8. In: *International Conference on Pests and Diseases of Importance to the Oil Palm Industry. Fostering Global Cooperation in Instituting Quarantine Shield*, 18-19 May 2004. Kuala Lumpur, Malaysia.
- Flood, J., Cooper, R., Rees, R., Potter, U. and Hasan, Y. (2000). Some latest R&D on *Ganoderma* diseases in oil palm. Presented at IOPRI/MPOB seminar:

Advances in the Controlling of Devastating Disease of Oil Palm (Ganoderma) in South East Asia, November 2000. Jogjakarta, Indonesia.

- Flood, J., Hasan, Y., Rees, R., Potter, U. and Cooper, R. (2010). Some latest R&D on *Ganoderma* diseases in oil palm. Presented at Second International Seminar Oil Palm Diseases: *Advances in Ganoderma Research and Management*, 31 May 2010. Yogyakarta, Indonesia.
- Friedel, C. (1987). Sur des matieres grasses trouvees dans les tombes Egyptiennes d'abydos. *Comptes Rendus de l'Academie des Sciences*, 124: 648-659.
- Fujii, H., Hayasaka, T., Yokoyama, K. and Ando, H. (1999). Effect of silica gel application to a nursery bed of rice on rooting ability and early growth of rice plants. *Japanese Society Soil Science and Plant Nutrition*, 70: 785-790.
- Gait, C. F. (2011). Management of *Ganoderma* disease in oil palm plantations. *The Planter*, 87(1022): 325-339.
- George, S. T., Chung, G. F. and Zakaria, A. (1996). Updated results on trunk injection of fungicides for the control of *Ganoderma* basal stem rot. In: *Proceedings of the 1996 PORIM International Palm Oil Congress-Agriculture*, 22-23 July 1996. PORIM, Bangi, Selangor.
- Gerasimos, F. K., Anastasia, P. T. and Dimitrios, G. G. (2013). Characterization of the biocontrol activity of *Pseudomonas fluorescens* strain X reveals novel genes regulated by glucose. *PLoS ONE*, 8(4): 61-80.
- Goh, K. J. and Härdter, R. (2003). General oil palm nutrition. In: Fairhurst, T. and Härdter, R. (Eds.), *Oil palm: Management for Large and Sustainable Yield* (pp. 191-228). Potash and Institute (PPI), Potash and Institute of Canada (PPIC) and International Potash institute (IPI).
- Gong, H., Zhu, X., Chen, K., Wang, S. and Zhang, C. (2005). Silicon alleviates oxidative damage of wheat plant in pots under drought. *Plant Science*, 169: 313-321.
- Grabber, J. H., Hatfield, R. D. and Ralph, J. (1998). Diferulate cross-links impede the enzymatic degradation of nonlignified maize walls. *Journal of the Science and Food Agriculture*, 77(2): 193-200.
- Graham, D. R. (1983). Effects of nutrients stress on susceptibility of plants to disease with particular reference to the trace elements. *Advances in Botanical Research*, 10: 221-276.
- Gross, G. G. (1980). The biochemistry of lignifications. *Advance in Botanical Research*, 8: 25.
- Haniff, M. H., Ismail, S. and Idris, A. S. (2005). Gas exchange responses of oil palm to *Ganoderma boninense* infection. *Asian Journal of Plant Science*, 4(4): 438-444.

- Hardon, J. J. (1995). Oil palm. In: Smartt, J. and Simmonds, N. W. (Eds.), *Evolution of Crop Plants* (pp. 395-399). London: Longman Scientific and Technical.
- Hartley, C. W. S. (1977). *The Oil Palm*. London: Longman.
- Hartley, C. W. S. (1988). *The Oil Palm* (pp. 761). New York: John Wiley and Sons.
- Hasan, Y. and Turner, P. D. (1998). The comparative importance of different oil palm tissues as infection sources for basal stem rot in replantings. *The Planter*, 74: 119-135.
- Hassan, M. A. and Shirai, Y. (2003). Palm biomass utilization in Malaysia for the production of bioplastic. http://www.biomass-asia-workshop.jp/presentation_files/21_AliHassan.pdf (Accessed on 10 October 2014).
- Hattori, T., Sonobe, K., Inanaga, S., An, P. and Morita, S. (2008). Effects of silicon on photosynthesis of young cucumber seedlings under osmotic stress. *Journal of Plant Nutrition*, 31(6): 37-41.
- Hayasaka, T., Fujii, H. and Ishiguro, K. (2008). The role of silicon in preventing appressorial penetration by the rice blast fungus. *Phytopathology*, 98: 1038-1044.
- Henriques, A. R., Chalfun-Junior, A. and Aarts, M. (2012). Strategies to increase zinc deficiency tolerance and homeostasis in plants. *Brazilian Journal of Plant Physiology*, 24: 3-8.
- Higuchi, T. (1985). Biosynthesis of lignin. In: Higuchi, T. (Ed.), *Biosynthesis and Biodegradation of Wood Components* (pp. 161-188). New York: Academic Press.
- Ho, C. T. and Hashim, K. (1997). Usefulness of soil mounding treatments in prolonging productivity of prime-aged *Ganoderma* infected palms. *The Planter*, 73: 239-244.
- Hollis, J. P. (1951). Bacteria in healthy potato tissue. *Phytopathology*, 41: 350-366.
- Hossain, M. T., Mori, R., Soga, K., Wakabayashi, K., Kamisaka, S., Fujii, S., Yamamoto, R. and Hoson, T. (2002). Growth promotion and an increase in cell wall extensibility by silicon in rice and some other Poaceae seedlings. *Journal of Plant Research*, 115: 23-27.
- Huber, D. M. (1980). The role of mineral nutrition in defense. In: Horsfall, J. G. and Cowling, E. B. (Eds.), *Plant Disease* (pp. 381-406). Academic Press.
- Huber, D. M. and Thompson, I. A. (2007). Nitrogen and plant disease. In: Datnoff, L. E., Elmer, W. H. and Huber, D. M. (Eds.), *Mineral Nutrition and Plant Disease* (pp. 31-44). St. Paul: The American Phytopathological Society.

- Huber, D. M. and Watson, R. D. (1974). Nitrogen form and plant disease. *Annual Review of Phytopathology*, 12: 139-165.
- Huber, D. M. and Wilhelm, N. S. (1988). The role of manganese in resistance to plant diseases. In: Graham, R. D., Hannam, R. J. and Uren, N. C. (Eds.), *Manganese in soils and plants (pp. 155-173)*. Dordrecht, The Netherlands: Kluwer Academic Publishers.
- Idris, A. S. (1999). Basal stem rot (BSR) of oil palm (*Elaeis guineensis* Jacq.) in Malaysia: factors associated with variation in disease severity. PhD Thesis. University of London, UK.
- Idris, A. S. (2009). Basal Stem Rot in Malaysia- Biology, Epidemiology, Economic Importance, Detection and Control. In: Idris, A. S. (Ed.), *International Workshop on Awareness, Detection and Control of Oil Palm Devastating Disease Emerging Diseases of Oil Palm- Phytoplasma Disease, Orange Spotting and Cercospora Leaf Spot: Biology, Epidemiology, Detection, Control and Future Research*, 6 November 2009. Malaysia.
- Idris, A. S. (2011). Further advances in oil Palm research. In: Basri, M. W., Choo, Y. M. and Chan, K. W. (Eds.), *Other Devastating Diseases of Oil Palm*, (pp. 522-542). MPOB, Malaysia.
- Idris, A. S., Ariffin, D. and Singh, G. (2000). Status of *Ganoderma* in oil palm. In: Flood J., Bridge, P. D. and Holderness, M. (Eds.), *Ganoderma: Diseases of Perennial Crops*. New York: CABI Publishing.
- Idris, A. S., Ismail, S. and Ariffin, D. (2004). Prolonging the productive life of *Ganoderma*-infected palms with hexaconazole. MPOB Information Series No. 221, MPOB TT No. 214.
- Idris, A. S., Ismail, S., Ariffin, D. and Ahmad, H. (2002). Control of *Ganoderma*-infected palm-development of pressure injection and field applications. MPOB Information Series No.148, MPOB TT No. 131.
- Idris, A. S., Mior, M. H. A. Z., Maizatul, S. M. and Kushairi, A. (2011). Survey on status of *Ganoderma* disease of oil palm in Malaysia 2009-2010. In: *Proceedings of the 1993 PORIM International Palm Oil Congress Fortifying and Energizing World*, 15-17 November. Palm Oil Research Institute of Malaysia, Bangi.
- Idris, A. S., Mohd Shukri, I., Norman, K., Kushairi, A., Choo, Y. M., Hanafi, M. M., Talib, R. and Sofian, M. Y. M. (2014). GanoCare™ - Reducing risk of *Ganoderma* infection in oil palm. MPOB Information Series No. 653, MPOB TT No. 545.
- Idris, A. S., Mohd Tayeb, D., Ariffin, D., Zulkifli, D., Zulkifli, M., Mohd Rafii, Y. and Afandi, M. (2001). Morphology and pathogenicity of *Ganoderma* basal stem rot of oil palm obtained from Sabah and Sarawak. In: *Proceedings of the*

- Malaysian Science and Technology Congress*, 24-26 September 2001. Kota Kinabalu, Sabah.
- Ilias, G. N. M. (2000). *Trichoderma* and its efficacy as a bio-control agent of basal stem rot of oil palm (*Elaeis guineensis* Jacq.). Unpublished doctoral dissertation. Universiti Putra Malaysia.
- Inanaga, S., Okasaka, A. and Tanaka, S. (1995). Does silicon exist in association with organic compounds in rice plant? *Japan Journal of Soil Science and Plant Nutrition*, 11: 111-117.
- Jitendra, D. and Nikhilesh, K. (2013). Antagonistic and plant growth promoting potentials of indigenous endophytic bacteria of soybean. *Current Research in Microbiology and Biotechnology*, 1(2): 62-69.
- Jollands, P. (1983). Laboratory investigations on fungicides and biological agents to control three diseases of rubber and oil palm and their potential applications. *Tropical Pest Management*, 29: 33-38.
- Jones, J. B. and Huber, D. M. (2007). Magnesium and plant disease. In: Datnoff, L. E., Elmer, W. H. and Huber, D. M. (Eds.), *Mineral Nutrition and Plant Disease* (pp. 95-100). St. Paul: APS Press.
- Kashyap, R. K. (2009). *Soil Fertility, Fertilizer and Integrated Nutrient Management*. Jaipur, India: Oxford.
- Kaufman, P. B., Takeoka, Y., Carlson, T. J., Bigelow, W. C., Jones, J. D., Moore, P. H. and Gbosbeh, N. S. (1979). Studies on silica deposition in sugarcane, using scanning electron microscopy, energy dispersive X-ray analysis, neutron activation analysis and light microscopy. *Phytomorphology*, 29: 185-93.
- Khairudin, H. (1990). Basal stem rot of oil palm: Incidence, etiology and control. Master of Agricultural Science Thesis. Faculty of Agriculture, University Pertanian Malaysia.
- Khairudin, H. (1993). Basal stem rot of oil palm caused by *Ganoderma boninense*. An update. In: Jalani, B. S., Ariffin, D., Rajanaidu, N., Mohd Tayed, D., Paranjothy, K., Mohd Basri, W., Henson, I. E. and Chong, K. C. (Eds.), *Proceedings of the 1993 PORIM International Palm Oil Congress "Update and Vision" Agriculture*, (pp. 735- 738). Palm Oil Research Institute of Malaysia, Kuala Lumpur, Malaysia.
- Kim, S. G., Kim, K. W., Park, E. W. and Choi, D. (2002). Silicon-induced cell wall fortification of rice leaves: a possible cellular mechanism of enhanced host resistance to blast. *Phytopathology*, 92: 1095-1103.
- Kinge, T. R. and Mih, A. M. (2011). *Ganoderma ryvardense* sp. nov. associated with basal stem rot (BSR) disease of oil palm in Cameroon. *Mycosphere*, 2(2): 179-188.
- Klopper, J. W., Wei, G. and Tuzun, S. (1992). Rhizospore population dynamics and internal colonization of cucumber by plant growth- promoting rhizobacteria

- which induce systemic resistance to *Colletotrichum orbiculare*. In: Tjamos, E. S. (Ed.), *Biological Control of Plant Disease* (pp. 185-191). New York: Plenum Predd.
- Kobayashi, D. Y. and Palumbo, J. D. (2000). Bacterial endophytes and their effects on plants and uses in agriculture. In: Bacon, C. W. and White, J. F. (Eds.), *Microbial Endophytes* (pp. 199-236). New York: Dekker.
- Kommedahl, T. and Windels, C. E. (1981). Introduction of microbial antagonists to specific courts of infection: seeds, seedlings and wounds. In: Beemster, A. B. R., Bollen, G. J., Gerlagh, M., Ruissen, M. A., Schippers, B. and Tempel, A. (Eds.), *Biotic Interaction and Soil-Borne Diseases* (pp. 121-127). Netherlands Society of Plant Pathology.
- Kosuge, T. (1969). The role of phenolics in hist responses to infection. *Annual Reviews of Phytopathology*, 7: 195-222.
- Kranz, J. (1988). Measuring plant disease. In: Kranz, J. and Rotem, J. (Eds.), *Experimental Techniques in Plant Disease Epidemiology* (pp. 35-50). New York: Springer-Verlag.
- Krupinsky, J. M., Bailey, K. L., McMullen, M. P., Gossen, B. D. and Turkington, T. K. (2002). Managing plant disease risk in diversified cropping systems. *Agronomy Journal*, 94: 198-209.
- Lamb, T. G., Tonkyn, D. W. and Kluepfel, D. A. (1996). Movement of *Pseudomonas aureofaciens* from the rhizosphere to aerial plant tissue. *Canadian Journal of Microbiology*, 42: 1112-1120.
- Legros, S., Mialet-Serra, I., Caliman, J. P., Siregar, F. A., Clement-Vidal, A., Fabre, D. and Dingkuhn, M. (2009). Phenology, growth and physiological adjustments of oil palm (*Elaeis guineensis*) to sink limitation induced by fruit pruning. *Annals of Botany*, 104: 1183-1194.
- Liang, Y., Sun, W., Si, J. and Romheld, V. (2005). Effects of foliar and root-applied silicon on the enhancement of induced resistance to powdery mildew in *Cucumis sativus*. *Plant Pathology*, 54: 678-685.
- Lim, T. K., Chung, G. F. and Ko, W. H. (1992). Basal stem rot of oil palm caused by *Ganoderma boninese*. *Plant Pathology Bulletin*, 1: 147-152.
- Lin, C. C., Chen, L. M. and Liu, Z. H. (2005). Rapid effect of copper on lignin biosynthesis in soybean root. *Plant Science*, 168: 855-861.
- Liu, L., Klopper, J. W. and Tuzun, S. (1995). Induction of systemic resistance in cucumber against bacterial angular leaf spot by plant growth-promoting rhizobacteria. *Journal of Phytopathology*, 85: 843-847.
- Lo, C. T., Nelson, E. B. and Harman, G. E. (1997). Improved biocontrol efficacy of *Trichoderma harzianum* for foliar phases of turf diseases by use of spray applications. *Plant Disease*, 81: 1132-1138.

- Maathuis, F. J. and Diatloff, E. (2013). Roles and functions of plant mineral nutrients. In: *Plant Mineral Nutrients* (pp. 1-21). Humana Press.
- Mark, S. R., Steven, L. R. and Joshua, H. F. (2012). Nitrogen fertilization and growth regulator impacts on tuber deformity, rot and yield russet potatoes. *International Journal of Agronomy*, 20(12): 1-7.
- Marschner, H. (1995). Relationship between mineral nutrition and plant disease and pests. In: *Mineral Nutrition of Higher Plants 2nd Edition* (pp. 436-460). New York: Academic Press.
- Matsukuma, S., Okuda, T. and Watanabe, J. (1994). Isolation of *Actinomycetes* from pine litter layers. *Actinomycetology*, 8: 57-65.
- Mielke, T. (2012). *Oil World Annual, Vol. 1*. Hamburg, Germany: ISTA Mielke GmbH.
- Miller, R. N. G., Holderness, M., Bridge, P. D., Chung, G. F. and Zakaria, M. H. (1999). Genetic diversity of *Ganoderma* in oil palm plantings. *Plant Pathology*, 48: 593-603.
- Monties, B. and Fukushima, Y. (2001). Occurrence, function and biosynthesis of lignins. In: Hofrichter, M. and Steinbüchel, A. (Eds.), *Biopolymers, Vol. 1 Lignins, Humic Substances and Coal* (pp. 1-64). Weinheim, Germany: Wiley-VCH.
- Mousavi, S. R., Galavi, M. and Rezaei, M. (2013). Zinc (Zn), importance for crop production-a review. *International Journal of Agronomy and Plant Production*, 4(1): 64-68.
- Moyer, C., Peres, N. A., Datnoff, L. E., Simonne, E. H. and Deng, Z. (2008). Evaluation of silicon for managing powdery mildew on gerbera daisy. *Journal of Plant Nutrition*, 31(12): 2131-2144.
- MPOB (2008). Statistic 2006/2007. <http://www.mpob.gov.my> (Accessed on 16 April 2015).
- MPOB (2014). *Annual Research Review 2014*. Malaysian Palm Oil Board, Bangi.
- Muchow, R. C. (2009). Effect of leaf nitrogen and water regime on the photosynthetic capacity of kenaf (*Hibiscus cannabinus* L.) under field conditions. *Australian Journal of Agricultural Research*, 41(5): 845-852.
- Naik, P. R., Raman, G., Narayanan, K. D. and Sakthivel, N. (2008). Assessment of genetic and functional diversity of phosphate solubilizing fluorescent *Pseudomonads* isolated from rhizospheric soil. *Biomedcentral Microbiology*, 8: 230.
- Najihah, N. I., Hanafi, M. M., Idris, A. S. and Hakim, M. A. (2015). Silicon treatment in oil palms confers resistance to basal stem rot disease caused by *Ganoderma boninense*. *Crop Protection*, 67: 151-159.

- Nashwa, A. S., Shamas, N. R., Mohamad, S. M. and Ahmed, S. E. (2013). Formulations of *Bacillus* spp. and *Pseudomonas fluorescens* for biocontrol of cantaloupe root rot caused by *Fusarium solani*. *Journal of Plant Protection Research*, 53(3): 295-300.
- Nazeeb, M., Barakabah, S. S. and Loong, S. G. (2000). Potential of high intensity oil palm plantings in diseased environments. *The Planter*, 76: 699-710.
- Ning, D., Song, A., Fan, F., Li, Z. and Liang, Y. (2014). Effects of slag-based silicon fertilizer on rice growth and brown-spot resistance. *Plos one*, 9: e102681.
- Noorhaida, S. and Idris, A. S. (2009). *In vitro*, colonization and nursery evaluation of endophytic fungi as biological control of *Ganoderma boninense*, causal agent of basal stem rot disease of oil palm. In: *Proceedings of PIPOC 2009*, 14 November 2009. KLCC, Kuala Lumpur.
- Nur Ain Izzati, M. Z. and Abdullah, F. (2008). Disease suppression in *Ganoderma* infected oil palm seedlings treated with *Trichoderma harzianum*. *Plant Protection Science*, 44(3): 101-259.
- Nur Sabrina, A. A., Sariah, M. and Zaharah, A. R. (2012). Suppression of basal stem rot progress in oil palm (*Elaeis guineensis*) after copper and calcium supplementation. *Pertanika Journal of Tropical Agricultural Science*, 35(S): 13-24.
- Okazaki, T., Takahashi, K., Kizuka, M. and Enokita, R. (1995). Studies on *Actinomycetes* isolated from plant leaves. *Annual Report Sankyo Research Laboratory*, 47: 97-106.
- Paterson, R. R. M., Sariah, M. and Lima, N. (2009). The feasibility of producing oil palm with altered lignin content to control *Ganoderma boninense*. *Journal of Phytopathology*, 157: 649-656.
- Peltonen, J., Kittila, S., Peltonen-Sainio, P. and Karjalainen, R. (1991). Use of foliar-applied urea to inhibit the development of *Septoria nodorum* in spring wheat. *Crop Protection*, 10: 260-264.
- Petrini, O. (1990). Fungal endophyte of tree leaves. In: Andrews, J. and Hirano, S. S. (Eds.), *Microbial Ecology of Leaves* (pp. 179-197). New York: Springer Verlag.
- Pilotti, C. A. (2005). Stem rot of oil palm caused by *Ganoderma boninense*: pathogen biology and epidemiology. *Mycopathologia*, 159: 129-137.
- Pintro, J. C. and Taylor, G. J. (2005). Calcium requirement in the background nutrient solution on growth of wheat plants using the relative addition rate technique. *Journal of Plant Nutrition*, 28: 551-565.
- Potash and Phosphate Institute (PPI) (1988). Phosphorus nutrition improves plant disease resistance. In: Potash and Phosphate Institute (Ed.), *Better Crops with Plant Food* (pp. 22-23). Atlanta: Potash and Phosphate Institute.

- Prabhu, A. S., Fageria, N. K. and Huber, D. M. (2007). Potassium nutrition and plant diseases. In: Datnoff, L. E., Elmer, W. H. and Huber, D. M. (Eds.), *Mineral Nutrition and Plant Disease*. St Paul, MN: American Phytopathological Society.
- Purba, R. Y., Purba, A. R. and Sipayung, A. (1994). Uji resistensi beberapa persilangan kelapa sawit *D x P* terhadap *Ganoderma boninense* PAT. *Bulletin PPKS*, 2: 81-88.
- Purseglove, J. W. (1975). *Tropical Crops: Monocotyledons (pp. 609)*. Harlow, England: English Language Book Society Longman.
- Ramasamy, S. (1972). Cross-infectivity and decay ability of *Ganoderma* species parasitic to rubber, oil palm and tea. Bachelor Degree. University of Malaya.
- Rankine, I. and Fairhurst, T. H. (1999). Management of phosphorus, potassium and magnesium in mature oil palm. *Better Crop International*, 13(1).
- Rankine, I. R. and Fairhurst, T. H. (1998). *Field Handbook: Oil Palm Series Volume 3-Mature*. Singapore: Potash & Phosphate Institute.
- Rao, V., Lim, C. C., Chia, C. C. and Teo, K. W. (2003). Studies on *Ganoderma* spread and control. *The Planter*, 79(929): 367-383.
- Rebitanim, N. A., Rebitanim, N. Z. and Tajudin, N. S. (2015). Impact of silicon in managing important rice diseases: blast, sheath blight, brown spot and grain discoloration. *International Journal of Agronomy and Agricultural Research*, 6(3): 71-85.
- Rebitanim, N. Z., Ghani, W. A. W. A. K., Rebitanim, N. A. and Salleh, M. A. M. (2013). Potential application of wastes from energy generation particularly biochar in Malaysia. *Renewable and Sustainable Energy Review*, 21: 694-702.
- Rees, R. W., Flood, J., Hasan, Y. and Cooper, R. M. (2007). Low soil temperature and root-inoculum contact enhance *Ganoderma* infection of oil palm: implications for late disease appearance in plantations and screening for disease resistance. *Plant Pathology*, 56: 862-870.
- Rees, R. W., Flood, J., Hasan, Y., Wills, M. A. and Cooper, R. M. (2012). *Ganoderma boninense* basidiospores in oil palm plantations: evaluation of their possible role in stem rots of *Elaeis guineensis*. *Plant Pathology*, 61: 567-578.
- Reuter, D. J. and Robinson, J. B. (1986). *Plant Analysis an Interpretive Manual*. Melbourne, Australia: Inkata Press.
- Rodrigues, F. A., McNally, D. J., Datnoff, L. E., Jones, J. B., Labbe, C., Benhamou, N., Menzies, J. G. and Belanger, R. R. (2004). Silicon enhances the accumulation of diterpenoid phytoalexins in rice: a potential mechanism for blast resistance. *Phytopathology*, 94: 177-183.

- Rodrigues, F. Á., Vale, F. X. R., Korndorfer, G. H., Prabhu, A. S., Datnoff, L. E., Oliveira, A. M. A. and Zambolim, L. (2003). Influence of silicon on sheath blight of rice in Brazil. *Crop Protection*, 22: 23-29.
- Rodrigues, K. F. (1996). Fungal endophytes of palms. In: Redlin, S. C. and Carris, L. M. (Eds.), *Endophytic Fungi in Grasses and Woody Plants: Systematics, Ecology and Evolution* (pp. 121-132). St. Paul, Minnesota, USA: APS Press.
- Rodrigues, M. L., Nimrichter, L., Oliveira, D. L., Frases, S., Miranda, K., Zaragoza, O., Alvarez, M., Nakouzi, A., Feldmesser, M. and Casadevall, A. (2007). Vesicular polysaccharide export in *Cryptococcus neoformans* is a eukaryotic solution to the problem of fungal trans-cell wall transport. *Eukaryotic Cell*, 6: 48-59.
- Rodriguez, H. and Fraga, R. (1999). Phosphate solubilizing bacteria and their role in plant growth promotion. *Biotechnology Advances*, 17: 319-339.
- Rong, L. H., Guo-ping, W., Xiao-hong, L., Chu-long, Z. and Fu-cheng, L. (2009). Antagonistic bioactivity of an endophytic bacterium isolated from *Epimedium brevicornu* Maxim. *African Journal of Biotechnology*, 8(2): 191-195.
- Sanderson, F. R., Pilotti, C. A. and Bridge, P. D. (2000). Basidiospores: their influence on our thinking regarding a control strategy for basal stem rot of oil palm. In: Flood, J. (Ed.), *Ganoderma Diseases of Perennial Crops* (pp. 113-120). UK: CABI Publishing.
- Sapak, Z., Sariah, M. and Ahmad, Z. A. M. (2008). Effect of endophytic bacteria on growth and suppression of *Ganoderma boninense* infection in oil palm. *International Journal of Agricultural Biology*, 10: 127-132.
- Sariah, M. and Zakaria, H. (2000). The use of soil amendments for the control of basal stem rot of oil palm seedlings. In: Flood, J. (Ed.), *Ganoderma Diseases of Perennial Crops* (pp. 89-100). UK: CABI Publishing.
- Sariah, M., Choo, C. W., Zakaria, H. and Norihan, M. S. (2005). Quantification and characterization of *Trichoderma* spp. from different ecosystems. *Mycopathologia*, 159: 113-117.
- Sariah, M., Joseph, H. and Zakaria, H. (1997). Suppression of basal stem rot of oil palm seedlings by calcium nitrate. *The Planter*, 73(856): 359-361.
- Sathya, B., Indu, H., Seenivasan, R. and Geetha, S. (2010). Influence of seaweed liquid fertilizer on the growth and biochemical composition of legume crop *Cajanus cajan*. *Journal of Phytology*, 2(5): 50-63.
- Sattar, A., Cheema, M. A., Basra, S. M. A. and Wahid, A. (2013). Optimization of source and rate of soil applied silicon for improving the growth of wheat. *Pakistan Journal of Agricultural Science*, 50(1): 63-68.

- Sayer, J., Ghazoul, J., Nelson, P. and Boedhihartono, A. K. (2012). Oil palm expansion transforms tropical landscapes and livelihoods. *Global Food Security*, 1: 114-119.
- Schutte, K. H. (1967). The influence of boron and copper deficiency upon infection by *Erysiphe graminis* D. C. the powdery mildew in wheat var. Kenya. *Plant Soil*, 27: 450-452.
- Semangun, H. (1990). *Penyakit-Penyakit Tanaman Perkebunan di Indonesia*. Yogyakarta: Gadjah Mada University Press.
- Sharma, M. (2007). Challenges facing the Malaysian palm oil industry-multi-pronged strategies for raising oil yield, productivity and profitability. *The Planter*, 83(981): 797-833.
- Sharma, M. and Tan, Y. P. (1990). Performance of the *Elaeis oleifera* x *Elaeis guineensis* (OG) and their back-crosses. In: *Proceedings of the 1989 PORIM International Palm Oil Conference (Agriculture)*. Palm Oil Research Institute of Malaysia, Bangi.
- Sheng, X. F. and He, L. V. (2006). Solubilization of potassium bearing minerals by a Wild-type strain of *Bacillus edaphicus* and its mutants and increased potassium uptake by wheat. *Canadian Journal of Microbiology*, 52: 66-72.
- Shuit, S. H., Tan, K. T., Lee, K. T. and Kamaruddin, A. H. (2009). Oil palm biomass as a sustainable energy source: a Malaysian case study. *Energy*, 34: 1225-1235.
- Shukla, A. N. and Uniyal, K. (1989). Antagonistic interactions of *Ganoderma lucidum* (lyss.) Karst. against some soil microorganisms. *Current Science*, 58: 265-267.
- Simeh, M. A. and Fairuz, M. K. (2009). An overview of Malaysia palm oil market share in selected markets. *Oil Palm Industry Economic Journal*, 9(1): 29-36.
- Simoglou, B. K. and Dordas, C. (2006). Effect of foliar applied boron, manganese and zinc on tan spot in winter durum wheat. In: *Proceedings of the 12th Congress of Mediterranean Phytopathological Union*, 11-15 June 2006. Rhodes Island Hellas.
- Sinclair, W. A., Lyon, H. H. and Johnson, W. T. (1987). *Diseases of Trees and Shrubs*. Ithaca, New York: Cornell University Press.
- Singh, G. (1990). *Ganoderma* – the scourge of oil palm in coastal areas. *The Planter*, 67 (786): 421-444.
- Singh, R. P., Ibrahim, M. H., Norizan, E. and Iliyana, M. S. (2010). Composting of waste from palm oil mill: a sustainable waste management practice. *Review of Environmental Science and Biotechnology*, 9: 331-344.
- Stangoulis, J. C. R. and Graham, R. D. (2007). Boron and plant disease. In: Datnoff, L. E., Elmer, W. H. and Huber, D. M. (Eds.), *Mineral Nutrition and Plant Disease (pp. 207-214)*. St. Paul, MN: The American Phytopathological Society.

- Sticknothe, H. and Schuchardt, F. (2011). Life cycle assessment of two palm oil production systems. *Biomass and Bioenergy*, 35: 3976-3984.
- Sturz, A. V., Christie, B. R. and Nowak, J. (2000). Bacterial endophytes: potential role in developing sustainable system of crop production. *Critical Reviews Plant Sciences*, 19: 1-30.
- Sugimoto, T., Watanabe, K., Yoshida, S., Aino, M., Irie, K., Matoh, T. and Biggs, A. R. (2008). Select calcium compounds reduce the severity of *Phytophthora* stem rot of soybean. *Plant Disease*, 92: 1559-1565.
- Sumathi, S., Chai, S. P. and Mohamed, A. R. (2008). Utilization of oil palm as a renewable energy in Malaysia. *Renewable and Sustainable Energy Reviews*, 12: 2404-2421.
- Sun, W., Zhang, J., Fan, Q., Xue, G., Li, Z. and Liang, Y. (2010). Silicon enhanced resistance to rice blast is attributed to silicon-mediated defense resistance and its role as physical barrier. *European Journal of Plant Pathology*, 128: 39-49.
- Sundram, S. (2005). Performance of *Trichoderma harzianum* Rifai. as a biological control agent for basal stem rot of oil palm (*Elaeis guineensis* Jacq.) caused by *Ganoderma boninense* Pat. Master of Science Thesis. Faculty of Science, University Putra Malaysia.
- Sundram, S. (2012). Assessment of *Ganoderma* infection in oil palm (*Elaeis guineensis* Jacq.) by pre-inoculation of arbuscular mycorrhiza fungi and endophytic bacteria. PhD Thesis. Institute of Tropical Agriculture, Universiti Putra Malaysia.
- Sundram, S. (2013). The effects of *Trichoderma* in surface mulches supplemented with conidial drenches in the disease development of *Ganoderma* basal stem rot in oil palm. *Journal of Oil Palm Research*, 24(3): 314-325.
- Sundram, S. and Idris, A. S. (2009). *Trichoderma* as a biological agent against *Ganoderma* in oil palm. MPOB Information Series, No. 463, MPOB TT No. 422.
- Sundram, S., Meon, S., Seman, I. A. and Othman, R. (2014). Application of arbuscular mycorrhizal fungi with *Pseudomonas aeruginosa* UPMP3 reduces the development of *Ganoderma* basal stem rot disease in oil palm seedlings. *Mycorrhiza*, 25: 387-397.
- Sundram, S., Sariah, M., Idris, A. S. and Radziah, O. (2011). Symbiotic interaction of endophytic bacteria with arbuscular mycorrhizal fungi and its antagonistic effect on *Ganoderma boninense*. *The Journal of Microbiology*, 49(4): 551-557.
- Susanto, A. (2009). Basal stem rot in Indonesia. Biology, economic importance, epidemiology, detection and control. In: *Proceedings of International Workshop on Awareness, Detection and Control of Oil Palm Devastating Diseases*, November 2009. Kuala Lumpur Convention Center, Malaysia.

- Susanto, A., Sudharto and Purba, R. Y. (2002). Superior biocontrol agents of *Ganoderma boninense* pathogen of basal stem rot disease in oil palm plantations. In: *International Oil Palm Conference*. Indonesian Oil Palm Research Institute, Bali, Indonesia.
- Susanto, A., Sudharto, P. S. and Purba, R. Y. (2005). Enhancing biological control of basal stem rot disease (*Ganoderma boninense*) in oil palm plantations. *Mycopathologia*, 159: 153-157.
- Syed, R. A., Law, I. H. and Corley, R. H. V. (1982). Insect pollination of oil palm: introduction, establishment and pollinating efficiency of *Elaeidobius kamerunicus* in Malaysia. *The Planter*, 58: 547-561.
- Tan, C. J., How, K. C., Lohmia, P. P., Ismet, A., Getha, K., Seki, T. and Vikineswary, S. (2002). Bioactivity of selected actinomycetes against *Ganoderma boninense*. *Asia Pacific Journal of Molecular Biology and Biotechnology*, 10(2): 119-125.
- Tan, K. S. (1983). *The Botany of Oil Palm, Casual Papers on Oil Palm*. Kuala Lumpur: Incorporated Society of Planters.
- Tandon, H. L. S. and Sekhon, G. S. (1989). Potassium research and agricultural production in India. *Potash Review*, 1: 1-11.
- Tayeb, M. D. and Hamdan, A. B. (1999). Relation of fertilizer nutrients to *Ganoderma*. In: *Proceedings of the 1999 PORIM International Palm Oil Conference Malaysia*, (pp. 422-453). Palm Oil Research Institute of Malaysia, Bangi, Selangor, Malaysia.
- Tayeb, M. D., Idris, A. S. and Haniff, M. H. (2003). Reduction of *Ganoderma* infection in oil palm through balanced fertilization in peat. In: *Proceedings of the Agriculture Conference of 2003 PIPOC*, 24-28 August 2003, (pp. 193-219). Malaysian Palm Oil Board, Bangi, Putrajaya, Malaysia.
- Texeira, A. F., Andrade, A. B., Ferrarese-Filho, O. and Ferrarese, M. L. L. (2006). Role of calcium on phenolic compounds and enzymes related to lignifications in soybean [*Glycine max* L.] root growth. *Plant Growth Regulator*, 76: 69-76.
- Thompson, A. (1931). Stem-rot of the oil palm in Malaya. *Bulletin Department of Agriculture*, 6: 23.
- Thompson, I. A. and Huber, D. M. (2007). Manganese and plant disease. In: Datnoff, L. E., Elmer, W. H. and Huber, D. M. (Eds.), *Mineral Nutrition and Plant Disease* (pp. 139-153). St Paul, Minnesota, USA: APS Press.
- Tinker, P. B. (1976). Soil requirements of the oil palm. In: Corley, R. H. V., Hardon, J. J. and Wood, B. J. (Eds.), *Oil Palm Research (Developments in Crop Science [1])* (pp. 165-181). Amsterdam: Elsevier.
- Tomlinson, P. B. and Metcalfe, C. R. (1961). *Anatomy of The Monocotyledons* (pp. 453). Oxford: Clarendon Press.

- Torres, G. A., Sarria, G. A., Varon, F., Coffey, M. D., Elliot, M. L. and Martinez, G. (2010). First report of bud rot caused by *Phytophthora palmivora* on African oil palm in Colombia. *The American Phytopathological Society*, 94(9): 1163.1-1163.1.
- Treu, R. (1998). Macro fungi in oil palm plantations of South East Asia. *Journal of Genetic Mycology*, 12: 10-14.
- Turner, P. D. (1965). The oil palm and *Ganoderma* IV. Avoiding disease in the plantings. *The Planter*, 41: 331-333.
- Turner, P. D. (1968). The use of surgery as a method of treating basal stem rot in oil palms. *The planter*, 44: 303-308.
- Turner, P. D. (1981). *Oil Palm Diseases and Disorders* (pp. 88-110). Oxford University Press.
- Turner, P. D. and Gillbanks, R. A. (2003). *Oil Palm Cultivation and Management* (pp. 633). Kuala Lumpur, Malaysia: The Incorporated Society of Planters.
- Uchida, R. S. (2000). *Essential Nutrients for Plant Growth: Nutrient Functions and Deficiency Symptoms. Plant Nutrient Management in Hawaii's Soils* (pp. 31-55). University of Hawaii at Manoa: College of Tropical Agriculture and Human Resources, Manoa.
- Usherwood, N. R. (1980). The effects of potassium on plant disease. In: Potash and Phosphate Institute (Ed.), *Potassium for Agriculture: A Situation Analysis* (pp. 151-164). Atlanta: Potash and Phosphate Institute.
- Van Loon, L. C., Bakker, P. A. H. M. and Pieterse, C. M. J. (1998). Systemic resistance induced by rhizosphere bacteria. *Annual Review of Phytopathology*, 36: 453-483.
- Van Peer, R. and Schippers, B. (1988). Plant growth responses to bacterization with selected *Pseudomonas* spp. strains and rhizosphere microbial development in hydroponic cultures. *Canadian Journal of Microbiology*, 35: 456-463.
- Varghese, G., Chew, P. S. and Lim, J. K. (1975). Biology and chemically assisted biological control of *Ganoderma*. In: *Proceeding of International Rubber Conference*. Kuala Lumpur.
- Veresoglou, S. D., Barto, E. K., Meneses, G. and Rillig, M. C. (2013). Fertilization affects the severity of disease caused by fungal plant pathogen. *Plant Pathology*, 62: 961-969.
- Volk, R. J., Kahn, R. P. and Weintraub, R. L. (1958). Silicon content of the rice plant as a factor influencing its resistance to infection by the blast fungus, *Piricularia oryzae*. *Phytopathology*, 48: 121-178.
- Wafaa, M. H., Abdel-latif and Faten, M. (2001). Interaction between vesicular arbuscular mycorrhizae and antagonistic biocontrol micro-organisms on

- controlling root-rot disease incidence of *Geranium* plants. *Journal of Biological Science*, 1: 1147-1153.
- Wakefield, E. M. (1920). Diseases of the oil palm in West Africa. *Kew bulletin*, 306-308.
- Wallace, A. and North, C. P. (1962). Metal chelates and virus disease. In: Wallace, A. (Ed.), *A Decade of Synthetic Chelating Agents in Inorganic Plant Nutrition* (pp. 142-145). Ann Arbor, MI: Edward Brothers.
- Walter, M. H. (1992). Regulations of lignification in defense. In: Boller, T. and Meins, F. (Eds.), *Plant Gene Research: Genes Involved in Plant Defense* (pp. 327-352). Vienna: Springer.
- Wan, H. H. (2007). *Ganoderma* disease of oil palm in Sabah. *The Planter*, 83(974): 299-313.
- Wang, S. W. and Galleta, G. J. (1998). Foliar application of potassium silicate induces metabolic changes in strawberry plants. *Journal of Plant Nutrition*, 21: 157-167.
- Whipps, J. M. and Lewis, D. H. (1981). *Effects of Disease in the Physiology of the Plant*. Cambridge University Press.
- Whitesides, S. K. and Spotts, R. A. (1991). Frequency, distribution and characteristics of endophytic *Pseudomonas syringae* in pear trees. *Phytopathology*, 81: 453-457.
- Wijesekera, H. T. R., Wijesundera, R. L. C. and Rajapakse, C. N. K. (1996). Hyphal interactions between *Trichoderma viridae* and *Ganoderma boninense* Pat. the cause of coconut root and bole rot. *Journal of the National Science Sri Lanka*, 24(3): 217-219.
- Wilcove, D. S. and Koh, L. P. (2010). Addressing the threats to biodiversity from oil-palm agriculture. *Biodiversity Conservation*, 19: 999-1007.
- Willats, W. G. T., McCartney, L., Mackie, W. and Knox, J. P. (2001). Pectin cell biology and prospects for functional analysis. *Plant Molecular Biology*, 47: 9-27.
- Wittenbach, V. A., Lin, W. and Herbert, R. R. (1982). Vacuolar localization of proteases and degradation of chloroplasts in mesophyll protoplasts from senescing primary wheat leaves. *Plant Physiology*, 69: 98-102.
- Yeo, A. R., Flowers, S. A., Rao, G., Welfare, K., Senanayake, N. and Flowers, T. J. (1999). Silicon reduces sodium uptake in rice (*Oryza sativa* L.) in saline conditions and this is accounted for by a reduction in the transpirational bypass flow. *Plant Cell Environment*, 22: 559-565.

- Zaiton, S., Sariah, M. and Zainal Abidin, M. A. (2006). Isolation and characterization of microbial endophytes from oil palm roots: implication as biocontrol agents against *Ganoderma*. *The Planter*, 82: 587-597.
- Zaiton, S., Sariah, M. and Zainal Abidin, M. A. (2008). Effect of endophytic bacteria on growth and suppression of *Ganoderma* infection in oil palm. *International Journal of Agricultural Biology*, 10: 127-132.
- Zakaria, Z. A., Sufian, A. S., Ramasamy, K., Ahmat, N., Sulaiman, M. R. and Arifah, A. K. (2010). *In vitro* antimicrobial activity of *Muntingia calabura* extracts and fractions. *African Journal of Microbiology Research*, 4: 304-308.
- Zinniel, D. K., Lambrecht, P., Harris, N. B. and Feng, Z. (2002). Isolation and characterization of endophytic colonizing bacteria from agronomic crops and prairie plants. *Applied Environmental Microbiology*, 68: 2198-2208.