FUNCTIONALLY GRADED PLGA-NANO APATITE-LAURIC ACID BIOCOMPOSITE MEMBRANE FOR POTENTIAL CLINICAL APPLICATIONS

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To my dearest mother.....

Mrs. Jannanayagam

For being a mentor, friend and pillar of strength

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ABSTRACT

Bone healing is a challenge in orthopaedics and dentistry. An occlusive membrane is used for the reconstruction of bone defects in guided bone regeneration (GBR) technique. Infection is the major cause for GBR membrane failure in which multiple antibiotics have been used to prevent bacterial colonisation in regenerative clinical practice. An anti-infective membrane with alternative antimicrobial agent to substitute antibiotics is paramount to overcome the incidence of bacterial resistance In this study, a composite membrane was developed by and side-effects. incorporating lauric acid (LA), a naturally derived antimicrobial substance. Poly(lactic-co-glycolic acid) (PLGA) based composite membrane was successfully fabricated using a combination of solvent casting-thermally induced phase separation (TIPS)-solvent leaching technique. The triple-layered membrane structure was attained via solvent casting of the composite solutions which then immediately phase separated by freezing at -18±1°C for 24 h. Then, the solvent in phase separated membrane was removed by immersing in precooled water at 3±1°C for 26 h, after which the membrane was air dried at 25°C for 3 days. The triple-layered construct of the PLGA composite membrane was developed with a gradient structure of LA and non-stoichiometric nanoapatite (NAp), to deliver the antimicrobial and osteconductive properties, respectively. The surface morphology and phase composition of the membrane were examined using scanning electron microscopy (SEM) and X-ray diffraction (XRD), respectively. The resulting graded membrane consisted of small pore size layer-1 containing 10wt% NAp + 1-3wt% LA, an intermediate labyrinth layer-2 with 20-50wt% NAp + 1wt% LA, and a large pore size layer-3 containing 30-100wt% NAp without LA. The existence of chemical interaction between PLGA, NAp and LA was identified using Fourier transform infrared spectrophotometry (FTIR) analysis. The synergistic effects of 10-30wt% NAp and 1wt% LA in dry membranes demonstrated higher tensile strength $(0.61\pm0.17 \text{ MPa})$ and elastic modulus $(23.15\pm6.19 \text{ MPa})$. However, a more pliable behavior with a decrease in elastic modulus (12.50± 4.32MPa) was observed in 3wt% LA added membrane compared to the pure PLGA (20.17 ± 2.21 MPa). The addition of LA resulted in a plasticizing effect at 3wt% due to weak intermolecular interactions in PLGA chains, caused by LA (-OH) and PLGA (C-O) bondings. These results were corroborated by the FTIR peak shift (1-3 cm⁻¹) and glass transition temperature (T_g) reduction as detected using differential scanning calorimeter (DSC). The composite membrane retained its structural integrity with only 22% weight loss after incubation for 24 weeks in phosphate buffered saline (PBS), which indicates its potential use as a physical barrier. The 1-3wt% LA loaded composite membranes had good cell viability toward mouse fibroblasts and showed increased bacterial reduction with increased LA loadings against S. aureus. These results demonstrate the potential of LA loaded biocomposite membrane to provide anti-infective surfaces, useful in clinical applications.

ABSTRAK

Penyembuhan tulang adalah satu cabaran dalam bidang ortopedik dan pergigian. Pertumbuhan semula tulang berpandukan (GBR) telah digunakan untuk pembinaan semula kecacatan tulang dengan menggunakan membran penghalang. Jangkitan adalah punca utama kegagalan membran tersebut di mana beberapa antibiotic telah digunakan untuk menghalang pertumbuhan bacteria dalam amalan klinikal. Agen antibakteria alternatif adalah perlu untuk mengatasi kesan sampingan dan rintangan bakteria yang dihasilkan oleh antibiotik. Dalam kajian ini, membran komposit telah dibangunkan melalui penggabungan asid laurik (LA) yang mempunyai sifat antibakteria. Membran komposit berasaskan asid poli(laktik-coglycolic) (PLGA) telah berjaya direka dengan menggunakan gabungan teknik-teknik pelarut tuangan-pemisahan fasa haba teraruh-larut lesap pelarut. Struktur membran tiga-lapis telah dihasilkan melalui pelarut tuangan komposit yang telah melalui pemisahan fasa haba teraruh pada suhu -18±1°C selama 24 jam. Kemudian, pelarut membran telah dibuang dengan merendamkannya dalam air sejuk pada suhu 3±1°C selama 26 jam. Setelah itu, membran telah dikeringkan di udara pada 25°C selama 3 Membran komposit PLGA tiga-lapis ini telah difabrikasi dengan struktur hari kecerunan melalui penambahan LA dan apatitnano bukan stoikiometrik (NAp) yang memainkan peranan sebagai antimikrob dan penggalak pertumbuhan tulang. Morfologi permukaan dan fasa komposisi membran telah diperiksa dengan menggunakan mikroskopi elektron imbasan (SEM) dan pembelauan sinar-X (XRD). Membran ini terdiri daripada lapisan-1 dengan saiz liang kecil yang mengandungi 10% berat NAp + 1-3% berat LA, lapisan-2 sebagai lapisan perantaraan dengan 20-50% berat NAp + 1% berat LA dan akhirnya lapisan-3 dengan saiz liang besar yang mengandungi 30-100% berat NAp tanpa LA. Kewujudan interaksi kimia antara PLGA, NAp dan LA telah dikenalpasti dengan menggunakan analisis spektrometer inframerah (FTIR). Kesan sinergi diantara 10-30% berat NAp dan 1% berat LA dalam membran komposit kering menunjukkan kekuatan tegangan $(0.61 \pm 0.17 \text{ MPa})$ dan modulus elastik (23.15±6.19 MPa) yang tinggi manakala membran mudah bentuk diperolehi dengan penurunan dalam modulus elastik (12.50±4.32 MPa) selepas penambahan 3% berat LA berbanding membran PLGA tulen (20.17±2.21 MPa). Penambahan 3% berat LA mengakibatkan kesan liat disebabkan interaksi lemah dalam rantaian PLGA melalui ikatan LA (-OH) dan PLGA (-CO). Ini telah dibuktikan melalui perubahan puncak FTIR (1-3 cm⁻¹), dan juga penurunan suhu peralihan kaca (T_{o}) yang dikesan melalui kalorimeter pengimbas kebedaan (DSC). Membran komposit mengekalkan struktur integriti dengan penurunan berat sebanyak 22% selepas rendaman selama 24 minggu di dalam PBS dimana ianya mempunyai potensi sebagai penghalang fizikal. Membran komposit yang mengandungi 1-3% berat LA menunjukkan pertumbuhan sel-sel fibroblas tikus dan juga pengurangan bacteria S. aureus dengan peningkatan kandungan LA. Keputusan ini menunjukkan potensi membran komposit yang mengandungi LA sebagai membran anti-jangkitan untuk kegunaan dalam aplikasi klinikal.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGEMENTS	iv
	ABSTRACT	vi
	ABSTRAK	vii
	TABLE OF CONTENTS	viii
	LIST OF TABLES	xvii
	LIST OF FIGURES	XX
	LIST OF SYMBOLS	xxxiii
	LIST OF ABBREVIATIONS	xxxiv
	LIST OF APPENDICES	xxxvii
1	INTRODUCTION	1
	1.1 Background	1
	1.2 Problem statements	4
	1.3 Objectives of the study	5
	1.4 Research hypothesis	6
	1.5 Scope of the study	7
	1.6 Significance of the study	8
	1.7 Thesis outline	8
2	LITERATURE REVIEW	12
	2.1 Introduction	12
	2.2 Bone damage and tissue reconstruction	13

2.3	Alveo	lar bone l	loss and treatment modalities	14		
2.4	Princi	ples of guided bone regeneration				
2.5	Design criteria for GBR membrane					
	2.5.1	Space-n	naking properties	17		
	2.5.2	Cell-occ	clusiveness	17		
	2.5.3	Biocom	patibility	18		
	2.5.4	Tissue i	ntegration	18		
	2.5.5	Clinical	manageability	18		
2.6	Comm	nercial GI	3R barrier membranes for clinical			
	applic	ations		18		
	2.6.1	Non-res	orbable membranes	19		
	2.6.2	Natural	bioresorbable membranes	21		
	2.6.3	Synthet	ic bioresorbable membranes	22		
2.7	Interface tissue specific functional surface					
	layers in barrier membranes					
	2.7.1	Bioreso	rbable polymer-calcium phosphate			
		compos	ites as GBR membranes	24		
		2.7.1.1	Short chain saturated aliphatic			
			polyesters	27		
		2.7.1.2	Multiple ions substituted			
			non-stoichiometric nanoapatite	29		
	2.7.2	Antibio	tics loaded GBR barrier			
		membra	ines	32		
		2.7.2.1	Systemic versus local antibiotic			
			treatment	34		
		2.7.2.2	Lauric acid as a potential antimicrobial			
			agent for clinical use	36		
2.8	Functionally graded and layered membranes					
	for gu	ided bone	e regeneration	38		
	2.8.1	Techniq	ues for polymeric composite			
		barrier r	nembrane fabrication	40		
		2.8.1.1	Solvent casting	41		
		2.8.1.2	Thermally induced phase separation	42		
		2.8.1.3	Solvent leaching	43		

2.9	In vitr	o degrada	ation characteristics of PLGA	
	based	membrar	nes	43
2.10	The d	rug releas	e mechanism in PLGA based	
	memb	oranes		45
2.11	Antim	nicrobial e	efficacy studies on antibiotic	
	loaded	d GBR m	embranes	48
2.12	Bioco	mpatibili	ty assessment	48
2.13	Challe	enges in g	guided bone	
	regene	eration us	ing barrier membrane	49
RES	SEARC	CH METI	HODOLOGY	51
3.1	Introd	uction		51
3.2	Synth	esis and c	characterisation of multiple ions	
	substi	tuted non	-stoichiometric nanoapatite (NAp)	53
	3.2.1	Materia	ls for synthesis of NAp powder	54
	3.2.2	Synthes	is of nanoapatite powder: Effects of	
		tempera	ture, concentration and multiple	
		ions sub	ostitution	54
	3.2.3	Physico	-chemical characterisation	
		of the sy	ynthesised powders	59
		3.2.3.1	Qualitative and quantitative analysis	
			using X-ray diffraction (XRD)	59
		3.2.3.2	Functional group characterisation	
			using Fourier transform	
			infrared spectrophotometry (FTIR)	61
		3.2.3.3	Elemental analysis using	
			Inductively Coupled Plasma – Atomic	
			Emission Spectroscopy (ICP-AES)	62
		3.2.3.4	Carbon, Hydrogen, Nitrogen elemental	
			analysis	61
		3.2.3.5	Image analysis by Field Emission	
			Scanning Electron Microscopy (FESEM)	62
		3.2.3.6	Image analysis by Transmission Electron	
			Microscopy (TEM)	62

3

	3.2.3.7	Thermal analysis using	
		thermogravimetric and differential thermal	
		analyser (TGA-DTA)	63
	3.2.3.8	Measurement of specific surface area	
		by Brunauner-Emmett-Teller (BET)	
		gas adsorption method	63
	3.2.3.9	Particle size analysis	63
3.2.4	In vitro	cytotoxicity assay on synthesized	
	NHA aı	nd NAp powders	64
	3.2.4.1	Materials for in vitro cytotoxicity	
		evaluation	64
	3.2.4.2	Preparation of complete medium	65
	3.2.4.3	Initiating cryopreserved cells	65
	3.2.4.4	Subculturing adherent monolayer cells	
		from 25 cm^2 to 75 cm^2 flask	66
	3.2.4.5	Split suspension	67
	3.2.4.6	Determining cell number with	
		a hemocytometer and trypan blue	
		staining	67
	3.2.4.7	Preparation of powder sample extracts	68
	3.2.4.8	Alamar Blue assay	68
Devel	opment o	f triple layered composite membrane	
gradeo	l with LA	and NAp particles in PLGA matrix	70
3.3.1	Materia	ls for fabrication of composite	
	membra	ane	70
3.3.2	A prelir	ninary study on fabrication and	
	characte	erisation of triple-layered and graded	
	compos	ite membranes using solvent casting – therma	ally
	induced	phase separation (TIPS) – solvent	
	leaching	g techniques	71
	3.3.2.1	Identification of LA and NAp	
		presence on fabricated composite	
		membranes	74

3.3

		3.3.2.2	Reproducibility of membrane fabrication			
			using solvent casting – TIPS – solvent			
			leaching techniques	75		
		3.3.2.3	Morphological and chemical characterisati	on		
			of fabricated membranes using SEM and			
			Energy Dispersive X-ray Spectroscopy			
			(EDS)	76		
		3.3.2.4	Characterisation of LA in			
			composite membranes using FTIR	76		
	3.3.3	Fabricat	tion of optimised composite membranes			
		with var	rious PLGA, NAp and LA contents			
		for phys	sical, mechanical and biological			
		assessm	ents	77		
	3.3.4	Physico	-chemical and mechanical			
		evaluati	on of the fabricated composite			
		membr	anes	83		
		3.3.4.1	Morphological characterisation using			
			VPSEM	83		
		3.3.4.2	Phase analysis using XRD	83		
		3.3.4.3	Interpretation of functional groups in			
			composite membranes using FTIR	84		
		3.3.4.4	Differential scanning calorimetric (DSC)			
			studies	84		
		3.3.4.5	Dry and wet mechanical strength			
			evaluation	85		
3.4	In vitro hydrolytic degradation and lauric					
	acid re	elease pro	ofiles of composite PLGA membranes	87		
	3.4.1	Materia	ls for in vitro degradation and LA release			
		studies		87		
	3.4.2	In vitro	degradation of composite PLGA			
		and pur	e PLGA membranes	87		
	3.4.3	In vitro	lauric acid release studies of			
		compos	ite PLGA membranes	90		
		3.4.3.1	Extraction of LA from degradation			

			medium	91
		3.4.3.2	Loading efficiency of LA in composite	
			PLGA membranes	92
		3.4.3.3	Derivatization of extracted lauric acid	93
		3.4.3.4	Quantification of LA using Reversed	
			Phase High-Performance Liquid	
			Chromatography (HPLC)	94
	3.4.4	Mathem	natical modeling to determine LA	
		release	mechanism	95
		3.4.4.1	Higuchi model	96
		3.4.4.2	Ritger-Peppas model	96
		3.4.4.3	First order kinetic model	97
		3.4.4.4	Zero order kinetic model	97
3.5	Quant	itative in	vitro antimicrobial efficacy assay	
	on LA	loaded c	omposite membranes	98
	3.5.1	Materia	ls for in vitro antimicrobial efficacy assay	98
	3.5.2	Bacteria	al culture and maintenance	98
	3.5.3	Determi	nation of mid-log exponential growth	
		phase of	f bacteria	99
	3.5.2	Antimic	robial efficacy assay	99
8.6	In vitr	o cytotox	icity assay on fabricated membranes	101
	3.6.1	Materia	ls for in vitro cytotoxicity assay	101
	3.6.2	Preparat	tion of complete medium	102
	3.6.3	Cell cul	ture and maintenance	102
	3.6.4	Preparat	tion of membrane sample extracts	103
	3.6.5	Alamar	Blue Assay	103
RES	SULTS	AND DI	SCUSSION	105
4.1	Introd	uction		105
4.2	Phase	evaluatio	n, physical and chemical characteristics,	
	eleme	ntal analy	sis, morphology and biological	
	evalua	ation of th	e synthesised as-prepared NHA	
	and	NAp pow	vders	106
	4.2.1	Phase co	omposition, crystallite size, crystallinity	

		and latti	ce parameters evaluation using XRD	107
	4.2.2	Morpho	logical evaluation using TEM	112
	4.2.3	Physica	l and chemical characterisation	
		using T	GA-DTA analysis	114
	4.2.4	Element	tal analysis	117
	4.2.5	Chemic	al and structural characterisation	121
	4.2.6	Therma	l stability	124
	4.2.7	Cytotox	icity assay on as-prepared apatite	
		powders	5	130
	4.2.8	Summar	ry of findings on synthesis and	
		characte	erization of nanoapatite powders	132
4.3	Develo	opment a	nd characterisation of triple-layered	
	and gr	aded con	posite PLGA membranes	133
	4.3.1	A prelin	ninary study on the design, processing	
		conditio	ns and effects of NAp and LA in	
		PLGA r	nembranes	134
		4.3.1.1	The design and processing conditions	
			of triple-layered and graded composite	
			PLGA membranes using solvent	
			casting-TIPS-solvent leaching	
			technique	134
		4.3.1.2	Morphological characterisation on the	
			effects of NAp and LA addition on	
			the formation of composite PLGA	
			membranes	137
		4.3.1.3	Identification of LA and NAp on	
			PLGA matrices	147
		4.3.1.4	Reproducible fabrication and	
			characterisation of triple layered	
			PLGA composite membranes with graded	1
			composition	151
	4.3.2	Fabric	ation and, morphological,	
		chemi	cal and structural characterisation	
		of the	optimised composite membrane	157

		4.3.2.1 Phase composition of membrane	
		Structure	169
		4.3.2.2 Chemical characteristics of	
		PLGA-NAp-LA components in	
		composite membrane	173
		4.3.2.3 Thermal transition of phases present	
		in composite membrane	180
4.4.	Mech	anical evaluation of composite membranes	
	in dry	y and wet state	184
	4.4.1	Mechanical properties of composite	
		membranes in dry state	184
	4.4.2	Mechanical properties of composite	
		membranes in wet state	190
4.5.	In vitr	o degradation of triple-layered and graded	
	comp	osite membranes containing NAp and LA in	
	PLGA	A matrices	194
	4.5.1	Weight loss in composite membranes	195
	4.5.2	Water uptake in composite membranes	210
4.6.	In vitr	o quantification of loading yield, delivery	
	profile	e and release mechanism of LA	216
	4.6.1	Quantification of LA loading yield in	
		composite membranes	216
	4.6.2	The in vitro release profile of LA in	
		simulated physiological solution	219
	4.6.3	The in vitro release mechanism of LA in	
		simulated physiological solution	221
4.7	In vitr	ro antimicrobial efficacy assay on LA	
	loaded	d composite membranes	224
	4.7.1	Growth curves to determine mid-log	
		exponential phase of bacteria	225
	4.7.2	In vitro antimicrobial efficacy assay	226
4.8	Cyte	otoxicity assay on NAp and LA added PLGA	
	com	posite membranes	233
4.9	Sum	nmary of overall findings	236

5	CO]	NCLUSION AND FUTURE RECOMMENDATIONS	5 238
	5.1	Conclusion	238
	5.2	Limitations and Future recommendations	239
REFEI	RENC	ΈS	242
Append	lices A	A-I	265-310

LIST OF TABLES

TABLE NO.	TITLE	PAGE
2.1	List of commercially available non-resorbable GBR	
	barrier membranes.	20
2.2	List of commercially available synthetic bioresorbable	
	membranes.	23
2.3	Drawbacks of antibiotics in systemic administration and	
	localised release systems.	35
3.1	Molar concentration of precursors used in the synthesis.	55
3.2	Freezing time for the formation of layered membranes.	
	Composition of PLGA, NAp and LA varied in layer 1 (L1),	
	layer 2 (L2) and layer 3 (L3) of the membranes.	72
3.3	Composition of NAp and LA added PLGA membranes used in	1
	reproducibility studies.	75
3.4	Varied composition of PLGA, NAp and LA in layer 1 (L1),	
	layer 2 (L2) and layer 3 (L3) of the optimised membranes.	78
3.5	The composition of membrane samples tested for	
	mechanical strength in dry and wet conditions.	86
3.6	Composition of membranes used for in vitro degradation	
	test and lauric acid release studies.	88
3.7	Types of membranes used for extraction of LA and as	
	control.	91
3.8	Membranes with various LA contents for loading	
	efficiency studies.	92
3.9	The HPLC gradient elution profile using acetonitrile	
	and water based mobile phase to quantify LA release in	
	buffer medium.	95

4.1	Effect of various synthesis parameters on crystallite size,	
	crystallinity, specific surface area and lattice parameters	
	of as-prepared stoichiometric HA (NHA) and ions	
	substituted apatite (NApF1 & NApF2) powders.	108
4.2	Molar concentrations of the precursors used in the	
	synthesis of nanoapatite compared to actual molar	
	concentrations obtained in the as-prepared powders	
	by varying the (Ca & P) equimolar reactant	
	concentrations and synthesis temperature. Reaction I	
	involves synthesis at 37±2 °C whereas in reaction II, NAp	
	initially synthesised at 37 \pm 2 °C which then increased to 85 \pm 2 °C	С
	until completion of synthesis reaction.	118
4.3	Molar concentrations of the precursors used in the	
	synthesis of nanoapatite compared to actual molar	
	concentrations obtained in the as-prepared powders by	
	reducing the substituents concentration (less than Table 4.2)	
	and using 1.5M (Ca & P) reactant concentrations.	119
4.4	Repeated batches of NApF2 powders synthesized using	
	1.5 M (Ca & P) reactant concentrations indicating	
	comparable reproducibility.	121
4.5	Average temperature changes during layering, TIPS	
	and solvent leaching steps within 24 h of each	
	membrane fabrication.	136
4.6	The photograph images of L1 and L2 surfaces of fabricated	
	membranes containing various amounts of PLGA, NAp	
	and LA.	139
4.7	The photograph images of fabricated membranes	
	containing various amounts of NAp and LA in PLGA.	
	PLGA was increased in L1 of membranes to produce less	
	porous structure.	145
4.8	SEM micrographs and processing conditions of reproducible	
	pure PLGA membranes.	153

4.9	SEM micrographs and processing conditions of reproducible	
	10 - 30 wt% of NAp + 1 wt% of LA added PLGA composite	
	membranes.	154
4.10	SEM micrographs and processing conditions of reproducible	
	10 - 30 wt% of NAp + 2 wt% of LA added PLGA composite	
	membranes.	155
4.11	SEM micrographs and processing conditions of reproducible	
	10 - 30 wt% of NAp + 3 wt% of LA added PLGA composite	
	membranes.	156
4.12	Glass transition temperature of pure PLGA and composite	
	membranes fabricated using various NAp and LA contents in	
	PLGA matrices.	181
4.13	Mechanical properties of triple-layered membranes in dry	
	and wet state. Data are presented as mean \pm SD, n=6.	186
4.14	Kinetic parameters of LA release from composite membrane	
	using various mathematical modeling.	223

LIST OF FIGURES

FIGURE NO	. TITLE	PAGE
1.1	LA incorporation into barrier membrane as an antimicrobial	
	agent for adjunct treatment in GBR procedures to inhibit	
	bacterial infection.	7
1.2	Representation of thesis outline.	11
2.1	(a) An adequate bone volume (height and width) is	
	a prerequisite for successful implant treatment.	
	(b) Barrier membrane and bone graft as bone substitute	
	materials are placed to accelerate bone formation.	
	(c) Development of final prosthesis after the	
	formation of new bone.	15
2.2	The principle of guided bone regeneration using barrier	
	membranes to mechanically occlude soft tissue invasion	
	and to retain blood clot in a secluded space. Bone growth	
	occurs through bone cells migration from the surrounding	
	original bone.	16
2.3	Structure of lactic acid, glycolic acid and	
	poly (lactic-co-glycolic acid) (PLGA).	28
2.4	Structure of lauric acid.	37
2.5	Functionally graded and layered GBR barrier membrane.	40
3.1	The schematic illustration of LA and NAp incorporation	
	into PLGA, forming composite membranes by combined	
	techniques of solvent casting-TIPS-solvent leaching.	52
3.2	Process flow of experimental methods involved in the	
	fabrication and evaluation of composite membranes.	53
3.3	Differences between synthesis reaction method I and II.	
	In reaction method I (a), the synthesis process was carried	
	In reaction method I (a), the synthesis process was carried	

	out and maintained at a lower temperature until the reaction	
	is completed. In reaction method II (b), initially the complete	
	ions addition was conducted at a lower temperature and	
	subsequently maintained at an elevated temperature until	
	the synthesis process is completed.	56
3.4	Process flow for the synthesis of multiple ions substituted	
	non-stoichiometric nanoapatite (NAp). (a)-(c) The PO_4^{3-} , Na ⁺	
	and CO_3^{2-} ionic solutions were added slowly into the basic	
	suspension containing Ca^{2+} , Mg^{2+} and K^{+} precursors while	
	vigorously stirring and heating it, (d) the precipitation product	
	was filtered and washed using DDI water, (e) the dried cake	
	was crushed, (f) crushed cake was ground to fine powders	
	and (g) sieved through 500 μ m and 20-50 μ m mesh size.	58
3.5	The refrigerator thermometer placed in the freezing	
	compartment to measure the temperature changes while	
	placing composite solutions to induce phase separation.	73
3.6	Membrane fabrication steps via solvent casting and TIPS.	
	(a-d) Weighed PLGA, LA and NAp separately for L1, L2	
	and L3, (e) mixed PLGA, LA and NAp in a vial,	
	(f-g) sonicated composite solution, (h) solvent casting,	
	and (i) cast solvent subjected to TIPS.	81
3.7	Washing steps for phase separated composite membranes.	
	(a) Frozen and phase separated solution placed in cool water,	
	(b-c) zoomed view of step (a) showing placement of petri dish	
	(containing frozen composite solution) in a glass beaker filled	
	with cool water, (d) membrane separated from petri dish and	
	placed in a fresh cool water, (e) zoomed view of step (d) and	
	finally room air dried composite membrane showing (f) L3	
	and (g) L1.	82
4.1	Triple layered composite PLGA membrane containing LA	
	and NAp promoting bone cells growth while preventing	
	bacteria and fibroblast cells.	106
4.2	XRD patterns of the as-prepared ions substituted apatite powder	
	(NApF2-1.5M) synthesised at 37±2 °C (reaction method I) and	

	37 and 85±2 °C (reaction method II).	107
4.3	XRD patterns of as-prepared ions substituted apatite (NApF2)	
	synthesised at 37 and 85±2 °C (reaction method II) using	
	1.0M, 1.5M and 2.0M equimolar precursors.	110
4.4	XRD patterns of as-prepared stoichiometric HA (NHA)	
	and ions substituted apatite (NApF1 & NApF2) powders	
	synthesised at 37 and 85±2 °C (reaction method II) using	
	1.5M equimolar precursors.	111
4.5	TEM micrographs of as prepared HA synthesised at 37 ± 2 °C	
	using reactant concentration (a) 1.0M, (b) 1.5M and (c-d) 2.0M	
	at low magnification (a,b,c) and high magnification (d),	
	respectively.	112
4.6	TEM micrographs of as-prepared NApF2 synthesised at	
	37 & 85±2 °C using reactant concentration 1.5 M at low	
	magnification on three different spots (a)-(c) and	
	high magnification (d), respectively.	113
4.7	TGA and DTA curves of the stoichiometric HA (NHA)	
	powders synthesised at 37±2 °C using 1.0 M, 1.5 M and	
	2.0 M reactants and dried at 80 °C.	115
4.8	TGA and DTA curves of NHA, NApF1 and NApF2 powders	
	initially synthesised at 37 \pm 2 °C and then increased to	
	85 ± 2 °C (reaction II) using 1.5M reactants and dried	
	at 80 °C. The samples denoted as 37 and 85 ± 2 °C.	117
4.9	FTIR spectra of as prepared stoichiometric HA (NHA)	
	and ionic substituted apatite (NApF1 & NApF2) powders	
	synthesised at 37 & 85 \pm 2 °C (reaction method II) using 1.5 M	
	equimolar precursors.	122
4.10	SEM micrograph for the morphology of (a) stoichiometric	
	HA (NHA); ions substituted nanoapatite (b) NApF1 and	
	(c) NApF2 powders synthesised at 37 and 85 ± 2 °C using	
	1.5 M reactant concentration (30,000× magnification).	124
4.11	XRD patterns of stoichiometric HA (NHA) and ionic	
	substituted apatite (NApF1 & NApF2) powders synthesised	
	at 37 & 85±2 °C (reaction method II) using 1.5M equimolar	

	precursors after heat-treatment at 900 °C in CO ₂ controlled	
	atmosphere.	125
4.12	XRD patterns of stoichiometric HA (NHA) and ionic	
	substituted apatite (NApF1 & NApF2) powders synthesised	
	at 37 & 85±2 °C (reaction method II) using 1.5M equimolar	
	precursors after sintering at 1250 °C in air.	126
4.13	FTIR spectra of stoichiometric HA (NHA) and ionic	
	substituted apatite (NApF1 & NApF2) powders synthesised	
	at 37 & 85 ± 2 °C (reaction method II) using 1.5 M equimolar	
	precursors after heat-treatment at 900 °C in CO ₂ controlled	
	atmosphere.	128
4.14	FTIR spectra of stoichiometric HA (NHA) and ionic	
	substituted apatite (NApF2) powders synthesised at	
	37 & 85±2 °C (reaction method II) using 1.5M equimolar	
	precursors after sintering at 1250 °C in air.	130
4.15	Representative 24-well plates for qualitative AB colour	
	changes. Colour change from blue to red indicates	
	presence of live cells whereas unchanged blue colour	
	indicates presence of dead cells after treatment with extracts.	
	(a) NHA and (b) NApF2 powder extracts, and	
	(c) positive control (phenol solution extracts).	131
4.16	Cytotoxicity assay results on L929 mouse fibroblast	
	cells viability in response to different extract concentrations	
	of NHA and NApF2 powder extracts. The data are presented	
	as means \pm SEM values of two independent	
	experiments $(n = 2)$.	132
4.17	SEM micrographs of L1 of composite membranes containing	
	10-30wt% NAp + 1wt% LA and 10-100wt% NAp + 1wt% LA	
	graded in (a) 7, (b) 9, (c) 11 and (d) 13wt% of PLGA matrices.	
	L1 of pure PLGA membranes were compared as control. (e) The	
	representative EDS spectrum of the composite membranes	
	taken on L1.	141

4.18	SEM micrographs of L3 of 10-30wt% NAp + 1wt% LA	
	and 10-100wt% NAp + 1wt% LA graded membranes in	
	(a) 7, (b) 9, (c) 11 and (d) 13wt% of PLGA matrices. L3	
	of pure PLGA membranes were compared as control. (e) The	
	representative EDS spectrum of the composite membranes	
	taken on L3.	142
4.19	SEM micrographs of L1 of (a) 15, (b) 17, (c) 20 and	
	(d) 23wt% PLGA membranes.	144
4.20	Photographs of (a) L1 and (b) L3 of PLGA membranes	
	with graded composition of 10-30 wt% NAp + 3 wt%	
	LA and the representative SEM micrographs for	
	(c) L1, (d) L3 and (e) cross section of membranes.	147
4.21	(A) Representative SEM micrographs of PLGA membranes	
	(a)-(c) and 3 wt% of LA added composite PLGA membranes	
	(d)-(f) on L1 surface. The representative micrographs	
	of (a) & (d) as-prepared membranes, and membranes	
	after immersion in ethanol for (b) & (e) 30 s and (c) & (f) 5min,	
	respectively with magnification 100 x. Insets show	
	high magnification (1000 x) view of the respective	
	membranes. Crossed marks (x) indicate peeled-off spots.	
	(B) FTIR spectra of 3 wt% of LA added (a) as-prepared	
	membrane; and membranes after immersion in ethanol	
	for (b) 30s and (c) 5 min; and (d) as-received LA and	
	(e) pure PLGA membrane.	149
4.22	SEM micrographs of 3 wt% LA + 10-30 wt% NAp added	
	PLGA composite membranes taken on (a) L1 and (b) L3	
	of as-prepared membranes, (c) 5 min immersed L3	
	surface of the membrane; and their respective EDS spectra.	
	Crossed (x) marks indicate EDS points.	151
4.23	Representative EDS spectra of the L3 of composite membranes	
	(Table 4.9 – 4.11 (c, f, i)) containing (a) 1 wt%, (b) 2 wt% and	
	(c) 3 wt% LA and 10-30 wt% of NAp, confirming the	
	presence of NAp particles within the membrane.	157

4.24	SEM micrographs of L1, L3 and cross-section of (a) pure PLGA membrane (S105) and (b) 1wt% (S98),	
	(c) 2wt% (S99), (d) 3wt% (S100) of LA incorporated	
	triple layered membranes containing 10-30 wt% of NAp	
	in 9-20 wt% of PLGA matrices. (e) The representative	
	EDS spectrum of the composite membrane.	159
4.25	SEM micrographs of L1, L3 and cross-section of	
	(a) pure PLGA membrane (S105) and (b) 1wt% (S106),	
	(c) 2 wt% (S107), (d) 3 wt% (S108) of LA incorporated	
	triple layered membranes containing 10-100 wt% of NAp	
	in 9-20 wt% of PLGA matrices. (e) The representative	
	EDS spectrum of the composite membranes.	161
4.26	SEM micrographs of L1, L3 and cross-section of	
	(a) pure PLGA membrane (S181) and (b) 1 wt% (S183),	
	(c) 2 wt% (S185), (d) 3 wt% (S187) of LA incorporated	
	triple layered membranes containing 10-30 wt% of NAp in	
	9-17 wt% of PLGA matrices. (e) The representative EDS	
	spectrum of composite membranes.	163
4.27	SEM micrographs of L1, L3 and cross-section of	
	(a) pure PLGA membrane (S181) and (b)1 wt% (S193),	
	(c) 2 wt% (S191), (d) 3 wt% (S189) of LA incorporated	
	triple layered membranes containing 10-100 wt% of NAp	
	in 9-17 wt% of PLGA matrices. (e) The representative EDS	
	spectrum of composite membranes.	164
4.28	High magnification SEM micrographs of L1 (left) and L3	
	(right) of (a,b) 9-20 wt% of pure PLGA matrices (S105),	
	(c,d) 3 wt% LA + 10-30 wt% NAp added in 9-20 wt% of	
	PLGA matrices (S100), (e,f) 3 wt% LA + 10-100 wt% of NAp	
	added in 9-20 wt% of PLGA matrices (S108). (g) The represent	ative
	EDS spectrum of NAp particles on composite membranes	
	as indicated by the black arrows in (c) and (e).	166

 (right) of (a,b) 9-17wt% of pure PLGA matrices (S181), (c,d) 3 wt% LA + 10-30wt% NAp added in 9-17wt% of PLGA matrices (S187), (e,f) 3wt% LA + 10-100wt% of NAp added in 9-17wt% of PLGA matrices (S189). (g) The representative EDS spectrum of NAp particles as indicated by the white arrows in (d) and (f). 168 4.30 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-30wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S98), (c) 2wt% (S99) and (d) 3wt% (S100) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S98), (f) 2wt% (S99) and (g) 3wt% (S100) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks, (j) LA. L1 and L3 were incorporated with 10 and 30wt% of NAp, respectively. 4.31 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-100wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S106), (c) 2wt% (S107) and (d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks, (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), (e) 1 wt% (S110), (f) 2 wt% (S102) and (g) 3 wt% (S115) 	4.29	High magnification SEM micrographs of L1 (left) and L3	
 matrices (S187), (e,f) 3wt% LA + 10-100wt% of NAp added in 9-17wt% of PLGA matrices (S189). (g) The representative EDS spectrum of NAp particles as indicated by the white arrows in (d) and (f). 168 4.30 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-30wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S98), (c) 2wt% (S99) and (d) 3wt% (S100) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S98), (f) 2wt% (S99) and (g) 3wt% (S100) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite (1) peaks, (j) LA. L1 and L3 were incorporated with 10 and 30wt% of NAp, respectively. 4.31 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-100wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S106), (c) 2wt% (S107) and (d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite (1) peaks, (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		(right) of (a,b) 9-17wt% of pure PLGA matrices (S181),	
 in 9-17wt% of PLGA matrices (S189). (g) The representative EDS spectrum of NAp particles as indicated by the white arrows in (d) and (f). 168 4.30 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-30wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S98), (c) 2wt% (S99) and (d) 3wt% (S100) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S98), (f) 2wt% (S99) and (g) 3wt% (S100) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite (■) peaks, (j) LA. L1 and L3 were incorporated with 10 and 30wt% of NAp, respectively. 171 4.31 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-100wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S106), (c) 2wt% (S107) and (d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite (■) peaks, (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		(c,d) 3 wt% LA + 10-30wt% NAp added in 9-17wt% of PLGA	
 spectrum of NAp particles as indicated by the white arrows in (d) and (f). 168 4.30 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-30wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S98), (c) 2wt% (S99) and (d) 3wt% (S100) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S98), (f) 2wt% (S99) and (g) 3wt% (S100) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks, (j) LA. L1 and L3 were incorporated with 10 and 30wt% of NAp, respectively. 4.31 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-100wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S106), (c) 2wt% (S107) and (d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks, (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		matrices (S187), (e,f) 3wt% LA + 10-100wt% of NAp added	
 in (d) and (f). 168 4.30 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-30wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S98), (c) 2wt% (S99) and (d) 3wt% (S100) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S98), (f) 2wt% (S99) and (g) 3wt% (S100) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks, (j) LA. L1 and L3 were incorporated with 10 and 30wt% of NAp, respectively. 4.31 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-100wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S106), (c) 2wt% (S107) and (d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks, (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		in 9-17wt% of PLGA matrices (S189). (g) The representative E	DS
 4.30 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-30wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S98), (c) 2wt% (S99) and (d) 3wt% (S100) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S98), (f) 2wt% (S99) and (g) 3wt% (S100) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite (1) peaks, (j) LA. L1 and L3 were incorporated with 10 and 30wt% of NAp, respectively. 4.31 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-100wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S106), (c) 2wt% (S107) and (d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite (1) peaks, (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		spectrum of NAp particles as indicated by the white arrows	
 L1 of 10-30wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S98), (c) 2wt% (S99) and (d) 3wt% (S100) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S98), (f) 2wt% (S99) and (g) 3wt% (S100) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite (■) peaks, (j) LA. L1 and L3 were incorporated with 10 and 30wt% of NAp, respectively. 4.31 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-100wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S106), (c) 2wt% (S107) and (d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite (■) peaks, (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		in (d) and (f).	168
 composite membrane containing (b) 1wt% (S98), (c) 2wt% (S99) and (d) 3wt% (S100) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S98), (f) 2wt% (S99) and (g) 3wt% (S100) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks, (j) LA. L1 and L3 were incorporated with 10 and 30wt% of NAp, respectively. 4.31 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-100wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S106), (c) 2wt% (S107) and (d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks, (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 	4.30	XRD patterns for (a) pure PLGA membrane (S105);	
 (c) 2wt% (S99) and (d) 3wt% (S100) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S98), (f) 2wt% (S99) and (g) 3wt% (S100) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks, (j) LA. L1 and L3 were incorporated with 10 and 30wt% of NAp, respectively. 171 4.31 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-100wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S106), (c) 2wt% (S107) and (d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks , (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 172 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		L1 of 10-30wt% of NAp added in 9-20wt% of PLGA	
 layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S98), (f) 2wt% (S99) and (g) 3wt% (S100) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks, (j) LA. L1 and L3 were incorporated with 10 and 30wt% of NAp, respectively. 4.31 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-100wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S106), (c) 2wt% (S107) and (d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks , (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		composite membrane containing (b) 1wt% (S98),	
 patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S98), (f) 2wt% (S99) and (g) 3wt% (S100) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks, (j) LA. L1 and L3 were incorporated with 10 and 30wt% of NAp, respectively. 171 4.31 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-100wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S106), (c) 2wt% (S107) and (d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks, (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		(c) 2wt% (S99) and (d) 3wt% (S100) of LA. The opposite	
 (e) 1wt% (S98), (f) 2wt% (S99) and (g) 3wt% (S100) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks, (j) LA. L1 and L3 were incorporated with 10 and 30wt% of NAp, respectively. 171 4.31 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-100wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S106), (c) 2wt% (S107) and (d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks, (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		layers of L1, i.e., L3 were not introduced with LA. The XRD	
 LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite (■) peaks, (j) LA. L1 and L3 were incorporated with 10 and 30wt% of NAp, respectively. 171 4.31 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-100wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S106), (c) 2wt% (S107) and (d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite (■) peaks, (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		patterns of L3 were the opposite layers of L1 added with	
 (i) magnified region for NAp showing apatite (■) peaks, (j) LA. L1 and L3 were incorporated with 10 and 30wt% of NAp, respectively. 171 4.31 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-100wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S106), (c) 2wt% (S107) and (d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite (■) peaks , (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		(e) 1wt% (S98), (f) 2wt% (S99) and (g) 3wt% (S100) of	
 (j) LA. L1 and L3 were incorporated with 10 and 30wt% of NAp, respectively. 4.31 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-100wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S106), (c) 2wt% (S107) and (d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite (■) peaks, (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		LA in triple layered membranes containing (h) NAp;	
 NAp, respectively. 4.31 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-100wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S106), (c) 2wt% (S107) and (d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks, (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		(i) magnified region for NAp showing apatite (\blacksquare) peaks,	
 4.31 XRD patterns for (a) pure PLGA membrane (S105); L1 of 10-100wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S106), (c) 2wt% (S107) and (d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks , (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		(j) LA. L1 and L3 were incorporated with 10 and 30wt% of	
 10-100wt% of NAp added in 9-20wt% of PLGA composite membrane containing (b) 1wt% (S106), (c) 2wt% (S107) and (d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite (■) peaks, (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		NAp, respectively.	171
 membrane containing (b) 1wt% (S106), (c) 2wt% (S107) and (d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks, (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 	4.31	XRD patterns for (a) pure PLGA membrane (S105); L1 of	
 (d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks, (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		10-100wt% of NAp added in 9-20wt% of PLGA composite	
 were not introduced with LA. The XRD patterns of L3 were the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks, (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		membrane containing (b) 1wt% (S106), (c) 2wt% (S107) and	
 the opposite layers of L1 added with (e) 1wt% (S106), (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite () peaks, (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		(d) 3wt% (S108) of LA. The opposite layers of L1, i.e., L3	
 (f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite (■) peaks , (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		were not introduced with LA. The XRD patterns of L3 were	
 layered membranes containing (h) NAp; (i) magnified region for NAp showing apatite (■) peaks , (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		the opposite layers of L1 added with (e) 1wt% (S106),	
 region for NAp showing apatite (■) peaks , (j) LA. L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		(f) 2wt% (S107) and (g) 3wt% (S108) of LA in triple	
 L1 and L3 were incorporated with 10 and 100wt% of NAp, respectively. 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		layered membranes containing (h) NAp; (i) magnified	
respectively.1724.32FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112),		region for NAp showing apatite (■) peaks , (j) LA.	
 4.32 FTIR spectra of (a) as-received PLGA, (b) LA particles, (c) NAp powder and L1 surface of (d) pure PLGA (S112), 		L1 and L3 were incorporated with 10 and 100wt% of NAp,	
(c) NAp powder and L1 surface of (d) pure PLGA (S112),			172
	4.32	FTIR spectra of (a) as-received PLGA, (b) LA particles,	
(e) 1 wt% (S110), (f) 2 wt% (S102) and (g) 3 wt% (S115)		(c) NAp powder and L1 surface of (d) pure PLGA (S112),	
		(e) 1 wt% (S110), (f) 2 wt% (S102) and (g) 3 wt% (S115)	
of LA added composite membranes containing		of LA added composite membranes containing	

	10-30 wt% of NAp in 9-20 wt% of PLGA matrices.	175
4.33	FTIR spectra of (a) NAp powder and L3 surface of	
	(b) pure PLGA (S112) membrane (c) 1 wt% of LA (S117),	
	(d) 2 wt% of LA (S118) and (e) 3 wt% of LA (S119) added	
	composite membranes containing 10-100 wt% of NAp in	
	9-20 wt% of PLGA matrices.	177
4.34	The illustration of possible interaction mechanisms between	
	PLGA-NAp-LA composite systems presenting (a) hydrogen	
	bonding between PLGA-NAp-LA and (b) ionic bonding	
	between NAp-LA.	179
4.35	DSC thermogram of LA.	180
4.36	DSC thermograms of pure PLGA membrane (S195),	
	1 wt% LA (S197), 2 wt% LA (S199) and	
	3 wt% LA (S201) added composite membranes	
	containing 10-30 wt% of NAp in 9-20 wt% of PLGA	
	matrices.	181
4.37	DSC thermograms of pure PLGA membrane (S195),	
	1 wt% LA (S203), 2 wt% LA (S205) and 3 wt% LA (S207)	
	added composite membranes containing 10-100 wt% of NAp	
	in 9-20 wt% of PLGA matrices.	182
4.38	Representative stress-strain curves of the 10-30 wt% of	
	NAp added 9-20 wt% of PLGA composite membranes	
	with different LA contents. Only one typical plot for each	
	membrane is shown.	185
4.39	Tensile strength of 1-3 wt% of LA added composite	
	membranes containing 10-30 wt% and 10-100 wt% NAp	
	in 9-20 wt% of PLGA matrices. Data are presented as	
	mean \pm SD, n=6.	187
4.40	Representative stress-strain curves of the 10-100 wt% of	
	NAp containing 9-20 wt% of PLGA composite	
	membranes with different LA contents. Only one typical	
	plot for each membrane is shown.	189
4.41	Tensile strength of 1-3 wt% of LA added composite	
	membranes containing 10-30 wt% and 10-100 wt% NAp	

	in 9-17 wt% of PLGA matrices. Data are presented	
	as mean \pm SD, n=6.	192
4.42	The weight loss of 9-20 wt% of pure PLGA and composite	
	triple layered membranes. The composites loaded with	
	1 wt% of LA and NAp varied at 10-30 wt% and 10-100 wt%.	
	Data are presented as mean \pm SD, $n = 3$.	196
4.43	The weight loss of 9-20 wt% of pure PLGA and composite	
	triple layered membranes. The composites loaded with	
	2 wt% of LA and NAp varied at 10-30 wt% and 10-100 wt.	
	Data are presented as mean \pm SD, $n = 3$.	197
4.44	The weight loss of 9-20 wt% of pure PLGA and composite	
	triple layered membranes. The composites loaded with	
	3 wt% of LA with NAp varied at 10-30 wt% and 10-100 wt%.	
	Data are presented as mean \pm SD, $n = 3$.	198
4.45	SEM micrographs of L3 surfaces of pure PLGA and	
	composite membranes (a) before immersion and after	
	immersion for (b) 4 weeks, (c) 12 weeks and	
	(d) 24 weeks in PBS. The PLGA content is 9-20 wt%. The	
	PLGA content is 9-20 wt%. (e) The EDS spectrum of the	
	region marked by rectangle taken on 10-100 wt% of NAp	
	added composite membrane as in (d).	199
4.46	SEM micrographs of L3 surface morphology of pure	
	PLGA (9-20 wt%) membrane, (a) before immersion	
	and after immersion for (b) 4 weeks and (c) 24 weeks in	
	PBS.	200
4.47	SEM micrographs of L1 surfaces of pure PLGA and	
	composite membranes (a) before immersion and after	
	immersion for (b) 4 weeks, (c) 12 weeks and (d) 24 weeks	
	in PBS. The PLGA content is 9-20 wt%. (e) The EDS	
	spectrum of the region marked by rectangle taken on	
	10-100 wt% of NAp added composite membrane as in (c).	201
4.48	The weight loss of 9-20 wt% of pure PLGA and	
	composite triple layered membranes. The composites	
	loaded with 10-30 wt% of NAp and LA varied at 1-3 wt%.	

	Data are presented as mean \pm SD, n=3.	202
4.49	The weight loss of 9-20 wt% of pure PLGA and composite	
	triple layered membranes. The composites loaded with	
	10-100 wt% of NAp and LA varied at 1-3 wt%.	
	Data are presented as mean \pm SD, n=3.	203
4.50	The weight loss of 9-17 wt% of pure PLGA and composite	
	triple layered membranes. The composites loaded with	
	3 wt% of LA and NAp varied at 10-30 wt% and	
	10-100 wt%. Data are presented as mean \pm SD, n=3.	204
4.51	SEM micrographs of L1 surface morphology of composite	
	PLGA (9-17 wt%) membranes with 10-100 wt% of NAp	
	and 3wt% LA, (a) before immersion and (b) after immersion	
	for 24 weeks in PBS.	204
4.52	The pH change of 9-20 wt% of pure PLGA and composite	
	triple layered membranes. The composites loaded with	
	10-30 wt% of NAp and LA varied at 1-3 wt%.	
	Data are presented as mean \pm SD, n=3.	207
4.53	The pH change of 9-20 wt% of pure PLGA and composite	
	triple layered membranes. The composites loaded with	
	10-100 wt% of NAp and LA varied at 1-3 wt%.	
	Data are presented as mean \pm SD, n=3.	207
4.54	The pH change of 9-17 wt% of pure PLGA and composite	
	triple layered membranes. The composites loaded with	
	3 wt% of LA and NAp varied at 10-30 wt% and 10-100 wt.	
	Data are presented as mean \pm SD, n=3.	209
4.55	The pH change of 9-20 wt% of pure PLGA and composite	
	triple layered membranes. The composites loaded with	
	3 wt% of LA and NAp varied at 10-30 wt% and 10-100 wt%.	
	Data are presented as mean \pm SD, n=3.	209
4.56	The water uptake of 9-20 wt% of pure PLGA and composite	
	triple layered membranes. The composites loaded with	
	10-30 wt% of NAp and LA varied at 1-3 wt%.	
	Data are presented as mean \pm SD, n=3.	210

4.57	The water uptake of 9-20wt% of pure PLGA and composite	
	triple layered membranes. The composites loaded with	
	10-100wt% of NAp and LA varied at 1-3wt%.	
	Data are presented as mean \pm SD, n=3.	211
4.58	The water uptake of 9-20 wt% of pure PLGA and composite	
	triple layered membranes. The composites loaded with	
	1 wt% of LA and NAp varied at 10-30 wt% and	
	10-100 wt%. Data are presented as mean \pm SD, n=3.	212
4.59	The water uptake of 9-20 wt% of pure PLGA and	
	composite triple layered membranes. The composites	
	loaded with 2 wt% of LA and varied at 10-30 wt% and	
	10-100 wt% of NAp. Data are presented as mean \pm SD, n=3.	213
4.60	The water uptake of 9-20 wt% of pure PLGA and	
	composite triple layered membranes. The composites	
	loaded with 3 wt% of LA and varied at 10-30 wt% and	
	10-100 wt% of NAp. Data are presented as mean \pm SD, n=3.	213
4.61	The water uptake of 9-17 wt% of pure PLGA and	
	composite triple layered membranes. The composites	
	loaded with 3 wt% of LA and varied at 10-30 wt% and	
	10-100 wt% of NAp. Data are presented as mean \pm SD, n=3.	214
4.62	The highest UV absorption intensity of derivatized	
	LA at the concentration of 2000 μ g/mL and	
	(b) the corresponding linear calibration standard curve.	
	Data represents mean±SD of three replicates.	218
4.63	The cumulative release percentage of LA from 3 wt%	
	of LA loaded composite membranes containing 10-30 wt%	
	of NAp in 9-20 wt% of PLGA matrices. Data represents	
	mean±SD of two replicates.	220
4.64	The cumulative amount of LA released from the	
	formulated membrane fitted to Ritger-Peppas model.	221
4.65	The cumulative amount of LA released from the	
	formulated membrane fitted to Higuchi model.	222
4.66	The cumulative amount of LA released from the	
	formulated membrane fitted to First order kinetic	

	model.	222
4.67	The cumulative amount of LA released from the	
	formulated membrane fitted to Zero order kinetic	
	model.	223
4.68	Growth curve of S. aureus (ATCC 6538) grown at	
	37 °C in TSB. Data represents means \pm SEM	
	of three independent experiments $(n = 3)$.	225
4.69	Growth curve of P. aeruginosa (ATCC 9027) grown	
	at 37 °C in TSB. Data represents means \pm SEM of	
	three independent experiments $(n = 3)$.	226
4.70	Antimicrobial activity of 1, 2, and 3 wt% of LA	
	incorporated composite membranes compared to pure	
	PLGA control group against S. aureus. The number of	
	viable microbes on the membranes after 24 h was	
	obtained using colony counting formation method.	
	Data represents means \pm SEM of two	
	independent experiments $(n = 2)$.	227
4.71	Antimicrobial activity. The recovery of S. aureus	
	on LA incorporated membranes after 24 h of incubation	
	at 37 °C. The number of viable microbes on (a) PLGA	
	membrane, and (b) 1 wt%, (c) 2 wt% and (d) 3 wt% of	
	LA incorporated membranes, was obtained using colony	
	counting formation method. The representative TSA	
	plates for all three replicates show S.aureus colony	
	formation after incubation. The TSA plates (e) represent	
	colony formation of recovered inoculums after serially	
	diluted to 10^{-1} (left plate) and 10^{-2} (right plate).	
	At higher dilution (10^{-2}) , fewer colonies were formed	
	on plates which indicate that serial dilutions of the	
	recovered inoculums were performed appropriately.	229
4.72	Antimicrobial activity of 1, 2, and 3 wt% of LA incorporated	
	composite membranes compared to pure PLGA control	
	group against P. aeruginosa. The number of viable	

	microbes on the membranes after 24 h was obtained using	
	colony counting formation method. Data represents	
	means \pm SEM of two independent experiments ($n = 2$).	
	After 24 h of incubation, the bacteria colonies had	
	increased by 10^2 in all samples compared to initial	
	loading of <i>P. aeruginosa</i> which demonstrated	
	growth of bacteria after incubation period.	230
4.73	The recovery of P. aeruginosa on LA incorporated	
	membranes after 24 h of incubation at 37 °C. The number	
	of viable microbes on (a) PLGA membrane, and (b) 1 wt%,	
	(c) 2 wt% and (d) 3 wt% of LA incorporated membranes,	
	was obtained using colony counting formation method.	
	The representative TSA plates for all three replicates	
	show formation of P. aeruginosa colonies after incubation.	
	The TSA plates (e) represent colony formation of recovered	
	inoculums after serially diluted to 10 ⁻³ (left plate) and	
	10^{-4} (right plate). At higher dilution (10^{-4}), fewer colonies	
	were formed on plates which indicate that serial dilutions	
	of the recovered inoculums were performed appropriately.	
	The bacteria growth on LA added composite membranes	
	was prominently higher than pure PLGA membranes as	
	more dilutions were performed beyond 10^{-1} and 10^{-2} .	231
4.74	Representative 24-well plates for AB colour changes	
	assessment. Cells exposed to extracts of (a) PLGA,	
	(b) 1 wt% LA, (c) 2 wt% LA, (d) 3 wt% LA and	
	(e) phenol solution.	234
4.75	Cytotoxicity assay results of the pure PLGA and	
	LA added composite membranes. Data are	
	means \pm SEM of two independent experiments (<i>n</i> =2).	235

LIST OF SYMBOLS

cfu	-	Colony forming unit
d	-	Interplanar spacing
Exo	-	Exothermic
k	-	drug release kinetic constant
m_0	-	Initial weight
m_1	-	Wet weight
m ₂	-	Dry weight
Μ	-	Molar
M_t	-	Amount of drug released at time t
M_{∞}	-	Total amount of drug released
n	-	Diffusional exponent
R^2	-	Correlation coefficient
rad	-	Radian
t	-	time
Tg	-	Transition temperature
X _c	-	Crystallinity
X _s	-	Crystallite size
θ	-	Diffraction Angle
λ	-	Wavelength of X-ray beam

LIST OF ABBREVIATIONS

AMP	-	Antimicrobial peptide
ATCC	-	American Type Culture Collection
ASTM	-	American Society for Testing and Materials
ATR	-	Attenuated total reflectance
BET	-	Brunauer – Emmet – Teller
CaP	-	Calcium phosphate
CHN	-	Carbon, Hydrogen, Nitrogen elemental analysis
DSC	-	Differential scanning calorimetry
DDI	-	Double distilled de-ionised
DMSO	-	Dimethyl sulfoxide
DTA	-	Differential thermal analysis
d-PTFE	-	Dense polytetrafluoroethylene
ECACC	-	European Collection of Cell Cultures
e-PTFE	-	expanded PTFE
et al.	-	and others
FDA	-	US Food and Drug Administration
FESEM	-	Field Emission Scanning Electron Microscope
FGM	-	Functionally graded membrane
FTIR	-	Fourier Transform Infrared spectrophotometry
F. nucleatum	-	Fusobacterium nucleatum
FWHM	-	Full width at half maximum
GBR	-	Guided bone regeneration
HA	-	Hydroxyapatite
HPLC	-	High Performance Liquid Chromatography
HSF	-	Human Skin Fibroblast cells
ICP-AES	-	Inductively Coupled Plasma-Atomic Emission
		Spectroscopy
i.e.	-	that is

IC ₈₀	-	Inhibition concentration at 80% killing
ICDD	-	International Centre for Diffraction Data
ISO	-	International Organisation for Standardisation
LA	-	Lauric acid
L1	-	Layer 1
L2	-	Layer 2
L3	-	Layer 3
MEM	-	Minimum Essential Medium
MePEG	-	Methoxypoly(ethyleneglycol)
MIC	-	Minimum inhibition concentration
MTT	-	3-(4,5-Dimethylthiazol-2-yl)-2,5-
		diphenyltetrazolium bromide
NAp	-	Non-stoichiometric nanoapatite
NApF1	-	Non-stoichiometric nanoapatite Formulation 1
NApF2	-	Non-stoichiometric nanoapatite Formulation 2
NHA	-	Stoichiometric nanohydroxyapatite
OFP	-	Open Flap Debridement
OD	-	Optical density
PBS	-	Phosphate buffered saline
PCL	-	Polycaprolactone
PDL	-	Periodontal ligament
PDLLA	-	poly(DL-lactic) acid
P. gingivalis	-	Porphyromonas gingivalis
PGA	-	Polyglycolic acid
P. intermedia	-	Prevotella intermedia
PLA	-	Polylactic acid
PLLA	-	poly(L-lactic) acid
PLGC	-	poly (L-lactide-co-glycolide-ɛ-caprolactone)
PLCL	-	poly (L-lactide-co-ɛ-caprolactone)
PU	-	Polyetherurethane
PTFE	-	polytetrafluoroethylene
rpm	-	revolution per minute
SEM	-	Scanning Electron Microscopy
SD	-	Standard Deviation

TEM	-	Transmission Electron Microscopy
TGA	-	Thermogravimetric analysis
TIPS	-	Thermally induced phase separation
UV	-	Ultraviolet
UV-Vis	-	Ultraviolet-Visible
wt%	-	Weight percentage
XRD	-	X-ray Diffraction
β-ΤСΡ	-	β -tricalcium phosphate
3D	-	3 dimensional

LIST OF APPENDICES

APPENDICES	TITLE	PAGE
А	List of publications	265
A1	Published article 1	266
A2	Published article 2	268
A3	Published article 3	270
A4	Published article 4	272
B1	Calculation for the preparation of $1.0 \text{ M Ca}(\text{OH})_2$	
	and H ₃ PO ₄ reactants for the synthesis of NHA-1.0M.	274
B2	Calculation for the preparation of $1.5 \text{ M Ca}(\text{OH})_2$	
	and H ₃ PO ₄ reactants for the synthesis of NHA-1.5M.	275
В3	Calculation for the preparation of $2.0 \text{ M Ca}(\text{OH})_2$	
	and H ₃ PO ₄ reactants for the synthesis of NHA-2.0M.	276
B4	Calculation for the preparation of 1.0 M Ca(OH) ₂ ,	
	H ₃ PO ₄ reactants and ionic solutions for the synthesis	
	of NApF1 and NApF2-1.0M.	277
B5	Calculation for the preparation of 1.5 M Ca(OH) ₂ ,	
	H ₃ PO ₄ reactants and ionic solutions for the synthesis	
	of NApF1 and NApF2-1.5M.	278
B6	Calculation for the preparation of $2.0 \text{ M Ca}(\text{OH})_2$,	
	H ₃ PO ₄ reactants and ionic solutions for the synthesis	
	of NApF1 and NApF2-2.0M.	279
C1	Lattice parameters calculation for as prepared	
	stoichiometric nanohydroxyapatite (NHA) powder	
	synthesized using reaction method I (37±2°C).	280
C2	Lattice parameters calculation for as prepared	
	stoichiometric nanohydroxyapatite (NHA) powder	

	synthesized using reaction method II (37&85±2°C).	281
C3	Lattice parameters calculation for as prepared	
	non-stoichiometric nanoapatite (NApF1) powder	
	synthesized using reaction method I (37±2°C).	282
C4	Lattice parameters calculation for as prepared	
	non-stoichiometric nanoapatite (NApF1) powder	
	synthesized using reaction method II (37&85±2°C).	283
C5	Lattice parameters calculation for as prepared	
	non-stoichiometric nanoapatite (NApF2) powder	
	synthesized using reaction method I (37±2°C).	284
C6	Lattice parameters calculation for as prepared	
	non-stoichiometric nanoapatite (NApF2) powder	
	synthesized using reaction method II (37&85±2°C).	285
C7	Lattice parameters calculation for NHA, NApF1 and	
	NApF2 powders sintered at 900°C in CO2.	286
C8	Lattice parameters calculation for NHA powders	
	sintered at 900°C in CO ₂ .	287
C9	Lattice parameters calculation for NApF1 powders	
	sintered at 900°C in CO _{2.}	288
C10	Lattice parameters calculation for NApF2 powders	
	sintered at 900°C in CO ₂ .	289
C11	Lattice parameters calculation for NHA, NApF1	
	and NApF2 powders sintered at 1250°C in air.	290
C12	Crystallite size (Xs in nm) of as prepared	
	nanohydroxyapatite powders determined using	
	Scherrer equation.	291
C13	Fraction of crystalline phase (Xc) of the as prepared	
	NHA, NApF1 and NApF2 powders.	292
D1	Dry and wet tensile strength of 10-30 wt% of NAp	
	containing 1-3 wt% of LA added PLGA (9-20wt%)	
	composite membranes.	293
D2	Dry and wet tensile strength of 10-100 wt% of NAp	
	containing 1-3 wt% of LA added PLGA (9-20wt%)	
	composite membranes.	294

D3	Dry and wet tensile strength of 10-30 wt% of NAp	
	containing 1-3 wt% of LA added PLGA (9-17wt%)	
	composite membranes.	295
D4	Dry and wet tensile strength of 10-100 wt% of NAp	
	containing 1-3 wt% of LA added PLGA (9-17wt%)	
	composite membranes.	296
E1	Weight loss measurements and weight loss difference	
	in post-immersed membranes added with 1-2wt% of	
	LA and varied with 10-30 wt% and 10-100wt% NAp.	297
E2	Weight loss measurements and weight loss difference	
	in post-immersed membranes added with 3 wt% of LA	
	and varied with 10-30 wt% and 10-100wt% NAp.	298
E3	Weight loss measurements of 9-20 wt% PLGA membranes	
	loaded with 10-30wt% and 10-100wt% of NAp and LA	
	varied at 1-3 wt%.	299
E4	pH measurements of 9-20 wt% membranes containing	
	10-30 wt% and 10-100 wt% NAp and LA varied at 1-3wt%.	300
E5	pH measurements of 9-17 wt% and 9-20 wt% membranes	
	containing 10-30 wt% and 10-100 wt% NAp and LA loaded	
	at 3wt%.	301
F1	Comparison of water uptake in membranes containing	
	10-30wt% and 10-100wt% of NAp and LA varied at 1-3wt%.	302
F2	Comparison of water uptake in membranes containing	
	LA loaded at 1 & 2wt% and NAp varied at 10-30wt%	
	and 10-100wt%.	303
F3	Comparison of water uptake in membranes containing	
	LA loaded at 3wt% and NAp varied at 10-30wt% and	
	10-100wt% in 9-20 wt% and 9-17wt% of PLGA matrices.	304
G1	Data for standard calibration curve and loading	
	efficiency studies.	305
G2	Calculations for LA release from 3wt% of LA	
	loaded composite membrane containing 10-30wt% of	
	NAp in 9-20wt% of PLGA matrices (Sample A).	306

G3	Calculations for LA release from 3wt% of LA	
	loaded composite membrane containing 10-30wt% of	
	NAp in 9-20wt% of PLGA matrices (Sample B).	307
G4	Calculations for average LA release from 3wt% of LA	
	loaded composite membrane containing 10-30wt% of	
	NAp in 9-20wt% of PLGA matrices (Sample A+B).	308
Н	Data for S. aureus and P. aeruginosa growth inhibition against	
	LA concentration.	309
Ι	Data for cell viability of membrane samples.	310

CHAPTER 1

INTRODUCTION

1.1 Background

Rapid bone defect filling with normal bone is a challenge in the fields of orthopaedic and dentistry [1]. The bone has limited regeneration capability due to insufficient blood supply, large defects and invasion of highly proliferative nonosteogenic tissues that can impair bone repair [2,3]. Bone grafting is an established treatment to restore bone tissue. However, problems such as redundant fibrous connective tissue growth surrounding implanted bone graft and the movement of bone graft particles are still remain to be solved [1]. GBR has become an area of increasing interest in bone restorative procedures for guiding bone healing and regeneration [2,3] due to its success in curing cranial, maxillofacial and alveolar bone defects [4,5]. The concept of GBR is to cover the bone defect using a barrier membrane that enhances new bone ingrowth while preventing the ingrowth of fibrous tissue into the grafted site [6]. Hence, the bone regenerative approaches using GBR membranes have been extensively investigated to reveal their clinical potential [7,8,9].

GBR membranes have been widely studied as they are useful for bone repair in oral and maxillofacial surgery where limited mechanical loading exists [5,10]. The commercially available GBR membranes are made of non-resorbable and resorbable polymers. The non-resorbable polytetrafluoroethylene (PTFE) membranes have exhibited significant disadvantages such as requirement for second surgery and increased risk of infection leading to early removal of the membrane [9]. Collagen based resorbable membranes are widely used in clinical therapies. Since majority collagen membranes are animal derived, these membranes carry the risk of potential transmission of infectious agents, including the inappropriate immune responses in patients [7]. The synthetic resorbable membranes have found widespread use in clinical medicine as they are totally degradable, thus not requiring second surgery [8,9]. Poly(lactic-co-glycolic acid) (PLGA) is a FDA approved synthetic resorbable material and widely used in GBR applications [11,12]. Nonetheless, an inflammatory reaction by the accumulation of acidic degradation products in resorbable membranes has been reported [4,13]. The combination of calcium phosphate (CaP) with resorbable polymeric membranes is expected to neutralize the acidic degradation products from the membranes; which is intended to overcome inflammatory reaction in vivo [13,14,12,15,16]. Moreover, CaP particles in polymeric membranes has been also reported to improve structural integrity, flexibility and bone regeneration in vivo [17,15,18,14]. The aforementioned studies emphasises the need for incorporation of CaP particles to improve physical and mechanical properties of the resorbable polymeric membrane.

Currently, biomaterial-associated infection is regarded as a devastating complication in clinical surgery. Therefore, anti-infective biomaterials need to be developed as the main strategy to prevent infection in clinical applications [19]. A bacteria-free environment is highly important to regenerate bone tissues in GBR strategies [20]. Recently, the antibiotics incorporated GBR membranes have been developed for local delivery of antimicrobial agents [21]. Nonetheless, the increasing bacterial resistance prompted the development of alternative antimicrobial agent incorporated GBR membranes [22,23,20,24]. In light of this, a naturally derived antimicrobial agent to substitute the use of antibiotics is sought after to develop a new antimicrobial membrane for clinical applications.

The antimicrobial properties of naturally found fatty acids have been recognized for many years. Lauric acid (LA) is naturally found in coconut oil [25] and has been recognized to possess broad-spectrum with effective antimicrobial activity against gram-positive bacteria [26,27]. Unlike antibiotics, fatty acids and their derivatives have diverse modes of action that appear to be non-specific and

development of resistance to these compounds has not been reported [28]. It is suggested that LA kills Gram-positive bacteria by separating their inner and outer membranes, resulting in cytoplasmic disorganization of the bacteria [25]. Thus, it is envisaged that incorporating LA in composite membranes for anti-infective bone regeneration purposes could possibly overcome clinical complications caused by the administration of antibiotics.

The development of functionally graded and multiple layered membrane is to enhance the features required for GBR, namely a combination of physical, mechanical, biological and antimicrobial properties [13,23]. Also, the incorporation of functional gradients in a multilayered membrane structure offers the possibilities to overall usefulness to the membrane. Solvent casting technique offers the formation of layered membrane structure [16] whereas porous network formation is attainable through thermally induced phase separation (TIPS) [29] of the polymeric materials. The presence of residual toxic organic solvent is a major concern in solvent based fabrication technique. Thus, it is vital to include solvent removal step to reduce possible toxicity by solvent residues in fabricated membranes [30]. In this study, a new modified solvent casting-TIPS-solvent leaching technique is proposed to fabricate triple layered and graded composite PLGA membrane. Collectively, it is suggested that a new combination of CaP nanoparticles and LA as an antimicrobial agent being graded and layered in PLGA matrices can potentially function as an antimicrobial barrier membrane. This thesis will advance the knowledge in the area of antimicrobial composite membrane development for potential use in cranial, maxillofacial and dental applications. A new technique to establish the fabrication of multilayered and graded composite membrane utilizing solvent casting-TIPS-solvent leaching technique will be developed in this study. The fabrication and structural properties of the triple-layered PLGA membrane, graded with various amounts of LA and CaP nanoapatite will be studied. The effects of LA and CaP addition on the physical, chemical, mechanical, biological and antimicrobial properties of the PLGA composite membrane will also be explored. This membrane will deliver antimicrobial and osteoconductive properties by the incorporation of LA and CaP nanoapatite, respectively.

1.2 Problem statements

The major concerns in GBR surgical intervention are the problems related to the increasing bacterial resistance and side effects caused by antibiotics [31,32]. Multiple antibiotics are currently used to protect the bone defect from bacterial invasion, increasing the risks of bacterial resistance and side effects [33,22,31]. Hence, an alternative antimicrobial agent to substitute antibiotics is sought after. LA has been exhibiting effective antimicrobial activity against gram-positive bacteria that eliminates the need for multiple antibiotics to prevent bacteria colonization [26,27]. Therefore, the incorporation of antimicrobial LA in the composite membrane and its controlled release is proposed to circumvent the above mentioned drawbacks.

Apart from antimicrobial property, other important membrane characteristics such as surface morphology, pore size, membrane degradability, mechanical properties and cytocompatibility should be equally evaluated. Hence, appropriate materials selection and membrane design for GBR applications are highly indispensable for a successful bone defect treatment [7]. Poly(lactic-co-glycolic acid) (PLGA) is a FDA approved synthetic resorbable material which is widely used in GBR applications [11,12]. However, the accumulations of acidic degradation products from the synthetic bioresorbable membranes have been reported to cause inflammatory reaction in vivo [8,9]. Hence, the combination of synthetic polymers with CaP has been reported to neutralize the acidic degradation products from the polymers using ionic interactions [13,14,12,15,16]. Moreover, CaP incorporation improves structural integrity, flexibility and bone regeneration of the resorbable membranes [17,15,18,14]. Therefore, the current clinical disadvantage of using pure synthetic polymeric material as a GBR membrane could be overcome by incorporating CaP particles to reduce the potential inflammatory reactions. Thus, in this study, multiple ions substituted nanoapatite (NAp) powder which has close resemblance to natural bone mineral composition will be synthesized and incorporated into the PLGA matrices to form composite membranes.

Incorporating multiple additives in a composite membrane is a challenge as it requires the development of multilayered and graded membrane structure [13,16,34]. In order to address GBR applications, two functional surface layers are required. One of the surfaces with porous morphology allows bone ingrowth whereas the other dense surface prevents fibrous tissue penetration [16,13]. Therefore, in this study a triple-layered composite membrane with new combination of porous/dense layers will be developed. The NAp particles and LA will be graded in each layer to deliver osteoconductive and antimicrobial properties, respectively.

In order to develop a multilayered and graded composite membrane, an appropriate technique is indispensable to achieve the desired membrane structure. Currently, solvent casting [16] and TIPS [29] techniques have been employed to fabricate composite membranes. However, there are two disadvantages of using solvent casting method: i) toxic organic solvents application [15,18] that requires critical attention especially on its exposure in biomedical applications, ii) CaP particles can spontaneously precipitate from the polymer solution due to poor affinity and can cause non-uniform dispersion of CaP in polymer matrix [18]. Hence, these drawbacks could be overcome by freezing the CaP dispersed polymer matrix structure through TIPS technique. Moreover, solvent removal from the fabricated membrane is another important step to reduce toxic solvent residues [30,35]. Hence, in this study, composite membranes will be fabricated utilizing a new combination of solvent casting-TIPS-solvent leaching technique to address the formation of layered and graded membrane, dispersed with CaP particles and removal of toxic solvent from the membrane. The new modified technique is envisaged to form a composite membrane with graded porous/dense structure that has functional gradients, i.e., NAp and LA.

1.3 Objectives of the study

This work explores a novel fabrication technique, structure and design of a polymer-ceramic composite membrane incorporating LA as an antimicrobial agent. The goal is to design a functionally graded triple layered barrier membrane with

antimicrobial property using solvent casting-TIPS-solvent leaching techniques. In order to achieve the main objective, the following specific objectives were executed.

- a) To synthesise multiple ions substituted non-stoichiometric nanoapatite (NAp) powder.
- b) To establish a combined solvent casting-TIPS-solvent leaching techniques for the formation of triple-layered PLGA composite membranes graded with LA and NAp powder.
- c) To determine the physical, chemical, mechanical and in vitro degradation properties of the membrane.
- d) To evaluate the cytocompatibility and antimicrobial efficacy of the membrane.

1.4 Research hypothesis

It is possible to achieve an antimicrobial composite membrane by incorporating antimicrobial agents, in order to prevent biomaterial-associated infection in GBR applications. Therefore, it is envisaged that incorporating LA in the composite membrane could impart antimicrobial property which could prevent bacterial infection associated to the membrane. Furthermore, a resorbable composite membrane is desired to achieve less in vivo inflammation by reducing acidic degradation products through the addition of CaP particles [8,9]. Moreover, the combination of synthetic resorbable membranes with CaP is expected to deliver improved mechanical strength to the composite membranes [17,15,18,14]. Hence, in this study, it is hypothesised that varying the NAp and LA contents in PLGA matrices can significantly alter the physico-chemical, mechanical and antimicrobial properties of the membrane.

The GBR membrane is designed to have a smooth surface on one face to inhibit soft tissue penetration while the opposite porous face is capable of accommodating bone tissue ingrowth in vivo [16,36]. The dense/porous network formation through TIPS [29] technique is easily attainable whereas a multilayered membrane structure via solvent casting and the removal of solvent [30] could translate a safer membrane fabrication technique for clinical practice. The solvent casting-TIPS-solvent leaching technique will be used to test the hypothesis that one can tailor the properties of the different layers to form a functionally graded composite membrane to retain its structural, dimensional and mechanical properties for bone regeneration. Figure 1.1 demonstrates the importance of incorporating LA in composite membrane which may prevent bacterial infection on the membrane surface. In addition, formation of dense membrane surface also excludes fibroblast penetration into the barrier membrane.

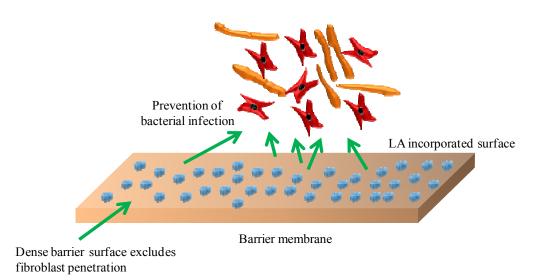


Figure 1.1: LA incorporation into barrier membrane as an antimicrobial agent for adjunct treatment in GBR procedures to inhibit bacterial infection.

1.5 Scope of the study

A new design of triple-layered and graded PLGA composite membrane has been fabricated. The triple layered membrane is comprised of PLGA matrix, graded with non-stoichiometric NAp and LA at each layer. PLGA with a lactic acid to glycolic acid ratio of 85:15 degrade over 2–6 months [37] and have the ability to deliver drugs locally in a controlled manner. These properties are making it suitable for use as a GBR barrier membrane. Besides improving mechanical strength of the membranes, the incorporation of CaP particles should be merely targeted for its osteoconductivity and hydrophilic nature to enhance bone growth into the polymer surfaces [38]. NAp powder is synthesized by introducing substituents within 1.84wt% (Na), 1.46wt% (Mg), 0.06wt% (K) and 4.80wt% (CO_3^{2-1} to closely mimic natural bone apatite. The NAp powder is incorporated to enhance bioactivity and osteoconductivity of the membrane. LA is added to introduce antimicrobial properties to the composite membrane to prevent bacterial infection as it is known to possess effective antimicrobial activity against gram-positive bacteria [26,27]. The composite membrane is fabricated by employing a modified solvent casting-TIPSsolvent leaching technique. The solvent casting facilitated lamination of multiple layers of graded LA and NAp in PLGA matrices whereas TIPS used to form porous/dense layers in the membrane structure. Solvent leaching is performed to remove toxic solvent residues.

1.6 Significance of the study

LA, as a substitute for antibiotics is identified and incorporated in the composite membrane which is to be used as a potential antimicrobial membrane for clinical applications. Prevention of bacterial infection is a promising strategy whereby LA imparts antimicrobial activity on the membrane surface. This would render an antimicrobial barrier membrane appropriate for adjunctive treatment in guiding bone regeneration. This work also reports the fabrication of PLGA-NAp-LA composite membrane using solvent casting-TIPS-solvent leaching technique. This new technique largely eliminates the solvent residue in the fabricated membrane through solvent leaching step using water as the exchanging medium.

1.7 Thesis outline

Chapter 1 is the introduction to the study of this thesis. The entire outline of the thesis is illustrated in Fig. 1.2.

Chapter 2 describes the review of literatures related to the development and application of commercially available GBR membrane that has been related to its profound improvement through current research to overcome clinically reported shortcomings. Moreover, selection criteria for PLGA, LA and NAp are also reviewed to ensure the fabricated composite membrane is more likely to possess appropriate physical, structural, dimensional, mechanical, antimicrobial and biological properties for potential use in bone regeneration procedures.

Chapter 3 deals with the materials and methods used to investigate the appropriate parameters, experimental set-up, test conditions, characterization using analytical equipment and material evaluation involved in the fabrication and evaluation of the composite membranes. The synthesis of NAp powder is reported in the first part of the chapter. Subsequently, the development of PLGA based NAp-LA composite membrane through a new fabrication technique using solvent casting-TIPS-solvent leaching is reported. This is followed by the development of methods to test on the membrane's properties such as physico-chemical, mechanical, in vitro degradation profile over six months duration, quantification of LA release and finally, LA release mechanism; since the effects of NAp and LA additions in the PLGA membranes are highly imperative to meet the design criteria of membranes for GBR applications.

Chapter 4 elaborates the outcome of NAp synthesis, fabrication of composite membranes, degradation profiles for composite membranes, mechanical evaluation of membranes in dry and wet condition, released LA concentration and its release Synthesis of NAp with the highest substitutent composition, the mechanism. morphology of triple layered membrane, phase composition, physical changes in amorphous/crystalline state of LA, interaction mechanisms between PLGA-NAp-LA in composite membranes, weight loss and water absorption of membranes, and finally the quantification of LA release and its release mechanism from composite membranes for sufficient antimicrobial effects while maintaining its cytocompatibility are discussed. The cytocompatibility of synthesized NAp powder and composite membranes along with antimicrobial evaluation on the effects of LA addition in composite membranes were discussed.

Chapter 5 concludes structural, dimensional and mechanical integrity of the layered and graded composite membrane. The effects of LA and NAp addition on physico-chemical, mechanical and antimicrobial properties are also described.

Publications and presentations at conferences: This section forms part of the thesis, which described the synthesis of NAp powder and the fabrication of composite membranes published in peer reviewed impact factor journals and presented at international conferences as listed in Appendix A.

CHAPTER 1:Introduction

- Background.
- Problem statement.
- Objectives of the study.
- · Research hypothesis.
- Scope of the study.
- Significance of the study.

CHAPTER 2: Literature review

- Bone damage and tissue reconstruction.
- Alveolar bone loss and treatment modalities.
- Principles of guided bone regeneration.
- Design criteria for GBR membrane.
- Comparison between types of commercially available membranes.
- Antimicrobial properties of GBR membranes.
- Functionally graded and layered composite GBR membranes.
- Membrane fabrication techniques.
- In vitro degradation characteristics of PLGA based membranes.
- The drug release mechanism in PLGA based membranes.
- Challenges in GBR using barrier membranes

CHAPTER 3: Methodology

- Synthesis of NAp powder
- Fabrication and evaluation of composite membranes
- In vitro degradation of membranes
- •In vitro antimicrobial efficacy of membranes
- Cytotoxicity on composite membranes

CHAPTER 4: Results and discussion

- NAp composition with the highest ions substitution.
- Morphology of triple layered membrane, phase composition, physical changes in amorphous/ crystalline state of LA.
- Interaction mechanisms between PLGA-NAp-LA in composite membranes.
- Tensile strength, stiffness vs elasticity and elongation of composite membranes.
- In vitro weight loss and water absorption of membranes.
- Quantification and mechanism of LA release from composite membranes for sufficient antimicrobial effects while maintaining its cytocompatibility.

CHAPTER 5: Conclusion and future recommendations

- Concludes structural, dimensional and mechanical integrity of the membranes.
- The effects of LA and NAp addition on physicochemical, mechanical and antimicrobial properties.
- Recommended to improve tensile strength of membranes and to test against various types of bacteria.

Figure 1.2: Representation of thesis outline.

REFERENCES

- Cai, Y., Guo, J., Chen, C., Yao, C., Chung, S.-m., Yao, J., Lee, I.-s. and Kong, X. Silk fi broin membrane used for guided bone tissue regeneration. *Materials Science & Engineering C.* 2017. 70: 148-154.
- Pasetto, S., Herculano, R.D., Ereno, C., Silva, P., Graeff, C.F.O., Tavano, O., Baffa, O. and Kinoshita, A. Latex use as an occlusive membrane for guided bone regeneration. *Journal of Biomedical Materials Research A*. 2010. 932-939.
- Moura, J.M.L., Ferreira, J.F., Marques, L., Holgado, L., Graeff, C.F.O. and Kinoshita, A. Comparison of the performance of natural latex membranes prepared with different procedures and PTFE membrane in guided bone regeneration (GBR) in rabbits. *Journal of Materials Science: Materials in Medicine*. 2014. 25(9): 2111-2120.
- Fujihara, K., Kotaki, M. and Ramakrishna, S. Guided bone regeneration membrane made of polycaprolactone/calcium carbonate composite nanofibers. *Biomaterials*. 2005. 26(19): 4139-4147.
- Kellomäki, M., Niiranen, H., Puumanen, K., Ashammakhi, N., Waris, T. and Törmälä, P. Bioabsorbable scaffolds for guided bone regeneration and generation. *Biomaterials*. 2000. 21(24): 2495-2505.
- Yang, F., Both, S.K., Yang, X., Walboomers, X.F. and Jansen, J.A. Development of an electrospun nano-apatite/PCL composite membrane for GTR/GBR application. *Acta Biomaterialia*. 2009. 5(9): 3295-3304.
- Jung, R.E., Kokovic, V., Jurisic, M., Yaman, D., Subramani, K. and Weber, F.E. Guided bone regeneration with a synthetic biodegradable membrane: a comparative study in dogs. *Clinical Oral Implants Research*. 2011. 22(8): 802-807.
- 8. von Arx, T., Cochran, D.L., Schenk, R.K. and Buser, D. Evaluation of a prototype trilayer membrane (PTLM) for lateral ridge augmentation: an

- van Leeuwen, A.C., Huddleston Slater, J.J.R., Gielkens, P.F.M., de Jong, J.R., Grijpma, D.W. and Bos, R.R.M. Guided bone regeneration in rat mandibular defects using resorbable poly(trimethylene carbonate) barrier membranes. *Acta Biomaterialia*. 2012. 8(4): 1422-1429.
- Peltola, O.L.J., Asikainen, A.J., Noponen, J. and Mesim, K.A. Tyrosine derived polycarbonate membrane is useful for guided bone regeneration in rabbit mandibular defects. *Journal of Materials Science: Materials in Medicine*. 2005. 16: 753-758.
- Jazayeri, H.E., Tahriri, M., Razavi, M., Khoshroo, K., Fahimipour, F., Dashtimoghadam, E., Almeida, L. and Tayebi, L. A current overview of materials and strategies for potential use in maxillofacial tissue regeneration. *Materials Science & Engineering C.* 2017. 70: 913-929.
- Liao, S., Watari, F., Zhu, Y., Uo, M., Akasaka, T., Wang, W., Xu, G. and Cui, F. The degradation of the three layered nano-carbonated hydroxyapatite/collagen/PLGA composite membrane in vitro. *Dental Materials.* 2007. 23(9): 1120-1128.
- Bottino, M.C., Thomas, V. and Janowski, G.M. A novel spatially designed and functionally graded electrospun membrane for periodontal regeneration. *Acta Biomaterialia*. 2011. 7(1): 216-224.
- Kikuchi, M., Koyama, Y., Yamada, T., Imamura, Y., Okada, T., Shirahama, N., Akita, K., Takakuda, K. and Tanaka, J. Development of guided bone regeneration membrane composed of β-tricalcium phosphate and poly (llactide-co-glycolide-co-ε-caprolactone) composites. *Biomaterials*. 2004. 25(28): 5979-5986.
- Puppi, D., Chiellini, F., Piras, A.M. and Chiellini, E. Polymeric materials for bone and cartilage repair. *Progress in Polymer Science*. 2010. 35(4): 403-440.
- Liao, S., Wang, W., Uo, M., Ohkawa, S., Akasaka, T., Tamura, K., Cui, F. and Watari, F. A three-layered nano-carbonated hydroxyapatite /collagen /PLGA composite membrane for guided tissue regeneration. *Biomaterials*. 2005. 26(36): 7564-7571.

- Song, X., Ling, F., Ma, L., Yang, C. and Chen, X. Electrospun hydroxyapatite grafted poly(l-lactide)/poly(lactic-co-glycolic acid) nanofibers for guided bone regeneration membrane. *Composites Science and Technology*. 2013. 79: 8-14.
- Zhou, H., Lawrence, J.G. and Bhaduri, S.B. Fabrication aspects of PLA-CaP/PLGA-CaP composites for orthopedic applications: A review. *Acta Biomaterialia*. 2012. 8(6): 1999-2016.
- Campoccia, D., Montanaro, L. and Arciola, C.R. A review of the clinical implications of anti-infective biomaterials and infection-resistant surfaces. *Biomaterials*. 2013. 34(33): 8018-8029.
- Münchow, E.A., Albuquerque, M.T.P., Zero, B., Kamocki, K., Piva, E., Gregory, R.L. and Bottino, M.C. Development and characterization of novel ZnO-loaded electrospun membranes for periodontal regeneration. *Dental Materials.* 2015. 31(9): 1038-1051.
- Chen, D.W.C., Lee, F.Y., Liao, J.Y., Liu, S.J., Hsiao, C.Y. and Chen, J.K. Preclinical Experiments on the Release Behavior of Biodegradable Nanofibrous Multipharmaceutical Membranes in a Model of Four-Wall Intrabony Defect. *Antimicrobial Agents and Chemotherapy*. 2012. 57(1): 9-14.
- Ji, W., Wang, H., van den Beucken, J.J.J.P., Yang, F., Walboomers, X.F., Leeuwenburgh, S. and Jansen, J.A. Local delivery of small and large biomolecules in craniomaxillofacial bone. *Advanced Drug Delivery Reviews*. 2012. 64(12): 1152-1164.
- Bottino, M.C., Thomas, V., Schmidt, G., Vohra, Y.K., Chu, T.-M.G., Kowolik, M.J. and Janowski, G.M. Recent advances in the development of GTR/GBR membranes for periodontal regeneration—A materials perspective. *Dental Materials*. 2012. 28(7): 703-721.
- 24. de Breij, A., Riool, M., Kwakman, P.H.S., de Boer, L., Cordfunke, R.A., Drijfhout, J.W., Cohen, O., Emanuel, N., Zaat, S.A.J., Nibbering, P.H. and Moriarty, T.F. Prevention of Staphylococcus aureus biomaterial-associated infections using a polymer-lipid coating containing the antimicrobial peptide OP-145. *Journal of Controlled Release*. 2016. 222: 1-8.
- 25. Yang, D., Pornpattananangkul, D., Nakatsuji, T., Chan, M., Carson, D., Huang, C.-M. and Zhang, L. The antimicrobial activity of liposomal lauric

acids against Propionibacterium acnes. *Biomaterials*. 2009. 30(30): 6035-6040.

- Bergsson, G., Arnfinnsson, J., Steingrimsson, O. and Thormar, H. In Vitro Killing of Candida albicans by Fatty Acids and Monoglycerides. *Antimicrobial Agents and Chemotherapy*. 2001. 45(11): 3209-3212.
- Rouse, M.S., Rotger, M., Piper, K.E., Steckelberg, J.M., Scholz, M., Andrews, J. and Patel, R. In Vitro and In Vivo Evaluations of the Activities of Lauric Acid Monoester Formulations against Staphylococcus aureus. *Antimicrobial Agents and Chemotherapy*. 2005. 49(8): 3187-3191.
- Nobmann, P., Bourke, P., Dunne, J. and Henehan, G. In vitroantimicrobial activity and mechanism of action of novel carbohydrate fatty acid derivatives againstStaphylococcus aureusand MRSA. *Journal of Applied Microbiology*. 2010. 108: 2152-61.
- Ho, M.-H., Hsieh, C.-C., Hsiao, S.-W. and Van Hong Thien, D. Fabrication of asymmetric chitosan GTR membranes for the treatment of periodontal disease. *Carbohydrate Polymers*. 2010. 79(4): 955-963.
- Rowlands, A.S., Lim, S.A., Martin, D. and Cooper-White, J.J. Polyurethane/poly(lactic-co-glycolic) acid composite scaffolds fabricated by thermally induced phase separation. *Biomaterials*. 2007. 28(12): 2109-2121.
- Schwach-abdellaoui, K., Vivien-castioni, N. and Gurny, R. Local delivery of antimicrobial agents for the treatment of periodontal diseases. *European Journal of Pharmaceutics and Biopharmaceutics*. 2000. 50: 83-99.
- 32. Sweeney, L.C. Antibiotic resistance in general dental practice--a cause for concern? *Journal of Antimicrobial Chemotherapy*. 2004. 53(4): 567-576.
- Herrera, D., Matesanz, P., Bascones-Martínez, A. and Sanz, M. Local and Systemic Antimicrobial Therapy in Periodontics. *Journal of Evidence Based Dental Practice*. 2012. 12(3): 50-60.
- Sun, F., Zhou, H. and Lee, J. Various preparation methods of highly porous hydroxyapatite/polymer nanoscale biocomposites for bone regeneration. *Acta Biomaterialia*. 2011. 7(11): 3813-3828.
- Vaquette, C. and Cooper-White, J. A simple method for fabricating 3-D multilayered composite scaffolds. *Acta Biomaterialia*. 2013. 9(1): 4599-4608.

- Carlo Reis, E.C., Borges, A.P.B., Araújo, M.V.F., Mendes, V.C., Guan, L. and Davies, J.E. Periodontal regeneration using a bilayered PLGA/calcium phosphate construct. *Biomaterials*. 2011. 32(35): 9244-9253.
- 37. Owen, G.R., Jackson, J.K., Chehroudi, B., Brunette, D.M. and Burt, H.M. An in vitro study of plasticized poly(lactic-co-glycolic acid) films as possible guided tissue regeneration membranes: Material properties and drug release kinetics. *Journal of Biomedical Materials Research Part A*. 2010. 95A(3): 857-869.
- Lee, E.-J., Teng, S.-H., Jang, T.-S., Wang, P., Yook, S.-W., Kim, H.-E. and Koh, Y.-H. Nanostructured poly(ε-caprolactone)–silica xerogel fibrous membrane for guided bone regeneration. *Acta Biomaterialia*. 2010. 6(9): 3557-3565.
- 39. Yu, Z., Geng, J., Gao, H., Zhao, X. and Chen, J. Evaluations of guided bone regeneration in canine radius segmental defects using autologous periosteum combined with fascia lata under stable external fixation. *Journal of Orthopaedics and Traumatology*. 2015. 16(2): 133-140.
- Fu, S., Ni, P., Wang, B., Chu, B., Zheng, L., Luo, F., Luo, J. and Qian, Z. Injectable and thermo-sensitive PEG-PCL-PEG copolymer/collagen/n-HA hydrogel composite for guided bone regeneration. *Biomaterials*. 2012. 33(19): 4801-4809.
- Lee, Y.J., Lee, J.-H., Cho, H.-J., Kim, H.K., Yoon, T.R. and Shin, H. Electrospun fibers immobilized with bone forming peptide-1 derived from BMP7 for guided bone regeneration. *Biomaterials*. 2013. 34(21): 5059-5069.
- Lee, E.-J., Shin, D.-S., Kim, H.-E., Kim, H.-W., Koh, Y.-H. and Jang, J.-H. Membrane of hybrid chitosan–silica xerogel for guided bone regeneration. *Biomaterials*. 2009. 30(5): 743-750.
- 43. Jung, R.E., Zwahlen, R., Weber, F.E., Molenberg, A., van Lenthe, G.H. and Hammerle, C.H.F. Evaluation of an in situ formed synthetic hydrogel as a biodegradable membrane for guided bone regeneration. *Clinical Oral Implants Research.* 2006. 17(4): 426-433.
- 44. Nieminen, T., Kallela, I., Keränen, J., Hiidenheimo, I., Kainulainen, H., Wuolijoki, E. and Rantala, I. In vivo and in vitro degradation of a novel

bioactive guided tissue regeneration membrane. *International Journal of Oral and Maxillofacial Surgery*. 2006. 35(8): 727-732.

- 45. Schliephake, H., Dard, M., Planck, H., Hierlemann, H. and Jakob, A. Guided bone regeneration around endosseous implants using a resorbable membrane vs a PTFE membrane. *Clinical oral implants research*. 2000. 11(3): 230-41.
- Bottino, M.C., Kamocki, K., Yassen, G.H., Platt, J.A., Vail, M.M., Ehrlich, Y., Spolnik, K.J. and Gregory, R.L. Bioactive Nanofibrous Scaffolds for Regenerative Endodontics. *Journal of Dental Research*. 2013. 92(11): 963-969.
- Vasconcelos, M., Afonso, A., Branco, R. and Cavalheiro, J. Guided bone regeneration using osteopatite granules and polytetrafluoroethylene membranes. *Journal of Materials Science: Materials in Medicine*. 1997. 8: 815-818.
- Hitti, R.A. and Kerns, D.G. Guided Bone Regeneration in the Oral Cavity : A Review. *Pathology*. 2011. 5: 33-45.
- Ten Heggeler, J.M.A.G., Slot, D.E. and Van der Weijden, G.A. Effect of socket preservation therapies following tooth extraction in non-molar regions in humans: a systematic review. *Clinical Oral Implants Research*. 2011. 22(8): 779-788.
- Pilipchuk, S.P., Plonka, A.B., Monje, A., Taut, A.D., Lanis, A., Kang, B. and Giannobile, W.V. Tissue engineering for bone regeneration and osseointegration in the oral cavity. *Dental Materials*. 2015. 31(4): 317-338.
- De Boever, A.L. and De Boever, J.A. Guided bone regeneration around nonsubmerged implants in narrow alveolar ridges: a prospective long-term clinical study. *Clinical Oral Implants Research*. 2005. 16(5): 549-556.
- Rakhmatia, Y.D., Ayukawa, Y., Furuhashi, A. and Koyano, K. Current barrier membranes: Titanium mesh and other membranes for guided bone regeneration in dental applications. *Journal of Prosthodontic Research*. 2013. 57(1): 3-14.
- Retzepi, M. and Donos, N. Guided Bone Regeneration: biological principle and therapeutic applications. *Clinical Oral Implants Research*. 2010. 21(6): 567-576.
- 54. Hämmerle, C.H.F. and Jung, R.E. Bone augmentation by means of barrier membranes. *Periodontology 2000.* 2003. 33: 36-53.

- 55. Dumitrescu, A.L. Guided Tissue Regeneration Barriers. 2011. 1-71.
- Moses, O., Pitaru, S., Artzi, Z. and Nemcovsky, C.E. Healing of dehiscencetype defects in implants placed together with different barrier membranes: a comparative clinical study. *Clinical Oral Implants Research*. 2005. 16(2): 210-219.
- 57. Kim, Y.-K., Yun, P.-Y., Kim, S.-G. and Oh, D.S. In vitro scanning electron microscopic comparison of inner surface of exposed and unexposed nonresorbable membranes. *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology.* 2009. 107(6): e5-e11.
- Strietzel, F.P., Khongkhunthian, P., Khattiya, R., Patchanee, P. and Reichart,
 P.A. Healing Pattern of Bone Defects Covered by Different Membrane
 Types A Histologic Study in the Porcine Mandible. *Journal of Biomedical Materials Research Part B: Applied Biomaterials*. 2005. 35-46.
- 59. Donos, N., Kostopoulos, L. and Karring, T. Alveolar ridge augmentation using a resorbable copolymer membrane and autogenous bone grafts. An experimental study in the rat. *Clinical oral implants research*. 2002. 13(2): 203-213.
- 60. Belleggia, F. Treatment of an infected exposure of a dense polytetrafluoroethylene membrane in a vertical guided bone regeneration procedure : a protocol proposal. *Clinical Oral Implants Research*. 2014. 25 (Suppl. 10).
- Harzeler, M.R., Kohal, R.J., Naghshbandi, J., Mota, L.F., Conradt, J., Mote, L.F. and Conradt, J. Evaluation of a new bioresorbable barrier to facilitate guided bone regeneration around exposed implant threads An experimental study in the monkey. *Int. J. Oral Maxillofacial Surgery*. 1998. 27: 315-320.
- Coelho, P.G., Giro, G., Kim, W., Granato, R., Marin, C., Bonfante, E.A., Bonfante, S., Lilin, T. and Suzuki, M. Evaluation of collagen-based membranes for guided bone regeneration, by three-dimensional computerized microtomography. *Oral Surgery, Oral Medicine, Oral Pathology and Oral Radiology*. 2012. 114(4): 437-443.
- Gentile, P., Chiono, V., Tonda-Turo, C., Ferreira, A.M. and Ciardelli, G. Polymeric membranes for guided bone regeneration. *Biotechnology Journal*. 2011. 6(10): 1187-1197.

- Song, J.-h., Kim, H.-e. and Kim, H.-w. Collagen-Apatite Nanocomposite Membranes for Guided Bone Regeneration. *Journal of Biomedical Materials Research.* 2007. 248-257.
- 65. Landi, E., Tampieri, A., Mattioli-Belmonte, M., Celotti, G., Sandri, M., Gigante, A., Fava, P. and Biagini, G. Biomimetic Mg- and Mg,CO3substituted hydroxyapatites: synthesis characterization and in vitro behaviour. *Journal of the European Ceramic Society*. 2006. 26(13): 2593-2601.
- 66. Zhou, H. and Lee, J. Nanoscale hydroxyapatite particles for bone tissue engineering. *Acta Biomaterialia*. 2011. 7(7): 2769-2781.
- Park, J.K., Yeom, J., Oh, E.J., Reddy, M., Kim, J.Y., Cho, D.-W., Lim, H.P., Kim, N.S., Park, S.W., Shin, H.-I., Yang, D.J., Park, K.B. and Hahn, S.K. Guided bone regeneration by poly(lactic-co-glycolic acid) grafted hyaluronic acid bi-layer films for periodontal barrier applications. *Acta Biomaterialia*. 2009. 5(9): 3394-3403.
- Parent, M., Nouvel, C., Koerber, M., Sapin, A., Maincent, P. and Boudier, A.
 PLGA in situ implants formed by phase inversion: Critical physicochemical parameters to modulate drug release. *Journal of Controlled Release*. 2013. 172(1): 292-304.
- 69. Rezwan, K., Chen, Q.Z., Blaker, J.J. and Boccaccini, A.R. Biodegradable and bioactive porous polymer/inorganic composite scaffolds for bone tissue engineering. *Biomaterials*. 2006. 27(18): 3413-3431.
- Budyanto, L., Goh, Y.Q. and Ooi, C.P. Fabrication of porous poly(L-lactide) (PLLA) scaffolds for tissue engineering using liquid–liquid phase separation and freeze extraction. *Journal of Materials Science: Materials in Medicine*. 2008. 20(1): 105-111.
- Zamani, M., Morshed, M., Varshosaz, J. and Jannesari, M. Controlled release of metronidazole benzoate from poly ε-caprolactone electrospun nanofibers for periodontal diseases. *European Journal of Pharmaceutics and Biopharmaceutics*. 2010. 75(2): 179-185.
- Chen, S., Hao, Y., Cui, W., Chang, J. and Zhou, Y. Biodegradable electrospun PLLA/chitosan membrane as guided tissue regeneration membrane for treating periodontitis. *Journal of Materials Science*. 2013. 48(19): 6567-6577.

- Ranjbar-Mohammadi, M., Zamani, M., Prabhakaran, M.P., Bahrami, S.H. and Ramakrishna, S. Electrospinning of PLGA/gum tragacanth nanofibers containing tetracycline hydrochloride for periodontal regeneration. *Materials Science and Engineering: C.* 2016. 58: 521-531.
- Yüksel, E., Karakeçili, A., Demirtaş, T.T. and Gümüşderelioğlu, M. Preparation of bioactive and antimicrobial PLGA membranes by magainin II/EGF functionalization. *International Journal of Biological Macromolecules*. 2016. 86: 162-168.
- 75. Dorozhkin, S.V. Nanosized and nanocrystalline calcium orthophosphates. *Acta Biomaterialia*. 2010. 6(3): 715-734.
- Laurencin, D., Almora-Barrios, N., de Leeuw, N.H., Gervais, C., Bonhomme, C., Mauri, F., Chrzanowski, W., Knowles, J.C., Newport, R.J., Wong, A., Gan, Z. and Smith, M.E. Magnesium incorporation into hydroxyapatite. *Biomaterials*. 2011. 32(7): 1826-1837.
- 77. Kannan, S., Ventura, J.M.G. and Ferreira, J.M.F. Synthesis and thermal stability of potassium substituted hydroxyapatites and hydroxyapatite/βtricalciumphosphate mixtures. *Ceramics International*. 2007. 33(8): 1489-1494.
- Kothapalli, C., Wei, M., Vasiliev, A. and Shaw, M.T. Influence of temperature and concentration on the sintering behavior and mechanical properties of hydroxyapatite. *Acta Materialia*. 2004. 52(19): 5655-5663.
- Pretto, M., Costa, A.L., Landi, E., Tampieri, A., Galassi, C. Dispersing Behavior of Hydroxyapatite Powders Produced by Wet-Chemical Synthesis. *J. Am. Ceram. Soc.* 2003. 86(9): 1534-39.
- 80. Kannan, S., Vieira, S.I., Olhero, S.M., Torres, P.M.C., Pina, S., da Cruz e Silva, O.A.B. and Ferreira, J.M.F. Synthesis, mechanical and biological characterization of ionic doped carbonated hydroxyapatite/β-tricalcium phosphate mixtures. *Acta Biomaterialia*. 2011. 7(4): 1835-1843.
- Kim, S.R., Lee, J.H., Kim, Y.T., Riu, D.H., Jung, S.J., Lee, Y.J., Chung, S.C. and Kim, Y.H. Synthesis of Si, Mg substituted hydroxyapatites and their sintering behaviors. *Biomaterials*. 2003. 24(8): 1389-98.
- 82. Sprio, S., Tampieri, A., Landi, E., Sandri, M., Martorana, S., Celotti, G. and Logroscino, G. Physico-chemical properties and solubility behaviour of

multi-substituted hydroxyapatite powders containing silicon. *Materials Science and Engineering: C.* 2008. 28(1): 179-187.

- Landi, E., Tampieri, A., Celotti, G., Sprio, S., Sandri, M. and Logroscino, G. Sr-substituted hydroxyapatites for osteoporotic bone replacement. *Acta Biomaterialia*. 2007. 3(6): 961-969.
- Ramesh, S., Tan, C.Y., Tolouei, R., Amiriyan, M., Purbolaksono, J., Sopyan,
 I. and Teng, W.D. Sintering behavior of hydroxyapatite prepared from different routes. *Materials & Design*. 2012. 34: 148-154.
- 85. Sopyan, I., Ramesh, S. and Hamdi, M. Synthesis of nano sized hydroxyapatite powder using sol-gel technique and its conversion to dense and porous bodies. *Indian Journal of Chemistry*. 2008. 47(November): 1626-1631.
- Landi, E., Tampieri, A., Celotti, G. and Sprio, S. Densification behaviour and mechanisms of synthetic hydroxyapatites. *J. Euro. Ceram. Soc.* 2000. 20: 2377-2387.
- Uskoković, V. and Uskoković, D.P. Nanosized hydroxyapatite and other calcium phosphates: Chemistry of formation and application as drug and gene delivery agents. *Journal of Biomedical Materials Research Part B: Applied Biomaterials*. 2011. 96B(1): 152-191.
- Kuriakose, T.A., Kalkura, S.N., Palanichamy, M., Arivuoli, D., Dierks, K., Bocelli, G. and Betzel, C. Synthesis of stoichiometric nano crystalline hydroxyapatite by ethanol-based sol–gel technique at low temperature. *Journal of Crystal Growth*. 2004. 263(1-4): 517-523.
- Pang, Y.X. and Bao, X. Influence of temperature, ripening time and calcination on the morphology and crystallinity of hydroxyapatite nanoparticles. *Journal of the European Ceramic Society*. 2003. 23(10): 1697-1704.
- 90. Cacciotti, I., Bianco, A., Lombardi, M. and Montanaro, L. Mg-substituted hydroxyapatite nanopowders: Synthesis, thermal stability and sintering behaviour. *Journal of the European Ceramic Society*. 2009. 29(14): 2969-2978.
- Boanini, E., Gazzano, M. and Bigi, A. Ionic substitutions in calcium phosphates synthesized at low temperature. *Acta Biomaterialia*. 2010. 6(6): 1882-1894.

- 92. Lazic, S., Zec, S., Miljevic, N. and Milonjic, S. The effect of temperature on the properties of hydroxyapatite precipitated from calcium hydroxide and phosphoric acid. *Thermochimica acta*. 2001. 374: 13-22.
- Reise, M., Wyrwa, R., Müller, U., Zylinski, M., Völpel, A., Schnabelrauch, M., Berg, A., Jandt, K.D., Watts, D.C. and Sigusch, B.W. Release of metronidazole from electrospun poly(l-lactide-co-d/l-lactide) fibers for local periodontitis treatment. *Dental Materials*. 2012. 28(2): 179-188.
- 94. Xue, J., He, M., Niu, Y., Liu, H., Crawford, A., Coates, P., Chen, D., Shi, R. and Zhang, L. Preparation and in vivo efficient anti-infection property of GTR/GBR implant made by metronidazole loaded electrospun polycaprolactone nanofiber membrane. *International Journal of Pharmaceutics*. 2014. 475(1-2): 566-577.
- Sundararaj, S.C., Thomas, M.V., Peyyala, R., Dziubla, T.D. and Puleo, D.A. Design of a multiple drug delivery system directed at periodontitis. *Biomaterials*. 2013. 34(34): 8835-8842.
- 96. Lan, S.-F., Kehinde, T., Zhang, X., Khajotia, S., Schmidtke, D.W. and Starly,
 B. Controlled release of metronidazole from composite poly-εcaprolactone/alginate (PCL/alginate) rings for dental implants. *Dental Materials.* 2013. 29(6): 656-665.
- 97. Kitahara, T., Aoyama, Y., Hirakata, Y., Kamihira, S., Kohno, S., Ichikawa, N., Nakashima, M., Sasaki, H. and Higuchi, S. In vitro activity of lauric acid or myristylamine in combination with six antimicrobial agents against methicillin-resistant Staphylococcus aureus (MRSA). *International Journal of Antimicrobial Agents*. 2006. 27(1): 51-57.
- 98. Pragati, S., Ashok, S. and Kuldeep, S. Recent advances in periodontal drug delivery systems. *International Journal of Drug Delivery*. 2009. 1: 1-14.
- 99. Castillo, A., Liebana, J., Lopez, E., Baca, P., Liebana, M. and Castillo, F. Interference of antibiotics in the growth curves of oral streptococci. *International Journal of Antimicrobial Agents*. 2006. 27(3): 263-266.
- 100. Pornpattananangkul, D., Fu, V., Thamphiwatana, S., Zhang, L., Chen, M., Vecchio, J., Gao, W., Huang, C.-M. and Zhang, L. In Vivo Treatment ofPropionibacterium acnesInfection with Liposomal Lauric Acids. *Advanced Healthcare Materials*. 2013. 2(10): 1322-1328.

- 101. Huang, C.B., Alimova, Y., Myers, T.M. and Ebersole, J.L. Short- and medium-chain fatty acids exhibit antimicrobial activity for oral microorganisms. *Archives of Oral Biology*. 2011. 56(7): 650-654.
- 102. Kitahara, T., Koyama, N., Matsuda, J., Aoyama, Y., Hirakata, Y., Kamihira, S., Kohno, S., Nakashima, M. and Sasaki, H. Antimicrobial activity of saturated fatty acids and fatty amines against methicillin-resistant Staphylococcus aureus. *Biological & pharmaceutical bulletin.* 2004. 27(9): 1321-1326.
- 103. Amet, Y., Adas, F. and Berthou, F. High performance liquid chromatography of fatty acid metabolites Improvement of sensitivity by radiometric , fluorimetric and mass spectrometric methods. *Analytica Chimica Acta*. 2002. 465: 193-198.
- 104. Zhao, L., Hu, Y., Xu, D. and Cai, K. Surface functionalization of titanium substrates with chitosan-lauric acid conjugate to enhance osteoblasts functions and inhibit bacteria adhesion. *Colloids and Surfaces B: Biointerfaces.* 2014. 119: 115-125.
- Renouf-Glauser, A.C., Rose, J., Farrar, D. and Cameron, R.E. A degradation study of PLLA containing lauric acid. *Biomaterials*. 2005. 26(15): 2415-2422.
- 106. Teng, S.-H., Lee, E.-J., Wang, P., Shin, D.-S. and Kim, H.-E. Three-layered membranes of collagen/hydroxyapatite and chitosan for guided bone regeneration. *Journal of Biomedical Materials Research Part B: Applied Biomaterials.* 2008. 87B(1): 132-138.
- 107. Vaquette, C., Frochot, C., Rahouadj, R. and Wang, X. An innovative method to obtain porous PLLA scaffolds with highly spherical and interconnected pores. *Journal of Biomedical Materials Research Part B: Applied Biomaterials.* 2008. 86B(1): 9-17.
- Vaquette, C. and Cooper-White, J.J. Increasing electrospun scaffold pore size with tailored collectors for improved cell penetration. *Acta Biomaterialia*. 2011. 7(6): 2544-2557.
- Ji, C., Khademhosseini, A. and Dehghani, F. Enhancing cell penetration and proliferation in chitosan hydrogels for tissue engineering applications. *Biomaterials*. 2011. 32(36): 9719-9729.

- 110. Cho, W.J., Kim, J.H., Oh, S.H., Nam, H.H., Kim, J.M. and Lee, J.H. Hydrophilized polycaprolactone nanofiber mesh-embedded poly(glycolic-colactic acid) membrane for effective guided bone regeneration. *Journal of Biomedical Materials Research Part A*. 2009. 91A(2): 400-407.
- 111. Teng, S.-H., Lee, E.-J., Yoon, B.-H., Shin, D.-S., Kim, H.-E. and Oh, J.-S. Chitosan/nanohydroxyapatite composite membranes via dynamic filtration for guided bone regeneration. *Journal of Biomedical Materials Research Part A.* 2009. 88A(3): 569-580.
- 112. Sun, F., Lim, B.-K., Ryu, S.-C., Lee, D. and Lee, J. Preparation of multilayered film of hydroxyapatite and chitosan. *Materials Science and Engineering: C.* 2010. 30(6): 789-794.
- 113. Yang, Y., Zhao, J., Zhao, Y., Wen, L., Yuan, X. and Fan, Y. Formation of porous PLGA scaffolds by a combining method of thermally induced phase separation and porogen leaching. *Journal of Applied Polymer Science*. 2008. 109(2): 1232-1241.
- 114. Ho, M.-H., Kuo, P.-Y., Hsieh, H.-J., Hsien, T.-Y., Hou, L.-T., Lai, J.-Y. and Wang, D.-M. Preparation of porous scaffolds by using freeze-extraction and freeze-gelation methods. *Biomaterials*. 2004. 25(1): 129-138.
- Mu, C., Su, Y., Sun, M., Chen, W. and Jiang, Z. Fabrication of microporous membranes by a feasible freeze method. *Journal of Membrane Science*. 2010. 361(1-2): 15-21.
- 116. Wu, L. and Ding, J. In vitro degradation of three-dimensional porous poly(d,l-lactide-co-glycolide) scaffolds for tissue engineering. *Biomaterials*. 2004. 25(27): 5821-5830.
- 117. Orava, E., Korventausta, J., Rosenberg, M., Jokinen, M. and Rosling, A. In vitro degradation of porous poly(dl-lactide-co-glycolide) (PLGA)/bioactive glass composite foams with a polar structure. *Polymer Degradation and Stability*. 2007. 92(1): 14-23.
- Deb, S., Braden, M. and Bonfield, W. Water absorption characteristics of modified hydroxyapatite bone cements. *Biomaterials*. 1995. 16: 1095-1100.
- Tang, C.Y., Chen, D.Z., Yue, T.M., Chan, K.C., Tsui, C.P. and Yu, P.H.F. Water absorption and solubility of PHBHV/HA nanocomposites. *Composites Science and Technology*. 2008. 68(7-8): 1927-1934.

- 120. Yang, Y., Zhao, Y., Tang, G., Li, H., Yuan, X. and Fan, Y. In vitro degradation of porous poly(l-lactide-co-glycolide)/β-tricalcium phosphate (PLGA/β-TCP) scaffolds under dynamic and static conditions. *Polymer Degradation and Stability*. 2008. 93(10): 1838-1845.
- 121. Dorati, R., Colonna, C., Genta, I., Modena, T. and Conti, B. Effect of porogen on the physico-chemical properties and degradation performance of PLGA scaffolds. *Polymer Degradation and Stability*. 2010. 95(4): 694-701.
- 122. Fredenberg, S., Wahlgren, M., Reslow, M. and Axelsson, A. The mechanisms of drug release in poly(lactic-co-glycolic acid)-based drug delivery systems—A review. *International Journal of Pharmaceutics*. 2011. 415(1-2): 34-52.
- 123. Stevanović, M., Bračko, I., Milenković, M., Filipović, N., Nunić, J., Filipič, M. and Uskoković, D.P. Multifunctional PLGA particles containing poly(l-glutamic acid)-capped silver nanoparticles and ascorbic acid with simultaneous antioxidative and prolonged antimicrobial activity. *Acta Biomaterialia*. 2014. 10(1): 151-162.
- 124. Said, S.S., Aloufy, A.K., El-Halfawy, O.M., Boraei, N.A. and El-Khordagui, L.K. Antimicrobial PLGA ultrafine fibers: Interaction with wound bacteria. *European Journal of Pharmaceutics and Biopharmaceutics*. 2011. 79(1): 108-118.
- 125. Woodruff, M.A. and Hutmacher, D.W. The return of a forgotten polymer— Polycaprolactone in the 21st century. *Progress in Polymer Science*. 2010. 35(10): 1217-1256.
- 126. Ma, D. and McHugh, A.J. The interplay of membrane formation and drug release in solution-cast films of polylactide polymers. *International Journal* of Pharmaceutics. 2010. 388(1-2): 1-12.
- 127. Machín, R., Isasi, J.R. and Vélaz, I. Hydrogel matrices containing single and mixed natural cyclodextrins. Mechanisms of drug release. *European Polymer Journal*. 2013. 49(12): 3912-3920.
- 128. Wei, Z., Wang, C., Liu, H., Zou, S. and Tong, Z. Facile fabrication of biocompatible PLGA drug-carrying microspheres by O/W pickering emulsions. *Colloids and Surfaces B: Biointerfaces*. 2012. 91: 97-105.

- 129. Lao, L.L., Peppas, N.A., Boey, F.Y.C. and Venkatraman, S.S. Modeling of drug release from bulk-degrading polymers. *International Journal of Pharmaceutics*. 2011. 418(1): 28-41.
- Peppas, N.A. and Narasimhan, B. Mathematical models in drug delivery: How modeling has shaped the way we design new drug delivery systems. *Journal of Controlled Release*. 2014. 190: 75-81.
- Chou, A.H.K., LeGeros, R.Z., Chen, Z. and Li, Y. Antibacterial Effect of Zinc Phosphate Mineralized Guided Bone Regeneration Membranes. *Implant Dentistry*. 2007. 16(1): 89-100.
- Nerem, R. Principles of tissue engineering. In: R.L. RP Lanza, J Vancanti. ed. *The Challenge of Imitating Nature*. San Diego: Academic Press. 9-11; 2000.
- Zhang, M. Biomaterials and Tissue engineering. In: D. Shi. ed. Biocompatibility of materials. Heidelberg: Springer. 83-143; 2004.
- 134. Franks, K., Salih, V., Knowles, J. and Olsen, I. The effect of MgO on the solubility behavior and cell proliferation in a quaternary soluble phosphate based glass system. *Journal of Materials Science: Materials in Medicine*. 2002. 13: 549-556.
- Marques, A., Reis, R. and Hunt, J. The biocompatibility of novel starchbased polymers and composites: in vitro studies. *Biomaterials*. 2002. 23: 1471-1478.
- 136. Renouf-Glauser, A.C., Rose, J., Farrar, D.F. and Cameron, R.E. Comparison of the hydrolytic degradation and deformation properties of a PLLA-lauric acid based family of biomaterials. *Biomacromolecules*. 2006. 7(2): 612-617.
- 137. Li, Z.Y., Lam, W.M., Yang, C., Xu, B., Ni, G.X., Abbah, S.A., Cheung, K.M.C., Luk, K.D.K. and Lu, W.W. Chemical composition, crystal size and lattice structural changes after incorporation of strontium into biomimetic apatite. *Biomaterials*. 2007. 28(7): 1452-1460.
- 138. Lima, I.R.D., Alves, G.G., Soriano, C.A., Campaneli, A.P., Gasparoto, T.H., Schnaider, E., Junior, R. and Sena, D. Understanding the impact of divalent cation substitution on hydroxyapatite: An in vitro multiparametric study on biocompatibility. *Journal of Biomedical Materials Research Part A*. 2011. 351-358.
- 139. Bottino, M.C., Coelho, P.G., Henriques, V.A.R., Higa, O.Z., Bressiani, A.H.A. and Bressiani, J.C. Processing, characterization, and in vitro/in vivo

evaluations of powder metallurgy processed Ti-13Nb-13Zr alloys. *Journal of Biomedical Materials Research Part A*. 2009. 88(3): 689-696.

- 140. Stevanović, M.M., Škapin, S.D., Bračko, I., Milenković, M., Petković, J., Filipič, M. and Uskoković, D.P. Poly(lactide-co-glycolide)/silver nanoparticles: Synthesis, characterization, antimicrobial activity, cytotoxicity assessment and ROS-inducing potential. *Polymer*. 2012. 53(14): 2818-2828.
- 141. Tripathi, G., Gough, J.E., Dinda, A. and Basu, B. In vitro cytotoxicity and in vivo osseointergration properties of compression-molded HDPE-HA-Al₂O₃ hybrid biocomposites. *J Biomed Mater Res Part A*. 2013. 101A: 1539-1549.
- 142. Bonnier, F., Keating, M.E., Wróbel, T.P., Majzner, K., Baranska, M., Garciamunoz, A. and Blanco, A. Toxicology in Vitro Cell viability assessment using the Alamar blue assay: A comparison of 2D and 3D cell culture models. *Toxicology Invitro*. 2015. 29(1): 124-131.
- 143. Akiko Watabe, Y., Yamaguchi, T., Kawanishi, T. and Uchida, E. Target-cell specificity of fusogenic liposomes: Membrane fusion-mediated macromolecule delivery into human blood mononuclear cells. *Biochimica et Biophysica Acta*. 1999. 1416: 339-348.
- International Standard Organisation. *Tests for cytotoxicity: in vitro methods*. Switzerland, ISO-10993-5. 2009.
- 145. Coecke, S., Balls, M., Bowe, G., Davis, J., Cstraunthaler, G., Hartung, T., Hay, R., Merten, O., Price, A., Schectman, L., Stacey, G. and Stokes, W. Guidance on Good Cell Culture Practice, A Report of the Second ECVAM Task Force on Good Cell Culture Practice, *ATLA*. 2005. 33: 261-287.
- 146. Fentem, J., Curren, R., Liebsch, M. Guidance Document on Using In vitro Data to Estimate In vivo Starting Doses for Acute Toxicity. NIH Publication No. 01-4500. August 2001.
- 147. American Type Culture Collection (ATCC). American Type Culture Collection Animal Cell Culture Guide (tips and techniques for continuous cell lines). U.S. 2014.
- 148. Cadena-herrera, D., Lara, J.E.E.-d., Ramírez-ibañez, N.D., López-morales, C.A., Pérez, N.O., Flores-ortiz, L.F. and Medina-rivero, E. Validation of three viable-cell counting methods: Manual, semi-automated, and automated. *Biotechnology Reports*. 2015. 7: 9-16.

- 149. K.S. Louis, A.C.S., G.A. Levy. Comparison of manual versus automated trypan blue dye exclusion method for cell counting. In: Stoddart, M.J. ed. *Mammalian cell viability: Methods and Protocols, Methods in Molecular Biology*, New York: Springer Protocols. 7-12; 2015.
- 150. International Standard Organisation. Biological Evaluation of MedicalDevices—Part 12: Sample Preparation and Reference Materials. Switzerland, ISO-10993-12. 2002.
- 151. Dygai, A.M., Ogorodova, L.M., Psakhie, S.G., Belsky, Y.P., Belska, N.V., Danilets, M.G., Ligatcheva, A.A. and Churin, A.A. A study of the Cytotoxicity of a New Nonwoven Polymeric Fibrous Bandaging Material in Vitro. *Journal of Biomaterials and Nanobiotechnology*. 2011. 2(July): 234-238.
- 152. Tavares, D.D.S., Castro, L.D.O., Soares, G.D.D.A., Alves, G.G. and Granjeiro, J.M. Synthesis and cytotoxicity evaluation of granular magnesium substituted β-tricalcium phosphate. *Journal of Applied Oral Science*. 2013. 21(1): 37-42.
- 153. Tomida, M., Nakano, K., Matsuura, S. and Kawakami, T. Comparative examination of subcutaneous tissue reaction to high molecular materials in medical use. *European Journal of Medical Research*. 2011. 16: 249-252.
- 154. Dimitrievska, S., Petit, A., Ajji, A., Bureau, M.N., Yahia, L.H. and Montre,
 P.D. Biocompatibility of novel polymer-apatite nanocomposite fibers. *Journal of Biomedical Materials Research Part A*. 2008. 84A(1): 44-53.
- 155. Wang, B.L.-s., Chow, P.-y., Phan, T.-t., Lim, I.J. and Yang, Y.-y. Fabrication and Characterization of Nanostructured and Thermosensitive Polymer Membranes for Wound Healing and Cell Grafting. *Adv. Funct. Mater.* 2006. 16: 1171-1178.
- 156. Xue, M., Hu, H., Jiang, Y., Liu, J., He, H. and Ye, X. Biodegradable polymer-coated, gelatin hydrogel/bioceramics ternary composites for antitubercular drug delivery and tissue regeneration. *Journal of Nanomaterials*. 2012. (Volume 2012): Article ID 530978.
- 157. Fisher, Material Safety Data Sheet of Dimethyl Sulfoxide. UK: 2012.
- 158. American Society for Testing and Materials. Standard Test Method for in vitro Degradation Testing of Hydrolytically Degradable Polymer Resins and Fabricated Forms for Surgical Implants. U.S., F1635-11. 2011.

- 159. Durect LACTEL Absorbable Polymers. *Material Safety Data Sheet of Poly(lactic-co-glycolic) acid.* U.S.:2012.
- Loo, S.C.J., Tan, Z.Y.S., Chow, Y.J. and Lin, S.L.I. Drug Release From Irradiated PLGA and PLLA Multi-Layered Films. *Journal of pharmaceutical sciences*. 2010. 99(7): 3060-3071.
- 161. Mehta, A., Oeser, a.M. and Carlson, M.G. Rapid quantitation of free fatty acids in human plasma by high-performance liquid chromatography. *Journal of chromatography. B, Biomedical sciences and applications*. 1998. 719(1-2): 9-23.
- 162. Jaidev, L.R., Krishnan, U.M. and Sethuraman, S. Gemcitabine loaded biodegradable PLGA nanospheres for in vitro pancreatic cancer therapy. *Materials Science and Engineering: C.* 2015. 47: 40-47.
- 163. Simchi, A., Tamjid, E., Pishbin, F. and Boccaccini, A.R. Recent progress in inorganic and composite coatings with bactericidal capability for orthopaedic applications. *Nanomedicine: Nanotechnology, Biology, and Medicine*. 2011. 7(1): 22-39.
- 164. Raphel, J., Holodniy, M., Goodman, S.B. and Heilshorn, S.C. Multifunctional Coatings to Simultaneously Promote Osseointegration and Prevent Infection of Orthopaedic Implants. *Biomaterials*. 2016. 84: 301-314.
- 165. Hoben, H.J. and Somasegaran, P. Comparison of the Pour, Spread and Drop plate methods for enumeration of Rhizobium spp. in inoculants made from presterilized peat. *Applied and Environmental Microbiology*. 1982. 44(5): 1246-1247.
- Miles. A. A. and Misra, S.S. The estimation of the bactericidal power of blood. *Journal of Hygiene*. 1938. 38: 732-749.
- 167. American Society for Testing and Materials. Determining the activity of Incorporated Antimicrobial Agent(s) In Polymeric or Hydrophobic Materials. U.S., E2180-07. 2012.
- 168. Braghirolli, D.I., Steffens, D., Quintiliano, K., Acasigua, G.A.X., Gamba, D., Fleck, R.A., Petzhold, C.L. and Pranke, P. The effect of sterilization methods on electronspun poly(lactide-co-glycolide) and subsequent adhesion efficiency of mesenchymal stem cells. *Journal of Biomedical Materials Research - Part B Applied Biomaterials*. 2014. 102(4).

- 169. Cañadas, C., Alvarado, H., Calpena, A.C., Silva, A.M., Souto, E.B., García, M.L. and Abrego, G. In vitro, ex vivo and in vivo characterization of PLGA nanoparticles loading pranoprofen for ocular administration. *International Journal of Pharmaceutics*. 2016. 511: 719-727.
- 170. Rameshbabu, N., Sampath Kumar, T.S., Prabhakar, T.G., Sastry, V.S., Murty, K.V.G.K. and Prasad Rao, K. Antibacterial nanosized silver substituted hydroxyapatite: Synthesis and characterization. *Journal of Biomedical Materials Research Part A*. 2007. 80A(3): 581-591.
- 171. Panda, R.N., Hsieh, M.F., Chung, R.J. and Chin, T.S. FTIR, XRD, SEM and solid state NMR investigations of synthesized by hydroxide-gel technique. *Journal of Physics and Chemistry of Solids*. 2003. 64: 193-199.
- 172. Bogdanoviciene, I., Beganskiene, A., Tõnsuaadu, K., Glaser, J., Meyer, H.J. and Kareiva, A. Calcium hydroxyapatite, Ca10(PO4)6(OH)2 ceramics prepared by aqueous sol-gel processing. *Materials Research Bulletin*. 2006. 41(9): 1754-1762.
- Costescu, A., Pasuk, I., Ungureanu, F., Dinischiotu, A., Huneau, F., Galaup, S., Coustumer, P.L.E., Predoi, D. and Ftir, C. Physico-chemical properties of nano-sized hexagonal hydroxyapatite powder synthesised by sol-gel. *Journal* of Nanomaterials. 2010. 5(4): 989-1000.
- 174. Landi, E., Tampieri, A., Celotti, G., Vichi, L. and Sandri, M. Influence of synthesis and sintering parameters on the characteristics of carbonate apatite. *Biomaterials*. 2004. 25(10): 1763-1770.
- 175. Greish, Y.E., Sturgeon, J.L., Singh, A., Krogman, N.R., Touny, A.H., Sethuraman, S., Nair, L.S., Laurencin, C.T., Allcock, H.R. and Brown, P.W. Formation and properties of composites comprised of calcium-deficient hydroxyapatites and ethyl alanate polyphosphazenes. *Journal of Materials Science: Materials in Medicine*. 2008. 19(9): 3153-3160.
- 176. Gustavsson, J., Ginebra, M.P., Engel, E. and Planell, J. Ion reactivity of calcium-deficient hydroxyapatite in standard cell culture media. *Acta Biomaterialia*. 2011. 7(12): 4242-4252.
- 177. Koutsopoulos, S. Synthesis and characterization of hydroxyapatite crystals: a review study on the analytical methods. *Journal of Biomedical Materials Research Part A.* 2002. 62: 600-612.

- 178. Gibson, I.R., Ke, S., Best, S.M. and Bonfield, W. Effect of powder characteristics on the sinterability of hydroxyapatite powders. *Journal of materials science. Materials in medicine.* 2001. 12(2): 163-171.
- 179. Suetsugu, Y., Takahashi, Y., Okamura, F.P. and Tanaka, J. Structure analysis of A-type carbonate apatite by a single-crystal x-ray diffraction method. *Journal of Solid State Chemistry*. 2000. 155: 292-297.
- 180. El Feki, H., Savariault, J.M., Ben Salah, A. Structure refinements by the Rietveld method of partially substituted hydroxyapatite: Ca₉Na_{0.5}(PO₄)_{4.5}(CO₃)_{1.5}(OH)₂. Journal of Alloys and Compounds. 1999. 287: 114-120.
- Gibson, I.R. and Bonfield, W. Novel synthesis and characterization of an AB-type carbonate-substituted hydroxyapatite. *Journal of Biomedical Materials Research*. 2002. 59(4): 697-708.
- 182. Joschek, S., Nies, B., Krotz, R. and Göferich, A. Chemical and physicochemical characterization of porous hydroxyapatite ceramics made of natural bone. *Biomaterials*. 2000. 21(16): 1645-58.
- 183. Fu, X., Liu, Z., Xiao, Y., Wang, J. and Lei, J. Preparation and properties of lauric acid/diatomite composites as novel form-stable phase change materials for thermal energy storage. *Energy and Buildings*. 2015. 104: 244-249.
- 184. Hao, S., Wang, Y., Wang, B., Deng, J., Zhu, L. and Cao, Y. Formulation of porous poly(lactic-co-glycolic acid) microparticles by electrospray deposition method for controlled drug release. *Materials science & engineering. C, Materials for biological applications.* 2014. 39: 113-119.
- 185. Wei, J. and Li, Y. Tissue engineering scaffold material of nano-apatite crystals and polyamide composite. *European Polymer Journal*. 2004. 40(3): 509-515.
- 186. Jose, M., Thomas, V., Johnson, K., Dean, D. and Nyairo, E. Aligned PLGA/HA nanofibrous nanocomposite scaffolds for bone tissue engineering. *Acta Biomaterialia*. 2009. 5(1): 305-315.
- 187. Tanaka, H., Watanabe, T., Chikazawa, M., Kandori, K. and Ishikawa, T. TPD, FTIR, and Molecular Adsorption Studies of Calcium Hydroxyapatite Surface Modified with Hexanoic and Decanoic Acids. *Journal of Colloid and Interface Science*. 1999. 214: 31-37.

- 188. Zhou, S., Zheng, X., Yu, X., Wang, J., Weng, J., Li, X., Feng, B. and Yin, M. Hydrogen Bonding Interaction of Poly (D, L-Lactide)/ hydroxyapatite Nanocomposites. *Chemical Materials*. 2007. 19(20): 247-253.
- Ma, S., Chen, Z., Qiao, F., Sun, Y., Yang, X., Deng, X., Cen, L., Cai, Q., Wu, M., Zhang, X. and Gao, P. Guided bone regeneration with tripolyphosphate cross-linked asymmetric chitosan membrane. *Journal of Dentistry*. 2014. 42(12): 1603-1612.
- 190. Cao, N., Yang, X. and Fu, Y. Effects of various plasticizers on mechanical and water vapor barrier properties of gelatin films. *Food Hydrocolloids*. 2009. 23(3): 729-735.
- 191. Tarvainen, M., Sutinen, R., Peltonen, S., Mikkonen, H., Maunus, J., Vähä-Heikkilä, K., Lehto, V.-P. and Paronen, P. Enhanced film-forming properties for ethyl cellulose and starch acetate using n-alkenyl succinic anhydrides as novel plasticizers. *European Journal of Pharmaceutical Sciences*. 2003. 19(5): 363-371.
- 192. Vieira, M.G.A., da Silva, M.A., dos Santos, L.O. and Beppu, M.M. Naturalbased plasticizers and biopolymer films: A review. *European Polymer Journal.* 2011. 47(3): 254-263.
- 193. Duan, B., Wu, L., Yuan, X., Hu, Z., Li, X., Zhang, Y., Yao, K. and Wang, M. Hybrid nanofibrous membranes of PLGA/chitosan fabricated via an electrospinning array. *Journal of Biomedical Materials Research Part A*. 2007. 83A(3): 868-878.
- 194. Ebrahimian-Hosseinabadi, M., Ashrafizadeh, F., Etemadifar, M. and Venkatraman, S.S. Preparation and mechanical behavior of PLGA/nano-BCP composite scaffolds during in-vitro degradation for bone tissue engineering. *Polymer Degradation and Stability*. 2011. 96(10): 1940-1946.
- 195. Lu, L., Peter, S.J., Lyman, M.D., Lai, H.-l., Leite, S.M., Tamada, J.A., Uyama, S., Vacanti, J.P., Langer, R. and Mikos, A.G. In vitro and in vivo degradation of porous poly (DL-lactic-*co*-glycolic acid) foams. *Biomaterials*. 2000. 21: 1837-1845.
- Ramchandani, M., Pankaskie, M. and Robinson, D. The influence of manufacturing procedure on the degradation of poly(lactide-co-glycolide) 85:15 and 50:50 implants. *Journal of Controlled Release*. 1997. 43: 161-173.

- 197. Tarola, A.M., Girelli, A.M. and Lorusso, S. High Performance Liquid Chromatography Determination of Fatty Acids in Drying Oils Following Lipase Action. *Journal of Chromatographic Science*. 2012. 50(4): 294-300.
- 198. Ding, A.G. and Schwendeman, S.P. Determination of water-soluble acid distribution in poly(lactide-co-glycolide). *Journal of pharmaceutical sciences*. 2004. 93(2): 322-331.
- 199. Peppas, N.A. and Khare, A.R. Preparation, structure and diffusional behavior of hydrogels in controlled release. Advanced Drug Delivery Reviews. 1993. 11: 1-35.
- 200. Pastorino, D., Canal, C. and Ginebra, M.-P. Drug delivery from injectable calcium phosphate foams by tailoring the macroporosity–drug interaction. *Acta Biomaterialia*. 2015. 12: 250-259.
- Zuleger, S. and Lippold, B.C. Polymer particle erosion controlling drug release . I . Factors influencing drug release and characterization of the release mechanism. *International Journal of Pharmaceutics*. 2001. 217: 139-152.
- 202. Matsuo, M., Oogai, Y., Kato, F., Sugai, M. and Komatsuzawa, H. Growthphase dependence of susceptibility to antimicrobial peptides in Staphylococcus aureus. *Microbiology*. 2011. 157(6): 1786-1797.
- 203. Farokhi, M., Mottaghitalab, F., Shokrgozar, M.A., Ou, K.-L., Mao, C. and Hosseinkhani, H. Importance of dual delivery systems for bone tissue engineering. *Journal of Controlled Release*. 2016. 225: 152-169.
- 204. Kirkham, L.-a.S., Corscadden, K.J., Wiertsema, S.P., Currie, A.J. and Richmond, P.C. A practical method for preparation of pneumococcal and nontypeable Haemophilus influenzae inocula that preserves viability and immunostimulatory activity. *BMC Research Notes*. 2013. 6: 522.
- 205. Cells, H., Harris, L.G., Foster, S.J., Richards, R.G. and Harris, L. An introduction to staphylococcus aureus, and techniques for identifying and quantifying s.aureus adhesins in relation to adhesin to biomaterials: Review. *European Cells and Materials.* 2002. 4: 39-60.
- 206. Udekwu, K.I., Parrish, N., Ankomah, P., Baquero, F. and Levin, B.R. Functional relationship between bacterial cell density and the efficacy of antibiotics. *Journal of Antimicrobial Chemotherapy*. 2009. 63(4): 745-757.

- Dayrit, F.M. The Properties of Lauric Acid and Their Significance in Coconut Oil. *Journal of the American Oil Chemists' Society*. 2014. 92(1): 1-15.
- 208. Fischer, C.L., Drake, D.R., Dawson, D.V., Blanchette, D.R., Brogden, K.A. and Wertz, P.W. Antibacterial Activity of Sphingoid Bases and Fatty Acids against Gram-Positive and Gram-Negative Bacteria. *Antimicrobial Agents* and Chemotherapy. 2011. 56(3): 1157-1161.
- 209. Umerska, A., Cassisa, V., Matougui, N., Joly-Guillou, M.-L., Eveillard, M. and Saulnier, P. Antibacterial action of lipid nanocapsules containing fatty acids or monoglycerides as co-surfactants. *European Journal of Pharmaceutics and Biopharmaceutics*. 2016. 108: 100-110.