Journal of Scientific Agriculture 2017: 08-18 doi: 10.25081/jsa.2017.v1i0.23 http://updatepublishing.com/journals/index.php/jsa



#### **REGULAR ARTICLE**

# Salt Stress induced changes in growth, pigments and protein contents in two horse gram [*Macrotyloma uniflorum* (Lam.) Verdc] varieties

#### G. Kanagaraj\* and C. Sathish

PG and Research Department of Botany, Government Arts College for Men, Krishnagiri- 635001, Tamil Nadu, India.

#### Abstract

Growth parameters and photosynthetic pigments changes in horse gram were investigated under salinity of different concentrations (0, 40, 80. 120mM). The two horse gram varieties Paiyur-2 and CO-1 were sampling was done in young and fully matured leaves were taken from control and salinity treated plants on 15<sup>th</sup> Days After Treatment (DAT) and 30<sup>th</sup> DAT. Treatments were planted in pots. Growth parameters such as plant height, lea area, fresh and dry weight of the whole plants decreased in both varieties under salinity stressed condition. Photosynthetic pigments such as total chlorophyll, chlorophyll 'a' chlorophyll 'b' were significantly reduced in the salinity stressed leaves. Quantitative differences with response to salinity. Were also noticed in the content of soluble protein in two horse gram varieties. Our data revealed that Paiyur-2 maintained lower reduction of growth and higher contents of photosynthetic pigments as well as soluble protein content when compared to variety CO-1 during the adverse effect of salinity stress.

Key words: NaCl, Photosynthetic pigments, Growth, Protein, Horse gram

#### Introduction

Salinity is one of the major abiotic stresses in arid and semi-arid regions but salt-affected soils have been recorded in practically all the climatic regions where more than 800 million hectares of agricultural and or over 6% of the world surfaces are salt affected. Sodium chloride is the most soluble, pervasive, and superabundant salt in the world (FAO, 2000; Munns and Tester, 2008). Rapid population and subsequent food growth shortage especially in Asia and Africa and advancing salinity in arable land due to climate change have increased the importance of finding salt tolerant genotypes (Blumwald et al., 2004). Growth and potential yield are the main factors affected by salinity (Khalida and Da Silva, 2010). Plant growth will be reduced with the onset of salinity due to osmotic stress due

to lowering of the water potential or specific ions interaction on metabolic processes (Munns, 2002). The metabolic imbalances due to ionic toxicity, osmotic stress and nutritional deficiency may lead to oxidative stress (Hasaneen et al., 2009; Baatour et al., 2010). The major plant metabolisms like photosynthesis, protein synthesis, lipid and energy metabolism are being affected by salinity (Parida and Das, 2005; Desingh et al., 2006; Desingh and Kanagaraj, 2007).

Horsegram [*Macrotyloma uniflorum* (Lam.) Verdc] is a popular pulse, locally known as Gaheth belongs to the family Fabaceae that still remain an under exploited legume crop. Horsegram is one among the nutritious pulses and have many medicinal values. In India, horsegram is commonly known as *Kollu* 

Received 09 March 2017; Revised 12 April 2017; Accepted 12 April 2017; Published 13 April 2017

Email: kanagabot15@gmail.com

<sup>\*</sup>Corresponding Author

G. Kanagaraj

PG and Research Department of Botany, Government Arts College for Men, Krishnagiri- 635001, Tamil Nadu, India

<sup>&</sup>lt;sup>©</sup>This article is open access and licensed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/) which permits unrestricted, use, distribution and reproduction in any medium, or format for any purpose, even commercially provided the work is properly cited. Attribution — You must give appropriate credit, provide a link to the license, and indicate if changes were made.

(Tamil). Seeds are rich in protein and consumed in majority by poorest section of the society. It is native to the old world tropics. It was probably domesticated in India where its cultivation known since prehistoric times. It is one of the most important food legume being grown in almost all over the world including temperate and sub-tropical regions (Durga, 2012; Krishna, 2010). In India it is the most extensively grown pulse in south India. Apart from the nutritional components it contains iron, molybdenum and calcium (Bhokre, 2012). Seed contains carbohydrate (57%), protein (22%), dietary fibre (5%), fat (0.50%), calcium (287mg), phosphorous (311mg), iron (6.77mg), calories (321 Kcal) as well as vitamins like thiamine (0.4mg), riboflavin (0.2mg) and niacin (1.5mg) per 100g of dry matter (Gopalan et al., 2000; Bolbhat and Dhumal, 2012; Bhartiya et al., 2015). It also helps in lowering cholesterol levels and has a role as it contains antioxidants. Horse gram is famous for its medicinal uses because different parts of the plant are used for the treatment of heart disease, Asthma, bronchitis urinary discharges and eliminating kidney stones and water is prescribed for treating jaundice (Ghani, 2003).

In India, horse gram has a wide geographic distribution extending over a range of environmental conditions. However, as other crops in India, horse gram is also subjected to environmental stresses, particularly salinity. Although much information is available on the agronomics aspects of horse gram, very little is known about the effects of salinity on physiological and biochemical aspects of horse gram. The present study was undertaken to evaluate the salinity responses of two horse gram varieties [*Macrotuloma* uniflorum (Lam.) Verdc] usually used for cultivation.

### Materials and methods

### Plant material and growth conditions

The certified Horsegram [*Macrotyloma uniflorum* (Lam.) Verdc] seeds (Variety: CO–1, PAIYUR–2) were procured from Tamilnadu Agriculture University, Coimbatore and Paiyur. Seeds with inform size were selected and the plants were raised in pots containing red and clay soil and pH of the soil was 7.2 with EC of 0.2 dsm<sup>-1</sup>. After 20 days, seedlings were thinned and three plants of uniform vigor were maintained in each pot. Plants were grown under natural climatic conditions. The

maximum irradiance (PAR, 400-700nm) available during growth was 1800-2000  $\mu$ mol m<sup>-2</sup>s<sup>-1</sup> on a clear day. Daily maximum and minimum temperatures were 29-33°C and 20-22°C, respectively. Plants were watered for the first 20 days after germination.

### Salinity treatments

The seedlings were divided into four groups. One group of seedlings was maintained under non-salinized conditions which served as control plants. The watering solution for control plants consists of tap water and onestrength of Hoagland nutrients fourth (Hoagland and Arnon, 1950). Other three groups were salinized by irrigation daily to soil capacity (500 ml d<sup>-1</sup>) with the nutrient medium containing 40 mM, 80 mM and 120 mM NaCl. 40mM consider as a low salinity level, 80mM consider as a medium salinity level and 120mM salinity consider as a high salinity level. All the plants used in this study were of comparable size.

Sodium chloride used in this study was Laboratory AR grade Assay 99.8%, (Universal Laboratories Pvt. Ltd. Mumbai). Salt treatment was continued until each plant received the required mM NaCl. Care was taken for individual plants in each group received the pre-calculated concentrations of NaCl in full. Additional pots with plants were also maintained for control, as well as each salinity treatment for need of plant material.

### Sampling days

Young and fully matured leaves were taken from control and salinity treated plants on 15<sup>th</sup> Days After Treatment (DAT) and 30<sup>th</sup> (DAT) for all the experiments described below.

### **Growth components**

### Height of the plant

The height of the plant was measured with a measuring tap on 15<sup>th</sup> DAT and 30<sup>th</sup> DAT.

### Leaf area

The leaf area was calculated multiplying the length and breadth of the broadest regions of the leaf.

Leaf area = length  $\times$  breadth

### Fresh weight of the whole plant

Mature plants were carefully uprooted. The roots were washed, blotted and whole plant was weighed.

## Dry weight of the whole plant

Mature plants were carefully uprooted and the roots were washed, blotted and the whole plant was dried in an oven at 75-80° for 40 hours until a constant weight was obtained.

#### **Photosynthetic pigments**

#### Total chlorophyll

The total chlorophyll content of the leaves was estimated according to Arnon, (1949).

#### **Protein content**

Total leaf protein content was estimated by Lowry's method (1951) using Folin-Ciocalteu reagent.

### Statistical analysis

Data for each parameter analyzed by Two-Way ANOVA and significant differences between treatment mean and varieties were determined by using SPSS (version 15.0, SPSS, Chicago, IL, USA). Data are presented as the mean  $\pm$  SE of five independent determinations and significance was determined at the 95% confidence (P<0.05) limits.

#### **Results and discussion**

### Plant height

Plant height was decreased with increasing salinity levels (40mM, 80mM and 120mM) in all two horsegram varieties on all the sampling days (15<sup>th</sup>DAT (Days After Treatment) and 30<sup>th</sup> DAT) and it was shown in Fig. 1. Maximum plant height was recorded in the variety PAIYUR-2 (32.51 cm) under high salinity (120mM) on 30<sup>th</sup> DAT relative to control plants (60.46 cm respectively) while minimum plant height was recorded in CO-1 (29.13 cm) over the control plants (51.35 cm respectively). Currently, there are about 20 % of the world's cultivated land are affected by salinity (Zhu, 2001). Hence, salt stress is the major hindering factor for crop productivity (Munns, 2002; Perez-Tornero et al., 2009). In the present study, the values of plant height was lowered by increasing salinity and were more pronounced using the highest concentration of NaCl (120 mM) compared to untreated control plants of horsegram varieties (Fig. 1). However, lower reduction in plant height was observed PAIYUR-2 with high salinity on all the sampling days (15<sup>th</sup> DAT and 30<sup>th</sup> DAT), while significantly higher reduction was

recorded in CO-1 on all the sampling days under salinity stress. Reduction in growth under salinity has been reported in various plant species e.g. rice (Demiral and Turkan, 2006), tomato (Kaya et al., 2001), cotton (Kanagaraj and Desingh, 2009), Finger millet (Manikandan and Desingh, 2009a). The reduced plant height exposed to saline medium might be due to the continued effect of decreased shoot and root length, leaf number and leaf area (Maggio et al., 2007; Baatour et al., 2010). Our data on plant height suggest that variety PAIYUR-2 maintained its better height on all the sampling days under varying salinity levels compared to other varieties, indicating substantial salt tolerance.

### Leaf area

Salt stress affects the leaf area by reducing the leaf expansion (Abbruzzese et al., 2009; Wang and Nil, 2000; Cramer, 2003). This reduced leaf area minimizes the light interception and thereby photosynthesis (Parida and Das, 2005). In our study, all of plants varieties horsegram showed reduction in surface area of the leaves on exposure to salinity. Among the two varieties, PAIYUR-2 exhibited lower reduction of leaf area under salinity stress even on 30th DAT relative to control plants, while comparatively higher reduction of leaf area was observed in CO-1 under salinity stress. Cell expansion and division processes are affected by Salinity, which in turn results in the reduction in leaf area (Ticha, 1982; Curtis and Lauchli, 1987) and cell division (Hasegawa et al., 2000). Leaf area was measured in salinity treated and control plants of two horsegram varieties on two sampling days (Fig. 2). On 30<sup>th</sup> DAT, significantly higher reduction of leaf area was measured in the variety CO-1 (6.14 cm<sup>2</sup>) over the control plants (21.58 cm<sup>2</sup> respectively) with 120mM salinity stress, while lower reduction of leaf area was observed in the variety PAIYUR-2 (8.38 cm<sup>2</sup>) compared to control plants (26.23 cm<sup>2</sup> respectively). The results on leaf area clearly indicated that under all salinity levels, PAIYUR-2 recorded higher leaf area in the plants on all the sampling days, which is relative to higher photosynthetic rates.



Figure 1. Effect of varying levels of salinity on plant height of two horsegram varieties on  $15^{\text{th}}$  DAT(a),  $30^{\text{th}}$  DAT(b). Each value represents mean  $\pm$  SE of five independent determinations (p<0.05).



Figure 2. Variation of leaf area of two horse gram varieties on 15<sup>th</sup> DAT(a), 30<sup>th</sup> DAT(b) under varying levels salinity. Each value represents mean ± SE of five independent determinations (p<0.05).

# Fresh weight of the whole plant

The lowest fresh and dry weight of the whole plant was noted in CO-1 under all the levels of salt stress. The main reason for this is the reduced absorption and translocation with reduced biochemical metabolism under salinity (Parida and Das, 2005). Fresh weight of the whole plant was decreased with increasing salinity concentrations on all the sampling days in two horsegram varieties (Fig. 3). On 30<sup>th</sup> DAT, the significantly higher decrease in fresh weight of the whole plant was observed in CO-1 by 38% (8.12gram) with 120mM salinity relative to control plants (20.85gram respectively), while lowest decrease in fresh weight of the whole plant was recorded in PAIYUR-2 by 24% (10.43gram) over the control plants (26.65gram respectively). The present data on biomass revealed that horse gram variety PAIYUR-2 recorded higher biomass even under higher salinity level compared to other varieties which is directly related to growth and yield of the plant.

# Dry weight of the whole plant

The dry mass is mainly affected by salinity, the main reason stands in the reduction of leaf growth and decline in the rates of net photosynthesis and CO<sub>2</sub> assimilation (Sharkey, 1985; Aragao et al., 2005). Reduced growth under salt stress is due to osmotic reduction in availability of water and accumulation of ions (Marschner, 1995). Fig. 4 showed variation of the dry weight of the whole plant under salinity stress and it was decreased with increasing salinity levels on all the sampling days. Maximum reduction of dry weight of the whole plant was recorded in the variety CO-1 and it (3.45gram respectively) compared to was controls (5.73gram respectively) on 30<sup>th</sup> DAT with 120mM salinity, while minimum reduction of dry weight of the whole plant was noticed in the variety PAIYUR-2 and it was 3.86 gram respectively relative to control (6.25 gram respectively). The reduction in plant dry weight can be attributed to the reduced photosynthetic capacity of the leaves under salinity stressed conditions (Sanchez-Rodriguez et al., 1999).

### Total chlorophyll

In the current study, the total chlorophyll content of leaves averaged over two varieties indicated that it decreased significantly with the increase in salt concentration. Effect of salinity on total chlorophyll content was studied in two horsegram varieties and it was decreased with increasing salinity levels on all the sampling days as shown in Fig. 5. On 30<sup>th</sup> DAT highest total chlorophyll content was recorded in the variety PAIYUR-2 (0.68 mg/gfw) over to control plants (1.28 mg/gfw, respectively) under 120mM salinity stress, whereas low total chlorophyll content was observed in the variety CO-1 (0.57 mg/gfw) relative to controls (1.20 mg/gfw, respectively). Varieties differed significantly under salt stress treatments. However, highest total chlorophyll content under salinity stress was recorded in PAIYUR-2 on all the sampling days (15<sup>th</sup> DAT 30<sup>th</sup> DAT) even at high salinity and concentrations, whereas low level of total chlorophyll content was observed in CO-1 under salinity stress. Chlorophyll content is one of the major component of salt tolerance in crop plants (Srivastava et al., 1988). According to Hernandez et al., (1995) the elevation in chlorophyll degradation in salt sensitive pea cultivar was more when compared to tolerant one. NaCl reduces chlorophyll content in crop plants such as broad bean (Gadallah, 1999), cotton (Boyer, 1965) and rice (Sultana et al., 1999). The earlier reports in cucumber (Kaya et al., 2003) and tomato (Agong et al., 2003; Doganlar et al., 2010) supported our results. Chlorophyll reduction is correlated with increased production of chlorophyll-degrading enzyme, chlorophyllase and ion accumulation in leaves (Sultana et al., 1999).

# Chlorophyll 'a' and 'b'

On all the sampling days, Chlorophyll 'a' and 'b' content was decreased with increasing salinity levels in all the horsegram varieties (Figs. 6 & 7). On 30<sup>th</sup> DAT, with 120mM salinity treatment, lowest chl 'a' and chl 'b' content was observed in the variety CO-1 and it was 0.20 mg/gfw and 0.37 mg/gfw, respectively, over the controls (0.43 mg/gfw and 0.77 mg/gfw, respectively), while highest chl 'a' and chl 'b' content was recorded in the variety PAIYUR-2 and it was 0.23 mg/gfw, 0.45 mg/gfw, respectively, compared to control plants (0.45 mg/gfw and 0.83 mg/gfw, respectively). Photosynthetic pigments like chlorophylls 'a' and 'b' have major role in the photosynthetic efficiency (Taiz and Zeiger, 2006). In our study, salinity stress led to a decrease in chlorophyll 'a' and 'b' on all the sampling days (15<sup>th</sup> DAT and 30<sup>th</sup> DAT) and this effect increased consistently with increasing salinity levels as compared to nonstressed treatment. However, higher reduction of chlorophyll 'a' and 'b' was observed in CO-1 and lowest reduction was noticed in the variety PAIYUR-2 on all the sampling days with varying salinity levels. Similar results were reported in tomato (Mohammad et al., 1998), cotton (Meloni et al., 2001) under salinity stress. The present study on the pigment composition clearly showed that the variety PAIYUR-2 maintained high pigment content on all the sampling days than other horse gram varieties when subjected to salt stress.



Figure 3. Influence of varying levels of salinity on fresh weight of the whole plant of horse gram varieties on  $15^{\text{th}}$  DAT (a),  $30^{\text{th}}$  DAT (b). Each value represents mean  $\pm$  SE of five independent determinations (p<0.05).



Figure 4. Salinity stress effects on dry weight of the whole plant of horse gram varieties on  $15^{\text{th}}$  DAT (a),  $30^{\text{th}}$  DAT (b). Each value represents mean  $\pm$  SE of five independent determinations (p<0.05).

### **Protein content**

In this investigation, when NaCl concentration increased, a soluble protein in two horse gram variety was significantly changed. Comparatively, lower decrease of soluble protein content was observed in the leaves of horse gram variety PAIYUR-2 on all the sampling days even with high salinity levels as compared to controls. More decrease of soluble protein content was observed in the variety CO-1, on all the sampling days compared to control plants. High temperature, can salinity and drought stress cause denaturation and dysfunction of many proteins (Vinocur and Altman, 2005). Salinity treatment caused a depletion of protein from, shoot and root tissues of lentil (Misra and saxena, 2009) and Phaseolus aureus (Misra and Dwived, 1990). Protein content was decreased in leaves of all the two horse gram varieties with increasing salinity level on all the sampling days (Fig. 8). Under 120mM salinity stress, on 30<sup>th</sup> DAT, protein content was highly decreased in CO-1 by 50% (29.73 mg/gfw) over control plants the (58.57)mg/gfw, respectively), while low decrease of protein content was recorded in the variety Paiyur-2 by 26% (32.68 mg/gfw) relative to control plants (60.51 mg/gfw, respectively). The decrease in soluble proteins in leaves by salinity indicated that salinity might have promoted hydrolysis of protein resulting in an accumulation of proline particularly at high concentration of NaCl and or inhibited protein synthesis. The results in this study clearly indicates that even though the protein synthetic machinery of the horse gram varieties are affected by the salinity stress, variety Paiyur-2 maintained higher protein content on all the sampling days compared to other varieties. There will be difference in the response of plant species to salinity and even different cultivars within the same species (Munns, 2002; Borsani et al., 2003). Under saline conditions, there is a change in the pattern of gene expression and both qualitative and quantitative changes in protein synthesis (Amini and Ehsanpour, 2005; Xu et al., 2010).

#### Acknowledgements

The authors are highly thankful to Dr. P. Ravikumar, Professor and Head, for providing necessary facilities.

### **Author contributions**

All authors contributed equally in the study and preparation of article. All authors approved the final version of the manuscript for publication.



Figure 5. Changes of total chlorophyll content in leaves of horse gram varieties on 15<sup>th</sup> DAT (a), 30<sup>th</sup> DAT (b) under varying levels of salinity.

Each value represents mean  $\pm$  SE of five independent determinations (p<0.05).



Figure 6. Influence of varying levels of salinity on chlorophyll 'a' content in leaves of horse gram varieties on 15<sup>th</sup> DAT (a), 30<sup>th</sup> DAT (b). Each value represents mean  $\pm$  SE of five independent determinations (p<0.05).



Figure 7. Effect of varying levels of salinity on chlorophyll 'b' content in leaves of horse gram varieties on 15<sup>th</sup> DAT (a) 30<sup>th</sup> DAT (b). Each value represents mean  $\pm$  SE of five independent determinations (p<0.05).



Figure 8. Effect of different salinity levels on soluble protein

content in the leaf extracts of horse gram varieties on  $15^{\text{th}}$  DAT (a),  $30^{\text{th}}$  DAT (b). Each value represents mean  $\pm$  SE of five independent determinations (p<0.05).

## References

- Abbruzzese, G., Beritognolo, I., Muleo, R., Piazzai, M., Sabatti, M., Mugnozza, G. S., & Kuzminsky, E. (2009). Leaf morphological plasticity and stomatal conductance in three *Populus alba* L. genotypes subjected to salt stress. *Environmental and experimental Botany*, 66(3), 381-388.
- Agong, S. G., Kingetsu, M., Yoshida, Y., Yazawa, S., & Masuda, M. (2003).
  Response of tomato genotypes to induced salt stress. African Crop Science Journal, 11, 1-9.
- Amini, F., & Ehsanpour, A. A. (2005). Soluble proline, carbohydrates proteins, and Na+/K+changes in two tomato (Lycopersicon esculentum Mill.) cultivars vitro salt under in stress. American Journal of Biochemistry and Biotechnology, 1(4), 204-208.
- Aragão, M. E. F. D., Guedes, M. M., Otoch, M. D. L. O., Guedes, M. I. F., Melo, D. F. D., & Lima, M. D. G. S. (2005). Differential responses of ribulose-1, 5-bisphosphate carboxylase/oxygenase activities of two Vigna unguiculata cultivars to salt stress. Brazilian Journal of Plant Physiology, 17(2), 207-212.
- Arnon, D. I. (1949). Copper enzymes in isolated chloroplasts. Polyphenoloxidase in Beta vulgaris. *Plant physiology*, 24(1), 1-15.
- Baatour, O., Kaddour, R., Wannes, W. A., Lachaâl, M., & Marzouk, B. (2010). Salt effects on the growth, mineral nutrition, essential oil yield and composition of marjoram (*Origanum majorana*). Acta physiologiae plantarum, 32(1), 45 -51.
- Bejaoui, M. (1985). Intéractions entre NaCl et quelques Phytohormones sur la croissance du Soja: Interactions Between NaCl and Some Phytohormones on Soybean Growth. Journal of Plant Physiology, 120(2), 95-110.
- Bhartiya, A., Aditya, J. P., & Kant, L. (2015).
  Nutritional and remedial potential of an underutilized food legume horse gram (*Macrotyloma uniflorum*): A review. *Journal of Animal and Plant Sciences*, 25(4), 908-920.
- Bhatt, M. J., Patel, A. D., Bhatti, P. M., & Pandey, A. N. (2008). Effect of soil salinity on growth, water status and nutrient accumulation in seedlings of *Ziziphus*

mauritiana (Rhamnaceae). Fruit and Ornamental Plant Research, 16, 383-401.

- Bhokre, C., Ghatge, P. U., Machewad, G., & Rodge, A. (2012). Studies on preparation of buns fortified with germinated horse gram flour. *Open Access Scien. Rep*, *2*, 127.
- Blumwald, E., Grover, A., & Good, A. G. (2004, September). Breeding for abiotic stress resistance: challenges and opportunities. In New directions for a diverse planet". Proceedings of the 4th International Crop Science Congress, Brisbane, Australia.
- Borsani, O., Valpuesta, V., & Botella, M. A. (2003). Developing salt tolerant plants in a new century: a molecular biology approach. *Plant Cell, Tissue and Organ Culture*, 73(2), 101-115.
- Boyer, J. S. (1965). Effects of osmotic water stress on metabolic rates of cotton plants with open stomata. *Plant Physiology*, 40 (2), 229-234.
- Cramer, G. R. (2003). Differential effects of salinity on leaf elongation kinetics of three grass species. *Plant and Soil*, *253*(1), 233-244.
- Curtis, P. S., & Läuchli, A. (1987). The effect of moderate salt stress on leaf anatomy in *Hibiscus cannabinus* (Kenaf) and its relation to leaf area. *American Journal of Botany*, 538-542.
- Demiral, T., & Türkan, I. (2006). Exogenous glycinebetaine affects growth and proline accumulation and retards senescence in two rice cultivars under NaCl stress. *Environmental and Experimental Botany*, 56(1), 72-79.
- Desingh, R., & Kanagaraj, G. (2007). Influence of salinity stress on photosynthesis and antioxidative systems in two cotton varieties. *General and Applied Plant Physiology*, 33(3-4), 221-234.
- Desingh, R., Jutur, P. P., & Reddy, A. R. (2006). Salinity stress-induced changes in photosynthesis and antioxidative systems in three casuarina species. *Journal of Plant Biology*, *33*, 155-161.
- Doganlar, Z. B., Demir, K., Basak, H., & Gul, I. (2010). Effects of salt stress on pigment and total soluble protein contents of three different tomato cultivars. *African Journal of Agricultural Research*, *5*(15), 2056-2065.

- Durga, K. K. (2012). Variability and divergence in horse gram (Dolichos). *Journal of Arid Land*, 4(1), 71-76.
- FAO, A. (2000). Global Network on Integrated Soil Management for Sustainable Use of Salt affected Soils19-12. http://www.fao.org/ag/agl/agll/spush
- Gadallah, M. A. A. (1999). Effect of proline and glycine-betaine on *Vicia faba* in response to salt stress. *Biologia Plantarum, 42,* 247-249.
- Ghani, A. (1998). *Medicinal plants of Bangladesh: chemical constituents and uses*. Asiatic society of Bangladesh. 460p.
- Gopalan, C., Ramasastri, B. V., & Balasubramanian, S. G. (2000). Nutritive value of Indian foods. Revised and updated by Rao BSN, Deosthale YB, Pant KC.
- Gorham, J., Papa, R., & Aloy-Lleonart, M. (1994). Varietal differences in sodium uptake in barley cultivars exposed to soil salinity or salt spray. *Journal of experimental botany*, *45*(7), 895-901.
- Greenway, H., & Munns, R. (1980). Mechanisms of salt tolerance in nonhalophytes. *Annual review of plant physiology*, *31*(1), 149-190.
- Hoagland, D. R., & Arnon, D. I. (1950). The water-culture method for growing plants without soil. *Circular*. *California Agricultural Experiment Station*, *347* (2nd edit).
- Hasaneen, M. N. A., Younis, M. E., & Tourky, S. M. N. (2009). Plant growth, metabolism and adaptation in relation to stress conditions XXIII. Salinity-biofertility interactive effects on growth, carbohydrates and photosynthetic efficiency of lactuca sativa. *Plant Omics Journal*, 2(2), 60-90.
- Hasegawa, P. M., Bressan, R. A., Zhu, J. K., & Bohnert, H. J. (2000). Plant cellular and molecular responses to high salinity. *Annual Review of Plant Biology* 51(1), 463-499.
- Hernandez, J. A., Olmos, E., Corpas, F. J., Sevilla, F., & Del Rio, L. A. (1995). Saltinduced oxidative stress in chloroplasts of pea plants. *Plant Science*, *105*(2), 151-167.
- Kanagaraj, G. and Desingh, R. (2009). Salinityinduced changes of biochemical constituents and ABA in two cotton varieties. *Geobios*, *36*, 281-284.
- Kanagaraj, G, Manikandan, K. and Desingh, R. (2009). Growth and carbohydrate

metabolism of two cotton varieties under salinity stress. *Plant Archives*, *9*, 413-415.

- Kaya, C., Higgs, D., Ince, F., Amador, B.M., Cakir, A. and Sakar, E. (2003).
  Ameliorative effects of potassium phosphate on salt stressed pepper and cucumber. *Journal of Plant Nutrition 26*, 807-820.
- Kaya, C., Kirnak, H. and Higgs, D. (2001). Enhancement of growth and normal growth parameters by foliar application of potassium and phosphorus in tomato cultivars growth at high (NaCl) salinity. *Journal of Plant Nutrition*, 24, 357-367.
- Khafagy, M. A., Arafa, A. A. and El-Banna, M.
  F. (2009). Glycinebetaine and ascorbic acid can alleviate the harmful effects of NaCl salinity in sweet pepper. *Australian Journal of Crop Science*, *3*, 257-267.
- Krishna, K. R. (2010). Legume agro ecosystems of south India: nutrient dynamics, ecology and productivity. Brown Walker Press, Florida, USA. 372-382 pp.
- Lowry, O. H., Rose Brough, N. J., Farr, A. L. and Randall, R. J. (1951). Protein measurement with the Folin phenol reagent. *Journal of Biological Chemistry*, 193, 265-275.
- Lutts, S., Kinet, J.M. and Bouharmont, T. (1996b). Effect of salt stress on growth, mineral nutrition and proline accumulation in relation to osmotic adjustment in rice (*Oryza sativa* L.) cultivars differing in salinity resistance. *Plant Growth Regulation*, 19, 207-218.
- Maggio, A. and Raimondi, G., Martinno, A. and De Pascale, S. (2007). Salt stress response in tomato beyond the salinity tolerance threshold. *Environmental and Experimental Botany*, 59, 276-282.
- Manikandan, K. and Desingh, R. (2009a). Effect of salt stress on growth, carbohydrate and proline content of two finger millet varieties. *Recent Research in Science and Technology*, *1*, 48-51.
- Marschner, M. (1995). Mineral nutrition of higher plants. 2<sup>nd</sup> ed. London, Academic Press. Limited, p 889.
- Meloni, D.A., Olive, M.A., Ruig, H.R. and Martinez, C.A. (2001). Contributing of proline and inorganic solutes to osmotic adjustment in cotton under salt stress. *Journal of Plant Nutrition*, 24, 599-612.
- Misra, N. and Dwived, U.N. (1990). Nitrogen assimilation in germinating *Phaseolus*

*aureus* seeds under saline stress. *Journal of Plant Physiology* 135, 719-724.

- Misra, N. and Saxena. P. (2009). Effect of salicylic acid on proline metabolism in lentil grown under salinity stress. *Plant Science*, *177*, 181-189.
- Morales, M.A., Sanchez-Blancó, M.J., Olmoss, E., Torrecillas, A. and Alarcon, J.J. (1998). Changes in the growth leaf water relations and cell ultrastructure in *Argyranthemum coronopifoluim* plants under saline conditions. *Journal of Plant Physiology*, 153, 174-180.
- Mohammad, M., Shibli, R., Ajouni, M. and Nimvi, L. (1998). Tomato root and shoot responses to salt stress under different level of phosphorous nutrition. *Journal of Plant Nutrition*, 21, 1667-1680.
- Munns R. (2002). Comparative physiology of salt and water stress. *Plant Cell Environment, 25, 239-250.*
- Munns R., Tester M. (2008). Mechanisms of salinity tolerance. *Annual Review of Plant Biology*, *59*, 651-681.
- Parida A.K. and Das, B. (2005). Salt tolerance and salinity effects on plants. *Ecotoxicology and Environmental Safety*, *60*, 324-349.
- Perez-Tornero, O., Tallon, C., Porras, I. and Navarro, J. (2009). Physiological and growth changes in micropropagated *Citrus macrophylla* explants due to salinity. *Journal of Plant Physiology*, *166*, 1923-1933.
- Ramoliya, P.J., Patel H.M. and Pandey A.N. (2006). Effect of salinization of soil on growth and nutrient accumulation in seedlings of *Prosopis cineraria*. *Journal of Plant Nutrition*, 29, 283-303.
- Rashid, P., Karmoker, J.L., Chakrabortty, S. and Sarker, B.C. (2004). The effect of accumulation and salinity on ion anatomical attributes in mungbean (Phaseolus radiatus L. cv. BARI-3) seedlings. International Journal of Agriculture and Biology, 6, 495-498.
- Rodriguez, H.G., Roberts, J., Jordan, W.R. and Drew, M.C. (1997). Growth, water relations and accumulation of organic and inorganic solutes in roots of maize seedling during salt stress. *Plant Physiology*, *113*, *881*-893.
- Sanchez-Rodriguez, J., Perez, P. and Martinez-Carrasco, R. (1999). Photosynthesis, carbohydrate levels and chlorophyll fluorescence-estimated intercellular CO<sub>2</sub> in

water-stressed *Casuarina equisetifolia* Forst & Forst. *Plant Cell and Environment*, *22*, 867-873.

- Sharkey, T.D. (1985). Photosynthesis in intact leaves of  $C_3$  plants; physics, physiology and rate limitation. *Botanical Reviews*, *51*, 53-105.
- Singh, A.K. and Dubey, R.S. (1995). Changes in chlorophyll a and b contents and activities of photosystem I and II in rice seedlings induced by NaCl. *Photosynthetica*, *31*, 489-499.
- Srivastava, T.P., Gupta, S.C., Lal, P., Muralia, P.N. and Kumar, A. (1988). Effect of salt stress on physiological and biochemical parameters of wheat. *Annals of Arid Zone*, 27, 197-204.
- Sultana, N., Ikeda, T. and Itoh, R. (1999). Effect of NaCl salinity on photosynthesis and dry matter accumulation in developing rice grains. *Environmental and Experimental Botany*, 42, 211-220.
- Taiz, L. and Zeiger, E. (2006). Plant physiology (Fourth Edition). Sinauer Associates, Inc., Publishers, Sunderland, USA. p.764.
- Ticha, I. (1982). Photosynthetic characteristics during ontogenesis of leaves. 7. Stomata density and sizes. *Photosynthetica*, *16*, 375-471.
- Vinocur, B. and Altman, A. (2005). Recent advances in engineering plant tolerance to abiotic stress: achievements and limitations. *Current Opinion in Biotechnology*, *16*, 123-132.
- Wang, Y. and Nil, N. (2000). Changes in chlorophyll, ribulose biphosphate carboxylase-oxygenase, glycine betaine content, photosynthesis and transpiration in *Amaranthus tricolor* leaves during salt stress. *Journal of Horticultural Science and Biotechnology*, *75*, 623-627.
- Xu, C., Sibicky, T. and Huang, B. (2010). Protein profile analysis of salt-responsive proteins in leaves and roots in two cultivars of creeping bentgrass differing in salinity tolerance. *Plant Cell Reports, 29,* 595-615.
- Zhu, J.K. (2001). Plant salt tolerance. *Trends Plant Science*, *6*, 66-72.