# Evaluation of cowhage (Mucuna pruriens L.) genotypes for growth, yield and quality characters in arecanut plantation under hill zone of Karnataka 

Vinay Patil, Sadashiv Nadukeri*, Shashikala Kolakar and K. Hima Bindu ${ }^{1}$<br>College of Horticulture, Mudigere-577 132, Karnataka, India<br>${ }^{1}$ ICAR-Indian Institute of Horticulture Research, Bengaluru- 560089, Karnataka, India

(Manuscript Received: 22-05-2018, Revised: 10-07-2019, Accepted: 15-07-2019)


#### Abstract

Cowhage (Mucuna pruriens L.) is leguminous medicinal plant grown in the tropics. Eight cowhage genotypes were used to study the performance of their growth, yield and quality characters in arecanut plantation under hill zone of Karnataka. Significant differences were recorded in genotypes with respect growth, yield and quality attributes. The genotype Arka Dhanvantari recorded the maximum vine length ( 282.03 cm ) and number of trifoliate leaves (71.03) at harvest. Maximum number of bunches per plant (6.47), stem girth $(0.93 \mathrm{~cm})$, pods per bunch (4.00), number of bunches per plant (4.67) and pod yield per plant ( 136.38 g ) was produced in genotype IIHR Selection-2. Genotype Arka Aswini exhibited maximum pod length ( 11.02 cm ) and pod width ( 1.89 cm ) over other genotypes. Maximum seed yield per plant $(96.13 \mathrm{~g})$, per plot ( 2.88 kg ) and per hectare ( 3384.56 kg ) was recorded in the genotype IIHR Selection-2. Genotype Arka Aswini exhibited maximum 100 seed weight ( 136.23 g ). Maximum L-DOPA content ( 5.17 \%) was recorded in genotype Arka Aswini which was followed by IIHR Selection-2 (4.69 \%). The genotype IIHR Selection-2 recorded maximum L-DOPA yield ( $4.52 \mathrm{~g} \mathrm{plant}^{-1}$ ).


Keywords: Cowhage, genotypes, L-DOPA, Mucuna pruriens, quality and yield

## Introduction

Cowhage (Mucuna pruriens L.) is an important annual twining herb climber, popular leguminous crop not only for its medicinal property but also grown as cover crop in tropical and subtropical areas of the world as it helps in nitrogen fixation. It belongs to the family 'Fabaceae', sub family Papilionaceae. It is commonly known as 'velvet bean', 'cowitch' and 'cowhedge' in English. Cowhage is not only cultivated in India but also grown as a commercial cover crop in rubber, arecanut and coconut plantations in Sri Lanka, Bangladesh, South East Asia and Malaysia. There are about 14 species spread over the foot hills of Himalayas, the plains of West Bengal, Madhya Pradesh, Karnataka, Kerala, Andhra Pradesh, Uttar Pradesh, Andaman and Nicobar Islands in India (Farooqi and Sreeramu, 2001).

Mисипа is known for medicinal properties as its all parts contains important active principles (Caius, 1989). The seeds of cowhage have attracted considerable attention since they are the source for the catecholic amino acid 3-(3, 4-dihydroxyphenyl)-L-alanine, also known as L-DOPA (Bell et al., 1971; Daxenbichler et al., 1972). L-DOPA, a neurotransmitter precursor, is used in the treatment of Parkinson's disease. No systematic information is available regarding the evaluation of available cowhage genotypes and released varieties for commercial cultivation as intercrop in arecanut plantations. Hence, this work was carried out to study the growth, yield and quality of different cowhage genotypes grown as an intercrop in arecanut plantations.

Table 1. Different genotypes of Mucuna pruriens used for the present study

| Sl. No. | Genotype | Colour of the seed | Source |
| :--- | :--- | :--- | :--- |
| 1. | Arka Aswini | Black | ICAR-IIHR, Bengaluru |
| 2. | Arka Dhanvantari | Black | ICAR-IIHR, Bengaluru |
| 3. | IIHR Selection-2 | White | ICAR-IIHR, Bengaluru |
| 4. | IIHR Selection -3 | White | ICAR-IIHR, Bengaluru |
| 5. | IIHR Selection-8 | Black with brown patches | ICAR-IIHR, Bengaluru |
| 6. | IIHR Selection-10 | White | ICAR-IIHR, Bengaluru |
| 7. | Farmer Selection-1 | Black with ash colour patches | Tarikere |
| 8. | Farmer Selection-2 | Black with brown blotches | KVK, Davangere |

## Material and methods

An experiment was designed to know the performance of cowhage (Mucuna pruriens L.) genotypes, grown as an intercrop in arecanut plantation under hilly zone of Karnataka State. The experiment was laid out in randomized complete block design (RCBD) with three replication and eight genotypes at College of Horticulture, Mudigere, during the year 2014-2015. Details of the genotypes used are provided in Table 1.

Each genotype was sown and maintained with spacing of 60 cm x 45 cm between row and plant, respectively. Seed sowing was taken up in the month of November 2014 and irrigation schedule and other cultural practices were followed as per the package of practices. Observations were recorded on five randomly tagged plants from each genotypes of each replication avoiding border plants. Observations were recorded and analyzed statistically for various growth and yield parameters such as number of pods bunch ${ }^{-1}$, number of bunches plant ${ }^{-1}$, pod length ( cm ), pod width (cm), pod yield (g plant ${ }^{-1}$ ), seed yield ( g plant $\mathrm{t}^{-1}$ ), seed yield ( $\mathrm{kg} \mathrm{plot}^{-1}$ ), seed yield ( $\mathrm{kg} \mathrm{ha}^{-1}$ ) and seed weight ( g 100 seeds ${ }^{-1}$ ) and quality parameters such as L-DOPA content (\%) and L-DOPA yield (g plant ${ }^{-1}$ ).

## Determination of L-DOPA content in seeds

The harvested seeds of cowhage were used for the determination of L-DOPA content by Ultra High Performance Liquid Chromatography (UHPLC) at ICAR-IIHR, Bengaluru. L-DOPA was extracted with 0.1 M orthophosphoric acid and estimation was done with Ultra-High Performance

Liquid Chromatography (UHPLC), using $5 \mu \mathrm{~m}$ Phenomenex C18 column, with sodium dihydrogen orthophosphate as mobile phase at pH 2.8. Retention time of sample was between 3.2-3.3 minutes. The percentage of L-DOPA was calculated by comparing the peak area of sample with peak area of the standard according to the formula given below:

L-Dopa $(\%)=\frac{\text { Test area }}{\begin{array}{c}\text { Standard } \\ \text { area }\end{array}} \times \frac{\begin{array}{c}\text { Standard } \\ \text { (L-Dopa) } \\ \text { weight }\end{array}}{\frac{\text { standard }}{\text { dilution }}} \times \frac{\begin{array}{c}\text { Test } \\ \text { solution }\end{array}}{\begin{array}{c}\text { Test } \\ \text { weight }\end{array}} \times \frac{98.5}{100} \times 100$

L-DOPA yield plant ${ }^{-1}$ was calculated by taking the L-DOPA content $(\%)$ and seed yield plant ${ }^{-1}(\mathrm{~g})$.

## Result and Discussion

## Growth parameters

Significant difference was observed in genotypes with respect to growth parameters. Vine length varied significantly among the genotypes at various stages of crop growth like 40 DAS, flowering and harvest stage (Table 2). Genotype Arka Dhanvantari was found to be vigorous in its growth habit at harvest with respect to vine length ( 282.03 cm ) and it was on par with genotypes IIHR Selection-2 ( 271.33 cm ) and IIHR Selection-10 ( 264.40 cm ), whereas, it was minimum in Farmer Selection-1 ( 227.67 cm ) which was less vigorous in growth. The difference in the vine length was mainly due to duration of the crop and genetic factor of the respective genotype as well as influence of the growing environmental conditions. This result is in close conformity with the Vadivel and Janardhan (2000a) in Mucuna pruriens L.,

Table 2. Performance of cowhage (Mucuna pruriens L.) genotypes for vegetative parameters

| Genotype | Vine length (cm) |  |  | Number of trifoliate leaves |  |  | Number of branches at flowering | Stem girth at harvest (cm) |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | 40 DAS | At flowering | At harvest | 40 DAS | At flowering | At harvest |  |  |
| T - Arka | 122.35 | 189.50 | 247.10 | 13.57 | 49.87 | 61.17 | 5.37 | 0.69 |
| Aswini |  |  |  |  |  |  |  |  |
| $\mathrm{T}_{2}$ - Arka | 112.34 | 226.00 | 282.03 | 15.80 | 59.40 | 71.03 | 6.03 | 0.91 |
| Dhanvantari |  |  |  |  |  |  |  |  |
| $\mathrm{T}_{3}$ - IIHR | 102.24 | 217.53 | 271.33 | 14.53 | 55.60 | 68.63 | 6.47 | 0.93 |
| Selection-2 |  |  |  |  |  |  |  |  |
| $\mathrm{T}_{4}$ - IIHR | 106.98 | 195.13 | 256.00 | 13.43 | 47.50 | 60.10 | 5.40 | 0.73 |
| Selection-3 |  |  |  |  |  |  |  |  |
| $\mathrm{T}_{5}$ - IIHR | 95.15 | 194.67 | 249.13 | 13.87 | 48.60 | 60.70 | 5.50 | 0.74 |
| Selection-8 |  |  |  |  |  |  |  |  |
| $\mathrm{T}_{6}$ - IIHR | 118.64 | 207.97 | 264.40 | 14.40 | 53.23 | 63.53 | 6.07 | 0.75 |
| Selection-10 |  |  |  |  |  |  |  |  |
| $\mathrm{T}_{7}$-Farmer | 85.32 | 179.87 | 227.67 | 12.30 | 45.73 | 56.70 | 4.53 | 0.61 |
| Selection-1 |  |  |  |  |  |  |  |  |
| $\mathrm{T}_{8}$-Farmer | 100.50 | 195.03 | 253.00 | 13.03 | 47.13 | 59.90 | 5.03 | 0.65 |
| Selection-2 |  |  |  |  |  |  |  |  |
| S.Em $\pm$ | 7.18 | 6.77 | 8.18 | 0.63 | 1.57 | 2.12 | 0.19 | 0.06 |
| CD@5\% | 21.78 | 20.53 | 24.80 | 1.91 | 4.78 | 6.44 | 0.58 | 0.18 |

DAS: Days after sowing

Sarada et al. (2005) in fenugreek and Das et al. (2014) in French bean for variability vine length.

Number of trifoliate leaves plant ${ }^{-1}$ was maximum (71.03) in genotype Arka Dhanvantari (Table 2). The increased number of leaves plant ${ }^{-1}$ recorded in Arka Dhanvantari might be due to increased vine length, number of branches plant ${ }^{-1}$, number of root nodules plant ${ }^{-1}$ and long duration of the genotype. The minimum (56.70) number of trifoliate leaves plant ${ }^{-1}$ was recorded in Farmer Selection-1. The variation in these parameters are due to decreased vine length, number of branches plant ${ }^{-1}$ and less number of root nodules plant ${ }^{-1}$ resulting in reduced growth and poor leaf production. Similar results have been also reported by Pugalenthi and Vadivel (2007) in Muсuna pruriens L. and Mamatha et al. (2010) in Mucuna utilis L .

## Yield parameters

The yield and yield parameters in cowhage genotypes found significant differences (Table 3). Number of pods bunch ${ }^{-1}$ was maximum (4.00) in the genotype IIHR Selection-2 which was on par with genotype IIHR Selection-10 (3.73) and Arka Dhanvantari (3.60), while the minimum (2.20) number of pods bunch ${ }^{-1}$ was recorded in genotype Farmer Selection-1. This difference may be due to the maximum inflorescence length, number of flowers inflorescence ${ }^{-1}$ and also higher assimilatory surface area due to the higher leaf area which in turn altered the canopy structure and might have lead to the stimulation of more number of pods bunch ${ }^{-1}$. Similar variation for number of pods in different genotypes was reported by Pugalenthi and Vadivel (2007) in Mucuna pruriens L. and Singh and Kaur (2007) in fenugreek.

Number of bunches plant ${ }^{-1}$ was maximum (4.67) in the genotype IIHR Selection-2 and was on par with genotypes Arka Dhanvantari (4.33) and IIHR Selection-10 (4.20) followed by IIHR Selection-8 (3.93). This increase in number of bunches plant ${ }^{-1}$ may be attributed to the vigorous growth of plants which accumulate higher photosynthates in the plants. Similar variation for number of bunches plant ${ }^{-1}$ in different genotypes were reported by Mamatha et al. (2010) in Mucuna utilis L.

The genotype Arka Aswini recorded maximum pod length $(11.02 \mathrm{~cm})$ and pod width $(1.89 \mathrm{~cm})$. This may be attributed to their genetic makeup and the different agro-ecological regions from which they have been originated. Pandey et al. (2011) in French bean, Islam et al. (2010) in hyacinth bean and Subramanian et al. (2005) in fenugreek observed similar variations in pod length.

The maximum pod yield plant ${ }^{-1}(136.38 \mathrm{~g})$ was recorded in the genotype IIHR Selection-2. It is
clearly visible that existence of relationship between production of number of pods bunch ${ }^{-1}$ and number of bunches plant ${ }^{-1}$ might have increased the pod yield plant ${ }^{-1}$. These results are in conformity with the results reported earlier in Mucuna pruriens L. (Gurumoorthi et al., 2003; Pugalenthi and Vadivel, 2007).

The genotype IIHR Selection-2 (96.13 g) recorded the higher seed yield plant ${ }^{-1}$ and it was on par with genotype IIHR Selection-10 ( 83.07 g ) followed by Arka Dhanvantari ( 74.73 g ). The increased seed yield plant ${ }^{-1}$ may be mainly due to the increased number of pods, pod length, pod width, leaf area plant ${ }^{-1}$ and also may be due to maximum nodulation capacity. The present results are in close conformity with the findings of Mamatha et al. (2010) in Mucuna utilis L. and Verma and Korla (2003) in fenugreek.

The seed yield plot ${ }^{-1}$ and hectare ${ }^{-1}$ was calculated in which significant difference among the

Table 3. Performance of cowhage (Mucuna pruriens L.) genotypes for yield parameters

| Genotype |  | No. of bunches plant ${ }^{-1}$ | Pod length (cm) | Pod width (cm) | $\begin{gathered} \text { Pod } \\ \text { yield } \\ \left(\mathrm{g} \text { plant } \mathrm{t}^{-1}\right) \end{gathered}$ | $\begin{gathered} \text { Seed } \\ \text { yield } \\ \left(\mathrm{g} \text { plant } \mathrm{t}^{-1}\right) \end{gathered}$ | Seed yield $\left(\mathbf{k g}\right.$ plot $\left.^{-1}\right)$ | $\begin{gathered} \text { Seed } \\ \text { yield } \\ \left(\mathrm{kg} \mathrm{ha}^{-1}\right) \end{gathered}$ | $\begin{gathered} 100 \\ \text { seed } \\ \text { weight }(\mathrm{g}) \end{gathered}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| $\mathrm{T}_{1}$ - Arka Aswini | 2.67 | 3.67 | 11.02 | 1.89 | 80.13 | 58.77 | 1.76 | 2068.99 | 136.23 |
| T2-Arka <br> Dhanvantari | 3.60 | 4.33 | 9.37 | 1.72 | 104.40 | 74.73 | 2.24 | 2631.13 | 88.05 |
| $\mathrm{T}_{3} \text { - IIHR }$ <br> Selection-2 | 4.00 | 4.67 | 11.01 | 1.85 | 136.38 | 96.13 | 2.88 | 3384.56 | 101.59 |
| T4- IIHR <br> Selection-3 | 2.80 | 3.80 | 9.05 | 1.69 | 85.70 | 60.77 | 1.82 | 2139.41 | 89.14 |
| $\mathrm{T}_{5}$ - IIHR <br> Selection-8 | 3.07 | 3.93 | 9.74 | 1.66 | 94.50 | 64.50 | 1.94 | 2270.85 | 112.74 |
| $\mathrm{T}_{6}-\mathrm{IIHR}$ <br> Selection-10 | 3.73 | 4.20 | 9.39 | 1.75 | 120.20 | 83.07 | 2.49 | 2924.53 | 117.05 |
| $\mathrm{T}_{7}$ - Farmer Selection-1 | 2.20 | 3.33 | 10.07 | 1.72 | 72.53 | 51.97 | 1.56 | 1829.45 | 119.85 |
| $\mathrm{T}_{8}$ - Farmer Selection-2 | 2.47 | 3.60 | 10.10 | 1.73 | 82.09 | 58.63 | 1.76 | 2064.30 | 122.90 |
| S. Em $\pm$ | 0.17 | 0.16 | 0.29 | 0.05 | 5.41 | 4.35 | 0.13 | 153.20 | 4.82 |
| CD@5\% | 0.51 | 0.47 | 0.88 | 0.14 | 16.41 | 13.20 | 0.40 | 464.70 | 14.61 |

genotypes was recorded. Higher seed yield (2.88 kg ) plot $^{-1}$ and hectare ${ }^{-1}(3384.56 \mathrm{~kg})$ was recorded in the genotype IIHR Selection-2, while minimum seed yield ( 1.56 kg ) plot $^{-1}$ and hectare ${ }^{-1}(1829.45 \mathrm{~kg})$ was in the genotype Farmer Selection-1. This increase in the seed yield might be due to the maximum number of pods, number of bunches, pod length, pod width and number of seeds plant ${ }^{-1}$. These findings are in agreement with the results reported by Das et al. (2014) in French bean.

The maximum weight of 100 seeds was recorded in Arka Aswini genotype ( 136.23 g ) and was on par with genotype Farmer Selection-2 $(122.90 \mathrm{~g})$ which may be due to large size of the seed and maximum recovery. The minimum weight $(88.05 \mathrm{~g})$ of 100 -seeds recorded in genotype Arka Dhanvantari could be attributed to the minimum number and weight of seeds. These results corroborate the findings of Gurumoorthi et al. (2003) and Mamatha et al. (2010) in Mucuna pruriens L .

## Quality parameters

Significant differences were observed in genotypes with respect to quality parameters (Table 4). L-DOPA content was found to be maximum ( $5.17 \%$ ) in the genotype Arka Aswini followed by genotypes IIHR Selection-2 (4.69 \%) and Arka Dhanvantari ( $4.51 \%$ ) while, the genotype Farmer Selection-2 had lowest L-DOPA content ( $3.68 \%$ ). This difference in L-DOPA content could be probably due to the variation in precursor compounds of L-DOPA present in the seeds and genetic factor of the individual genotype and environmental conditions. The similar results were also reported by Vadivel and Janardhan (2000b), Leelambika et al. (2010) and Mahesh and Sathyanarayana (2011) in Mucuna pruriens L.

The L-DOPA yield plant ${ }^{-1}$ was maximum in genotype IIHR Selection-2 (4.52 g plant ${ }^{-1}$ ) followed by genotypes IIHR Selection-10 ( $3.54 \mathrm{~g} \mathrm{plant}^{-1}$ ) and Arka Dhanvantari ( 3.36 g plant ${ }^{-1}$ ). This may be due to the maximum seed yield plant ${ }^{-1}$ and also L-DOPA (\%) had resulted in an increase of L-DOPA yield plant ${ }^{-1}$. Whereas, the lowest L-DOPA yield plant ${ }^{-1}$ was recorded in the genotype Farmer Selection-1 ( $1.94 \mathrm{~g} \mathrm{plant}^{-1}$ ). This might be due to the lowest seed yield plant ${ }^{-1}$ and L-DOPA (\%). This variation in

L-DOPA yield plant ${ }^{-1}$ is due to genotype character and environmental interactions. These findings are in close conformity with the results of Vadivel and Janardhan (2000a) and Archana and Renu (2011).

Table 4. Performance of cowhage (Mucuna pruriens L.) genotypes for quality parameters

| Genotype | L-DOPA <br> (\%) | L-DOPA yield <br> (g plant $\mathbf{t}^{-1}$ |
| :--- | :---: | :---: |
| $\mathrm{~T}_{1}$ - Arka Aswini | 5.17 | 3.03 |
| $\mathrm{~T}_{2}$ - Arka Dhanvantari | 4.51 | 3.36 |
| $\mathrm{~T}_{3}$ - IIHR Selection-2 | 4.69 | 4.52 |
| $\mathrm{~T}_{4}$ - IIHR Selection-3 | 4.23 | 2.58 |
| $\mathrm{~T}_{5}$ - IIHR Selection-8 | 4.04 | 2.60 |
| $\mathrm{~T}_{6}$ - IIHR Selection-10 | 4.25 | 3.54 |
| $\mathrm{~T}_{7}$ - Farmer Selection-1 | 3.75 | 1.94 |
| $\mathrm{~T}_{8}$ - Farmer Selection-2 | 3.68 | 2.15 |
| S.E m $\pm$ | 0.14 | 0.24 |
| CD @ 5\% | 0.41 | 0.71 |

## Conclusion

The present research on identification of suitable genotypes of cowhage in arecanut plantation under hill zone of Karnataka revealed that the genotype IIHR Selection-2 performed better in terms of number of bunches plant ${ }^{-1}$, number of pods bunch ${ }^{-1}$, pod yield and seed yield attributes followed by genotypes IIHR Selection-10 and Arka Dhanvantari. The L-DOPA content (\%) was found maximum in Arka Aswini while, highest L-DOPA yield plant ${ }^{-1}$ was obtained in IIHR Selection-2.

## References

Archana, P.R. and Renu, K. 2011. Quantitative determination of L-DOPA in seeds of Mucuna pruriens L. germplasm by high performance thin layer chromatography. Indian Journal of Pharmaceutical Sciences 73(4): 459-462.

Bell, E.A., Nulu, J.R. and Cone, C. 1971. L-DOPA andL-3-Carboxy-6,7- Dihydroxy - 1, 2, 3, 4 Tetrahydroisoquinoline, a new amino acid from seeds of Mucuna mutisiana. Phytochemistry 10:2191-2194.

Caius, J.F. 1989. The Medicinal and Poisonous Legumes of India. Scientific Publications, Jodhpur, India. 528p.

Das, R., Thapa, U., Debnath, S., Lyngdoh, Y.A. and Mallick, D. 2014. Evaluation of French bean (Phaseolus vulgaris L.) genotypes for seed production. Journal of Applied and Natural Science 6 (2): 594-598.
Daxenbichler, M.E., Vaneteen, C.H., Earle, F.R. and Tallent, W. H. 1972. L-DOPA recovery from Mисипа seed. Journal of Agriculture and Food Chemistry 20: 1046-1048.

Farooqi, A.A. and Sreeramu, B. S. 2001. Cultivation of Medicinal and Aromatic Crops. Universities Press (India) Ltd., Hyderabad. 647p.

Gurumoorthi, P., Senthil Kumar, S., Vadivel, V. and Janardhanan, K. 2003. Studies on agro botanical characters of different accessions of velvet bean collected from Western Ghats, South India. Tropical and Subtropical Agroecosystem 2: 105-115.

Islam, M.S., Rahman, M. M. and Hossain, T. 2010. Physicomorphological variation in hyacinth bean (Lablab purpureus L.). Bangladesh Journal of Agriculture Research 35 (3): 431-438.
Leelambika, M., Mahesh, S., Jaheer, M. and Sathyanarayana, N. 2010. Comparative evaluation of genetic diversity among Indian Mucuna species using morphometric, biochemical and molecular approaches. World Journal of Agriculture Science 6(5): 568-578.

Mahesh, S. and Sathyanarayana 2011. The genotype x environment interaction and stability analysis for L-DOPA trait in velvet bean (Mucuna pruriens L.) seeds. Indian Journal of Genetics 71(3): 279-282.

Mamatha, B.R., Siddaramappa, R. and Shivananda, T.N. 2010. Evaluation of Mucuna utilis germplasm for higher biomass production, active principle and seed yield. Journal of Medicinal Plant Research 4(13): 1297-1300.

Pandey, Y.R., Gautam, D.M., Thapa, R. B., Sharma, M.D. and Paudyal, K. P. 2011. Evaluation of pole-type french bean genotypes in the mid hills of western Nepal. Nepal Agriculture Research Journal 11: 80-86.

Pugalenthi, M. and Vadivel, V. 2007. Agrobiodiversity of eleven accessions of Mucuna pruriens (L.) DC. var. utilis (Wall. ex Wight) Baker ex Burck (velvet bean) collected from four districts of South India. Genetic Resources and Crop Evolution 54: 1117-1124.

Sarada, C., Giridhar Kaildasu and Hariprasada Rao, N. 2005. Performance of fenugreek genotypes (Trigonella foenumgraecum Linn.) in vertisols. Spice India 18 (2): 46-49.

Singh, P. and Kaur, K. 2007. Genetic evaluation of (Trigonella foenum graecum L.) for seed yield and quality attributes. Crop Improvement 34 (1): 90-94.

Subramanian, S., Rajeswari, E. and Chezhiyan, N. 2005. Evaluation of fenugreek germplasm for morphological and yield characters. South Indian Horticulture 3 (1-6): 172-174.

Verma, R. and Korla, B.N. 2003. Genetic variability in fenugreek (Trigonella foenum-graecum L.) grown under mid- hills of Himachal Pradesh. Journal of Spices and Aromatic Crops 12 (1): 60-62.

Vadivel, V. and Janardhanan, K. 2000a. Nutritional and antinutritional composition of velvet bean: An under-utilized food legume in South India. International Journal of Food Science and Nutrition 51:279-287.

Vadivel, V. and Janardhanan, K. 2000b. Preliminary agrobotanical traits and chemical evaluation of Muсипа pruriens (itching beans): a less-known food and medicinal legume. Journal of Medicinal and Aromatic Plant Sciences 22: 191-199.

