Journal of Experimental Sciences Vol. 2, Issue 6, Pages 01-03 [2011]



Regular Article FTIR spectroscoptc study and antifungal activity of the medicinal plant glory lily (*Gloriosa superba*)

S. Ravi^{1*}, S. Ashokkumar, K. Mallika¹, P. Kabilar, P. Paneerselvam¹, M. Gayathri²

¹Engineering Physics Section, Faculty of Engg. & Tech., Annamalai University, Annamalai Nagar, Tamil Nadu, India – 608 002; ²Department of Horticulture, Faculty of Agriculture, Annamalai University, Annamalai Nagar, Tamil Nadu, India – 608 002

Abstract

In this present study, the presence of the phyto compound (i.e.) Colchicine and other chemical constituents present in three different treated tuber and seed samples of Glory Lily (*Gloriosa superbd*) was confirmed using FTIR. An attempt has been made to correlate the extinction coefficient (K) values of all the samples. And also the samples were extensively studied for their antifungal activity against *Pseudomonas aeruginosa, Klebsiella phemoniae*, and *Salmonella typhi*. The results indicated that the Organic Manure treated samples were highly active against the three fungi.

Keywords: FTIR (Fourier Transform Infrared Spectrometer), Antifungal activity, *Pseudomonas aeruginosa, Klebsiella phemoniae* and *Salmonella typhi*

Introduction

Glory Lily (*Gloriosa superba* L) belongs to the family Liliaceae. It is an ancient medicinal plant in India. India was the first to use Ayurveda medicine, later as source colchicines and colchicoside [1]. Colchicine the main alkaloid of Gloriosa species [2], are a useful agent in the treatment of acute attacks of gout [3]. Traditionally, colchicine was used for its antimitotic properties [4] and treatment of familial Mediterranean fever [5].

The toxins of Gloriosa superba have an inhibitory action on cellular division resulting in diarrhea, depresent action on the bone marrow and alopecia. The technique based on Fourier Transform Infrared (FTIR) spectrometry was used to detect the presence of colchicines in tubers and seeds.

Thus the aim of this study was to confirm the presence of colchicines in Glory Lily by using FTIR technique and to investigate the anti-fungal activity against some selected fungi. And also an attempt has been to correlate the extinction coefficient (K) values of all the samples.

Materials and Methods

The three different treated samples of tuber and seed (Control (T_1) , Chemical Fertilizer (T2), and Organic Manure (T3)) of Glory Lily (Gloriosa superba) were collected from Jothy herbals, Jayamkondam, Perambalur District, TamilNadu.

The infrared spectra of these samples were recorded using FTIR in the range of 400-4000 cm- by the KBr pellet technique. It was carried out by the Instrumentation lab of Chemistry Department, Annamalai University. Anti-fungal activity of tuber and seed samples of Glory III1y *{Gloriosa superba}* was evaluated against *Pseudomonus aernginosa, Klebsiella pneumoriiae, Salmonella typhi.* The micro organisms were maintained on agar slants made of antibiotic assay medium A (Hi-Media Mumbai, India) making monthly transfers. Antifungal activity was evaluated by paper disc diffusion method. [6] It was carried out by RMMC & H, Annamalai University, Annamalai Nagar.

Results and Discussion

FT-IR Spectrum analysis

| Fig | 1 | shows | that | the | FTIR | spectra | of | different | treated |
|------|---|-------|------|-----|------|---------|----|-----------|---------|
| tube | r | and | se | ed | sam | ples | of | Glory | Lily. |

Fig 1: FT-IR spectra of different treated Tuber samples of Glory Lily under (a) Control (b) Chemical Fertilizer and (c) Organic Manure Treatment



Fig 2: FT-IR spectra of different treated Seed samples of Glory Lily under (a) Control (b) Chemical Fertilizer and (c) Organic Manure Treatment



The functional groups present in the samples were determined by FT-IR spectroscopy. The FT-IR spectrum confirmed the presence of colchicines in Glory Lily, Significant peaks were found at: 2926) cm¹ corresponding to CH₂ group, [7] (1646) cm^{*} attributed to Carbonyl

groups [8], and (1539) cm⁻¹ corresponding to amino acid groups [9], all of which confirms the presence of colchicines.

Table land Table 2 shows the Extinction Coefficient (K) values oftuberandseedsamplesofGloryLily.

| | | Extinction coefficient (K) cm ² / mg | | | | | | |
|----------|--|---|----------|--------------------------|----------|--------------------------|-------------|--|
| Absorpti | Tentative Assignments | T ₁ treatment | | T ₂ treatment | | T ₃ treatment | | |
| UI Ballu | | Sandy Soil | Red Soil | Sandy Soil | Red Soil | Sandy Soil | Red Soil | |
| 2922 | C-H (sym./asym) Vinyl ether | 0.04552 | 0.06840 | 0.06216 | 0.09182 | 0.12056 | 0.18376 | |
| 1646 | C=C (stret.) / Vinyl (ether) | 0.04552 | 0.06840 | 0.06216 | 0.09182 | 0.12056 | 0.18376 | |
| 1457 | N=N-O (antisymstret.) | 0.09808 | 0.18003 | 0.14242 | 0.14963 | 0.02889 | 0.31722 | |
| 1412 | C-N (stret.) / Amino acid III | 0.01303 | 0.02436 | 0.02165 | 0.02393 | 0.05749 | 0.17219 | |
| 845 | CH ₂ out of plane deformation | 0.02259 | 0.01949 | 0.02200 | 0.02624 | 0.02843 | 0.17219 | |
| 2925 | C-H (sym. / asym) / Vinyl ether | 0.03373 | 0.08825 | 0.05907 | 0.06586 | 0.06812 | 0.45513 | |
| 1646 | C=C (stret.) / Vinyl ether | 0.05023 | 0.08725 | 0.05933 | 0.06434 | 0.08222 | 0.36568 | |
| 1539 | N=N-O (antisymstret.) | 0.01188 | 0.02236 | 0.016518 | 0.04339 | 0.019627 | 0.05695 | |
| 1375 | CH ₃ deformation | 0.00739 | 0.01239 | 0.01287 | 0.01687 | 0.01082 | 0.06841 | |
| 871 | CH ₂ out of plane deformation | 0.01162 | 0.2819 | 0.00988 | 0.00934 | 0.00802 | 0.05568 | |

From the investigation it is seen that the cochicines level is the maximum in seed compared to the tubers of Glory Lily. In seed the

extinction Coefficient (K) value is 0.45513 \mbox{Cm}^2 / mg and in tuber 0.18376Cm²/mg.

Fig 2: Antifungal activity of Glory lily samples against Pseudomonas aeruginosa, Klebsiella pneumoniae, Salmonella typhi



Table 3 shows that the anti fungal activity of Glory Lily samples

| Table 3: The Anti-fungal activity of Glory Lily (G. Superba) sample |
|---|
|---|

| | | • | 5 5 1 | | | | |
|------------|------------------------|----------------|----------------|----------------|----------------|----------------|----------------|
| SI. No. | | Tuber | Tuber | | | | |
| | rungus Name | T ₁ | T ₂ | T ₃ | T ₁ | T ₂ | T ₃ |
| 1. | Pseudomonas aeruginosa | ++ | + | +++ | ++ | +++ | +++ |
| 2. | Klebsiella pneumoniae | + + | +++ | +++ | + | +++ | +++ |
| 3. | Salmonella typhi | + | ++ | +++ | ++ | +++ | + + + |

+ Low (3.0-3.5 mm) ++ Medium (4.5.5.5 mm)

+++ High (6.5.-7.5.mm)

The extracts of different treated tuber and seed samples of Glory Lily showed **antifungal** activity against Pseudomonas aeruginosa, Klebsiella pneumoniae, Salmonella typhi as determined by agar well and disc diffusion techniques. The results from the above studies may justify the use of plant in the treatment of certain skin diseases, infected wounds and also abscess [10, 11]. Many of the currently used anti-infective and anti neo plastic agents are natural products, initially isolated from plants. [12, 13]. Colchicine has abortifacient, anti-pyretic, anti-inflammatory and anti-leprotic properties [14]. On the basis of the present investigation it can be concluded that the root and seed of the Glory IIIy plant may be used as an anti-microbial agent. This may be due to the presence of highly active alkaloid compound, Colchicines and its derivatives.

References

G.Sivakumar and K.V.Krishnamurthy, Curr.Sci 78(1), 30(2000).

- G.Sivakumar, GloriosaSuperbaL. http://www.agrisupportonline.com /Articles/articles.htm, Melbourne, Australia
- K.Y.Kim, H.R.Schumacher, E.Hunsche, A.I.Wertheimer, and S.X.Kong, *Clin. Ther.*, 25(6), 1593(2003).
- S.Bergemann, R.Brecht, J.Buttner, D.Guenard, R.Gust, G.Seitz, M.T.Stubbs, and S.Thoret, Bioorg. *Med, Chem.*, 11, 1269(2003)
- V.Mijatovis,HompesPGA,andWoutersMGAJ,Eur.J.Obstet.Gynec. Reproduct.Biol.108, 171(2003).
- Jairaj, P., P. Khoohaswan, Y.Wongkrajang, P.Peungrisha, P. Suriyawong, S.Saraya and O.Ruangsonboon.1999. Anticough and antimicrobial activities of *Psidium guajava* Linn. Leaf extract. J.Ethnopharmacol, 67:203-212.

Sharma, Y.R.(1982). "Elementary organic spectroscopy", S.Chand & Company Ltd.

- Ramaswami, K., Perumal, R., and Pondurangan, S.V (1980). *CanJ. Spec*, 25,135.
- Rao, C.N.R.(1963). "Chemical applications of infrared spectroscopy", Academic Press, Newyork.
- Joshi, D.1993. Tribal remedies against Snake bites and Scorpion stings in Rajasthan. Glimpses Plant Res., 10:23-25.
- Singh, V.K.1993. Selected Indian Folk medicinal claims and their relevance in primary health care programme. *Glimpses Plant Res.*,

10:147-152.

- Poonkothai, MS.Hemaiswarya and D.Kavitha. 2005 a. Antibacterial activityof *Withania somnifera* a J. Microbial World, 7:97-99.
- Poonkothai, M., S.Hemaiswarya, D.Kavitha and K.Vallikkannu. 2005 b. Antibacterial activity of Gymnema sylvestre, Couroubita quianensis and withania somnifera. *Plant Arch.*, *5L2*1-26.
- Guhabakshi DN, Sensharma P, Paul DC (2001) In; A lexicon of medicinal plants in India. Naya prakash, Calcutta, Vol.2.pp 262-264.