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# Regular Article In vitro callus induction in rice (Oryza sativa L.)

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*In vitro* callus induction from embryos of matured seeds of four rice varieties *viz.*, ASD 16, ADT 43, Basmati 370, Pusa Basmati and Pokkali were studied. Observations on callus induction were carried out on six different callus induction medium having different concentration of 2,4-D *viz.*, 0.5, 1.0, 1.5, 2.0, 2.5 and 3.0 mg/l. The callus induction frequency varied from 58.33 % to 96.67 %. This study will be useful for selecting suitable callus induction medium for callus induction.

Key words: Rice, Embryo, Callus induction, 2,4-D

Rice (Oryza sativa L.) is the world most important cereal crop after wheat and maize. Rice has 24 species, of which 22 are wild and two viz. Oryza sativa and Oryza glabrrima are cultivated (Ray, 1985). It provides one-third of total dietary carbohydrate, especially in Asian countries and it is stable diet for more than three billion people, supplying 50 to 80 per cent of their daily calorie intake (Khush, 2005). A considerable improvement has been done through traditional rice breeding. Rice breeding has made significant progress towards higher yield, improved quality, disease greater resistance and other important characters agricultural of importance in the past and even in future, it will still play an important role (Sun et al., 1990).

Dehusked rice seed culture is a valuable technique to exploit somaclonal variation. But its application is limited by many factors which influence culture efficiency, such as plant genotype (Liu *et al.*, 1997), the culture

methods, the media (Sun *et al.*, 1990) and the culture conditions. Production of callus and its subsequent regeneration are the prime steps in crop plant to be manipulated by biotechnological means and to exploit somaclonal variation (Monirul Islam *et al.*, 2005). The objectives of this study were to find a suitable medium and culture condition for callus induction and this will also useful for callus based stress studies like salinity and drought.

## Materials and Methods

Mature seeds of five rice varieties *viz.*, ADT 43, ASD 16, Pusa Basmati, Basmati 370 and Pokkali were used. MS nutrient medium (Murashige and Skoog, 1962) was used as basal medium and it was solidified with 8.5 g/l agar. The agar was slowly dissolved in boiled distilled water without forming any clumps and three per cent sucrose used as carbon source in nutrient medium. For callus induction MS medium was supplemented

with different concentrations of 2, 4-D *viz.*, 0.5, 1.0, 1.5, 2.0, 2.5 and 3.0 mg/l and  $P^{H}$  of the medium was adjusted to 5.8.

Rice seeds were manually dehusked and washed with sterile water, and then the seeds were transferred to the laminar airflow chamber. The seeds were kept in 70 per cent ethanol for one minute. Then seeds were washed with sterile distilled water three times and were immersed in 0.1 per cent mercuric chloride for 15 minutes. Again the seeds are washed thoroughly three to four times with sterilized distilled water to remove all the trace of mercuric chloride and were blot dried using sterilized tissue paper. Surface sterilized seeds were cultured with the help of sterilized forceps into the test tube containing callus induction medium. Cultures were incubated in dark at 25±1°C. Callus induction was noticed within two weeks of inoculated cultures. Callus induction frequencies were recorded.

The frequency of callus induction was calculated according to the following formula: No. of seeds produced calli

Callus induction frequency (%) =  $\frac{1}{x + 1} x = 100$ 

No. of seeds inoculated

Varieties	Callus induction frequency (%)					
	M1	M2	M3	M4	M5	M6
ADT 43	80.00	70.00	91.33	92.00	83.33	71.33
ASD 16	81.67	84.17	94.17	96.67	86.67	80.83
Pusa Basmati	81.48	79.26	86.67	94.81	85.18	76.26
Basmati 370	80.83	81.67	93.33	94.17	89.17	77.50
Pokkali	75.00	63.33	71.67	83.33	76.67	58.00

### Table 1. Callus induction from five rice varieties

M1=MS + 0.5 mg/l 2,4-D; M2=MS + 1.0 mg/l 2,4-D; M3=MS + 1.5 mg/l 2,4-D; M4=MS + 2.0 mg/l 2,4-D; M5=MS + 2.5 mg/l 2,4-D; M6=MS + 3.0 mg/l 2,4-D

#### **Result and Discussion**

The effects of varieties on callus induction from dehusked rice seeds are shown in Table 1. Callus induction in rice was found highly variable and genotype specific. Among the five studied varieties, the variety ASD 16 produced 94.81 per cent callus from the inoculated seeds, which was higher than other four varieties. In all the treatments Pokkali had poor callus induction and ASD 16 had the best induction. This may be due to callusing efficiency was found to be genotype dependant. It was also confirmed by Rashid *et al.*, 2003 who reported that rice varieties

differed in degree of callusing. Rasheed et al., 2005 resulted that tissue culture generates a wide range of variation, which is resulted with incubation time and cultivar specific. induction For callus MS medium supplemented with different concentrations of 2,4-D was used. In that 2 mg/l 2,4-D showed high callus induction percentage in all the varieties and followed by 1.5 and 2.5 mg/l 2,4-D. It was also confirmed by Pandey et al., 1994; shankhdhar et al., 2002; Tam and Lang, 2003; Naqvi et al., 2005. Jaseela et al., 2009 reported that 60-100 per cent of the cultured seeds formed callus at all the concentrations of 2,4-D used and among the different auxin analogues used to induce somatic embryogenesis 2,4-D is the most efficient and therefore used in majority of embryogenic and tissue culture systems and also they proved 2 mg/l 2,4-D to be the most favorable for callus induction and callus proliferation. The role of 2,4-D in cell division is to increase the rate of cell division and this attributes to the increased amount of callus.

### References

- Jaseela, F., V.R. Sumitha and G.M. Nair. 2009. Somatic embryogenesis and plantlet regeneration in an agronomically important wild rice species *Oryza nivara*. *Asian J. Biotechnol*. 1(2): 74-78.
- Khush, G. S. 2005. What it will take to feed 5.0 billion rice consumers in 2030. *Plant Mol. Biol.*, 59: 1-6.
- Liu B. S., Chen, C.X., Yin, L.Q. and Zhang, J.J. 1997. Plant breeding and Genetics, *in vitro* Culture of plant the materials. Biotechnology, Agronomy Department, Shandong Agricultural University, 271018, China.
- Monirul Islam Md, Mahatalat Ahmed and Debabrata Mahaldar. 2005. *In Vitro* Callus Induction and Plant Regeneration in Seed Explants of Rice (*Oryza Sativa* L.). *Res. J. Agric. & Biol. Sci.* 1(1): 72-75.
- Murashige, T. and F. Skoog, 1962. A revised medium for rapid growth and bioassays with tobacco tissue cultures. *Physiologia Plantarum*, 15: 473- 497.
- Naqvi, S.M., S.Razia and H.Rashid. 2005. Tissue culture studies in *Oryza sativa* L. *cvs*. Basmati 385 and Super Basmati. *Pak. J. Bot.*, 37(4): 823-828.
- Pandey, S.K., B. Ramesh and P.K. Gupta. 1994. Study on effect of genotypes and culture medium on callus formation and

plant regeneration in rice (*Oryza sativa* L.). *Indian J. Genet.*, 54(3): 293-299.

- Rasheed, S, T. Fatima, T. Husnain, K. Bashir and S. Riazuddin. 2005. RAPD characterization of somaclonal variation in *indica* basmati rice. *Pak. J. Bot.* 37(2): 249-262.
- Rashid, H, F. Mohammad Abbasi and A. Quraishi. 2003. Plant Regeneration from Seed Derived Callus of three varieties of Basmati Rice. *Plant Tissue Cult.*, 13(1): 75-79.
- Ray, J.K., 1985. *Introduction to Botany of the Rice Plant.* 2nd Ed, p. 5. Rice Research Institute in India. Indian Council of Agricultural Research, New Delhi, India.
- Shankhdhar, D., S.C. Shankhdhar and R.C. Pant. 2002. Development of somatic embryos in rice. *Indian J. Plant Physiol.*, 7(3): 211-214.
- Sun, Z.R., P.C. Ni and Z.Z. Hung, 1990. Studies on the analysis of variance and major/minor factors of medium components influencing the efficiency of callus production ability. *Acta Argon. Sin.*, 16: 123–30.
- Tam, D.M. and N.T.Lang. 2003. In vitro selection for salt tolerance in rice. *Omonrice*,11: 68-73.